

Supplementary Material

Table S1. Primers used to link genomic contigs.

Name	Primer sequence (5' – 3')
B1	GAGGTGCCCTCTTCCTTTCT
B2	TGGTAACTTCCAATCACCTCT
B3	TTTACGTTTGTTAGAGCGAAGTC
B4	GGGACTTGAACCGCAACTTC
B5	CGTTCATGCAATGAAAACAGAAGC
B6	CGGCTGATTACACCTCCGTG
P1	AGTCTCTCTAGCCGGTATTAGT
P2	TGCCAAGACGAATACCTGACT
P3	GTGACGTACGAAGTATGTTCCCT
P4	GTA CTCCAGTGGCTCCTACG
P5	GGACATCATAGAAACATAATCTAACCTATC
P6	CATCAAATCTAAATCACTATACTCATATG

Table S2. GenBank accession numbers of protein sequences used to build hidden Markov models

Species	ccmC	ccmF	TatC	Rpl10	Rps4
<i>Andalucia godoyi</i>	YP_007890513.1	YP_007890514.1 AGH24008.1	YP_007890501.1	YP_007890474.1	YP_007890498.1
<i>Histiona aroides</i>	YP_007890585.1 AGH24079.1	YP_007890584.1 AGH24078.1	YP_007890573.1	YP_007890543.1	YP_007890570.1
' <i>Jakoba bahamiensis</i> '	YP_007890628.1	YP_007890629.1 AGH24123.1	YP_007890642.1	YP_007890649.1	YP_007890639.1 YP_007890639.1
<i>Jakoba libera</i>	YP_007890700.1	YP_007890699.1 AGH24193.1	YP_007890756.1	YP_007890732.1	YP_007890684.1
<i>Naegleria fowleri</i>	AOS85673.1 AOS85627.1 YP_007890053.1 AFP72326.1	AOS85658.1 AOS85612.1 YP_007890060.1 AFP72333.1	N/A	N/A	AOS85659.1 AOS85613.1 YP_007890062.1 AFP72335.1
<i>Naegleria gruberi</i>	AAG17810.1 NP_066532.1	AAG17817.1 NP_066539.1	AAG17818.1	N/A	AAG17819.1
<i>Reclinomonas americana</i>	NP_044790.1 AAD11905.1 AGH24367.1 AGH24434.1 AGH24300.1	NP_044789.1 AAD11904.1 AGH24366.1 AGH24433.1 AGH24299.1	AAD11893.1 NP_044778.1 AGH24355.1 AGH24422.1 AGH24288.1 O21266.1	NP_044748.1 AAD11863.1 AGH24325.1 AGH24392.1 AGH24258.1	NP_044775.1 AAD11890.1 O21263.1 AGH24352.1 AGH24419.1 AGH24285.1
' <i>Seculamonas ecuadoriensis</i> '	YP_007890768.1	YP_007890781.1 AGH24476.1	YP_007890761.1 AGH24456.1	YP_007890790.1	YP_007890767.1
<i>Tsukubamonas globosa</i>	N/A	N/A	N/A	N/A	YP_009004138.1

Table S3. Primers used to confirm RNA editing sites in amoeba BB2

Name	Forward primer sequence (5'-3')	Reverse primer sequence (5'-3')	Position on mtDNA
B1	GCCAGGAAGTGGTGGTATTCT	CCGCTATTCGACGCAAAAACA	30511-31294
Br5	AGTGCGACTAGTGCTGAACC	GATGCGATTGTTGAAGGCC	33489-34977
Br7	CGGCTACCGTCTAAACGAGG	GCAACTTCTTTGCAGTTTCCTG	35051-35692
Br9	GTTCTCAAGCGTTGGACCGT	AAGTGAAAATGCAAAATCGAAGTCA	36580-37216
Br13	TCTTTATGTTTCATTGCGCTATTTGT	ACGTTTTCCGTTCTTAAAAGTCCA	38703-40109
Br15	TGGACTIONTTAAGAACGGAAAACG	TCAAACATTCATTACACACAGCTT	40086-41253
Br21	AGGTTTGCCAAAGACGGTGA	TTCCACATACGTGGGTTCCGG	55272-55645
Br23	GTCCATGTAGGTACGCCGAA	AACGCCCTTCGCTATAAGCA	61595-62301

Table S4. Number of editing sites for each gene by nucleotide type.

Gene	Length (bp)	A	U	G	C	Total
<i>atp1</i>	2442	5	0	13	1	19
<i>atp3</i>	861	4	0	6	1	11
<i>atp6</i>	756	1	0	4	1	6
<i>atp8</i>	381	0	0	1	0	1
<i>atp9</i>	231	0	0	2	0	2
<i>cob</i>	1203	0	1	9	0	10
<i>cox1</i>	2016	1	1	23	2	27
<i>cox11</i>	411	0	0	1	0	1
<i>cox2</i>	744	0	0	7	1	8
<i>cox3</i>	1455	0	0	11	0	11
<i>nad1</i>	1281	0	0	11	1	12
<i>nad11</i>	2112	0	0	14	3	17
<i>nad2</i>	1533	0	3	8	0	11
<i>nad3</i>	465	0	0	3	0	3
<i>nad4</i>	1407	0	0	12	0	12
<i>nad4L</i>	345	0	0	3	0	3
<i>nad5</i>	1986	1	0	16	1	18
<i>nad6</i>	642	0	0	6	1	7
<i>nad7</i>	1203	0	0	12	2	14
<i>nad8</i>	483	1	0	4	1	6
<i>nad9</i>	669	2	0	2	0	4
<i>rpl10</i>	582	0	1	4	0	5
<i>rpl11</i>	516	1	0	2	0	3
<i>rpl14</i>	369	1	0	4	0	5
<i>rpl16</i>	396	0	0	3	0	3
<i>rpl2</i>	807	0	0	11	0	11
<i>rpl32</i>	177	0	0	0	0	0
<i>rpl5</i>	525	0	0	5	0	5
<i>rpl6</i>	687	0	0	7	0	7
<i>rps10</i>	648	0	0	2	0	2
<i>rps11</i>	2139	0	0	14	0	14
<i>rps12</i>	363	1	0	3	0	4
<i>rps13</i>	444	0	0	3	0	3
<i>rps14</i>	306	1	0	3	0	4
<i>rps19</i>	282	0	0	3	0	3
<i>rps2</i>	993	0	0	5	0	5
<i>rps3</i>	3030	1	0	13	1	15
<i>rps7</i>	930	0	0	7	0	7
<i>rps8</i>	396	0	1	2	0	3
<i>sdh2</i>	768	0	0	6	1	7

Gene	Length (bp)	A	U	G	C	Total
<i>orf164</i>	495	0	0	4	0	4
<i>orf242</i>	729	1	1	3	1	6
<i>orf3630</i>	10893	9	5	71	3	88
<i>orf925</i>	2778	2	1	12	0	15
<i>rnl</i>	2840	0	0	20	2	22
<i>rns</i>	1654	0	1	15	2	18
<i>trnA(ugc)</i>	73	0	0	0	0	0
<i>trnC(gca)</i>	71	0	0	1	0	1
<i>trnD(guc)</i>	74	0	0	1	0	1
<i>trnE(uuc)</i>	73	0	0	0	0	0
<i>trnF(gaa)_1</i>	72	0	0	1	0	1
<i>trnF(gaa)_2</i>	75	0	0	0	0	0
<i>trnG(ucc)</i>	71	0	0	1	0	1
<i>trnH(gug)</i>	74	0	0	0	0	0
<i>trnI(gau)</i>	73	0	0	1	0	1
<i>trnK(uuu)</i>	72	0	0	1	0	1
<i>trnL(caa)</i>	82	0	0	0	0	0
<i>trnL(gag)</i>	74	0	0	0	0	0
<i>trnL(uaa)</i>	84	0	0	1	0	1
<i>trnL(uag)</i>	74	0	0	0	0	0
<i>trnM(cau)_1</i>	72	0	1	0	0	1
<i>trnM(cau)_2</i>	73	0	0	0	0	0
<i>trnN(guu)</i>	73	0	0	0	0	0
<i>trnP(ugg)</i>	74	0	0	1	0	1
<i>trnQ(uug)</i>	72	0	0	0	0	0
<i>trnR(ucu)</i>	74	0	0	1	0	1
<i>trnS(gcu)</i>	85	1	0	0	0	1
<i>trnS(gga)</i>	85	0	0	0	0	0
<i>trnS(uga)</i>	82	0	0	0	0	0
<i>trnW(cca)</i>	74	0	0	0	0	0
<i>trnY(gua)</i>	83	0	0	1	0	1
intergenic region	83	1	0	0	0	1
Total		34	16	400	25	475

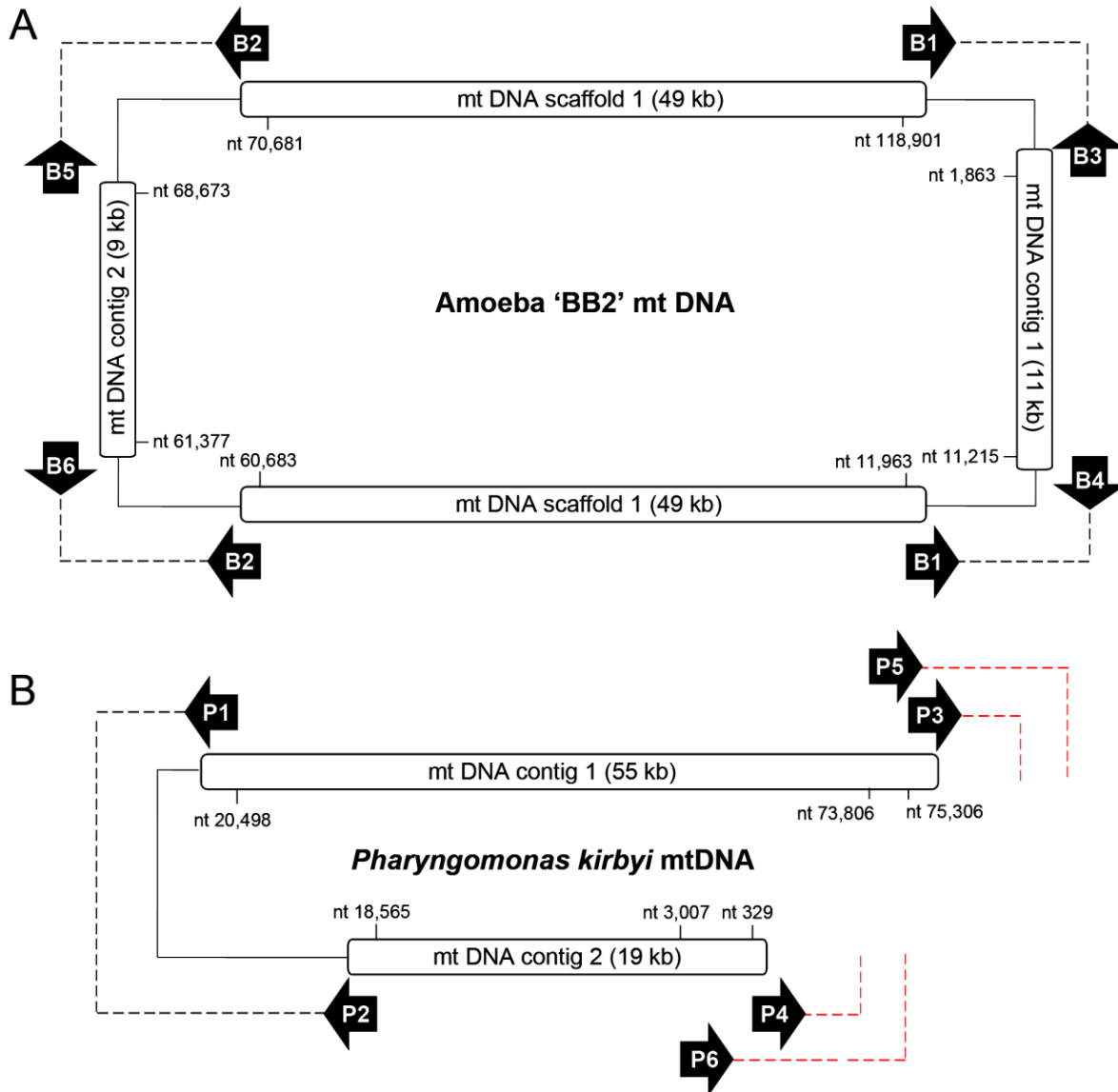


Figure S1. Schematic maps demonstrating how gaps in (A) amoeba 'BB2' and (B) *P. kirbyi* mitochondrial genomes were closed after sequencing of long-range PCR amplicons amplified with primers (black arrows, nucleotide numbers indicate the positions of binding sites) listed in Table S1. The set of PCR reactions that did not yield products is highlighted in red dashed lines.

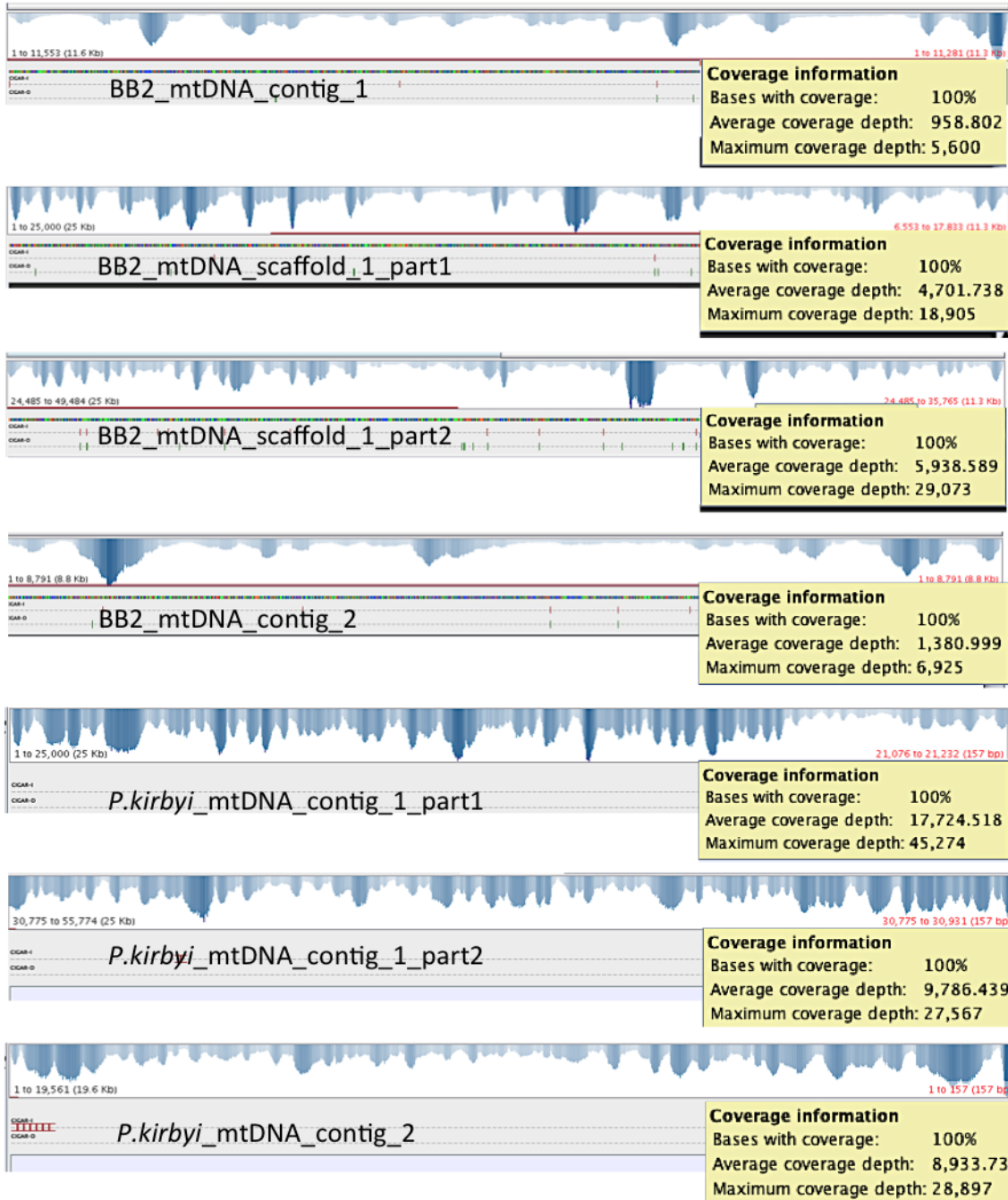


Figure S2. Assembly coverage plots for the mitochondrial genome of BB2 and *P. kirbyi* by contigs (shown in Figure S1).

	S10													Spc
<i>Rickettsia</i>	<i>rps10</i>	---	<i>rpl3</i>	<i>rpl4</i>	<i>rpl23</i>	<i>rpl2</i>	<i>rps19</i>	<i>rpl22</i>	---	<i>rps3</i>	<i>rpl16</i>	<i>rpl29</i>	<i>rps17</i>	<i>rpl14</i>
<i>Andalucia</i>	<i>rps10</i>	---	---	---	---	<i>rpl2</i>	<i>rps19</i>	---	---	<i>rps3</i>	<i>rpl16</i>	---	---	<i>rpl14</i>
<i>Tsukubamonas</i>	---	---	---	---	---	<i>rpl2</i>	<i>rps19</i>	---	---	<i>rps3</i>	<i>rpl16</i>	---	---	<i>rpl14</i>
<i>Amoeba</i> BB2	<i>rps10</i>	---	---	---	---	<i>rpl2</i>	<i>rps19</i>	---	<i>urf219</i>	<i>rps3</i>	<i>rpl16</i>	---	---	<i>rpl14</i>
<i>Pharyngomonas</i>	<i>rps10</i>	---	---	---	---	<i>rpl2</i>	<i>rps19</i>	---	---	<i>rps3</i>	<i>rpl16</i>	---	---	<i>rpl14</i>
<i>Naegleria</i>	<i>rps10</i>	<i>rpl11</i>	---	---	---	<i>rpl2</i>	<i>rps19</i>	---	---	<i>rps3</i>	<i>rpl16</i>	---	---	<i>rpl14</i>

	Spc										Alpha			
<i>Rickettsia</i>	<i>rpl24</i>	<i>rpl5</i>	<i>rps14</i>	<i>rps8</i>	<i>rpl6</i>	<i>rpl18</i>	<i>rps5</i>	<i>rpl30</i>	<i>rpl15</i>	<i>secY</i>	<i>adk</i>	<i>rps13</i>	<i>rps11</i>	<i>rpoA</i>
<i>Andalucia</i>	---	<i>rpl5</i>	<i>rps14</i>	<i>rps8</i>	<i>rpl6</i>	<i>rpl18</i>	---	---	---	---	---	<i>rps13</i>	<i>rps11</i>	<i>rpoA</i>
<i>Tsukubamonas</i>	---	<i>rpl5</i>	<i>rps14</i>	<i>rps8</i>	<i>rpl6</i>	---	---	---	---	---	---	<i>rps13</i>	<i>rps11</i>	---
<i>Amoeba</i> BB2	---	<i>rpl5</i>	<i>rps14</i>	<i>rps8</i>	<i>rpl6</i>	---	---	---	---	---	---	<i>rps13</i>	<i>rps11</i>	---
<i>Pharyngomonas</i>	---	<i>rpl5</i>	<i>rps14</i>	<i>rps8</i>	<i>rpl6</i>	---	---	---	---	---	---	<i>rps13</i>	<i>rps11</i>	---
<i>Naegleria</i>	---	<i>rpl5</i>	<i>rps14</i>	<i>rps8</i>	<i>rpl6</i>	---	---	---	---	---	---	<i>rps11</i>	<i>rps13</i>	---

Figure S3. Gene order comparison of *Discoba* mtDNA and α -proteobacteria *Rickettsia prowazekii* in the three contiguous ribosomal protein operons (S10-Spc-Alpha). Genes in reversed order are highlighted in green.

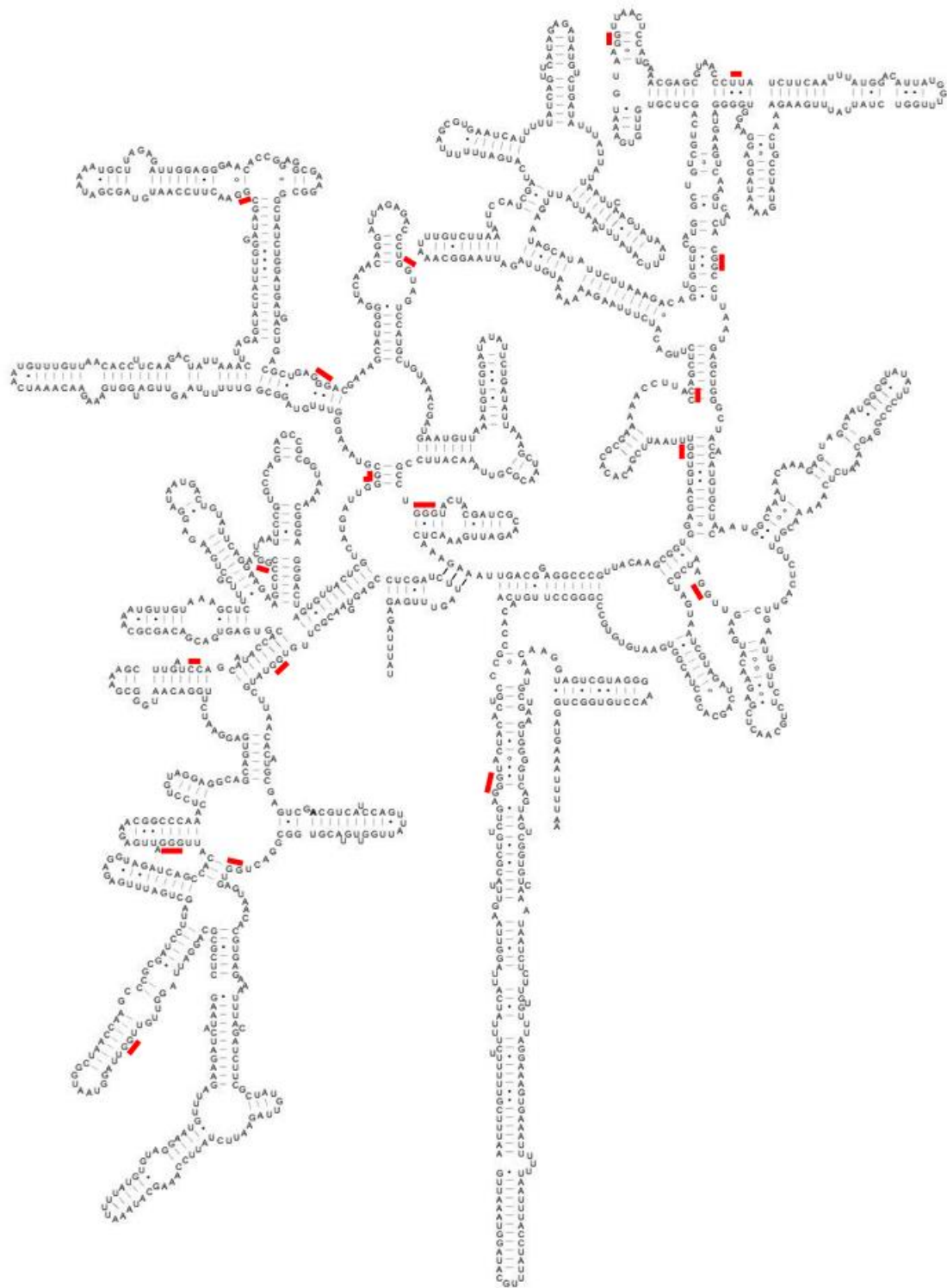


Figure S4. Predicted secondary structure of BB2 mitochondrial SSU rRNA. Regions including an editing site are indicated with red lines. For each highlighted region, only one nucleotide is inserted (the exact insertion sites are unknown since nucleotides are inserted next to one or more encoded nucleotides with the same identity).

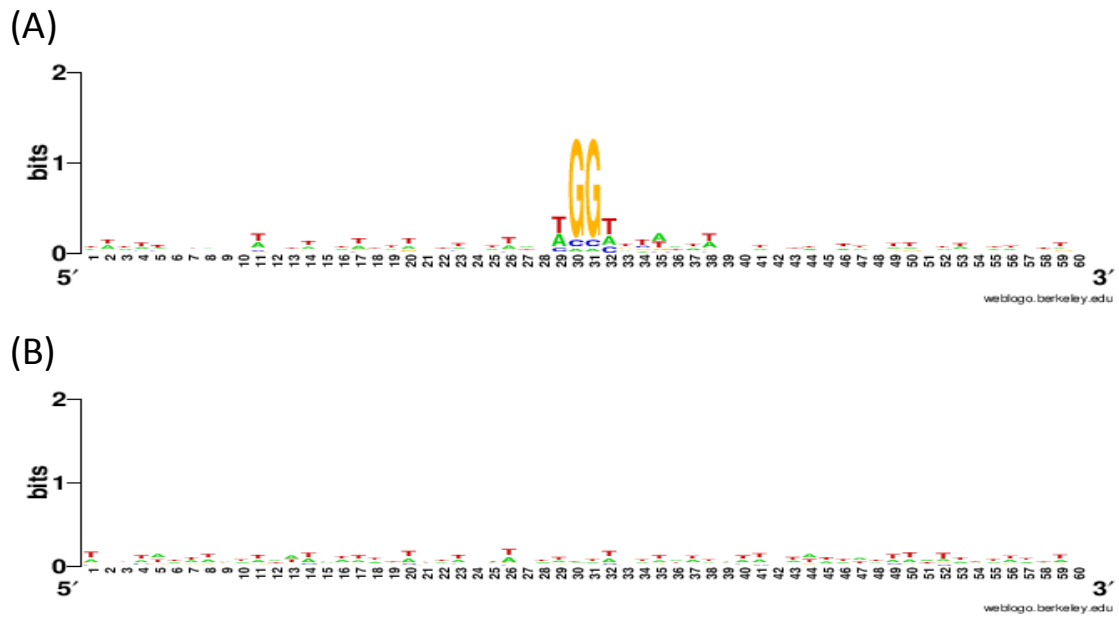


Figure S5. Sequence LOGO of (A) 311 60-nucleotide-long sequences near editing sites and (B) 328 random 60-nt-long sequences.