

Supplementary Figures

Supplementary Figure 1

Effect of vagotomy on spleen, blood and lung neutrophils, ELW, and phosphorylation of p65 NF- κ B and AKT1 in splenic neutrophils in LPS-induced ALI. Sham and vagotomized mice were IT challenged with LPS (5 mg/kg). 15 h after IT LPS, mice were killed to assess (A) neutrophils in the spleen; (B) blood neutrophils; (C) lung MPO (an index of lung neutrophil infiltration); ELW (D), and p-P65 NF- κ B (E) and p-AKT1^{ser473} levels (F) in splenic neutrophils. N = 4 in each group; * $P < 0.05$, Student's t test (A-B, D-F) and repeated measures two-way ANOVA was used for statistical analysis (C). Data are presented as mean \pm SD. Representative from two independent experiments.

Supplementary Figure 2

Identification of specificity of rabbit anti- $\alpha 7$ nAChR polyclonal antibody. Lung, blood, and BM cells were isolated from wildtype and *Chrna7*^{-/-} mice and labeled with Alexa Fluor® 488-rabbit anti- $\alpha 7$ nAChR antibody. The whole cell population was gated. Percentage of $\alpha 7$ nAChR⁺ cells was evaluated in lung (A-B), blood (C-D), and BM (E-F) cells. N = 3 in each group. Data are presented as mean \pm SD. Wildtype and *Chrna7*^{-/-} BM cells were labeled with Fluor-633 α -bungarotoxin (Biotium) and the whole cells were gated. The $\alpha 7$ nAChR⁺ cells in the BM cells were calculated (G).

Supplementary Figure 3

A-F. Changes of monocytes (Ly6C^{hi}Ly6G^{int}) and neutrophils (Ly6C^{int}Ly6G^{hi}) in the spleen, BM, and lung from PBS-challenged sham and vagotomized mice. Spleen, BM, and lung cells were isolated from sham + PBS (IT) and vagotomy + PBS (IT) mice at 24 h. The cells were labeled with fluorescent anti-Ly6C and Ly6G antibodies. The whole cell population was gated. Percentage of monocytes (Ly6C^{hi}Ly6G^{int}) and neutrophils (Ly6C^{int}Ly6G^{hi}) in the spleen (A-C), BM (D-F), and lung (G-I) were analyzed. N = 3 in each group. Data are presented as mean \pm SD.

J. Changes of lung MPO, an index of neutrophil infiltration. Sham and vagotomized (right cervical) wildtype mice were IT challenged with PBS and killed at 4, 12, and 24 h post PBS challenge. The lungs were excised and homogenized to measure MPO activity. N = 3 in each group. Data are presented as mean \pm SD.

Supplementary Figure 4

Representative photomicrographs of lung histology in *E. coli* pneumonia compared sham to vagotomized mice.

(A) Effect of vagotomy on infiltration of neutrophils in *E. coli* infected lungs. The sham and vagotomized mice were IT challenged with *E. coli* 2.5×10^6 cfu and then sacrificed at 12 and 24 h. The lungs were collected for immunohistochemistry. Red color: stained with anti-VE-cadherin antibody for labeling blood vessels; green color: labeled with anti-Gr1 antibody for detecting neutrophils. A: Arterioles; B: bronchus; V: vein. Scale bar 50 μ m; objective magnification $\times 20$.

(B) Effect of vagotomy on $\alpha 7$ nAChR⁺Gr1⁺ cells in *E. coli* infected lungs. The lung sections were incubated with primary rabbit anti- $\alpha 7$ nAChR and rat-anti-Gr1 antibodies then labeled with Fluoro488-anti rabbit or Fluor-594 anti rat secondary antibodies. Red color: Gr1⁺; green color: $\alpha 7$ nAChR⁺; objective magnification $\times 40$; Scale bar 20 μ m.

Supplementary Figure 5

Flow cytometry analysis of changes of $\alpha 7$ nAChR⁺CD11b⁺ cell population in peripheral blood from *E. coli*-infected sham, vagotomized and $\alpha 7$ nAChR-agonist supplemented vagotomized mice. Experimental setting was established in **Figure 4**. The blood cells were isolated and labeled with corresponding fluorescent antibodies. The whole cell population was divided into three gates: lymphocyte, monocyte, and PMN based on size and granularity of cells (**A**, **D**, **G**). The $\alpha 7$ nAChR⁺CD11b⁺ cells from each gate were subgated (**B**, **E**, **H**). The percentage of $\alpha 7$ nAChR⁺CD11b⁺ cells from each subgate was presented (**C**, **F**, and **I**). *P < 0.05, one-way ANOVA with *Bonferroni post hoc* test. Data are presented as mean \pm SD.

Supplementary Figure 6

Activation of $\alpha 7$ nAChR promotes phosphorylation of p-AKT1 at Ser473 site in granulocytes and suppresses cytokine production.

A-B. The wildtype and *Chrna7*^{-/-} splenic neutrophils were pretreated with $\alpha 7$ nAChR specific agonist-PHA568487 and then challenged them with LPS and separate cytoplasm and nucleus 1 h later. P-AKT1^{Ser473} was measured in cytoplasm (**A**) and nucleus (**B**) by ELISA. **C.** The supernatant was collected to measure CXCL2 levels by ELISA in LPS-challenged splenic neutrophils pretreated with different concentration of PHA568487. Representative of two experiments. **D.** The splenic neutrophils were pretreated with Wortmannin for 15 min and then challenged with LPS. The

supernatant was collected to measure CXCL2 levels by ELISA 4 h later. N = 8 in each group. ****P** < 0.01. **E.** The isolated splenic neutrophils were challenged with LPS (1 μ m). The supernatant was collected to measure CXCL2 and TNF- α levels by ELISA 4 h later. N = 3 in each group. ****P** < 0.01, ***P** < 0.05. Student's t test and data are presented as mean \pm SD.

Supplementary Figure 7

Deficiency of AKT1 worsens BAL profiles. The wildtype and *Akt1*^{-/-} mice were challenged with an IT *E. coli* (2.5×10^6 cfu). The mice were killed at 24 h after *E. coli* challenge. The BAL was collected to measure protein (**A**), *E. coli* colonies (**B**), and TNF- α level (**C**). N = 5 in each group. ***P** < 0.05, Student's t test. Data are presented as mean \pm SD.

Supplementary Figure 8

Effect of deletion of Chrna7 and vagotomy on BAL profiles and splenic p-AKT1, p-STAT3 and p-ERK levels in E. coli pneumonia. The wildtype, *Chrna7*^{-/-}, and vagotomized mice were respectively challenged with an intratracheal *E. coli* (2.5×10^6 cfu). The mice were killed at 24 h after *E. coli* challenge. The BAL was collected and spleens were harvested and homogenized. The supernatant of BAL was used to measure protein (**A**); *E. coli* cfu (**B**); TNF- α (**C**). The p-AKT1^{Ser473} (**D**), p-STAT3 (**E**) and p-ERK (**F**) levels in the supernatant of spleen homogenate were measured by ELISA. N = 4-5 in each group, ***P** < 0.05, Student's t test. Data are presented as mean \pm SD.

Supplementary Figure 9

A-D. Deletion of Chrna7 or Akt1 worsens acute lung infection and inflammation. The wildtype, *Chrna7*^{-/-}, or *Akt1*^{-/-} mice were respectively challenged with an intratracheal *E. coli* (2.5×10^6 cfu). The mice were killed at d1 and d2 after *E. coli* challenge to measure ELW (**A**). Blood was collected to analyze (**B**) blood neutrophils; (**C**) blood monocytes at d1. N = 4-5 in each group, ***P** < 0.05, ****P** < 0.01, student's t test. Data are presented as mean \pm SD. The lungs were harvested and homogenized to measure MPO activity in the supernatant at d1 (**D**). N = 5 in each group, ***P** < 0.05, Repeated Measures of Two-way ANOVA. Data are presented as mean \pm SD.

E. Change of survival in E. coli-infected wildtype, Akt1^{-/-}, and Chrna7^{-/-} mice. The wildtype, *Akt1*^{-/-}, and *Chrna7*^{-/-} mice were IT challenged with *E. coli* (5×10^6 cfu) and followed up for 5 days. N = 10-23 in each group. Data are presented as mean \pm SD. Log-rank test.

Supplementary Figure 10

A-B. Flow cytometry analysis of splenic CD4⁺ cells during *E. coli* pneumonia. Using the same experimental setting as Fig. 7A-D, the four groups of spleen cells were submitted to flow cytometric analysis. **A.** lymphocyte population was gated. **B.** Percentage of splenic CD4⁺ cells. N = 4 in each group, **P* < 0.05. Data are presented as mean ± SD.

C-D. p-AKT1 and CHAT expression in the *E. coli*-infected *Chrna7*^{-/-}*Itgam*^{-/-} spleens. Spleens were collected from *E. coli*-infected wildtype and *Chrna7*^{-/-}*Itgam*^{-/-} mice to perform western blotting at 24 h post *E. coli* IT infection. N = 3-4 in each group, **P* < 0.05. Data are presented as mean ± SD. Image J software was used for analyzing optical density.

Supplementary Figure 11

A-B. Effect of vagotomy and double deletion of *Chrna7* and *Itgam* on lung *Il22* and *Cxcl2* mRNA during *E. coli* pneumonia. The sham, vagotomized, and *Chrna7*^{-/-}*Itgam*^{-/-} mice were IT challenged with *E. coli* (2.5 × 10⁶ cfu). The wildtype mice receiving an IT PBS were used control. The mice were killed at 24 h after *E. coli* or PBS challenge. Lungs were excised to extract RNA for real-time PCR for *Il22* (**A**) and *Cxcl2* (**B**) mRNA. N = 3-4 in each group. Data are presented as mean ± SD.

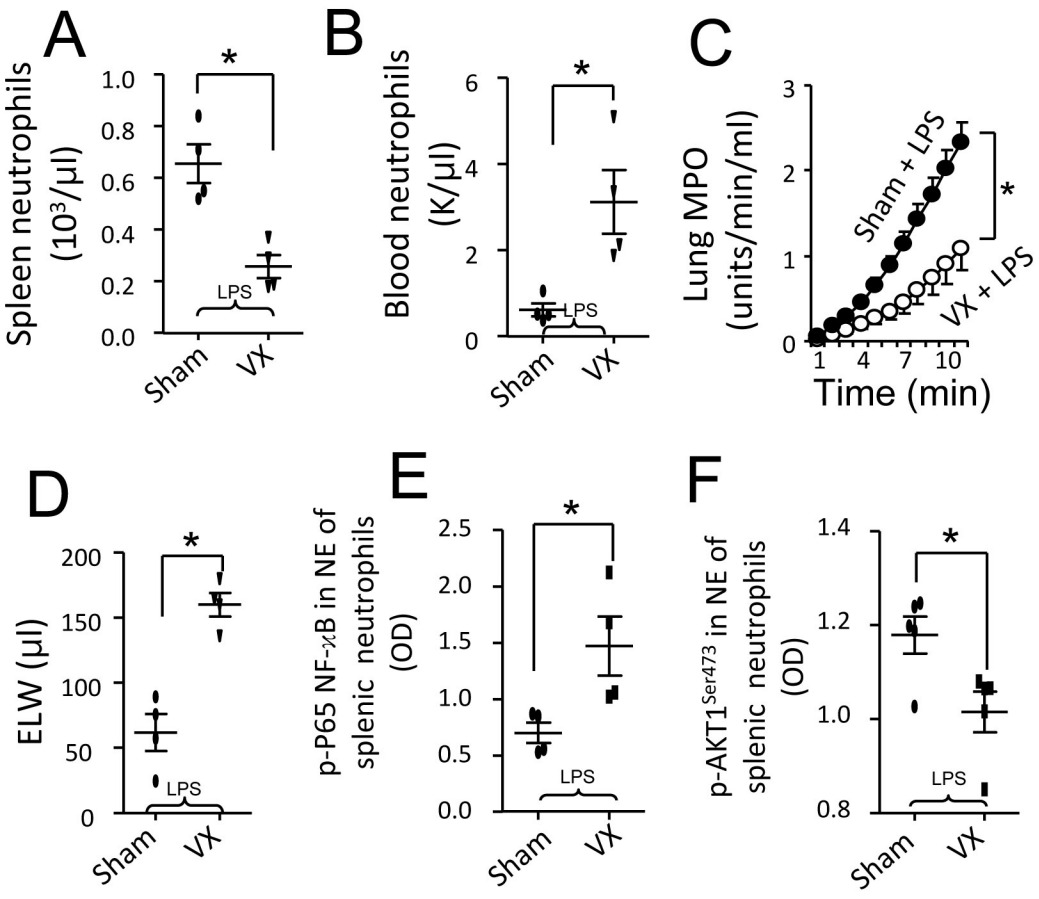
C-E. Effect of vagotomy and double deletion of *Chrna7* and *Itgam* on bone marrow monocytes and neutrophils during *E. coli* pneumonia. The sham, vagotomized, and *Chrna7*^{-/-}*Itgam*^{-/-} mice were IT challenged with *E. coli* (2.5 × 10⁶ cfu). The wildtype mice receiving an IT PBS were used control. The mice were killed at 24 h after *E. coli* or PBS challenge. BM cells were isolated and labeled with fluorescent anti-Ly6C and Ly6G antibodies. **C. Gating strategies for monocytes and neutrophils; D. Percentage of Ly6C^{hi}Ly6G^{int} monocytes; E. Percentage of Ly6C^{int}Ly6G^{hi} neutrophils.** N = 3-4 in each group. Data are presented as mean ± SD.

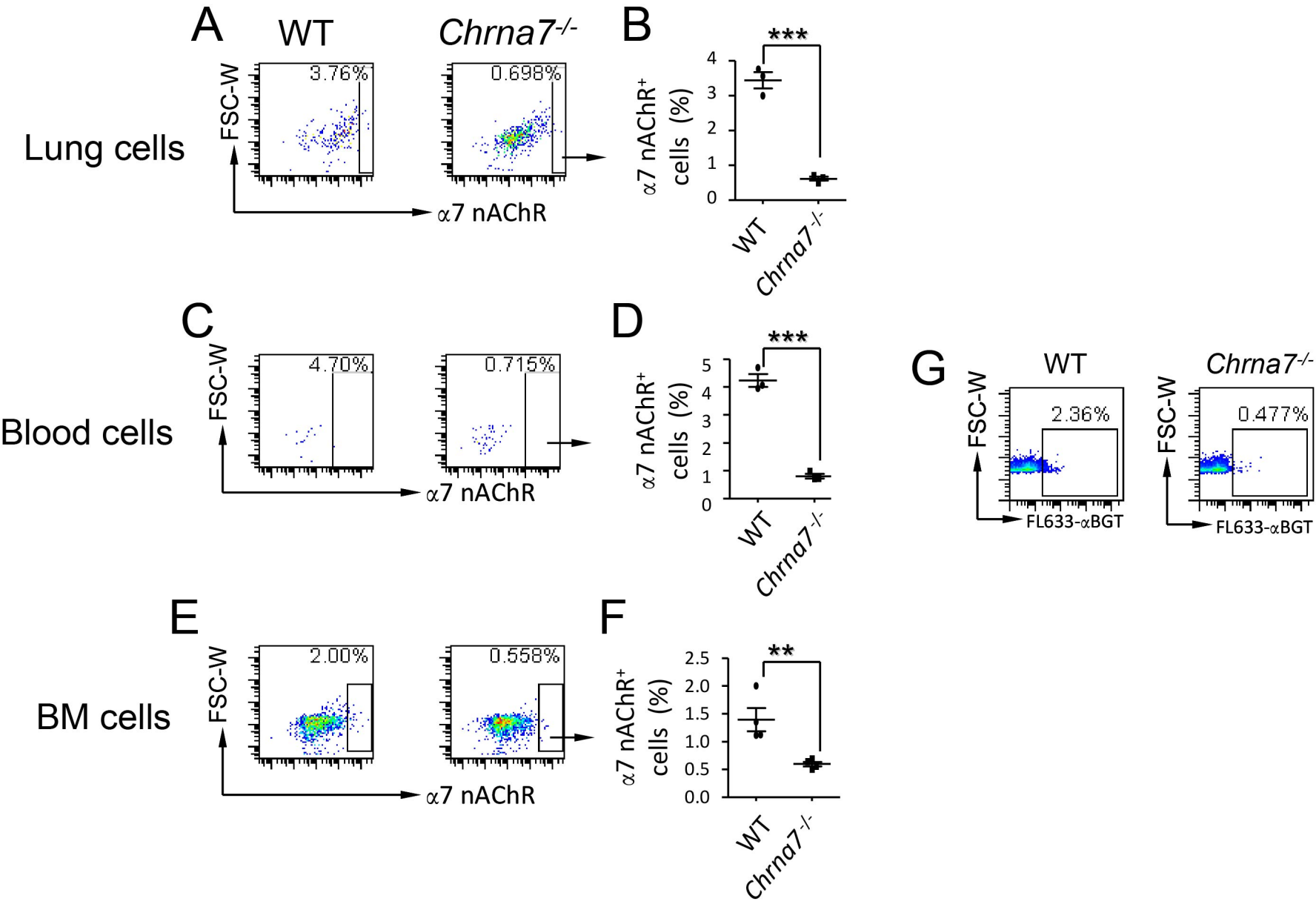
Supplementary Figure 12

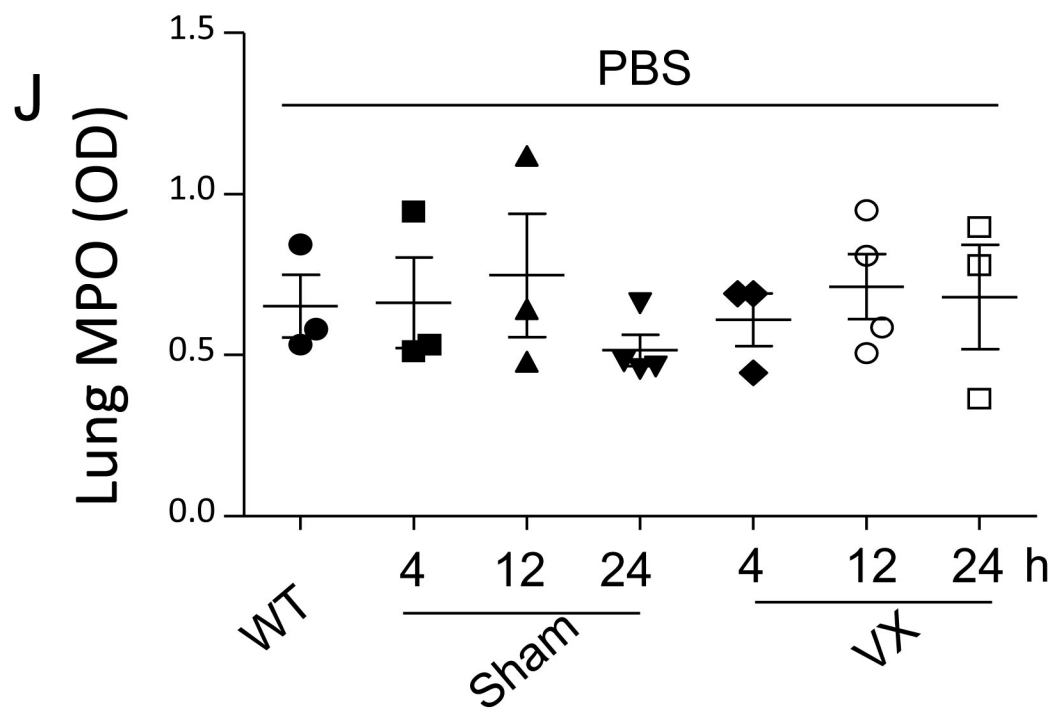
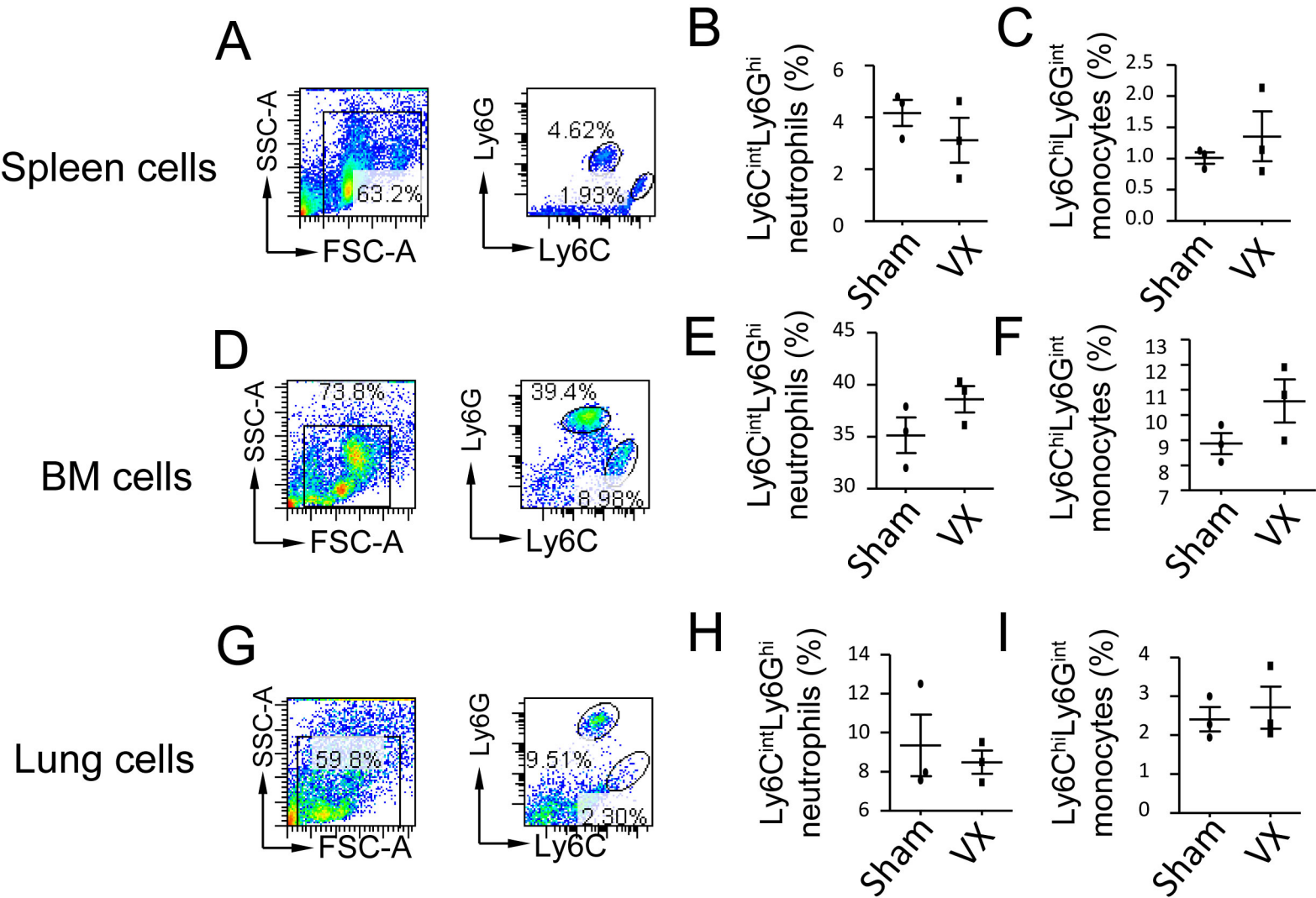
TNF-α production in LPS-challenged peritoneal macrophages. Peritoneal macrophages were isolated from wildtype, *Chrna7*^{-/-}, *Itgam*^{-/-}, and *Chrna7*^{-/-}*Itgam*^{-/-} mice. The mice were ip injected with 3 ml of 3% (w/v) Brewer thioglycollate medium into the peritoneal cavity. Peritoneal lavage was performed 3 days post thioglycollate injection. The purified wildtype, *Chrna7*^{-/-}, *Itgam*^{-/-}, and *Chrna7*^{-/-}*Itgam*^{-/-} macrophages were separately challenged with LPS (1 μM) for 4 h. TNF-α in the supernatant of media was measured by ELISA. N = 3 in each group. Data are presented as mean ± SD.

Supplementary Figure 13

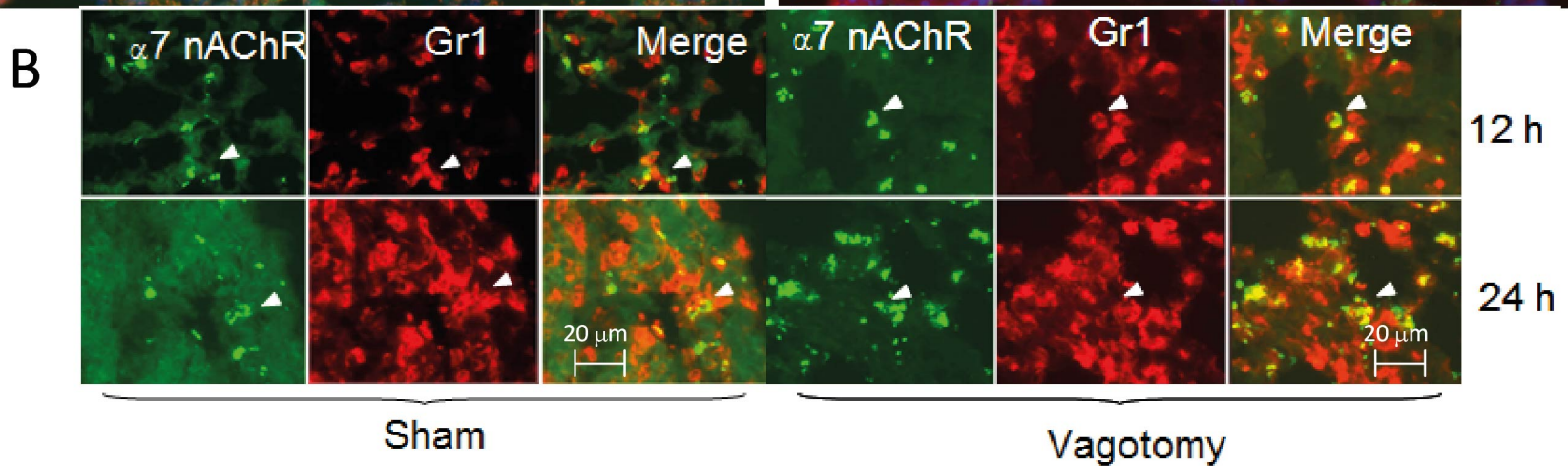
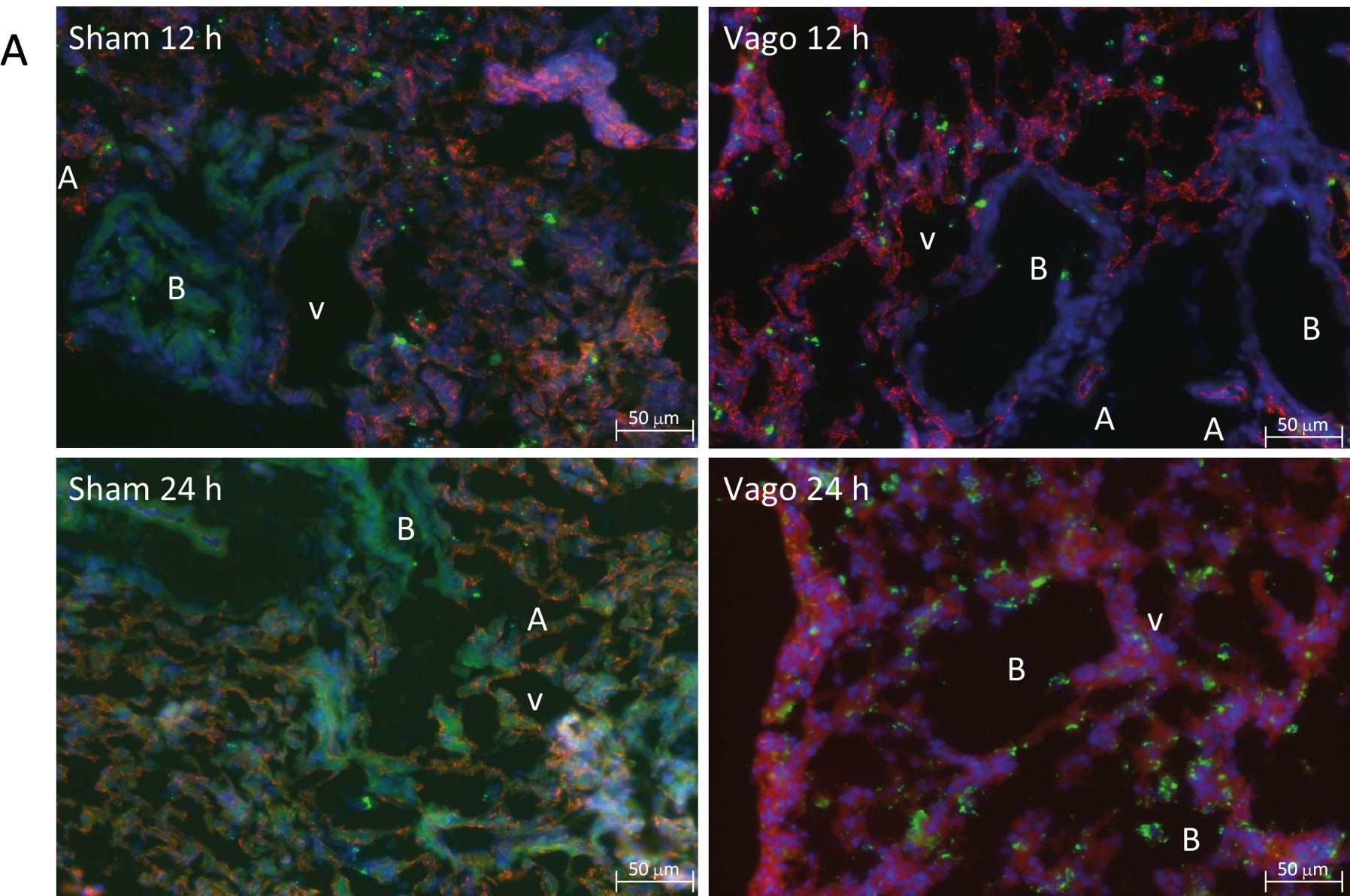
Vagal- $\alpha 7$ nAChR signals via AKT1 regulate $\alpha 7$ nAChR⁺CD11b⁺ cells by which dampen acute lung inflammatory responses. In the intact condition, vagus signals trigger release of acetylcholine in the spleen, and this event leads to phosphorylation of AKT1 in $\alpha 7$ nAChR⁺CD11b⁺ cells and confines these cells in the spleen. Disruption of vagal circuits reduces phosphorylation of AKT1 in splenic $\alpha 7$ nAChR⁺CD11b⁺ cells and facilitates spleen egress and lung recruitment of these cells. Without functional vagal circuits or $\alpha 7$ nAChR agonist stimulation, accumulated $\alpha 7$ nAChR⁺CD11b⁺ cells in the LPS or *E. coli* challenged lung lack of anti-bacterial propriety and propagate inflammatory responses.

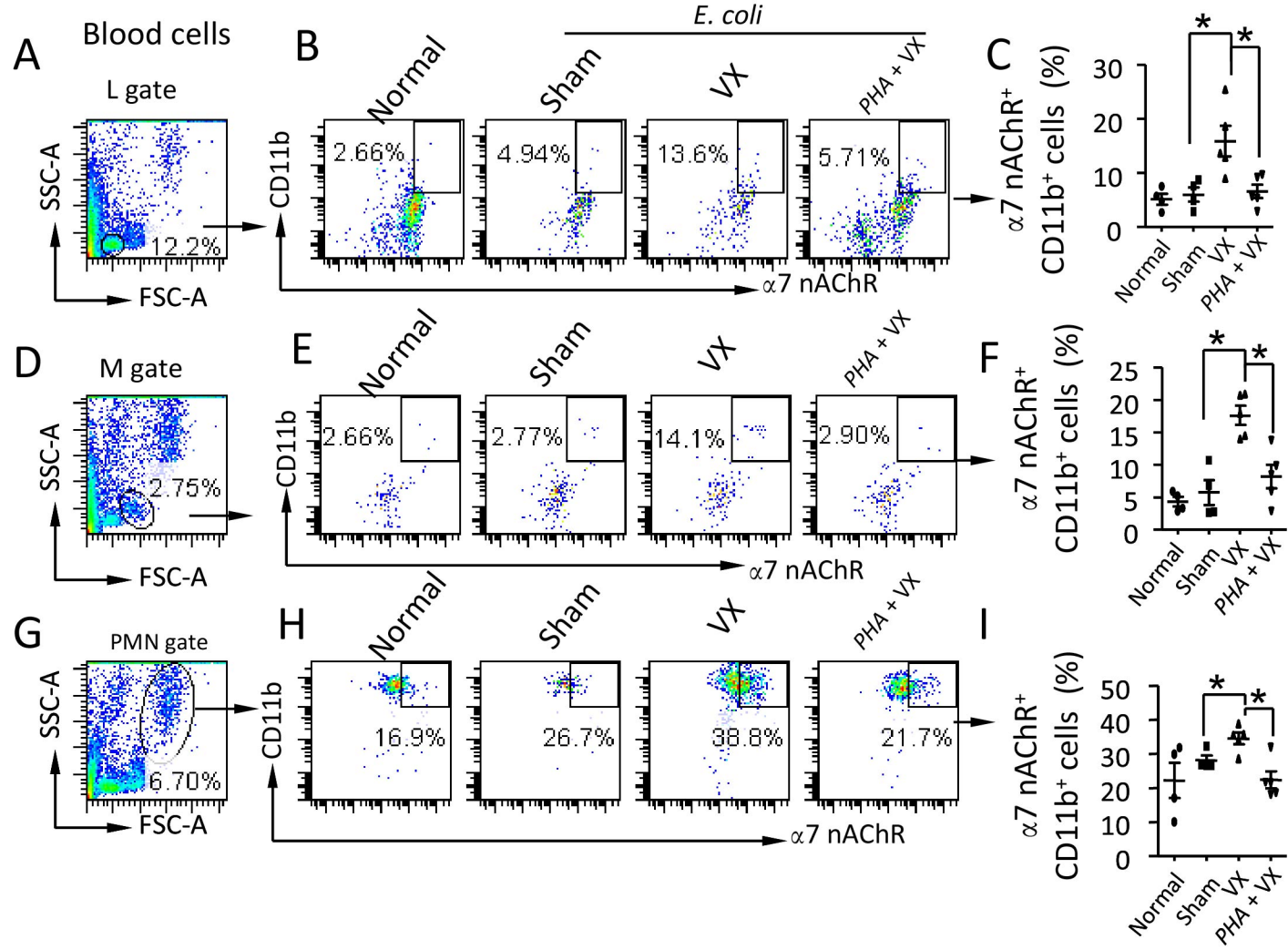


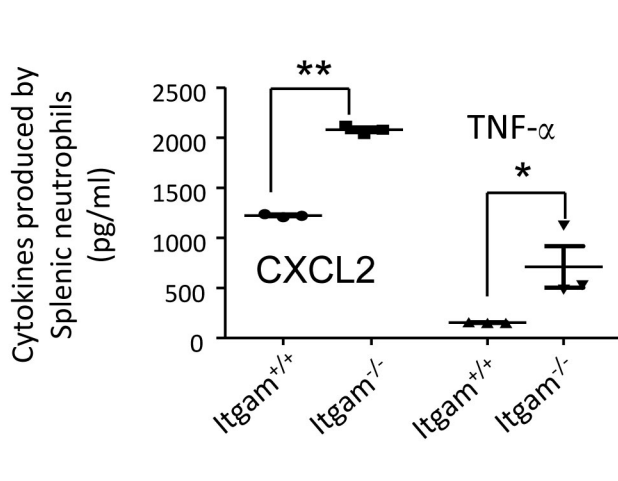
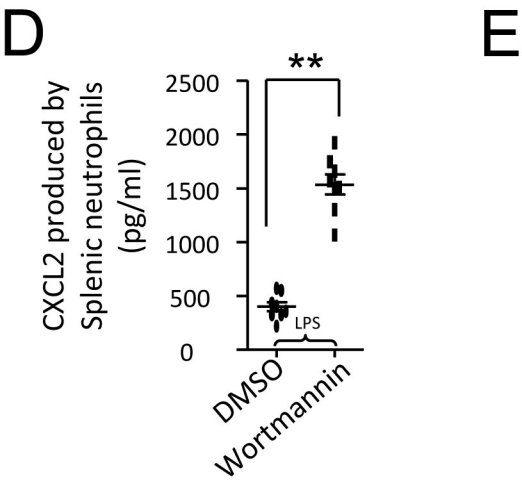
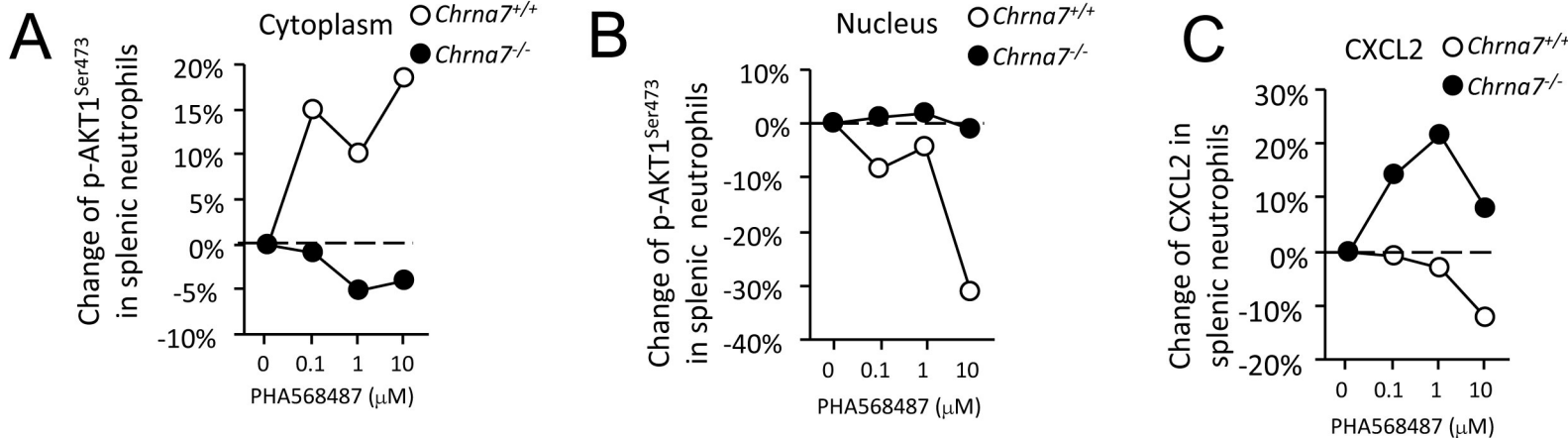




VE-cadherin/ $\alpha 7$ nAChR

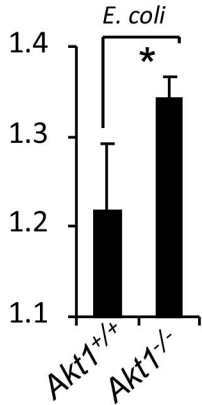
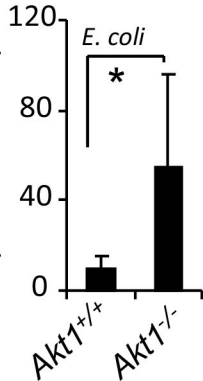
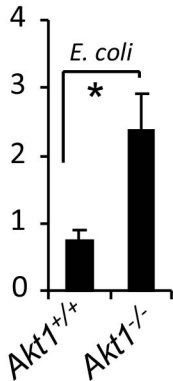


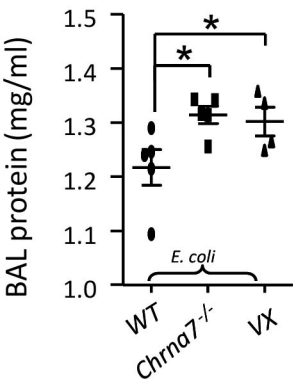
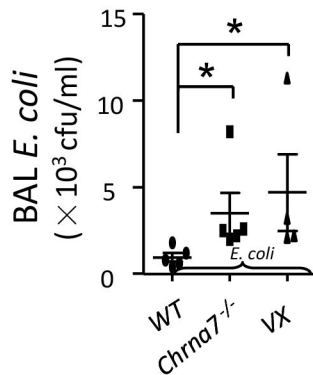
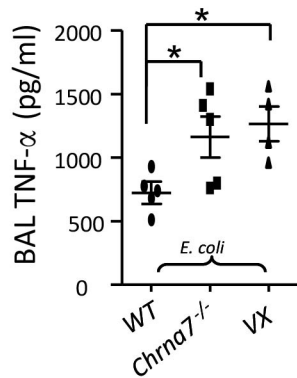
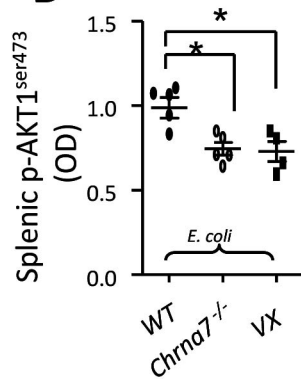
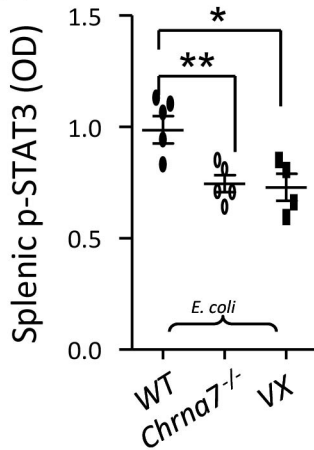
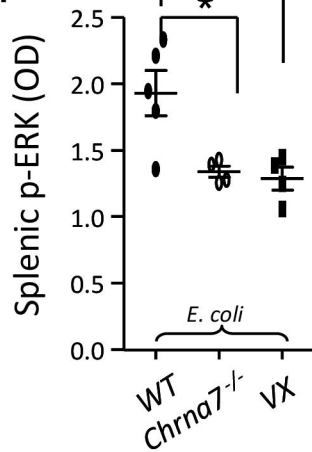


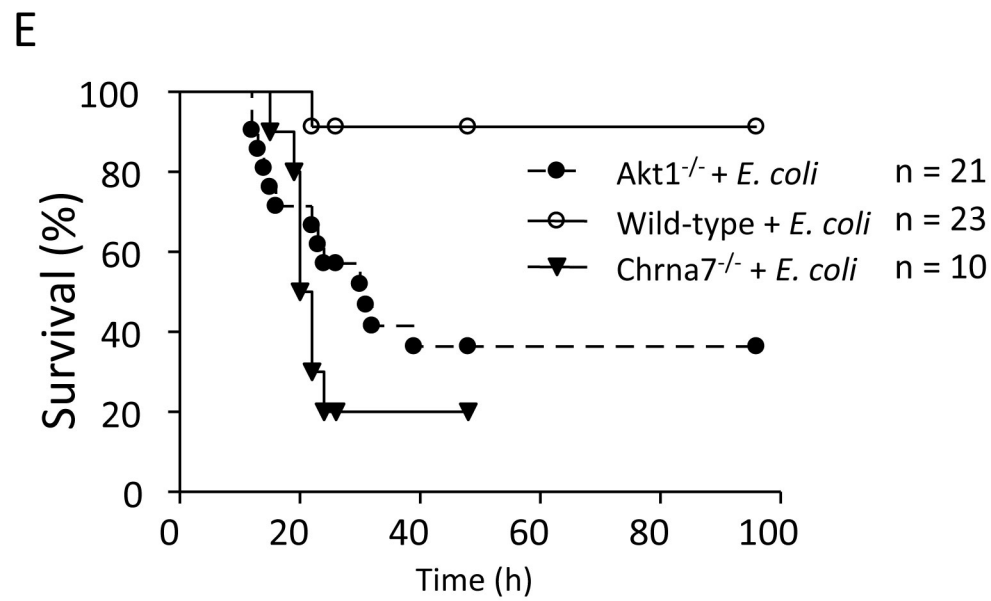
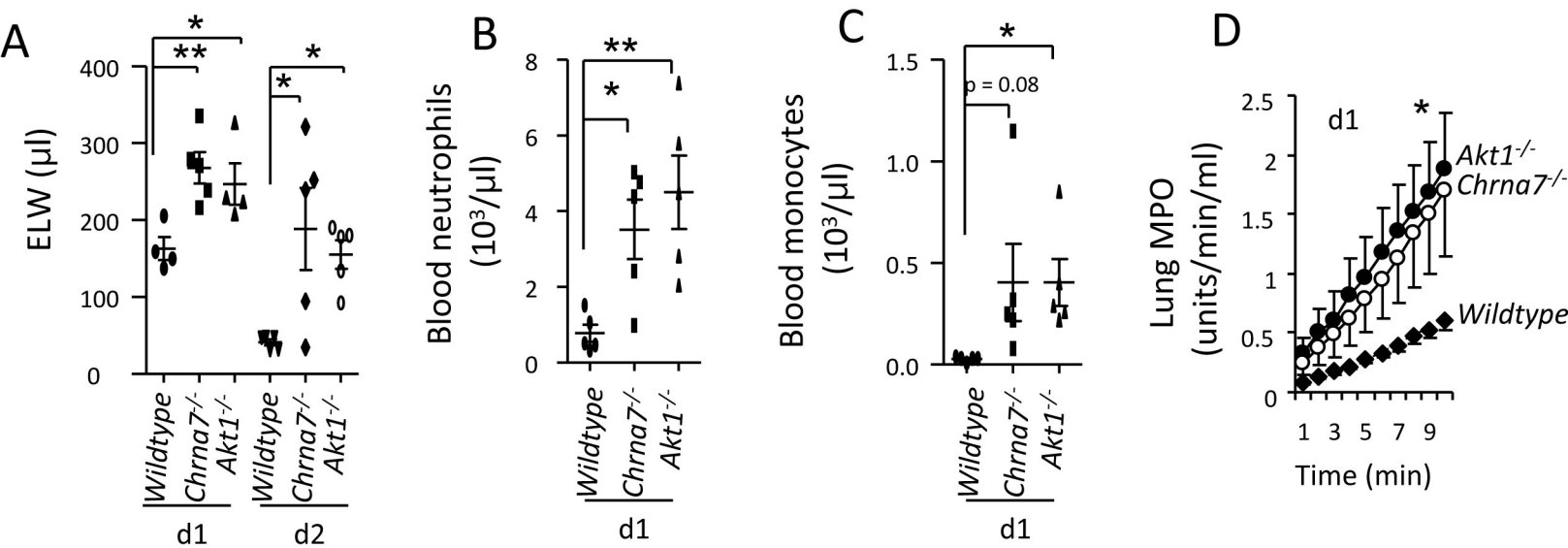


A

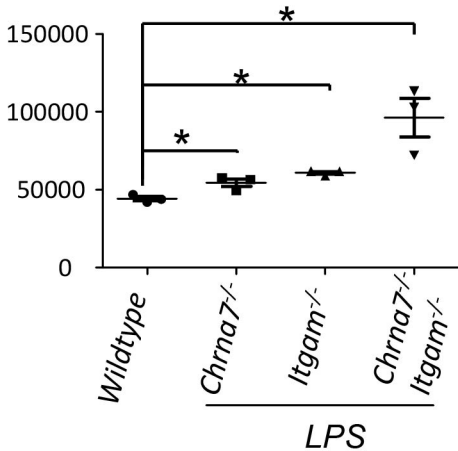
BAL protein (mg/ml)

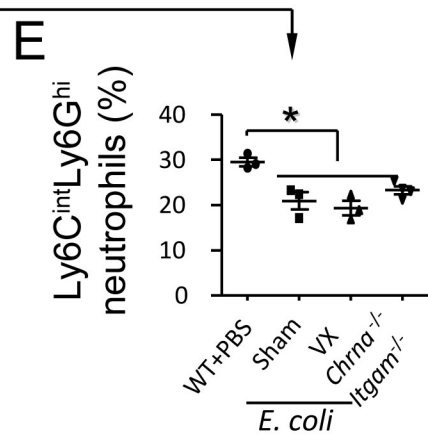
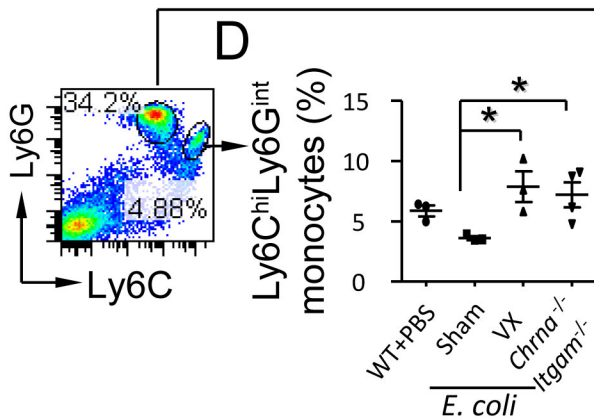
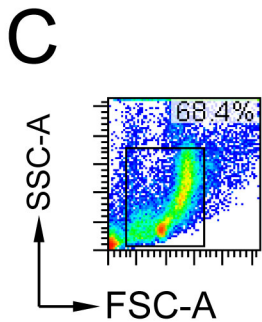
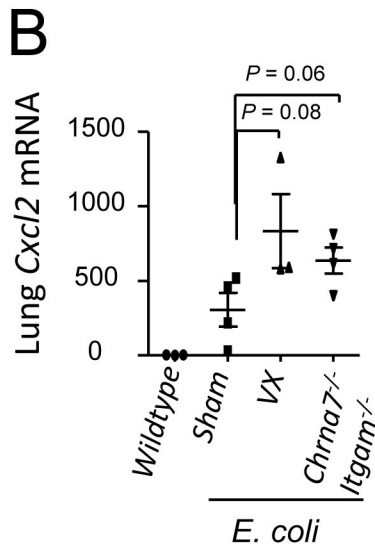
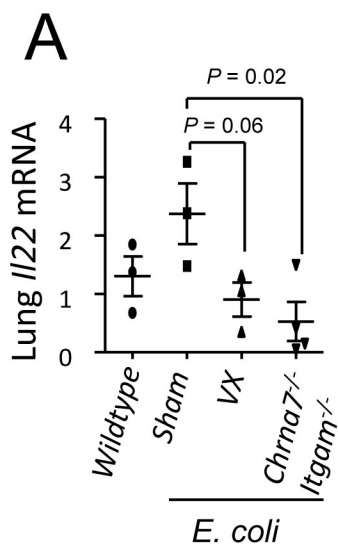
**B**BAL *E. coli* $(\times 10^3 \text{ cfu/ml})$ **C**BAL TNF- α $(\times 10^3 \text{ pg/ml})$ 

A**B****C****D****E****F**



TNF- α levels
(pg/ml)





TNF- α levels
(pg/ml)

