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Supplementary Figure 2. Inactivation of PTEN leads to defective sprouting angiogenesis. (a,b) Representative images of WT and PTEN^{-/-} embryoid bodies stained with CD31 (green). Scale bars, 200 μ m (a) and 150 μ m (b). (c-e) Quantitative analysis of embryoid bodies shown in a and b. (c) Sprouts width (n \geq 6). (d) Sprout length (n \geq 6). (e) Number of sprouts (n \geq 6). Errors bars are s.e.m. *P*-values **p<0.01, and ***p<0.001 were considered statistically significant. Statistical analysis was performed by nonparametric Mann Whitney's test.





PdgfbiCreER^{T2}: PTEN^{flox/flox}



Desmin: IB4



Supplementary Figure 3. PTEN does not regulate Notch target genes. (**a**,**b**) qPCR of Dll4, Hey1, Hes1 and Nrarp mRNA expression in the retinas of control and PTEN^{$i\Delta EC/i\Delta EC$} littermates at P7 ($n\geq 5$) (**a**) and WT and PTEN^{TG} ($n\geq 5$) (**b**). (**c**) Control and PTEN^{$i\Delta EC/i\Delta EC$} mECs were stimulated with Dll4 for 6 h, followed by qPCR analysis of Hest and Hey1 (n=4). (d,e) qPCR of Efnb2, Nr2f2, and Ephb4 mRNA expression in the retinas of control and PTEN^{iAEC/iAEC} littermates at P7 (n≥6) (d) and WT and PTEN^{TG} (n≥5) (e). mRNA levels were normalized by Hprt in **a**, **b**, **c**, **d** and **e**. (f) IB4 (Green) and desmin (Red) staining of control and PTEN^{$iAEC/iAEC} littermate retinas at P7 (n=3). Scale bars, 20 <math>\mu$ m. Errors</sup> bars are s.e.m. P-value ***p<0.001 were considered statistically significant. Statistical analysis was performed by nonparametric Mann Whitney's test.

Edu : Erg : IB4





Supplementary Figure 4. Notch negatively regulates EC proliferation. (a) WT mice were injected with Edu 2 h before isolating the retinas, followed by triple staining with IB4 (Blue), Erg (Red) and Edu (Green). Number of proliferative ECs was quantified at the sprouting front (20 μ m from the sprouting front line) and at the sub-front area (comprise the area enclosed 100 μ m below the sprouting front). Scale bars, 20 μ m. (b) Quantification of Edu positive cells at the sprouting front and at the sub-front area (n=8). (c) P6 WT mice were injected with Edu 2 h before isolating the retinas, followed by triple staining with IB4 (Red), Hes1 (Green) and Edu (Blue). Scale bars, 20 μ m. Islets show higher magnification of selected regions shown below. Scale bars, 10 μ m. (d) Quantification of Hes1/ Edu positive cells relative to Hes1 positive/ Edu negative cells in the vascular plexus (n=6). (e) P6 WT mice were injected with Edu (Blue). Islets show higher magnification of selected regions shown below. Yellow arrows and white asterisk indicate high PTEN expression/ Edu positive ECs and high PTEN expression/ Edu negative ECs respectively. White arrowhead indicates a pericyte. Scale bars, 20 μ m. (f) Quantification of PTEN expression levels vs. Edu positivity at the vascular front (n=4). Errors bars are s.e.m. *P*-values *p<0.05, and **p<0.01 were considered statistically significant. Statistical analysis was performed by nonparametric Mann Whitney's test.

0

С

е



b



Supplementary Figure 5. PTEN operates downstream of Notch *in vivo*. (a) Whole mount visualization of blood vessels by IB4 staining of control, $Jag1^{i\Delta EC/i\Delta EC}$, PTEN^{i\Delta EC/i\Delta EC} and PTEN^{i\Delta EC/i\Delta EC}, $Jag1^{i\Delta EC/i\Delta EC}$ littermates at P7. (b) Whole mount visualization of blood vessels by IB4 staining of WT and PTEN^{TG} P7 retinas treated with vehicle or DAPT (100 mg kg⁻¹). Scale bars, 20 µm.

b





PdgfbiCreER^{T2}; PTEN^{flox/flox}



Supplementary Figure 6. PTEN regulates endothelial cell proliferation *in vitro.* (a) Effect of pan PI3K inhibitor GDC-0941 (1 μ M) and VX680 (0.5 μ M) on control and PTEN^{iAEC/iAEC} mEC proliferation was assessed by MTS assay after treatment for 48 h. One representative of three experiments is shown. (b) Quantification of proliferation of WT and PTEN^{TG} mECs plated for 48 h, pulsed with BrdU for 2 h, and subjected to immunostaining analysis. Data shown are means of four independent experiments. (c) Immunoflorescence analysis of YFP tagged wild-type PTEN, PTEN^{C124S}, and PTEN^{K13,289K}, stained with BrdU (red) and DAPI. Images are representative staining of quantification shown in Fig. 5g. Scale bars, 25 μ m. White asterisk indicate strong YFP positive cells. Errors bars are s.e.m. *P*-values *p<0.05, were considered statistically significant. Statistical analysis was performed by nonparametric Mann Whitney's test.









Supplementary Figure 7. Lipid phosphatase dependent and independent roles of PTEN regulate angiogenesis. (a) pS6 staining of images shown in Fig. 6 a. Scale bars, 100 μ m. (b) Exponential growing *PdgfbiCreER*⁷²; Fzr1^{flox/flox} and Fzr1^{flox/flox} mECs were incubated with vehicle or 4-OHT for 24 h, 48 h, 72 h, and 96h followed by immunoblotting using the indicated antibodies (n=2). Molecular weight marker (kDa) is indicated. (c) Cre expression detected by X-gal staining (blue) in P7 retinas of *PdgfbiCreER*⁷²; Fzr1^{flox/flox}; Rosa26-R pups. 4-OHT was administered at P5 and P6 (n=3).

SFig. 1k



β**-actin**



SFig. 1I

PTEN

12



VE-cad



Fig. 3b-mECs



Supplementary Figure 8. Uncropped western blots related to Fig. 3, 5 and Supplementary 1, 7

Fig. 3b-HUVECs



Fig. 3j



Fig. 3k





Fig. 3l

PTEN





Hes1

β -actin



Fig. 5b





p-AKT





Geminin





β-actin

p-AKT



Fig. 5c





Geminin





Aurora



Fig. 5d

VE-cad

Geminin



Plk1

W SR

p-AKT



Aurora



β-actin



F:... **F**.

Fig. 5e



Geminin





β-actin







SFig. 7b



β-actin



Name	Forward Primer	Reverse Primer
1 (-2320 / -2181)	5'-GGTTGTTAACTCCTGCACAGC-3'	5'-AAAGCCACAGGCCATTACC-3'
2 (-2276 / -2136)	5'-GCAAAGTTTTATGGAGCCTCA-3'	5'-GCAGGCAACCTCTGAAGACT-3'
3 (-2000 / -1871)	5'-TGCTTCAACCTAGGTCTCGTT-3'	5'-CGACCCGCTTAAAGAGAATG-3'
4 (-1595 / -1418)	5'-GCAGGAGATACCCTCAAGCA-3'	5'-GCAAGCCAAAGGACTGAGAC-3'
5 (-1395 / -1286)	5'-CACCAGTTTGGGGGACTCTCT-3'	5'-GGGGAACTGGTTACACAAGC-3'
6 (-1223 / -1092)	5'-CAGGAAGGGTTGGGGTTC-3'	5'-TCACGTGTGTCCCTAGTTGG-3'
7 (-803 / -658)	5'-GGGAAAGATGCTCGACTCTC-3'	5'-ACGTGAACACATAGCCGTTG-3'

Supplementary Table 1. Human ChiP primers