

**Supplemental Information:**
**Table S1: Primers used in this work.** Relates to Experimental Procedures:

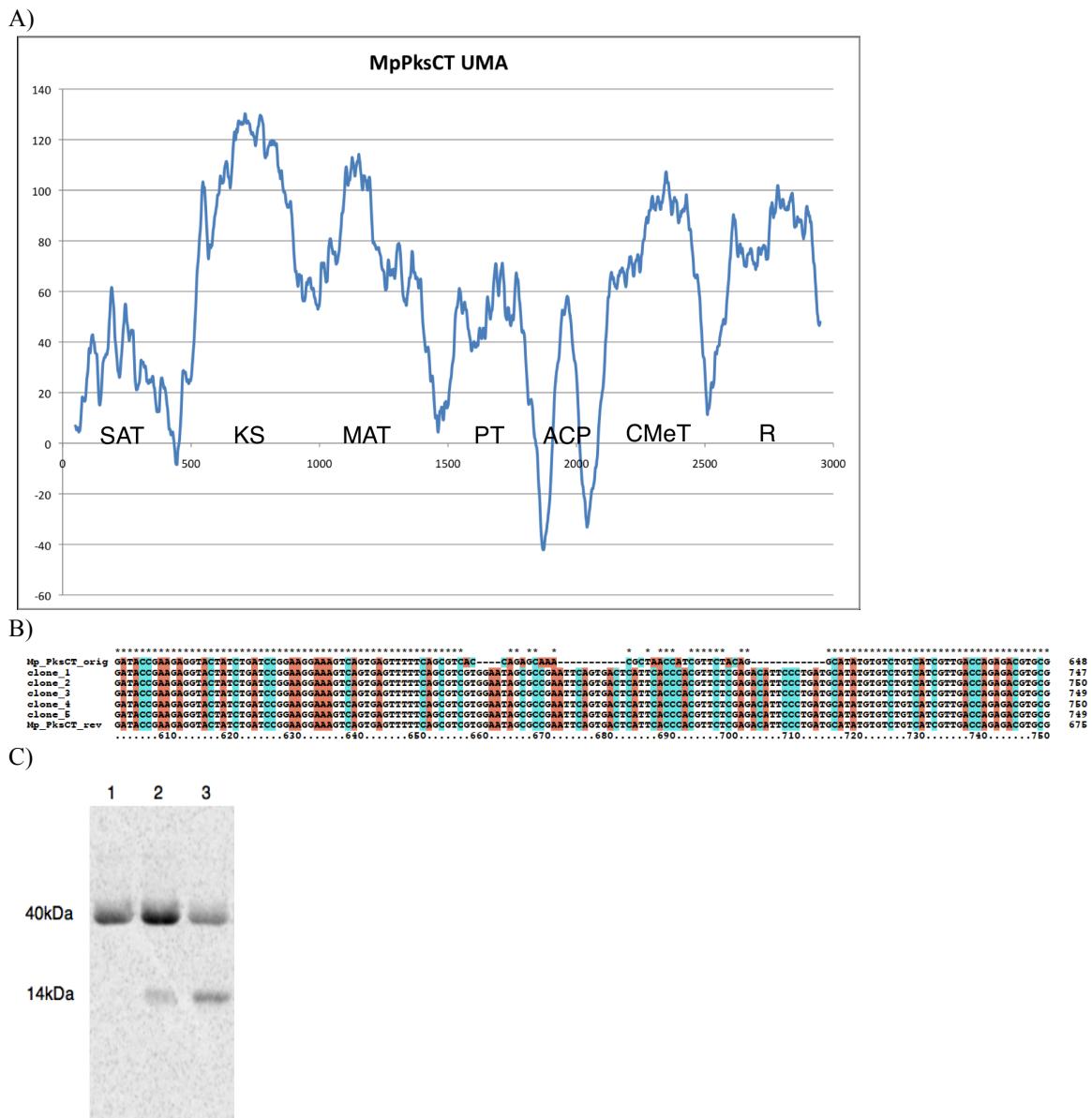
Primer Name	Sequence
MpPksCT-g5	ATGATTGACTCAACTTCGCACTCAAATCTGAG
MpPksCT-g3	TTAATCTAGAAATCCCATGGTCTTCATGC
MpPksCT-ex1-5	GCATATTAATGATTGACTCAACTTCGCACTC
MpPksCT-ex1-3.2	GTCAACGATGACAGACACATATGCATCAGGGAAATGTCTCGAGAACG
MpPksCT-ex2-5.2	CTCGAGACATTCCCTGATGCATATGTGTCTGTCATCGTTGAC
MpPksCT-ex2-3	GCATGCGCCGCATCTAGAAATCCCATGGTCTCCATG
MpPksCT-SAT-3	GCATGCGCCGCCACCGAGAAGCTGCC
MpPksCT-SKM3-3	GCATGCGCCGCAGAGCGCTGGATGCGTCTTG
MpPksCT-PT-5	GCATATTAATGACAGCAATTATCGAGGCACC
MpPksCT-PT-3	GCATGCGCCGCAGCGCTTACCGCAGTAGACG
MpPksCT-ACP-5	GCATATTAATGACACCTCCGACAAAGAAAGCGC
MpPksCT-ACP-3	GCATGCGCCGCACCGTTACCATTAAACGACACCATTAGTGC
MpPksCT-CMeT-5	GCATATTAATGGTCTTGTGAGCTAGGGGG
MpPksCT-CMeT-3	GCATGCGCCGCCTGTCAATGGGACCTGCACC
MpPksCT-CMeTAsel-5	CGAGGAGGCCAGACTGATTGATATCAACG
MpPksCT-CMeTAsel-3	CGTTGATATCAATCAGTCTGGCCTCCTCG
MpAct-5	GGAATTCTGCAGATTCTACAACGAACCTCCG
MpAct-3	GGAATTCTGCAGTCAGGGAGTTCATAGGAC
MpPksCT-CMeT-	GTCACTGATGTCTTGCCAAATCTCC
Y1955F-5	
MpPksCT-CMeT-	GGAGATTGGCAAAGACATCAGTGAC
Y1955F-3	
MpPksCT-CMeT-	GTCACTGATGTCGGGCCAAATCTCC
Y1955A-5	
MpPksCT-CMeT-	GGAGATTGGCCGCGACATCAGTGAC
Y1955A-3	
MpPksCT-CMeT-	CAAATTGTGTGGCGGCGACCCGG
H2067A-5	
MpPksCT-CMeT-	CCGGGTGCCGCCACACAATTG
H2067A-3	
MpPksCT-CMeT-	CAAATTGTGTGCAGGCGACCCGG
H2067Q-5	
MpPksCT-CMeT-	CCGGGTCGCCTGCACACAATTG
H2067Q-3	

**Figure S1, relates to Figures 1 and 2: Domain deconstruction, exon revision, and starter unit verification for PksCT.**

A) Sequence conservation, predicted secondary structure, and local hydrophobicity are combined to generate an UMA score for each position. Low scores indicate likely linker regions and were used to guide the domain boundaries for mono- or multi-domain expression constructs. For details on input sequences and algorithm settings, please see Supplemental Experimental Procedures.

B) Five independent clones amplified from a cDNA library generated from total RNA under citrinin producing conditions. Each clone exactly matched the FGENESH prediction for the exon 1:exon 2 boundary.

C) Radiolabel assay of acetyl starter unit transfer in PksCT. Lane 1: PksCT SAT, Lane 2: PksCT SAT + ACP without Sfp activation, Lane 3: PksCT SAT + ACP with Sfp activation. NR-PKS ACP domains are often partially phosphopantetheinylated during expression, accounting for the low ACP signal in lane 2.



**Table S2, relates to Figure 2: Plasmids used in this work:**

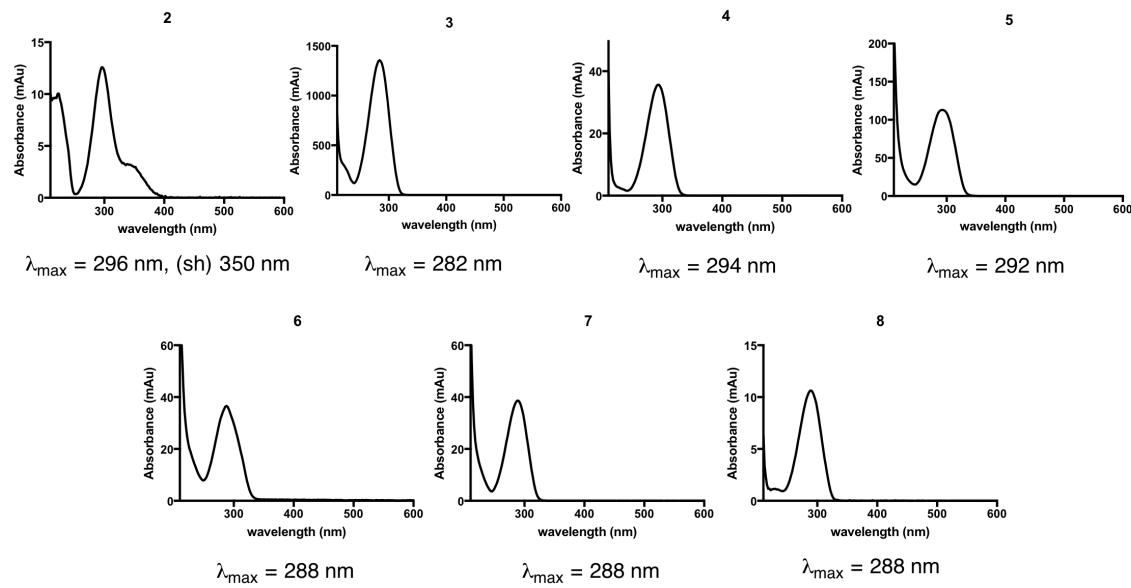
PksCT domain(s)	Sequence	Plasmid name	Parent vector	Tag	MW (kDa)
SAT	M1-G380	pEMpPksCT-SAT	pET-24a	C-His6	41.8
SAT-KS-MAT	M1-I1288	pEMpPksCT-SKM3	pET-24a	C-His6	144.0
PT	T1285-G1651	pEMpPksCT-PT	pET-24a	C-His6	41.5
ACP	T1652-G1779	pEMpPksCT-ACP	pET-24a	C-His6	15.0
CMeT	V1780-D2182	pEMpPksCT-CMeT	pET-24a	C-His6	46.6
CMeT-Y1955A	V1780-D2182	pEMpPksCT-CMeT-Y1955A	pET-24a	C-His6	46.6
CMeT-Y1955F	V1780-D2182	pEMpPksCT-CMeT-Y1955F	pET-24a	C-His6	46.6
CMeT-H2067A	V1780-D2182	pEMpPksCT-CMeT-H2067A	pET-24a	C-His6	46.6
CMeT-H2067Q	V1780-D2182	pEMpPksCT-CMeT-H2067Q	pET-24a	C-His6	46.6
CMeT-R	V1780-D2593	pEMpPksCT-CMeT-R	pET-24a	C-His6	92.7

**Table S3, relates to Figure 3: Crystallographic data collection and refinement statistics.**

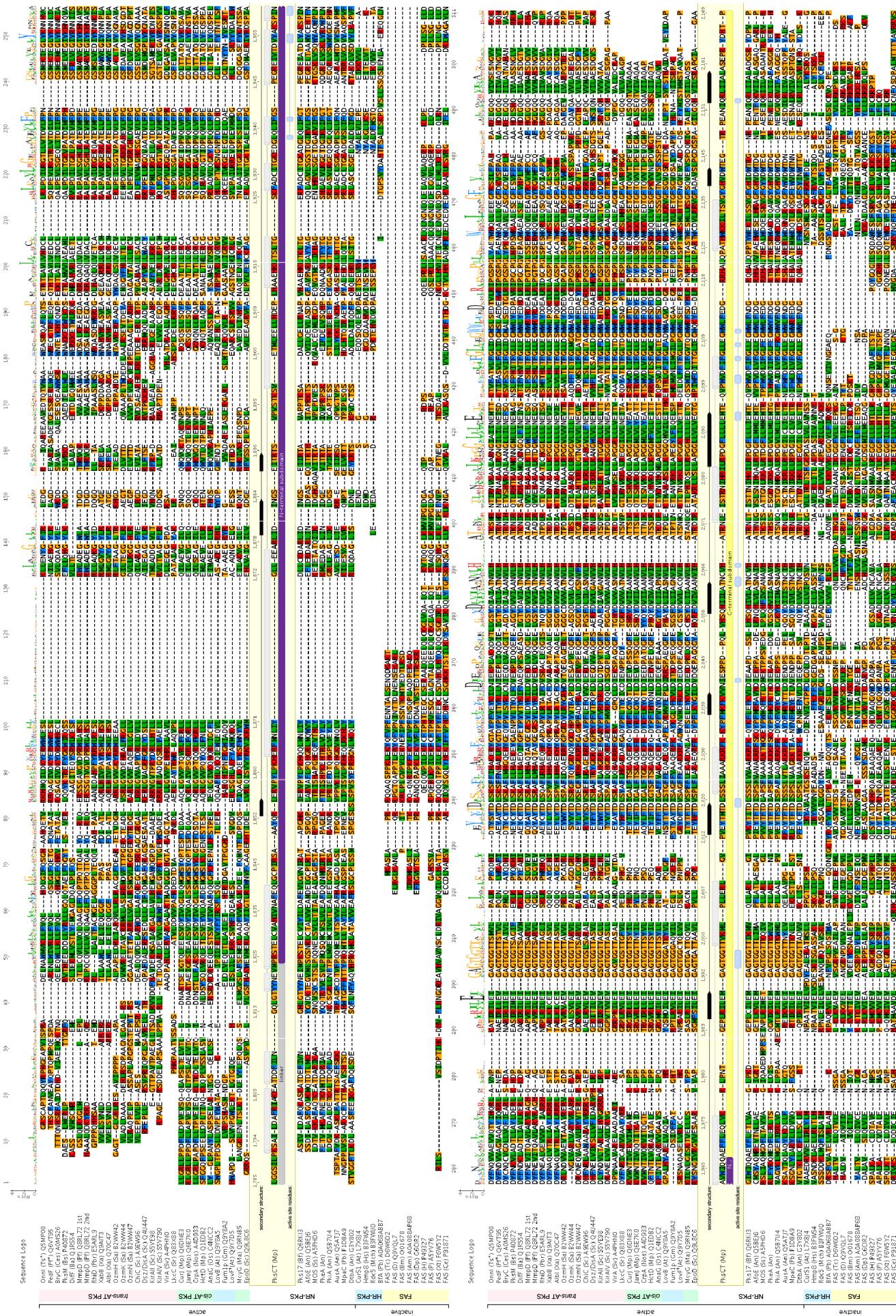
	CMeT native		CMeT SeMet
<b>Data collection</b>			
Space group	H3	H3 <sup>†</sup>	H3 <sup>†</sup>
Cell dimensions			
a, b, c (Å)	98.37, 98.37, 133.43	98.52, 98.52, 133.20	98.13, 98.13, 124.45
α, β, γ (°)	90.0, 90.0, 120.0	90.0, 90.0, 120.0	90.0, 90.0, 120.0
Wavelength (Å)	0.99998	1.90747	0.97929
Swipes	1	2	2
Resolution (Å)	71.80 – 1.65 <sup>‡</sup>	71.85 - 2.05	50.21 – 1.85
R <sub>merge</sub> (%)*)	8.0 (159.6)	8.4 (102.9)	13.5 (274.2)
I/σI*	10.61 (1.37)	15.62 (1.38)	13.75 (1.12)
CC <sub>1/2</sub> (%)*)	99.9 (76.0)	99.8 (71.1)	99.9 (57.1)
Completeness (%)*)	98.9 (94.3)	97.9 (81.7)	99.8 (99.7)
Redundancy*	6.6 (6.6)	8.5 (2.6)	10.4 (10.2)
Unique reflections*	57,453 (4,045)	59,253 (3,685)	76,184 (5,646)
<b>Refinement</b>			
Resolution (Å)	71.80 – 1.65		
R <sub>work</sub> / R <sub>free</sub>	0.19 / 0.22		
Ramachandran	99.48 / 0.00		
favored / outlier (%)			
No. atoms	6,512		
Protein	6,173		
Ligand	45		
Solvent	294		
B-factors	36.22		
Protein (Å <sup>2</sup> )	36.29		
Ligand (Å <sup>2</sup> )	26.84		
Solvent (Å <sup>2</sup> )	36.11		
R.m.s deviations			
Bond lengths (Å)	0.009		
Bond angles (°)	0.975		

The resolution cutoff was determined by CC<sub>1/2</sub> criterion (Karplus and Diederichs, 2012). \*, Highest resolution shell is shown in parenthesis. Datasets of native CMeT were acquired from the same crystal.

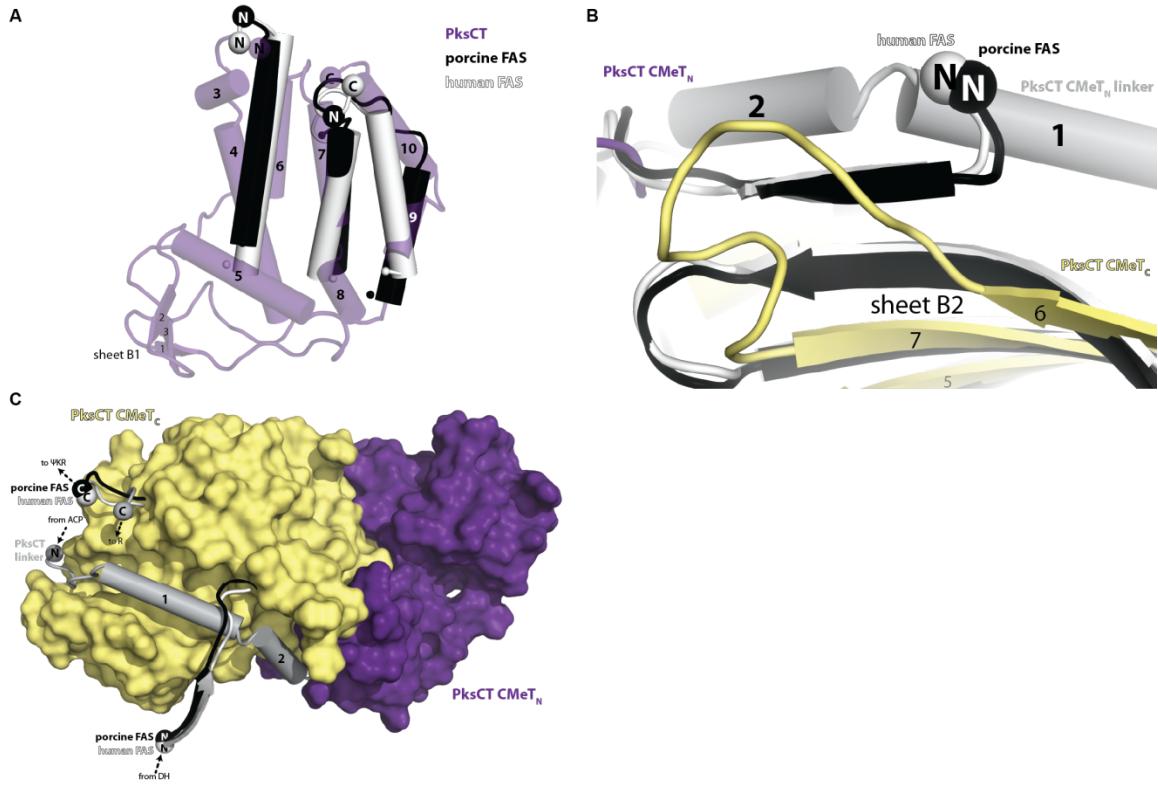
<sup>†</sup>Anomalous data processing was applied. <sup>‡</sup>A deprecated conservative resolution cutoff at I/σI=2.91 and R<sub>merge</sub>= 69.8% is reached at a resolution of 1.84 Å.

**Figure S2, relates to Figures 2 and 5A: UV-Vis spectra for compounds 2-8.****Table S4, relates to Figures 2 and 5A: Detected masses for compounds 2-8.**

#	Calculated [MH+] or [MNa+] m/z	Detected m/z (ppm)	Formula
2	237.1127	237.1121(2.53)	C <sub>13</sub> H <sub>17</sub> O <sub>4</sub>
3	127.0395	127.0391 (3.15)	C <sub>6</sub> H <sub>7</sub> O <sub>3</sub>
4	155.0708	155.0704 (2.58)	C <sub>8</sub> H <sub>11</sub> O <sub>3</sub>
5	211.0970	211.0970 (0.00)	C <sub>11</sub> H <sub>15</sub> O <sub>4</sub>
	233.0787	233.0790 (1.29)	C <sub>11</sub> H <sub>14</sub> O <sub>4</sub> Na
	169.0865	169.0859 (2.37)	C <sub>9</sub> H <sub>13</sub> O <sub>3</sub> [-C <sub>2</sub> H <sub>2</sub> O]
6	253.1075	253.1076 (0.40)	C <sub>13</sub> H <sub>17</sub> O <sub>5</sub>
	275.0895	275.0893 (0.73)	C <sub>13</sub> H <sub>16</sub> O <sub>5</sub> Na
	211.0970	211.0965 (2.73)	C <sub>11</sub> H <sub>15</sub> O <sub>4</sub> [-C <sub>2</sub> H <sub>2</sub> O]
	155.0708	155.0701 (4.51)	C <sub>8</sub> H <sub>11</sub> O <sub>3</sub> [-C <sub>5</sub> H <sub>6</sub> O <sub>2</sub> ]
7	141.0552	141.0553 (0.71)	C <sub>7</sub> H <sub>9</sub> O <sub>3</sub>
8	141.0552	141.0552 (0)	C <sub>7</sub> H <sub>9</sub> O <sub>3</sub>



**Figure S3, relates to Figure 4. Alignment of 51 CMeT domains from PKSs and FASs.** The alignment reveals a high sequence conservation of active CMeT domains, whereas inactive  $\Psi$ CMeT domains of FAS and some HR-PKS reveal deletions predominantly in the *N*-terminal subdomain and a low sequence conservation of active site residues. Sequence numbers, secondary structure elements, and active site residues correspond to PksCT. All sequences are labeled as protein name (organism abbreviation) Uniprot number and sorted according to their phylogenetic distance depicted in Figure 4. The sequence of PksCT corresponds to Supplemental Item 1. Multienzyme family classifications are indicated in colored groups. ( $\circ$ , endosymbiont of this organism;  $\ddagger$ , dикетид synthase)

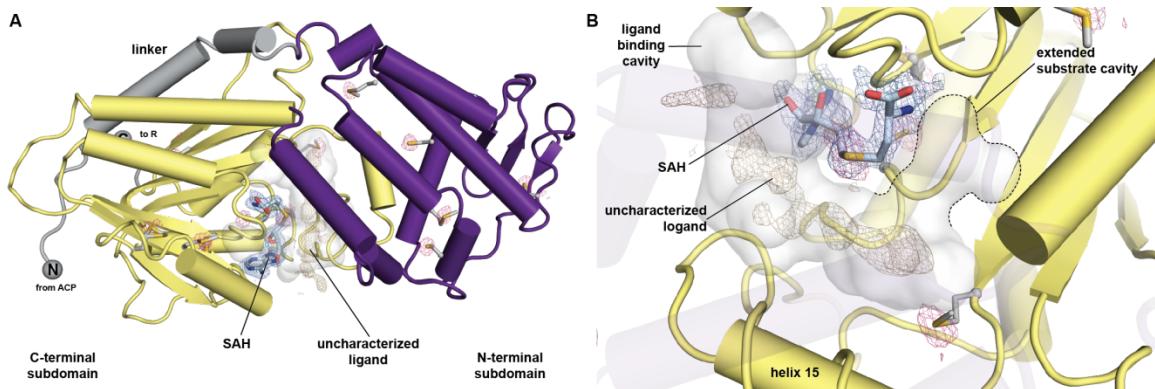


**Figure S4, relates to Figure 3. Structural comparison of PksCT CMeT with mammalian FAS  $\Psi$ CMeT domains.**

(A) A superposition of the truncated and largely disordered *N*-terminal subdomain of the animal FAS  $\Psi$ CMeT (porcine: black; human: white) with PksCT CMeT indicates a structural conservation of three helices, overlapping with PksCT CMeT helices 4, 7 and 9/10. Ends of disordered regions are indicated by small spheres. Secondary structures are assigned according to Figure 3B.

(B) In FAS, the linker connecting to the *N*-terminal subdomain contains a  $\beta$ -strand that extends the central  $\beta$ -sheet of the *C*-terminal subdomain. In PksCT CMeT, the position of this  $\beta$ -strand is occupied by helix 2 of the *N*-terminal linker leading into CMeT.

(C) In PksCT the *N*-terminal linker wraps in a surface groove around the *C*-terminal subdomain and positions the *N*- and *C*-terminal linker termini in close proximity to each other, whereas in FASs the *N*-terminal linker contacts this domain in a different location distant to the *C*-terminal linker terminus.



**Figure S5, relates to Figure 3A and 3C. Uncharacterized ligand and anomalous difference density.**

(A) The active site tunnel reveals significant  $F_O - F_C$  omit difference density at  $2.5 \sigma$  (brown) with a featured shape, which does not match components of the crystallization condition. Anomalous difference density at  $3.7 \sigma$  (pink), indicates the presence of sulfur in methionine and cysteine residues as well as in SAH, but no significant difference density as part of the ligand.

(B) The close up view on the active site reveals the expansion of the ligand's difference density along the central tunnel with its end in the junction to the extended substrate cavity. The same contour level of the SAH difference density (blue) suggests either a partial occupancy or various conformations of the less defined ligand. The extent of difference density corresponds approximately to an atomistic binding interface of  $500 \text{ \AA}^2$ .

**Item S1, relates to Figure 1: Revised *M. purpureus* PksCT sequence.** Residues encoded by alternate exon boundary are underlined:

MIDSTSHSNLRSKAFIFGPQDLSFDVRSFNKLHSQLQNHQWLDALASLPKLWDNFAASDQKVQQ  
 SNTGKLENLNAAWISSGVAPEEAFPLPVNLLSPLVVIGQLVEYMTFLKAAFPDLGKKHDLPISIKED  
TETGLCTGTLCAFACAVACSSIADIQHYGAVAARLAMLVGAIVDTEEVLSPEGKSVFSASWNSA  
EFSDFSTHVLETFDAYVSIVDQRRACTLASKKTAPAIIERLKQEGAHTVSIALSGRFHWKKHQD  
 AVSSLIQFCGLDPDLQLADATKMLPSRSSSDGQYITTGKLHELAIRAILEQSEWYKTCRISYLSK  
 FIMDDAAVICFGPERCMPPTLARKLGPRLTYVSEIDISSSRVPGQLGGTQKLNLTDLPDERIAVIG  
 MACRLPGAEDHEGFWEILKTGQSQHREVPEDRFGMATAWREADKRKWYGNFIDNYDTFDHKFF  
 KKSPREMASTDQPQHRLMLQVAYQAVEQSGYFRNNGTNRRIGCFMGVGNVDYEDNIACYPANAY  
 SATGNLKSFLAGKISHHFGWTGPSLTLDTACSSSVAIHQACRSILSGECCNGALAGGVNVITSPNW  
 YHNLAGASFLSPTGQCKPFDAGDGYCRGEVGAVFLKRLSSAIADGDQVFGVIASKVYQNQN  
 CTAITVPNAILSELFTDVVRQARLEPKDITLVEAHGTGAVGDPAEYDGIRAVFGGPIRSDVLSG  
 SVKGLVGHTECASGVVSLIKTLLMIQQGFIPPQASFSSINPSLNAKAEEKIEISTRLPKPWDAPFRAALI  
 NNYGASGSNASMVVTQPPNLTEPSTPLPGKYPFWISAFDQQSQLQSYVRRRLQFLEKHAADKNLS  
 VANLSFQVACQSNWSLPQALVFSASTKEELNRALASFEKGSTDFPSVQLPDKPVILCFGQVSTY  
 VGLDQEYVNSTAILRHYLDQCDAMCLSLGLQSIYPAIFQRSPIEDIVQLQTALFAMQYSCAKAWID  
 SGLKVASVVGHSFGELIALCVSNAVSLKDAVKMISGRARLIKERWGADKGSMAVEADLSDVEAL  
 LAKVKSQMGSETGLAIACYNASKSFTLAGPTKDVDHAENLLKNDPDFSGIRYKRLNVTFNAFHSV  
 VDALIDDLESLGQQGIRFKEPTIKLERATEQUESTSTLNANYVATHMRKPVFFAQVKRLSDKFPVAI  
 WLEAGSNSTITAMASRALGTSNSSFQAVNITSEGAFRFLCDTTVKLWKEGQKVSVWAHHRLQTPM  
 YTPVLLPPYQFEKSRHWMDLKVPBKPEASVQVAEQTAAEAPKGLTFVGYQDASQRSVRFRNV  
 TTEKFNRLLSGHIMANAAAVCPGMFQVEIALDALTSRPEFQARSFIPELHDRHYQPLVRDESRA  
 VWIEAHCPNAEGLVWNWKL~~TASDDKGSGSVHTSGTITFQAA~~DSVQKSEFEKLRRLIGRKRC  
 LLDSNVADDILQGRNIYRATTEVIDYKEIYRHTKIAGRDNESAGRVIKYDGETWLDTVLTDCFC  
 QVAGIFVNLM~~T~~TTKIDL~~S~~ERGIFICDGIDRWLRAPNAGSNNTPSQVYEVFALHHCESDSKYLSDVF  
 DARE GSLVEVALGISYQKV~~S~~ISGIRRVLSKGMPAGLQPQVPTSPA~~VAAIKTV~~SP~~PPV~~ADSP~~L~~VDGS  
 STAVSGTPPTKKAPKAPSVDITGKMREIICNLSGLEPDEVKDDSDLVELGIDSLMSMELAREV~~D~~LA~~F~~  
 KTTIDVTQLIDVTDFRSL~~VECMQRIL~~GIDNQEDNTYLAEGLNGHEGV~~TNGNAY~~HVN~~GTV~~GN  
 GNGVLFPELGG~~SILPK~~SAILDAFRIAKEATDDFILNGQLGTY~~YNEVMR~~STELCVAHIVNAF~~EQ~~QLGC  
 PIRSAAYQRLERVPYLPKHERFMNL~~IY~~GLLEEARLIDINGSEITRTSVP~~STK~~S~~VET~~M~~LE~~ELLHDE~~P~~  
 LHAAEHKLT~~S~~TGSKFADC~~ITG~~KEDGLQLIFGSPEGREIVTDVYAKSPINAVWIQQAEFF~~LE~~QLV  
 LPNTGEPLRILEMGAGTGGTTVKMLPLL~~ERL~~GVPVEY~~T~~MTD~~SS~~LIAARKRFKKY~~P~~FMKF  
 NIESPPDPQLVHSQHILATNCV~~H~~ATRNLEISTRNI~~H~~RILRPDG~~F~~LLLEMTEQ~~V~~PWVDF~~I~~FG~~L~~LEGW  
 WLFEDGRRHALQ~~PA~~THWKKIL~~T~~SVGYGHVD~~W~~TEGTRPEANIQRL~~I~~ALASEPRYD~~H~~TPQSLQ~~P~~VQ  
 VPLTDIAGRQEIIDTYIREYTEDF~~R~~ALPIP~~G~~IQQAVMP~~A~~PTGHC~~V~~L~~T~~GAT~~G~~SLGSHVV~~G~~YLSRLPN  
 VHTVVCLNRRSTV~~P~~ATIRQ~~E~~ALKVRG~~I~~SLDDNSRS~~K~~L~~V~~LEVETAKPLLGLP~~V~~ETYQ~~K~~L~~V~~NTATH  
 IVHSAWPM~~S~~LTRPIRGYESQFKVMQNL~~I~~NLAREVA~~AW~~RPVPFKFSFQFISSIGVVGYYPLRY~~G~~EIIAP  
 EETMTADSVLPVG~~Y~~AEAKLVCERML~~D~~ETLHQ~~Y~~PDR~~R~~PM~~A~~VRIA~~Q~~AG~~S~~T~~N~~GHWN~~P~~VEHFAFLI  
 KSSQTLKALPDFG~~S~~LSWP~~C~~V~~D~~V~~S~~ATL~~G~~ELLISNTTP~~S~~YHIENPSRQQWRKM~~V~~K~~T~~LAQSLD~~I~~PR  
 DGIIPFDQWIERVRN~~S~~SASINDNPARQL~~LE~~FFDQHFIRM~~S~~CGNL~~I~~LD~~T~~TKTREHSATL~~R~~ERGPVG~~G~~PL  
 VEKYISAWKTM~~G~~FLD

### Supplemental Procedures

#### UMA-guided domain dissection

UMAv2.18 was used with the BLOUSUM62 matrix. Secondary structure predictions were generated with ProteinPredict ((Yachdav et al., 2014), <https://www.predictprotein.org/>). Comparative sequences were curated from NCBI using the Conserved Domain Architecture Retrieval Tool (Geer et al., 2002) with PksCT as the query and included the following entries: XP\_001217248, ANID\_07903, ANID\_00523, CAN87161, XP\_001273475, XP\_002381902, EHA28237, XP\_001818926, XP\_001393501, CAK40124, XP\_660990.1, XP\_658638.1, AAR90253.1, BAE66025.1, XP\_001243185.1, CAK48487.1, XP\_001559289.1, XP\_002149737.1, XP\_002567553.1, XP\_002487778.1, XP\_002340070.1, XP\_002384396.1, XP\_002848394.1, XP\_003070229.1, XP\_003010590.1, XP\_001395291.2, EHA25844.1, EHA28237.1, and GAA92425.1.

### Supplemental References

Geer, L.Y., Domrachev, M., Lipman, D.J., and Bryant, S.H. (2002). CDART: protein homology by domain architecture. *Genome Res* 12, 1619-1623.

Yachdav, G., Kloppmann, E., Kajan, L., Hecht, M., Goldberg, T., Hamp, T., Honigschmid, P., Schafferhans, A., Roos, M., Bernhofer, M., et al. (2014). PredictProtein-an open resource for online prediction of protein structural and functional features. *Nucleic Acids Research* 42, W337-W343.