#### Supplementary information

# Kaposi's sarcoma-associated herpesvirus ORF34 is essential for late gene expression and virus production

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#### Supplementary Table S1: Primers for construction of BACmid/ expression plasmid and real-time PCR/ RT-qPCR

Primer name	Primer sequences $(5' \rightarrow 3')$
[BAC mutagenesis]	*a
BACmut-ORF34-3stop-F	$a act cccct gaac agette ct ett ggttte cca acg tgat g \underline{TAGTTAGATAGT} cccg agg tcc age ca at ctg TAGGGATAACAGGGTAATCGATTT according to the transformation of transformation$
BACmut-ORF34-3stop-R	$gcaaggccggccggccactgcagattggctggacctcggg \underline{ACTATCTAACTA} catcacgttggaaaccaagaGCCAGTGTTACAACCAATTAACC \\ eq:gcaggccggccggccggccggccggccggccggccggcc$
BACmut-ORF34-Rev-F	a act cccct gaac agette ctctt ggttte caacgt gat gcccg aggte cage caatet g TAGGGATAACAGGGTAATCGATTT
BACmut-ORF34-Rev-R	gcaaggccggccggccactgcagattggctggacctcgggcatcacgttggaaaccaagaGCCAGTGTTACAACCAATTAACC

#### [Cloning expression plasmid]

\*b

cloning_ORF34-F	acat <u>gaatte</u> ATGTTTGCTTTGAGCTCGC
cloning_ORF34-R	ctaagtcgacTTAGAGTTGGTTGAGTCCATTCTCC
cloning_ORF34D1-R	ttaagtcgacTTAAGTTGGACACCTTTGGGTC
cloning_ORF34D2-R	ttaagtcgacTTAAGTGTCGGAAGCATCTATG
cloning_ORF34D3-R	ttaagtcgacTTAAAGATCCCGTTTGGGAGC
cloning_ORF34D4-R	ttaagtcgacTTAAAGACCCTTCCAAGTTCCG
cloning_ORF34D5-R	taagtcgacTTAGTCCAGTGCGATTTCCTCAC
cloning_ORF18-F	catgaattcATGCTCGGAAAATACGTGTGTG
cloning_ORF18-R	taaacgcgtTTAAACCGCGTTGTTGTTAAAC
cloning_ORF23-F	cattetagaATGTTACGAGTTCCGGACGTGAAGGC
cloning_ORF23-R	taageggeegeTTAGACGGTCAATAAAGCG
cloning_ORF24-F	at <u>acgcgt</u> ATGGCAGCGCTCGAGG
cloning_ORF24-R	taa <u>tctaga</u> TTAGACCAGCGGACGGACGC
cloning_ORF30-F	catgaattcATGGGTGAGCCAGTGGATCCTG
cloning_ORF30-R	tgaacgcgtTCATTTCGCACCGGTGTC
cloning_ORF31-F	catgaattcATGTCACAAAACAGAAAGACTCTGC
cloning_ORF31-R	tagacgcgtCTACGTATCTTTCGTTGATAGC
cloning_ORF66-F	catt <u>etaga</u> ATGGCCCTGGATCAGCGCTGGGATC
cloning_ORF66-R	gagcggccgeTCAGGAGGAACACTTCCC

#### [qPCR/ RT-qPCR]

real-time_PCR_KSHV ORF11-F	TTGACAACACGCACCGCAAG
real-time_PCR_KSHV ORF11-R	AAAAATCAGCACGCTCGAGGAG
RT-qPCR_ORF72/vFLIP-F	CTGGACTTTTGGCACCACGAG
RT-qPCR_ORF72/vFLIP-R	AGCGCTGATAATAGAGGCGGG
RT-qPCR_ORF16-F	AGATTTCACAGCACCACCGGTA
RT-qPCR_ORF16-R	CCCCAGTTCATGTTTCCATCGC
RT-qPCR_ORF46-F	CACTGCTGCGATCCAGAGGATA
RT-qPCR_ORF46-R	GAACCTGACATTGCGGATCCAC
RT-qPCR_ORF59-F	AAGGGGAAGAAGTCGGTGGATG
RT-qPCR_ORF59-R	AACCAACCCGGGACTTTACACA
RT-qPCR_K8.1-F	TAAACGGGACCAGACTAGCAGC
RT-qPCR_K8.1-R	GTTTTCTGCGACCGGTGATACG
RT-qPCR_ORF26-F	AAGGGGAAGAAGTCGGTGGATG
RT-qPCR_ORF26-R	AACCAACCCGGGACTTTACACA
RT-qPCR_ORF25-F	TACGGTCTCAGGTATGAGCAGA
RT-qPCR_ORF25-R	GTAGATCCTCCGTGGTTACGAG
RT-qPCR_ORF27-F	GACGCATTTACCCGAACTCTAC
RT-qPCR_ORF27-R	TGCACATGACGTGTTAACCATA
RT-qPCR_ORF42-F	TCTCCAGGTGCTTGGTAAAGAT
RT-qPCR_ORF42-R	TAAGCTTCATAAGGCGATAGGC
RT-qPCR_ORF43-F	TCCTGTAAACGTCCCAGAGATT
RT-qPCR_ORF43-R	TGTCTCACCAACCAGATAAACG
RT-qPCR_ORF53-F	CTAAAACTATCCGCGGAACAAG
RT-qPCR_ORF53-R	GCCTCGATACTAGGTCACTGCT
RT-qPCR_ORF55-F	GAGGCAATACAGAAGTGGGTTC
RT-qPCR_ORF55-R	GCTCCAGTCCCTCTTAAACAAA
RT-qPCR_ORF65-F	GTTGTGAGAATGTCTGACGCCG
RT-qPCR_ORF65-R	GGTGGAATTCAAAGGCGGGATC
RT-qPCR_ORF68-F	TTCTGTCTATACGCCCCACACTG
RT-qPCR_ORF68-R	CCTCCTCTTCTGGAAAGTGCAGA
RT-qPCR_ORF52-F	ACAAAAGCAAGTGGACGATGCC
RT-qPCR_ORF52-R	CTCTTCGTCGCCTGTTATTGGC
RT-qPCR_ORF8-F	TGTATCCGCCAAGTTCGTAGGAG
RT-qPCR_ORF8-R	GCTCTTGTGGATGTTTACGGAGC
RT-qPCR_GAPDH-F	CATCAAGAAGGTGGTGAAGCAG
RT-qPCR_GAPDH-R	TGTCGCTGTTGAAGTCAGAGG
[CHIP qPCR]	
CUUD DOD ODE4(/47 E	

CHIP-qPCR_ORF46/4/-F	AGCCCCCTTCCGTAATATCTG
CHIP-qPCR_ORF46/47-R	TTTTCCGCGGAAGTATGTCG
CHIP-qPCR_K8.1-F	ACTCCCACCATGTTGAAGCTTG
CHIP-qPCR_K8.1-R	GGGATTTCTGTGCGAATCTGTG

\*a: Lowercase indicates homology sequence to KSHV BAC16, underlined uppercase indicates mutagenesis site, and uppercase indicates pEP-KanS sequence \*b: underlined lowercase indicates mutagenesis site, and uppercase indicates KSHV BAC16 ORF sequences

# Supplementary Fig. S1 (original data of Fig. 1b)



# Supplementary Fig. S2



# Supplementary Fig. S3 (original data of Figure 4a)



Blot: Myc

Whole cell extract | Pull-down: S-Agarose



Blot: S

# Supplementary Fig. S4 (original data of Figure 4b)









Whole cell extract



Blot: FLAG

Pull-down: S-Agarose

# Supplementary Fig. S5 (original data of Figure 4c)



Pull-down: S-Agarose

Blot: FLAG









Blot: FLAG

# Supplementary Fig. S6 (original data of Figure 4d)



Blot: FLAG

Pull-down: S-Agarose



Blot: S

# Supplementary Fig. S7 (original data of Figure 4e)



Blot: FLAG

Pull-down: S-Agarose



Blot: S

# Supplementary Fig. S8 (original data of Figure 4f)



Blot: Myc

#### Pull-down: S-Agarose





#### Pull-down: S-Agarose



Blot: Myc

#### Whole cell extract



Blot: Myc

#### Pull-down: S-Agarose



Blot: S

#### **Supplementary Fig. S9**



To establish iVero- $\Delta$ ORF34 cells stably expressing 3xFLAG-ORF34 protein, iVero- $\Delta$ ORF34 cells were transfected with 3xFLAG-tagged ORF34 plasmid, and then selected and maintained with media including 1.5 mg/mL G418. 3xFLAG-ORF34-expressing iVero- $\Delta$ ORF34 cells were treated with (or without) Dox and NaB for 72 hours and subjected to ChIP-qPCR. 3xFLAG-tagged ORF34 protein was Immunoprecipitated by anti-FLAG or control antibody, and precipitates including chromatin and viral DNA were subjected to SYBR green real-time PCR for measuring the amount of promoter DNA of ORF46/47 (E gene) or K8.1 (L gene). The levels of immunoprecipitated viral promoter were normalized to total input DNA.

# Supplementary Fig. S10 (original data of Figure 5b)



Pull-down: S-Agarose

Blot: Myc

Pull-down: S-Agarose



Blot: S



Blot: Myc

# Supplementary Fig. S11 (original data of Figure 5c)



Pull-down: S-Agarose













# Supplementary Fig. S12 (original data of Figure 5d)













Blot: FLAG

# Supplementary Fig. S13 (original data of Figure 5e)

Pull-down: S-Agarose



Blot: Myc

Pull-down: S-Agarose







Blot: Myc

# Supplementary Fig. S14 (original data of Figure 5f)

Blot: FLAG

Pull-down: S-Agarose







Blot: FLAG

Pull-down: S-Agarose