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Supplemental Information

2i Maintains a Naive Ground State in ESCs through Two Distinct Epigenetic Mechanisms

Ye-Ji Sim, Min-Seong Kim, Abeer Nayfeh, Ye-Jin Yun, Su-Jin Kim, Kyung-Tae Park, Chang-Hoon Kim, and Kye-Seong Kim

Supplemental information

Supplemental Materials and Methods

Cell culture and inhibitors

The mouse ESC lines J1 (ATCC SCRC-1010), CCE, and *Jmjd2c* KO were cultured on mitomycin C-treated CF1 MEF feeder cells or 0.1% gelatin-coated plates at 37°C with 5% CO₂ in Dulbecco's modified Eagle medium (DMEM; Hyclone) supplemented with 15% fetal bovine serum (FBS; Hyclone), 0.1 mM β-mercaptoethanol (Gibco), 100× penicillin/streptomycin (Gibco), 2 mM GlutaMAX (Gibco), 0.1 mM NEAA, and 1000 U/ml LIF (Millipore). We passaged the ESCs at a ratio of 1:20 every 3 days. We cultured 293T cells in DMEM supplemented with 10% FBS, 100× penicillin/streptomycin (Gibco), 2 mM GlutaMAX (Gibco), and 0.1 mM NEAA at 37°C with 5% CO₂. For the 2i culture condition, we cultured ESCs on 0.1% coated gelatin plates using the same ESC culture medium supplemented with Mek1 and Gsk3 inhibitors without LIF. We used the following inhibitors in our study: 1 μM PD0325901 (Stemgent, 04-0006-02), 3 μM CHIR99021 (Stemgent, 04-0004-02), 5 μM FR180204 (Millipore, 328010), 10 μM SU5402 (Abmole, M2194), 5 μM MG132 (Merck, 474790), 5 μM 5-Carboxyl-8-HQ (Merck, 420201), and 1–10 μM UNC0638 (Sigma-Aldrich, U4885).

Dot blot

We isolated genomic DNA (gDNA) using the Qiagen DNeasy Blood & Tissue Kit according to the manufacturer's protocol. We denatured the isolated gDNA in 0.8 M NaOH/20 mM EDTA for 10 min at 95°C. We then neutralized the samples with 2 M ammonium acetate (pH7.0) and serially diluted them twofold. We spotted the samples on a nitrocellulose membrane using a Bio-Dot apparatus (Bio-Rad). We washed the blotted membrane in 2×SSC buffer and exposed it to UV light for 2 min. After blocking the membrane with 5% skim milk for 1 hour, we incubated it overnight at 4°C with primary antibodies diluted in blocking solution. The primary antibodies were mouse anti-5-methylcytosine antibody (Abcam 1:500) and rabbit anti-5-hydroxymethylcytosine polyclonal antibody (Active Motif, 1:2000).

Quantitative reverse transcription-polymerase chain reaction (qRT-PCR)

We extracted total RNA with TRIzol reagent (Invitrogen) according to the manufacturer's protocol. We dissolved the RNA pellets in nuclease-free water and determined the RNA concentration of each sample with a NanoDrop. We reverse-transcribed 1 μg RNA into cDNA using SuperScript III Reverse Transcriptase (Invitrogen). We amplified the cDNA using IQ

SYBR Green Supermix (Bio-Rad) with appropriate PCR primer sets (Table S2). We performed real-time PCR assays on a CFX96 Touch real-time PCR detection system.

Table S2. qPCR primers used in the study

	Forward	Reverse
<i>L32</i>	CGCAAGTTCCTGGTCCACA	TGCTGCTCTTTCTACAATGGCT
<i>Jmjd2c</i>	ATGGATCGCAGATTGCAATGA	TTCCTCTCCCCTTGGATTACAT
<i>Oct4</i>	TAGCATTGAGAACCGTGTGAG	ACTTGATCTTTTGCCCTTCTGG
<i>Sox2</i>	TTTGCAAGCAACTTTTGTACAGTA	TCCTTCCTTGTTTGTAAACGGTC
<i>Dnmt3a</i>	CCATTACCACCAGGTCAAACCTCTA	AAGCGGCTCATGTTGGAGAC
<i>Dnmt3b</i>	ACAACCGTCCATTCTTCTGGAT	TTCATTCCGGGTAGGTTACC
<i>Dnmt3l</i>	AGGATGTCCGTGGCAGAGACTA	AGCTTGCTCCTGCTTCTGACTT
<i>Tet1</i>	CTTTCTCTGGTGTACCTGTTG	TATAGTGGCAGGACGTGGAGTT
<i>Tet2</i>	ATTCTCAGGAGTCACTGCATGTT	TACATAGGCAGCACGTGGAAC
<i>Prdm14</i>	TTCTTCACGTCCATGAGAGGC	TCTGATGTGTGTTCCGGAGTATGCT
<i>G9a</i>	AAGATCTACGGTTCACGCATT	TTGCACTTCTCAGAGCCACACT

Generation of mouse *Jmjd2c* KO ESCs

We transfected cells with pSpCas9(BB)-2A-GFP (PX458) or pSpCas9(BB)-2A-Puro (PX459) (Addgene #48138, 48139) to clone a *Jmjd2c*-guide sequence. We performed an insert/deletion assay using T7 endonuclease I (NEB). To generate *Jmjd2c* KO ESCs, we transfected CCE cells with pSpCas9 (BB) containing *Jmjd2c*-guide RNA and homologous recombination (HR) donor with a puromycin selection cassette. We incubated the cells with 3 μ g/ml puromycin to isolate puromycin-resistant colonies. We confirmed HR events by gDNA-mediated PCR to detect HR events in selected colonies. Finally, we confirmed the *Jmjd2c* KO cells by PCR, sequencing, and Western blot.

Western blot

To perform Western blot analysis, we washed the cells with PBS and lysed them with RIPA buffer (50 mM Tris-HCl pH 7.4, 150 mM of NaCl, 1% sodium deoxycholate, 1% Triton X-100, and 0.1% SDS). We used the cell lysates for Western blots and other experiments. We subjected 20 μ g lysate protein to SDS-polyacrylamide gel electrophoresis, transferred it to a PVDF membrane, and blotted it with appropriate antibodies. We detected proteins using HRP-conjugated secondary antibodies and ECL reagents. We visualized the bands using the ChemiDoc MP System (Bio-Rad).

GST pull down, *in vitro* kinase assay, ubiquitination assay, and *in vitro* methylation assay

For the pull-down analysis, we generated a mammalian GST expression vector system. We transfected GST-transgenes into 293T cells using a PEI transfection method. Two days after transfection, we washed cells with PBS and lysed them with RIPA buffer (50 mM Tris-HCl pH 7.4, 150 mM NaCl, 1% sodium deoxycholate, 1% Triton X-100, and 0.1% SDS). After clarifying cell lysates in a microcentrifuge, we added Glutathione Sepharose 4B and incubated them at 4°C overnight. We then washed the beads with RIPA buffer three times, added 1×SDS sample buffer, and performed Western blots.

We conducted an *in vitro* Mek1 kinase assay as follows. We transfected GST-JMJD2C and GST-MEK1 into 293T cells, isolated the proteins individually using glutathione beads, mixed them together, and incubated them in kinase buffer (25 mM Tris-HCl pH 7.5, 150 mM NaCl, 10 mM MgCl₂, 20 mM β-GP, 100 μM Na₃O₄V, 400 μM ATP, and 2 mM DTT) for 6 h at 37°C. We stopped the reaction by adding 2×SDS sample buffer and then performed Western blots. For ubiquitination assays, we transfected GST-JMJD2C and 5Myc-MEK1 with HA-Ub separately into 293T cells. We detected JMJD2C ubiquitination using anti-HA antibody after the pull-down assay. After washing the beads three times with RIPA buffer, we added 1×SDS sample buffer and performed Western blots.

We transfected GST-PRDM14, GST-DNMT1, GST-DNMT3A, and/or DNMT3B into 293T cells, individually isolated the proteins using glutathione beads, mixed them together, and incubated them in methylation buffer (50 mM Tris-HCl pH 8.8, 5 mM MgCl₂, and 4 mM DTT) with S-adenosyl-L-[methyl-³H] methionine (85 Ci/mmol from a 0.5 mCi/ml stock solution, Perkin-Elmer) for 6 hours at RT. We stopped the reaction by adding 2×SDS sample buffer and performed Western blots. We used the following antibodies: anti-JMJD2C (Abcam, ab85454 or Bethyl, A300-885A), anti-GAPDH (Cell signaling, 2118), anti-H3K4me3 (Millipore, 07-473), anti-H3K4me2 (Millipore, 07-030), anti-H3K9me3 (Abcam, ab8898), anti-H3K9me2 (Abcam, ab1220), anti-H3K27me3 (Millipore, 07-449), anti-H3K27me2 (Millipore, 07-451), anti-H3K36me3 (Abcam, ab9050), anti-H3K36me2 (Millipore, 07-369), anti-Histone3 (Abcam, ab1791), anti-OCT4 (Santa, sc-5279), anti-SOX2 (Abcam, ab97959), anti-Phospho-ERK1/2 (Cell Signaling, 9106), anti-ERK1/2 (Cell Signaling 9102), anti-4G10 (Millipore, 05-321), anti-GST (Abfrontier, LF-PA0189), anti-HA (Santa, sc-7392), anti-Myc (Santa, sc-40), anti-GFP (Santa, sc-9996), anti-DNMT3A (Novus, IMG-268A), anti-DNMT3B (Abcam, ab2851), anti-FLAG (Sigma, F3165), anti-TET1 (Abcam, ab156993), and anti-PRDM14 (Abcam, ab187881).

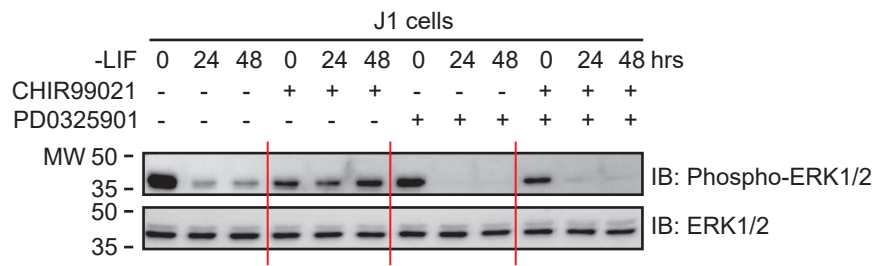


Figure S1

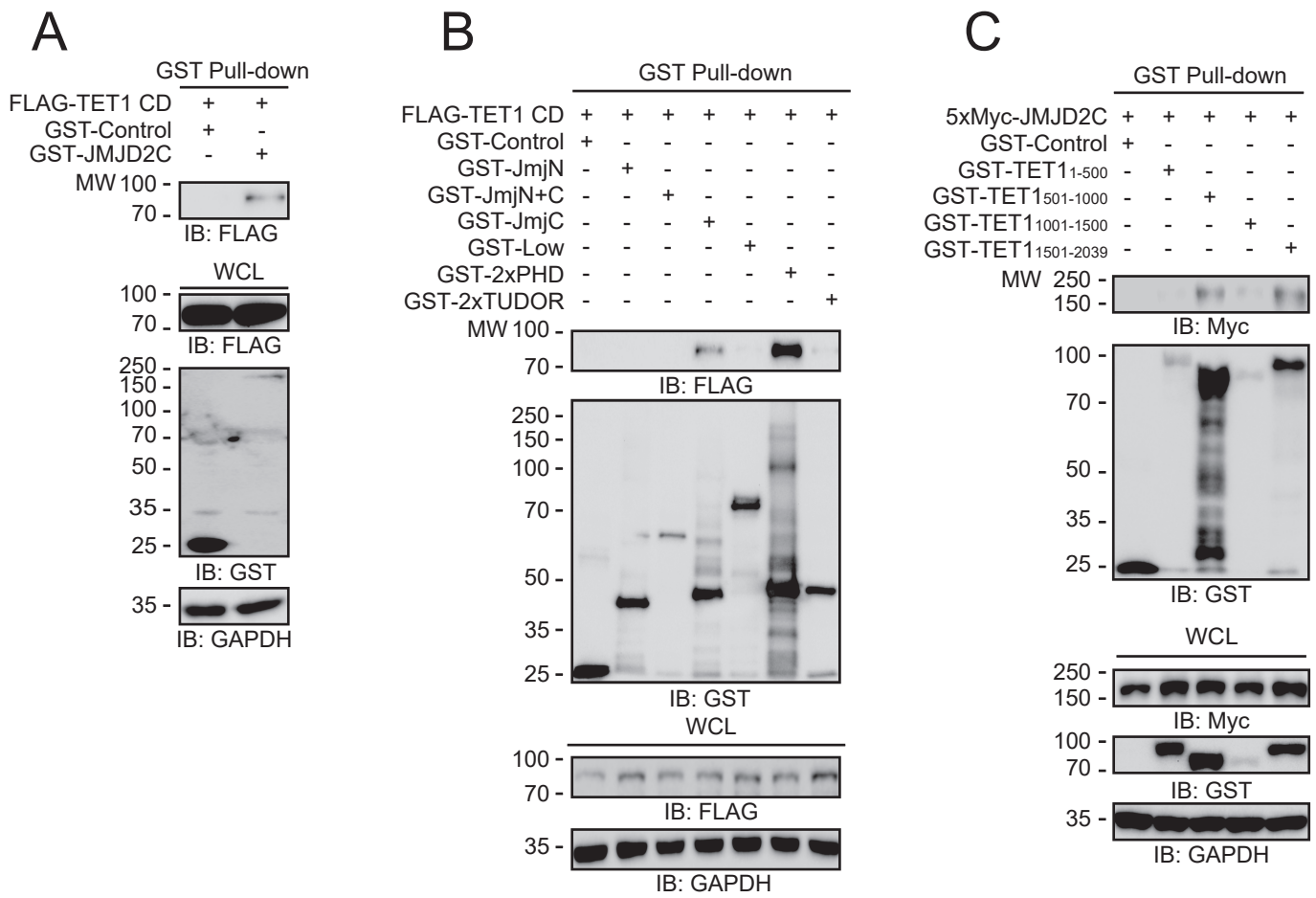


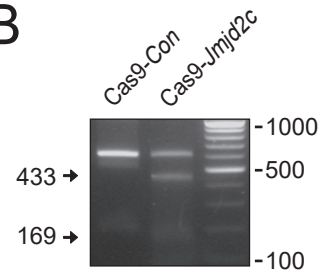
Figure S2

A

>mouse *Jmjd2c* genomic DNA sequence

GAGGGGGTAAATACCCCTTATCTCTATTTTGGAAATGTGGAAAAC**AACATTTGCGTGGCACA**
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Cas9-*Jmjd2c* target sequence : **AACATTTGCGTGGCACACGG**

B**C**

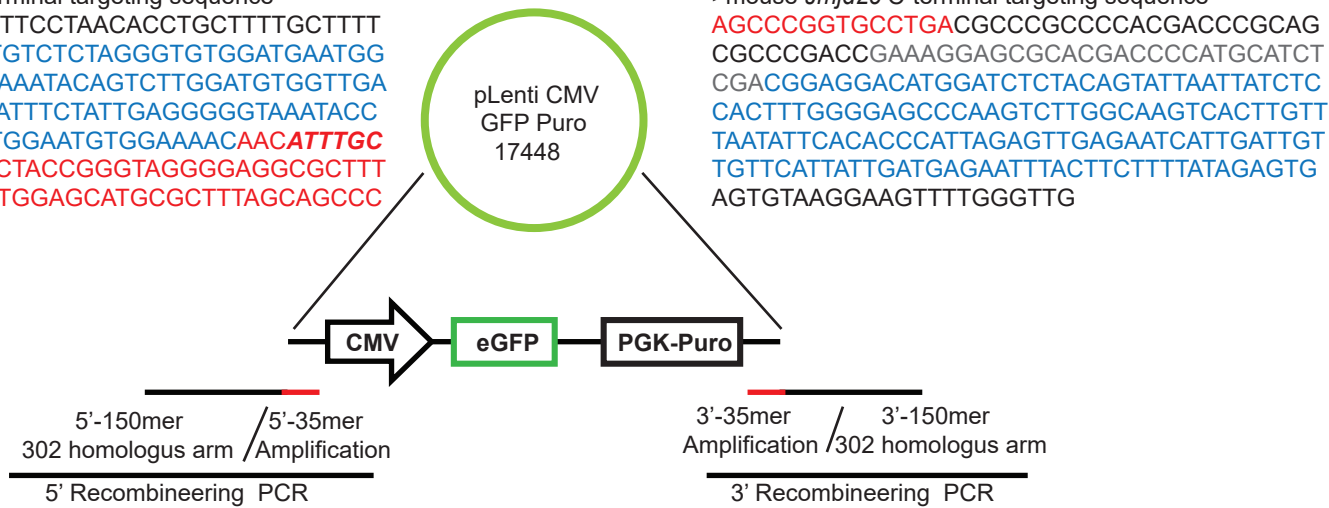
Addgene 17448 / pLenti CMV GFP Puro

>mouse *Jmjd2c* N-terminal targeting sequence

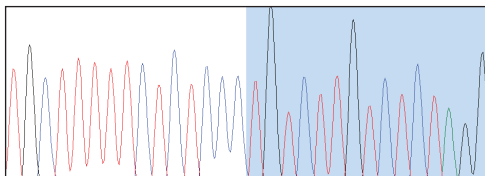
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AATATAGCTCGCTAAATACAGTCTTGGATGTGGTTGA
AGAAGAGTGTGGTATTTCTATTGAGGGGGTAAATACC
CCTTATCTCTATTTTGGAAATGTGGAAAACAACATTTGC****
GTGGCACAGAATTCTACCGGGTAGGGGAGGCGCTTT
TCCCAAGGCAGTCTGGAGCATGCGCTTTAGCAGCCC
CGCTGGGCAC

>mouse *Jmjd2c* C-terminal targeting sequence

AGCCCGGTGCCTGACGCCCCGCCCCACGACCCGCAG
 CGCCCCGACCGAAAAGGAGCGCACGACCCCATGCATCT
 CGACGGAGGACATGGATCTCTACAGTATTAATTATCTC
CACTTTGGGGAGCCCAAGTCTTGGCAAGTCACTTGT
 TAATATTCACACCATTAGAGTTGAGAATCATTGATTGT
 TGTTCAATTTGATGAGAATTTACTCTTTTATAGAGTG
 AGTGTAAAGGAAGTTTTGGGTTG



5'TGCTTTTTTCTCTCCCT**GTCTTTGTCTCTAGG**3'
 5'TGCTTTTTTCTCTCCCT**GTCTTTGTCTCTAGG**3'



5'CCCATGCATCTCGAC**GGAGGACATGGATC**3'
 5'**GATCCATGTCCTCCG**TCGAGATGCATGGGG3'

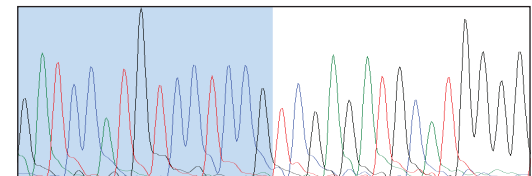
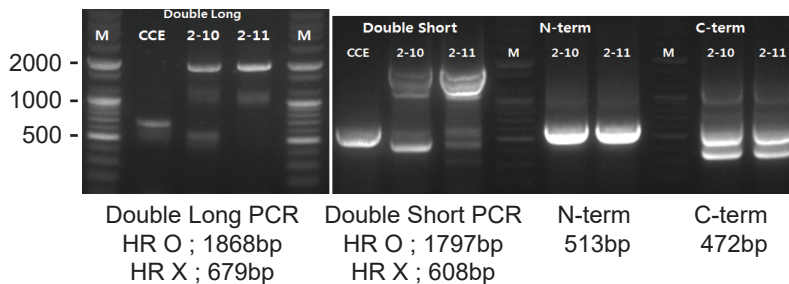
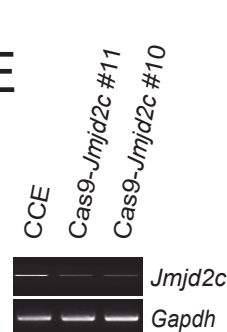
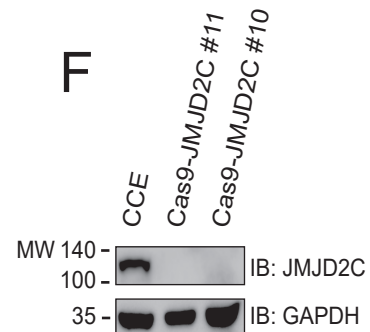
**D****E****F**

Figure S3

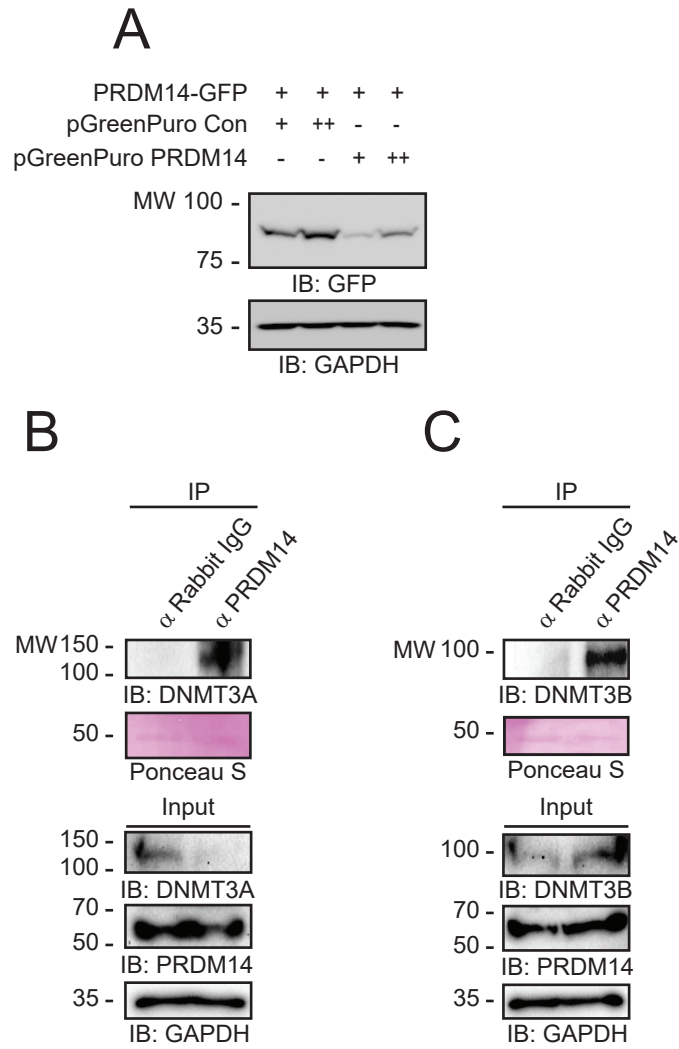


Figure S4

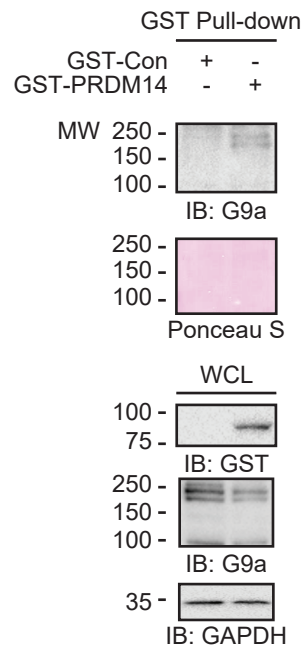
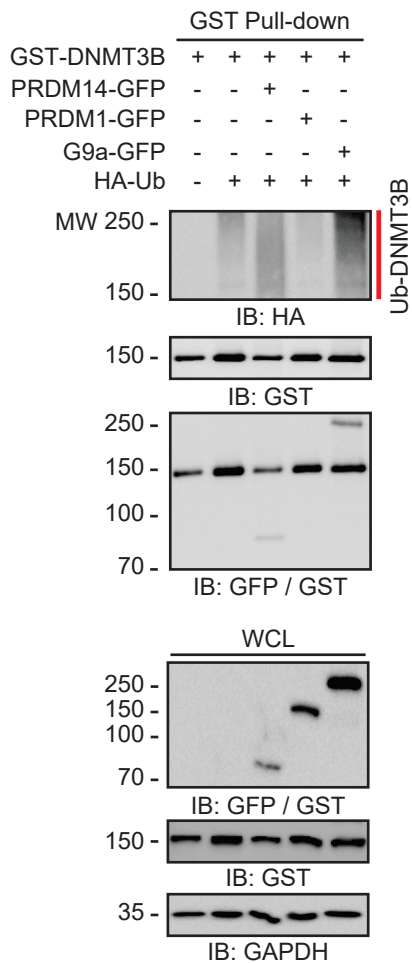
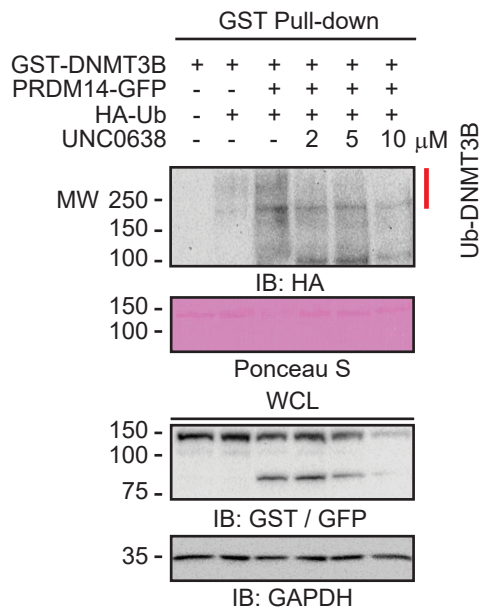


Figure S5

A



B



C

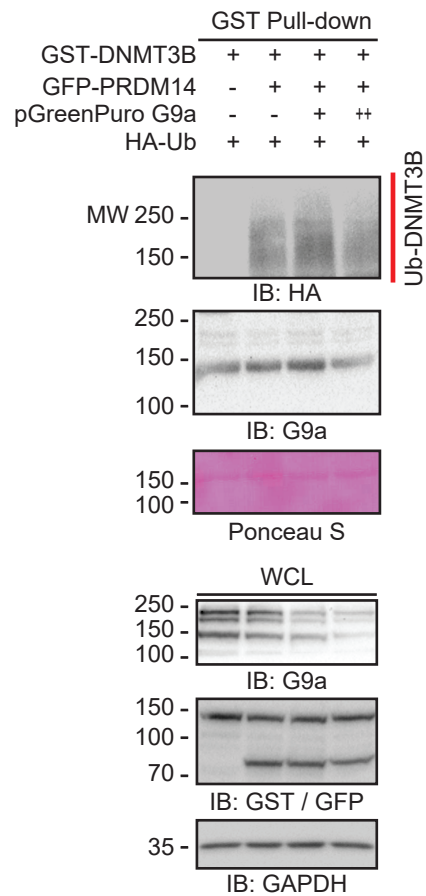


Figure S6

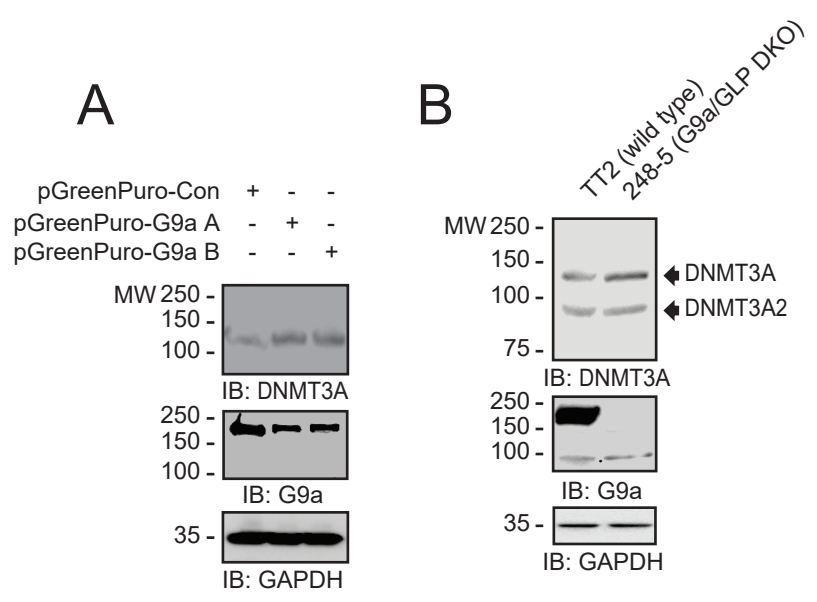


Figure S7

Supplemental figure legends

Figure S1 (related to Figure 1)

PD0325901, but not CHIR99021, specifically decreases phospho-ERK1/2 levels.

Figure S2 (related to Figure 3)

TET1 associates with JMJD2C.

(A) A pull-down assay shows that TET1 CD, a TET1 catalytic mutant, associates with JMJD2C. (B) A pull-down assay shows that TET1 CD binds to the JmjC and 2×PHD domains in JMJD2C. (C) A pull-down assay shows that JMJD2C associates with the CXXC and CD domains of TET1.

Figure S3 (related to Figure 3)

***Jmjd2c* KO strategy using CRISPR/Cas9 technology.**

(A) Mouse *Jmjd2c* genomic DNA sequences and a *Jmjd2c*-specific 20-bp guide RNA sequence. (B) T7 Endonuclease I assay shows that a *Jmjd2c*-specific guide RNA effectively targets *Jmjd2c* genomic DNA. (C) To disrupt genomic regions of the *Jmjd2c* gene through homologous recombination with an antibiotic puromycin cassette, we designed the primers containing the gene-specific homologous sequences (150 bp) with amplifying gene sequences (30 bp) for the PGK-Puro cassette using pLenti GFP Puro (Addgene Plasmid #17448) as a template. (D) Puromycin-resistant ESC colonies are determined by increasing PCR size due to the insertion of HR donor into *Jmjd2c* genomic DNA. To identify HR donor insertion by homologous recombination, 5' and 3' gene-specific primers were chosen from the adjacent upstream and downstream regions of the HR destination site in *Jmjd2c*. (E) *Jmjd2c*-KO clones have reduced *Jmjd2c* mRNA levels compared with control CCE cells, determined using RT-PCR assay. (F) *Jmjd2c*-KO clones do not express JMJD2C proteins as revealed by a Western blot.

Figure S4 (related to Figure 4)

PRDM14 associates with DNMT3A and DNMT3B.

(A) pGreenPuro *Prdm14*, a *Prdm14* shRNA, decreases PRDM14 expression

(B) Endogenous PRDM14 binds to DNMT3A in an immunoprecipitation assay. (C) Endogenous DNMT3B associates with PRMD14 in an immunoprecipitation assay.

Figure S5 (related to Figure 5)

GST-PRDM14 associates with endogenous G9a in a pull-down assay.

Figure S6 (related to Figure 5)

PRDM14-mediated G9a promotes DNMT3B ubiquitination.

(A) DNMT3B ubiquitination is increased in proportion to the expression of G9a > PRDM14 > Control > PRDM1 in a GST pull-down assay. (B) A GST pull-down assay shows that DNMT3B ubiquitination is increased by PRDM14, whereas UNC0638 inhibits a PRDM14-mediated DNMT3B degradation in a dose-dependent manner. (C) Silencing of endogenous G9a using pGreenPuro G9a reduces DNMT3B ubiquitination.

Figure S7 (related to Figure 7)

G9a/GLP deficiency causes an increase of DNMT3A protein levels.

(A) Western blots show that knockdown of G9a expression by pGreenPuro G9a-A and -B increases endogenous DNMT3A expression. (B) Western blots reveal that deficiency of G9a and GLP increases DNMT3A protein levels in the 248-5 ESC cell line compared with wild-type TT2 cells.

Supplemental table legend

Supplemental Table S1 (related to Figure 1E)

Quantitative gene expression data induced by combinatorial 2i treatments in ESCs

Bioinformatics data come from the GSE43597 in GEO DataSet.