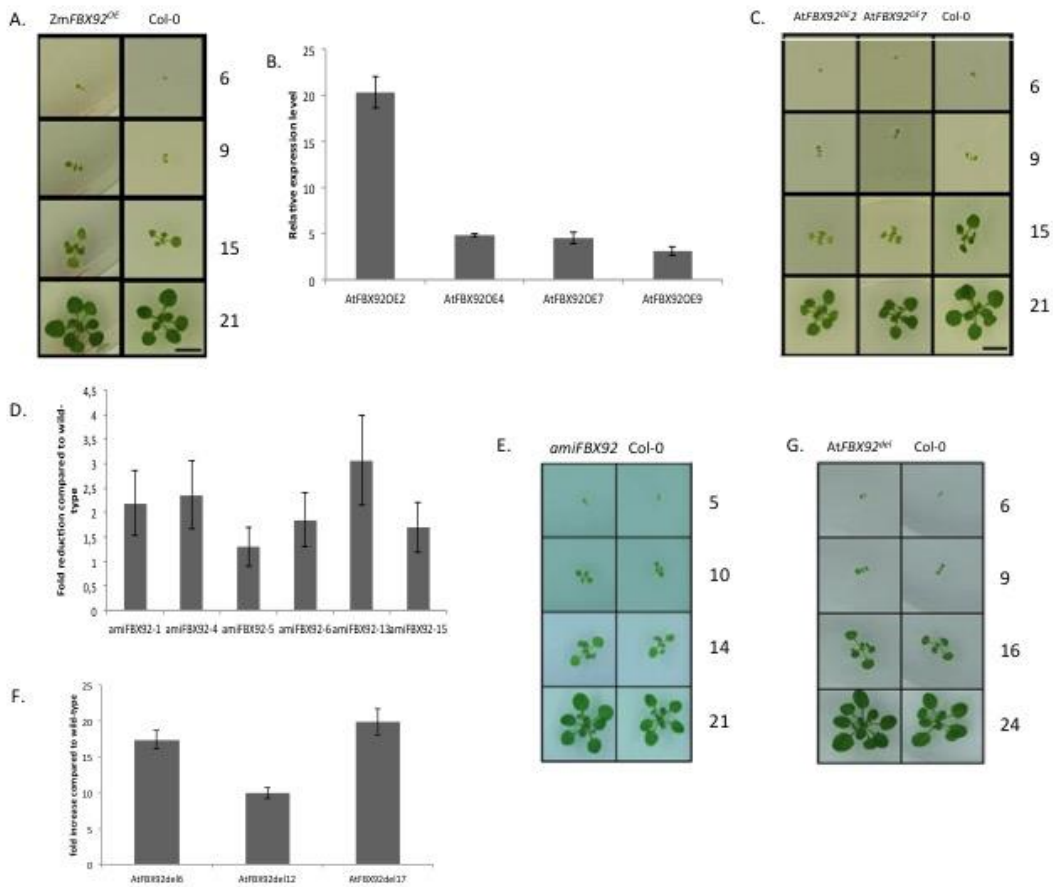
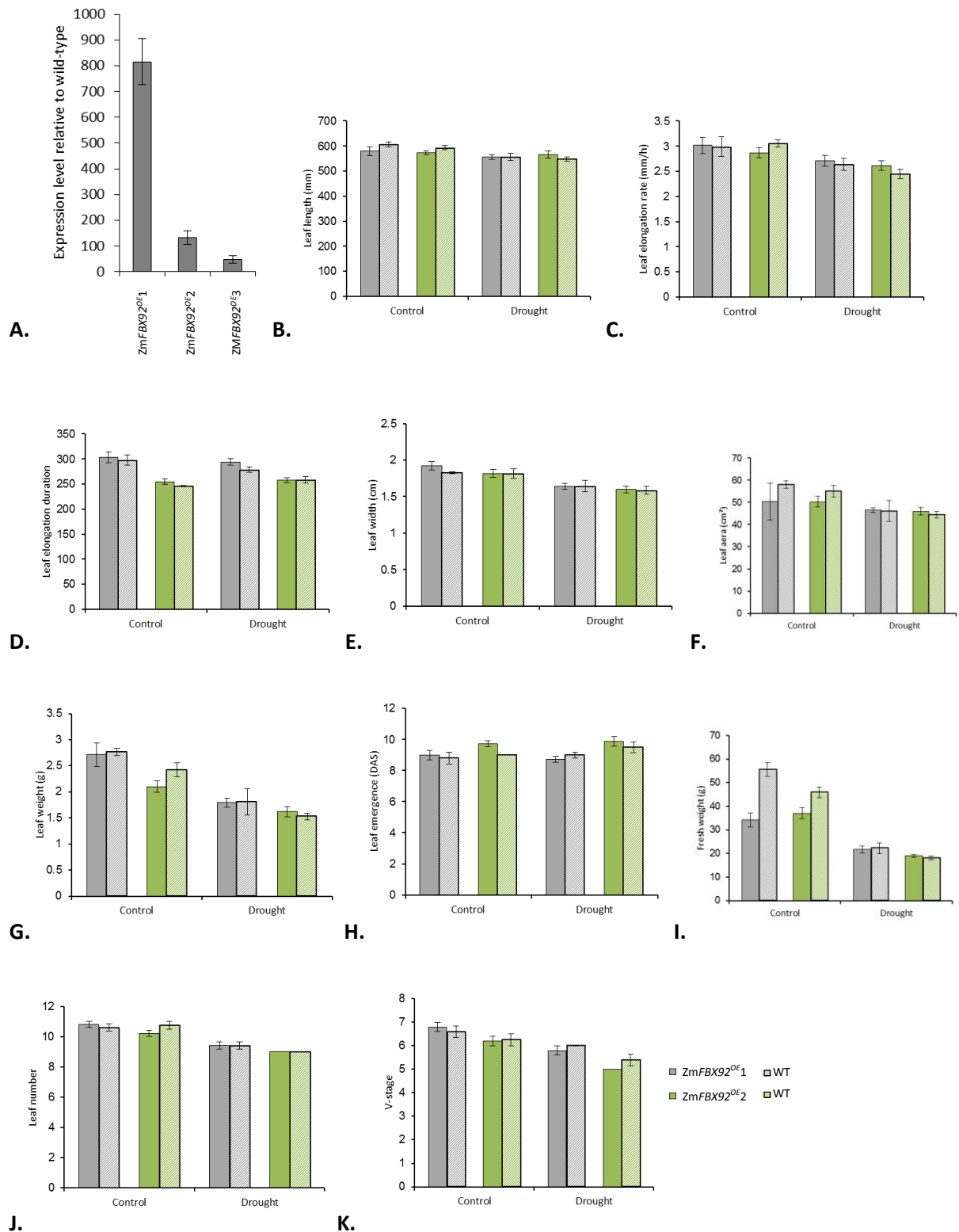


SUPPLEMENTARY DATA



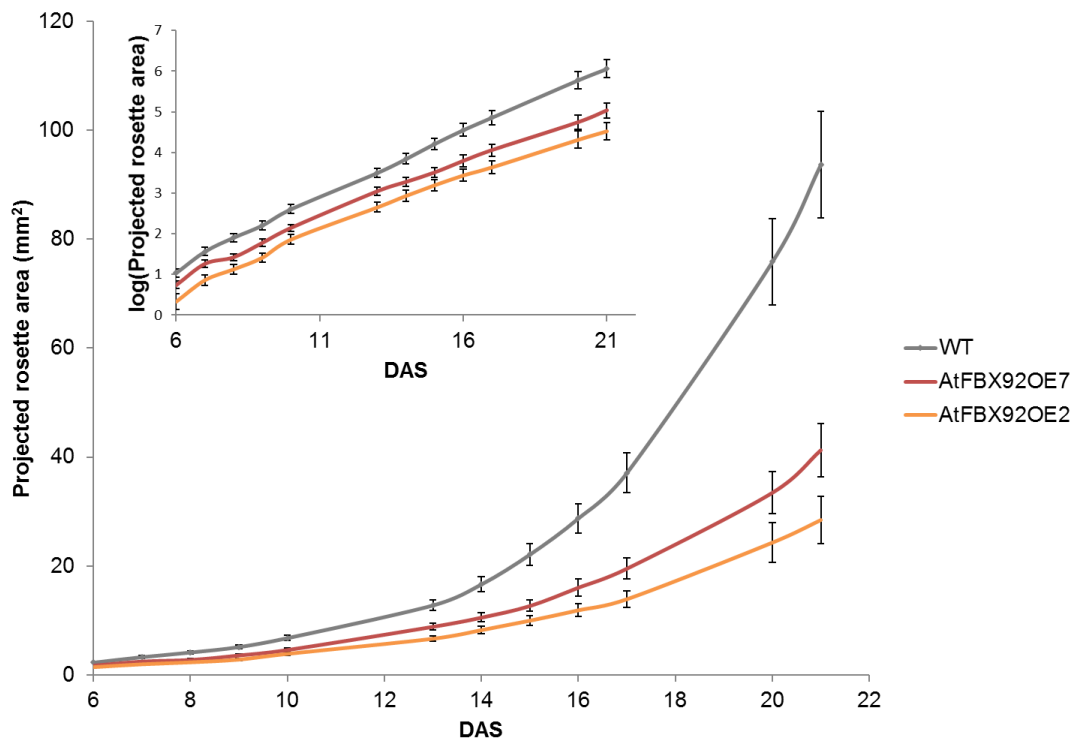
Supplementary Fig. S1. Representative pictures of ZmFBX92^{OE}, AtFBX92^{OE}, amiFBX92, AtFBX92^{del} and Col-0 grown *in vitro* and expression levels in mature leaves. (A) Pictures of ZmFBX92^{OE} and Col-0 plants grown *in vitro* at 6, 9, 15 and 21 DAS. Scale bar 1 cm. (B) qRT-PCR analysis of the AtFBX92 expression in mature leaves of four AtFBX92^{OE} lines relative to wild type. (C) Pictures of AtFBX92^{OE} and Col-0 plants grown *in vitro* at 6, 9, 15 and 21 DAS. Scale bar 1 cm. (D) qRT-PCR analysis of the AtFBX92 expression in mature leaves of six amiFBX92 lines relative to wild type. (E) Pictures of amiFBX92 and Col-0 plants grown *in vitro* at 5, 10, 14 and 21 DAS. Scale bar 1 cm. (F) qRT-PCR analysis of the AtFBX92 expression in mature leaves of three AtFBX92^{del} lines relative to wild type. (G) Pictures of AtFBX92^{del} and Col-0 plants grown *in vitro* at 6, 9, 16 and 24 DAS. Scale bar 1cm.



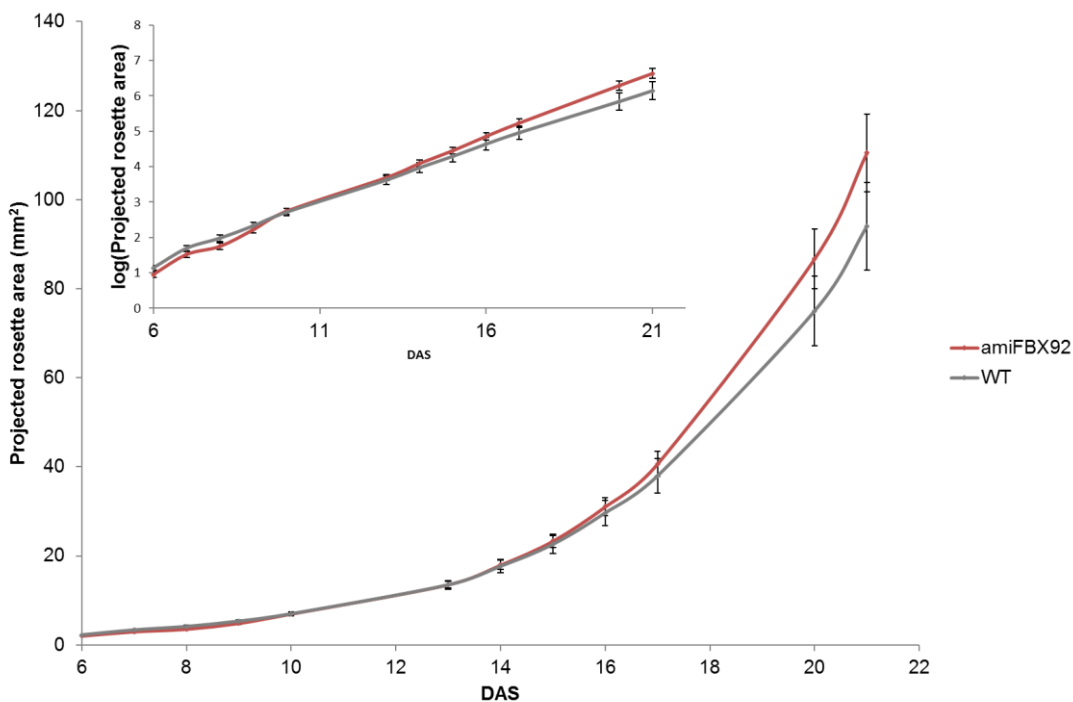
Supplementary Fig. S2. Effect of *ZmFBX92* overexpression on maize leaf size and shoot weight under control and mild drought conditions. (A) Expression level of *ZmFBX92* in mature leaves of *ZmFBX92*^{OE} plants relative to wild type as determined by qRT-PCR. Values were normalized against expression levels of housekeeping genes. Data are average fold change (transgenic versus wild-type plants) ± SE ($n_{\text{transgenic}} = 3$ and $n_{\text{wild-type}} = 9-20$). (B to H) Measurement of final leaf 4-related parameters: length (B), elongation rate (C), elongation duration (D), width (E), area (F), weight (G), and emergence

(H). Data are average \pm SE ($n_{\text{transgenic}} = 20$ and $n_{\text{wild-type}} = 20$. (I to K) Measurement of shoot-related parameters at seedling stage: fresh weight (I), leaf number (J) and V-stage (K). Data are average \pm SE ($n_{\text{transgenic}} = 20$ and $n_{\text{wild-type}} = 20$).

A.



B.



Supplementary Fig. S3. Effect of perturbed *AtFBX92* expression on rosette growth under mild osmotic stress conditions. (A) PRA of *AtFBX92*^{OE} lines and WT over time from 6 until 21 DAS. Plants were grown *in vitro* on medium containing 25 mM mannitol. Inset: PRA in log scale. Values represent the mean \pm SE ($n_{\text{transgenic lines}} = 26-32$, $n_{\text{WT}} = 40$). (B) PRA of *amiFBX92* and WT over time from 6 until 21 DAS. Plants were grown *in vitro* on medium containing 25 mM mannitol. Inset: PRA in log scale. Values represent the mean \pm SE ($n = 29$).

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                                10      20      30
ZmFBX92  -----MAEAAGGGPLLPEELVIWEILTRLPAKPLLRCR
                                ::  ::::  ::  ::  ::::  .  :
AtFBX92  MASEKSFKKRKITDDVDGVGVGGGLESLPED-IIADIFSRLPISSIARLM
                                10      20      30      40

                                40      50      60      70      80
ZmFBX92  AVCLSWRRRLTSDAKFLLAHRRRQPSLPLLTQGDRSTEDRRIDALDYRTG
                                ::  :::  ::  .  ::  ::  ::  :  .  ::
AtFBX92  FVCRSWRSVLTQHGR--LSSSSSPTKPCLLHCDSPINRGLHFLDLSEE
                                50      60      70      80      90

                                90      100     110     120
ZmFBX92  E-RLPVARTTGRTAANADDLNVLASCDGLLVL---IANGGLHICNPATRQ
                                :  .  .  :  :  :  .  .  .  :  :  :  :  :  :  :  :
AtFBX92  EKRIKTKKFTLRFASSMPEFDVVGSCNGLLCLSDSLYNDSLYLYNPFTTN
                                100     110     120     130     140

                                130     140     150     160     170
ZmFBX92  RAPLA----LLHGASCISALYPYRPSGSYRVAGALLRESRGRPRRVPRA
                                :  :  :  .  .  .  ::  .  :  :  .  :
AtFBX92  SLELPECSNKYHDQELVFGFGFHEMTKEYKVLKIVYFRGSSSNNGIYRG
                                150     160     170     180     190

                                180     190     200     210     220
ZmFBX92  HRRV----ARAQVRRRTSRAVGRRRGDGDADDVVLSPARRGRQDLLAA
                                :  .  .  :  .  .  :  :  :  :  :  :  :
AtFBX92  RGRIQYKQSEVQILTLSSKTT-----DQSLSWRSLGK----AP
                                200     210                               220     230

                                230     240     250
ZmFBX92  GKAARRQREGQQHARVRYSDRVLPATAVAPARGGPLP*-----
                                :  :  :  .  .  .  :  :  :
AtFBX92  YKFVKRSSEALVNGLHFVTE-----PRRHVPDRKFVSFDLEDEEFK
                                240     250                               260     270

ZmFBX92  -----
AtFBX92  RIPKPCDGLNRTNHRLVNLKGCLCAVVYGNYGKLDIWVMKTYGVKESWC
                                280     290     300     310     320

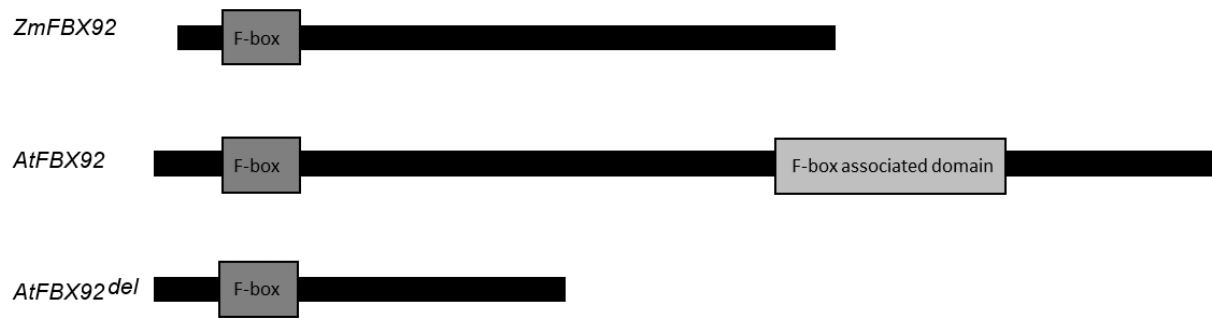
ZmFBX92  -----
AtFBX92  KEYSIGTYLPKGLKQNLDRPMWIWKNAENGKVVRVLCLLENGEILLEYKS
                                330     340     350     360     370

ZmFBX92  -----
AtFBX92  RVLVAYDPKLGKFKDLLFHGLPNWFHTVVHAGTLSWFDTPLDLW*
                                380     390     400     410

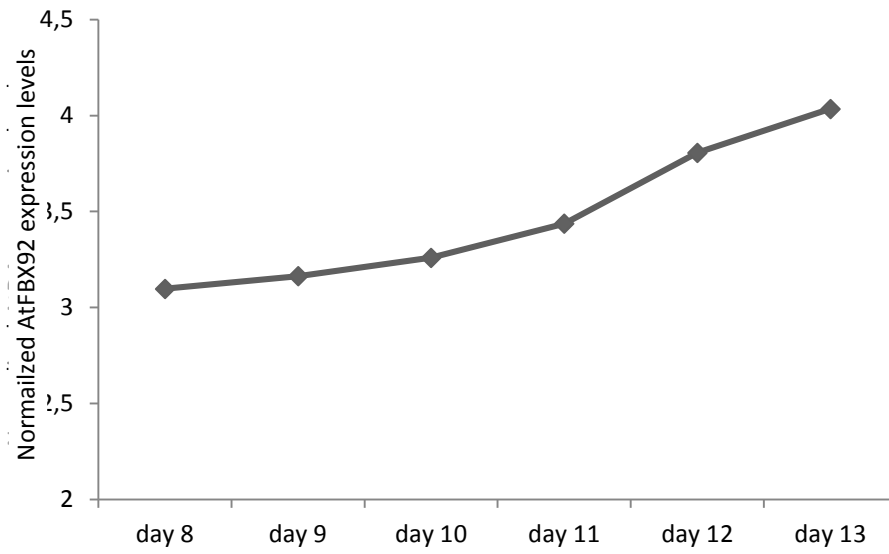
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Supplementary Fig. S4. Amino acid alignment of the *ZmFBX92* and *AtFBX92* proteins using EMBOSS Needle. Protein domains were determined using the Pfam database (Finn et al., 2014). The F-box

domains are indicated in yellow, the F-box associated domain of *AtFBX92* in red. Identical amino acids are indicated by “:”, similar amino acids by “.”.

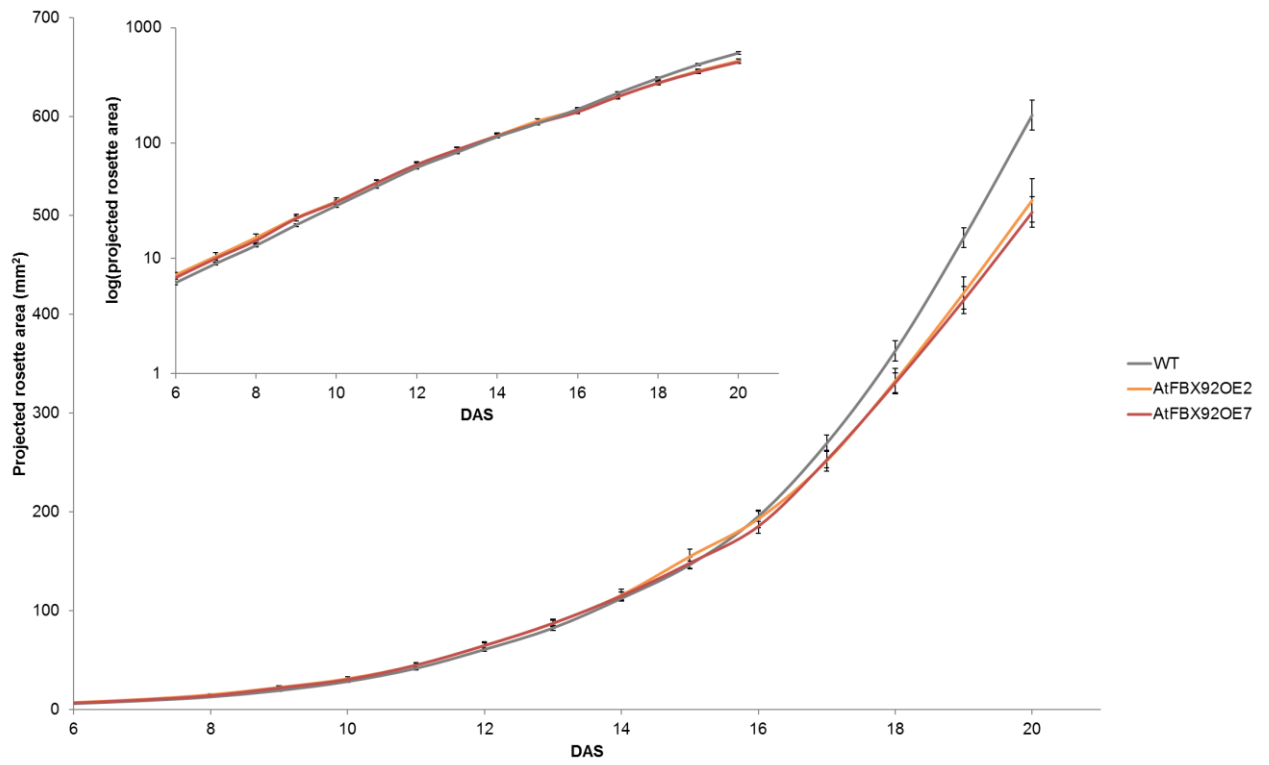


Supplementary Fig. S5. Amino acid sequence comparison of the ZmFBX92 and AtFBX92 proteins. Schematic representation of the protein domains in ZmFBX92, AtFBX92 and AtFBX92^{del} drawn to scale.

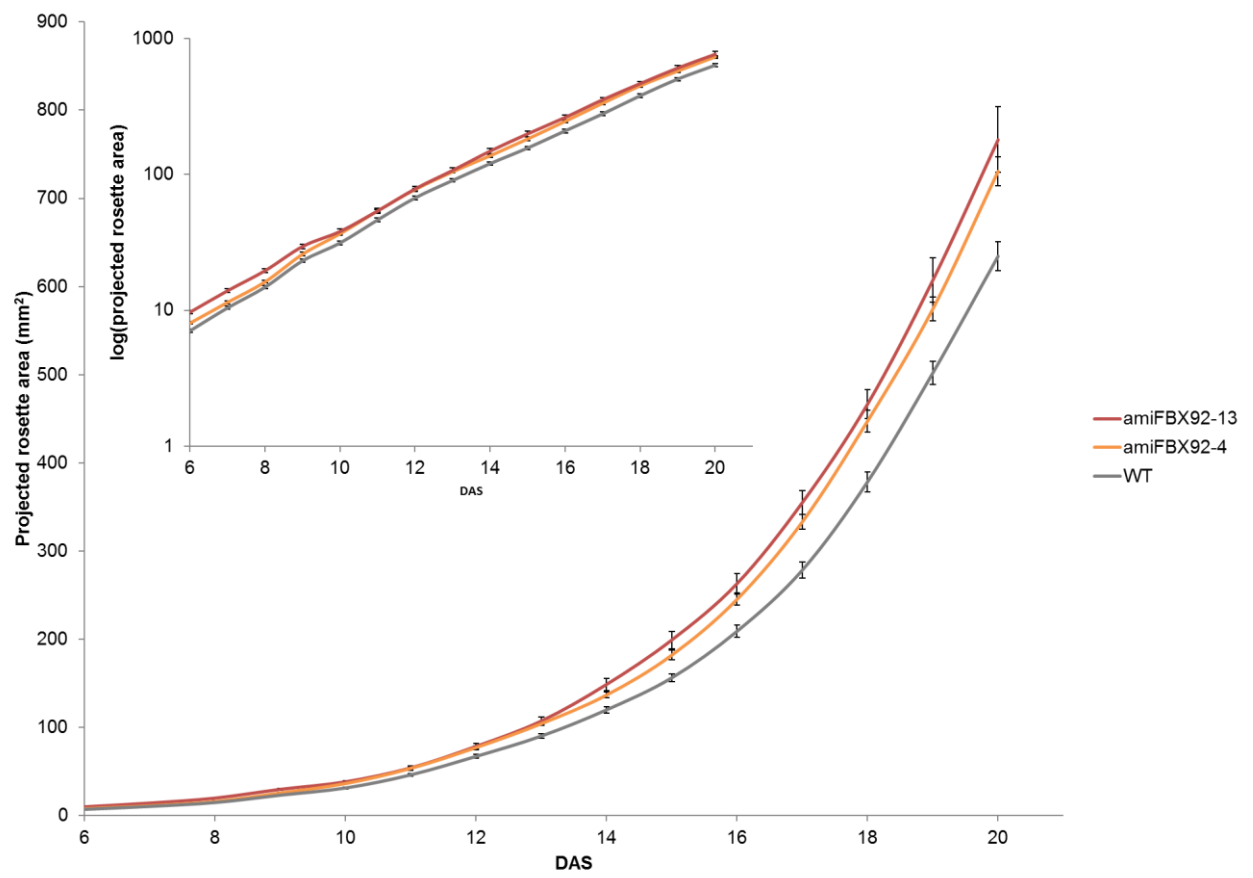


Supplementary Fig. S6. *AtFBX92* expression levels in leaf 3 during the subsequent phases of leaf development. Normalized expression levels as calculated from AGRONOMICS1 tiling arrays by Andriankaja et al. (2012).

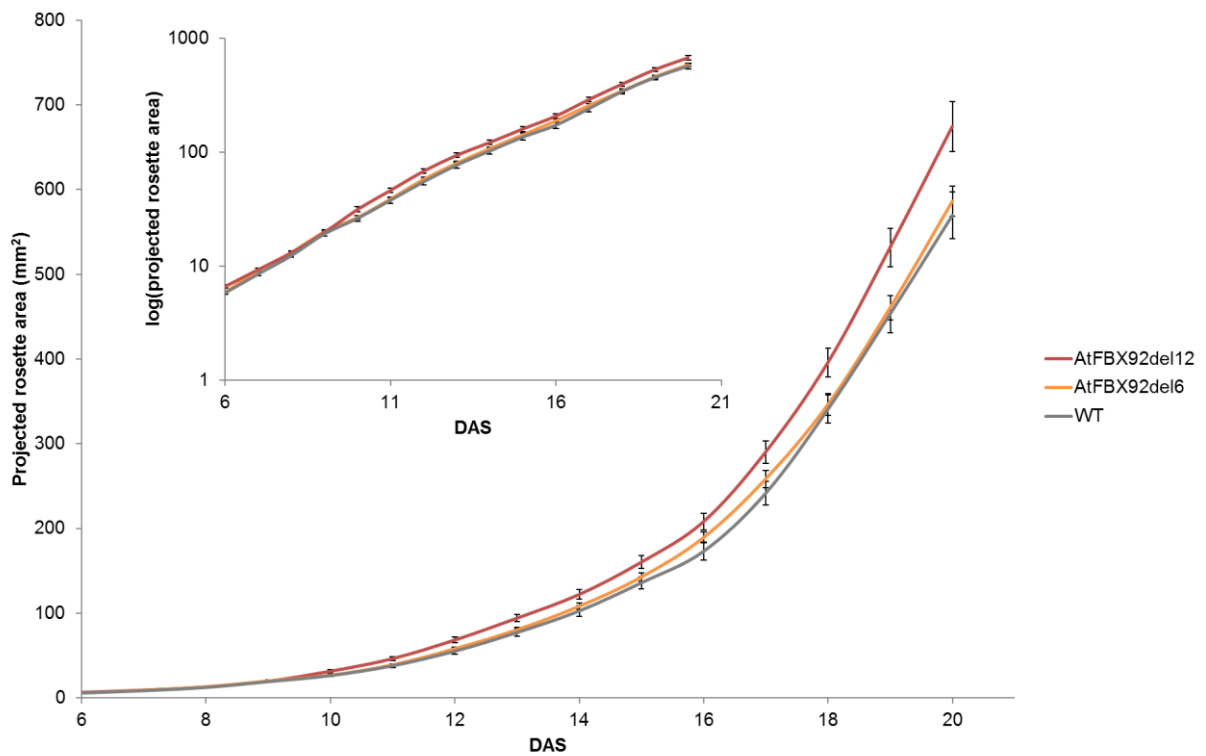
A.



B.

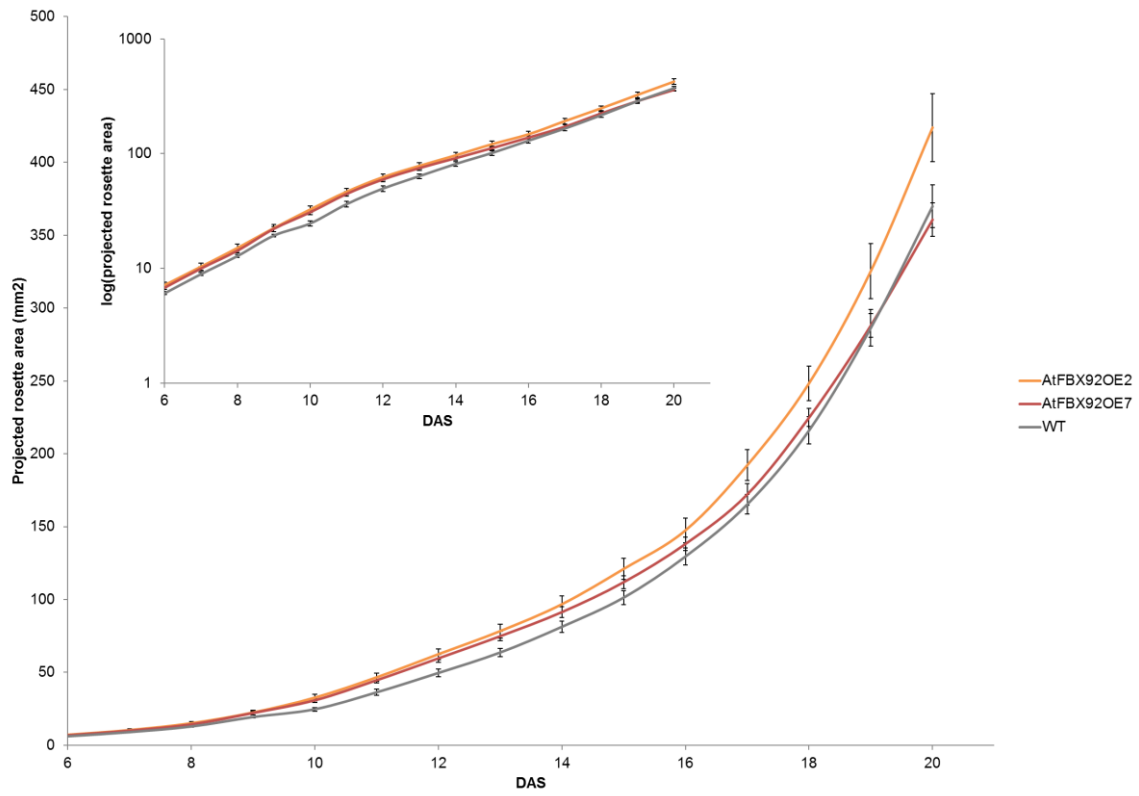


C.

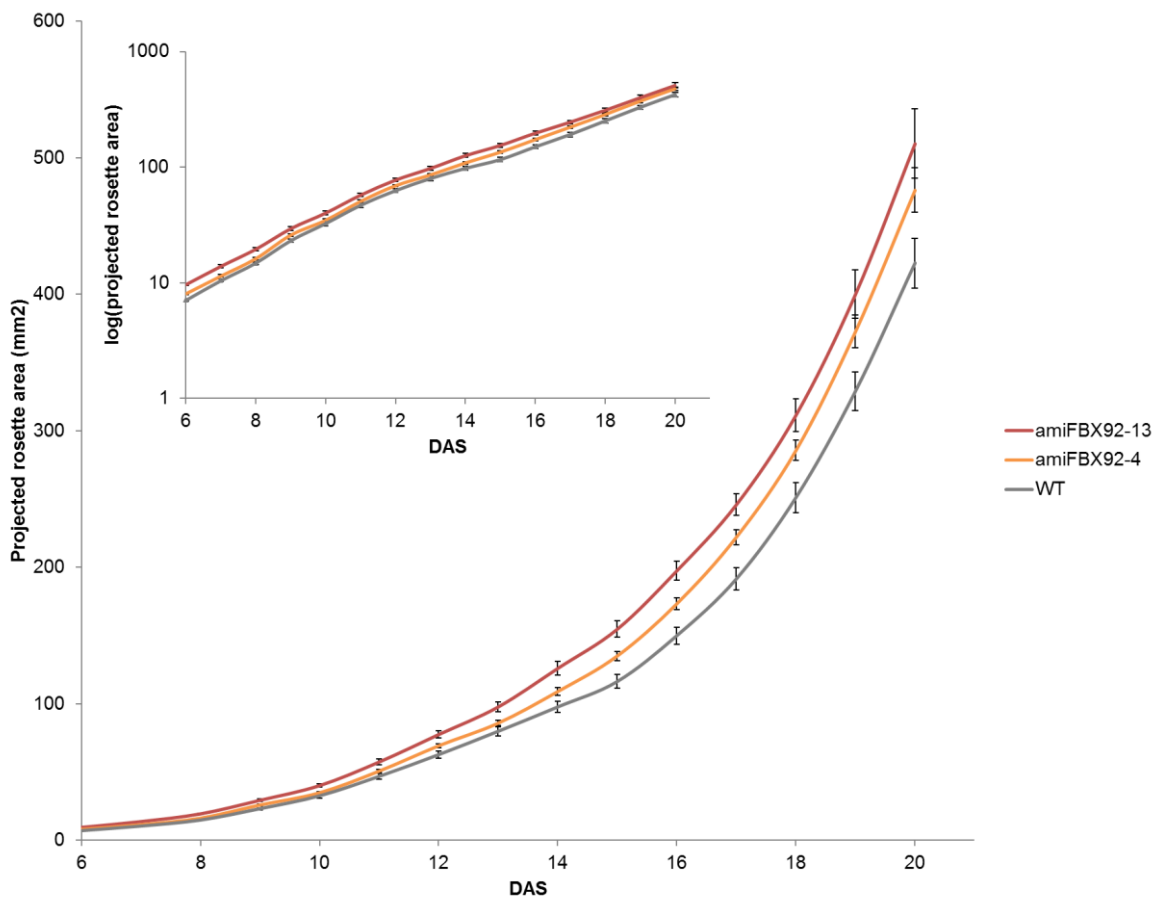


Supplementary Fig. S7. Effect of *AtFBX92* perturbation on rosette growth under standard conditions in soil. (A) PRA of *AtFBX92^{OE}* and WT plants grown in soil from 6 until 20 DAS. Plants were grown in standard. Inset: PRA in log scale. Values represent the mean \pm SE (n = 16). (B) PRA of *amiFBX92* and WT plants grown in soil from 6 until 20 DAS. Plants were grown in standard conditions. Inset: PRA in log scale. Values represent the mean \pm SE (n = 16). (C) PRA of *AtFBX92^{del}* and WT plants grown in soil from 6 until 20 DAS. Plants were grown in standard conditions. Inset: PRA in log scale. Values represent the mean \pm SE (n = 16).

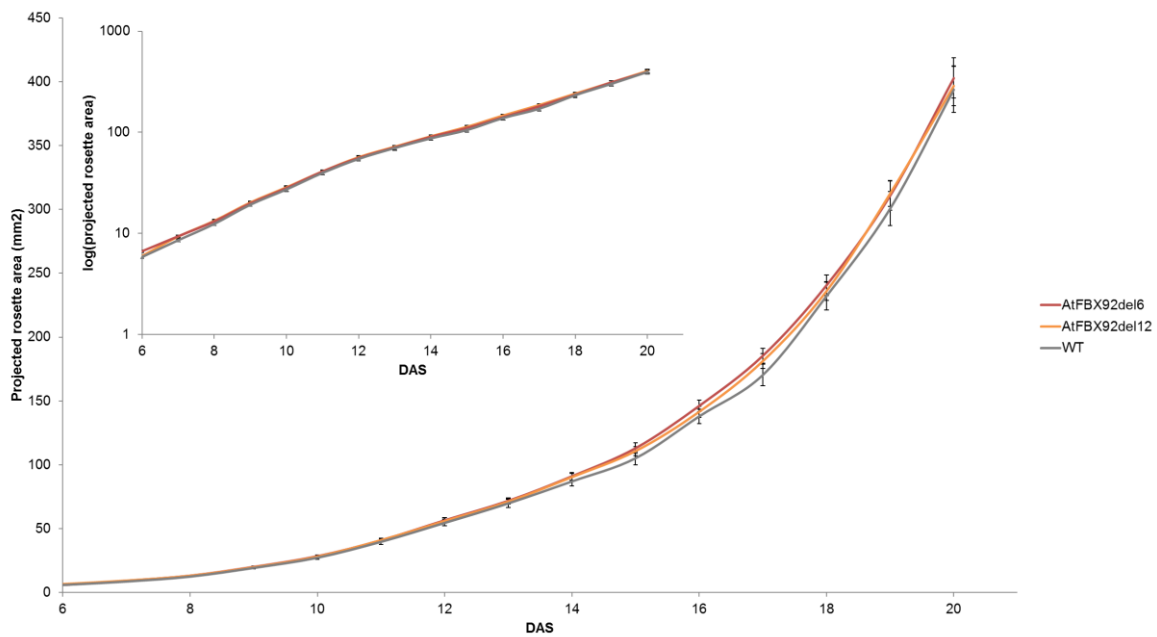
A.



B.

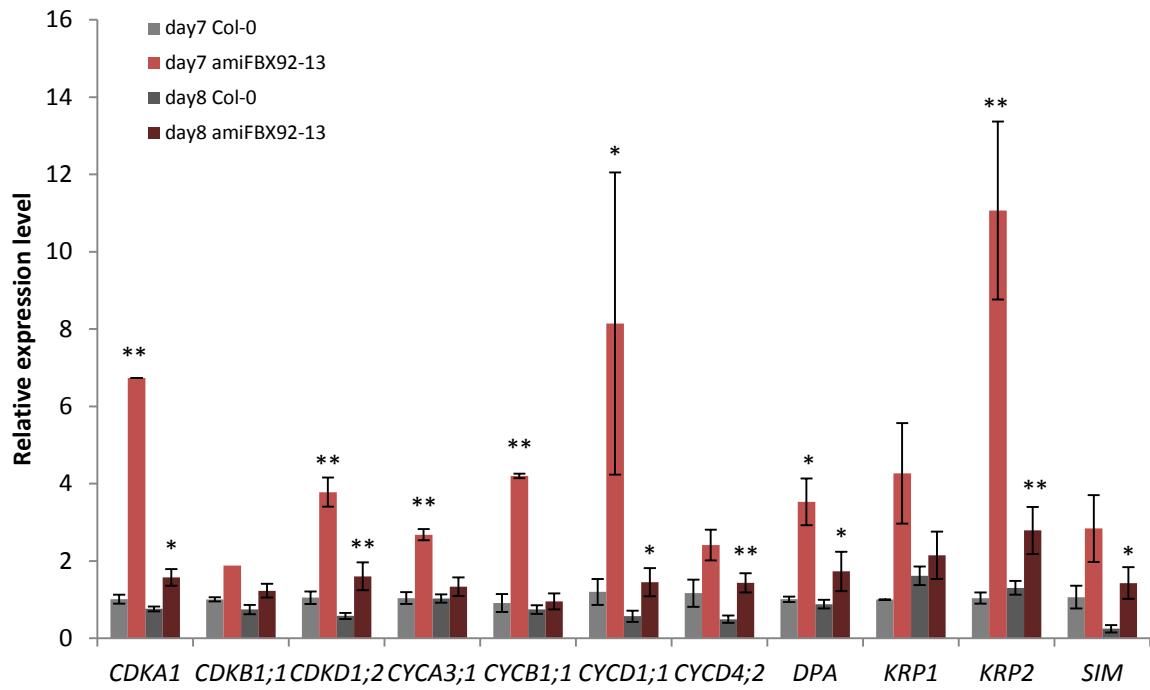


C.

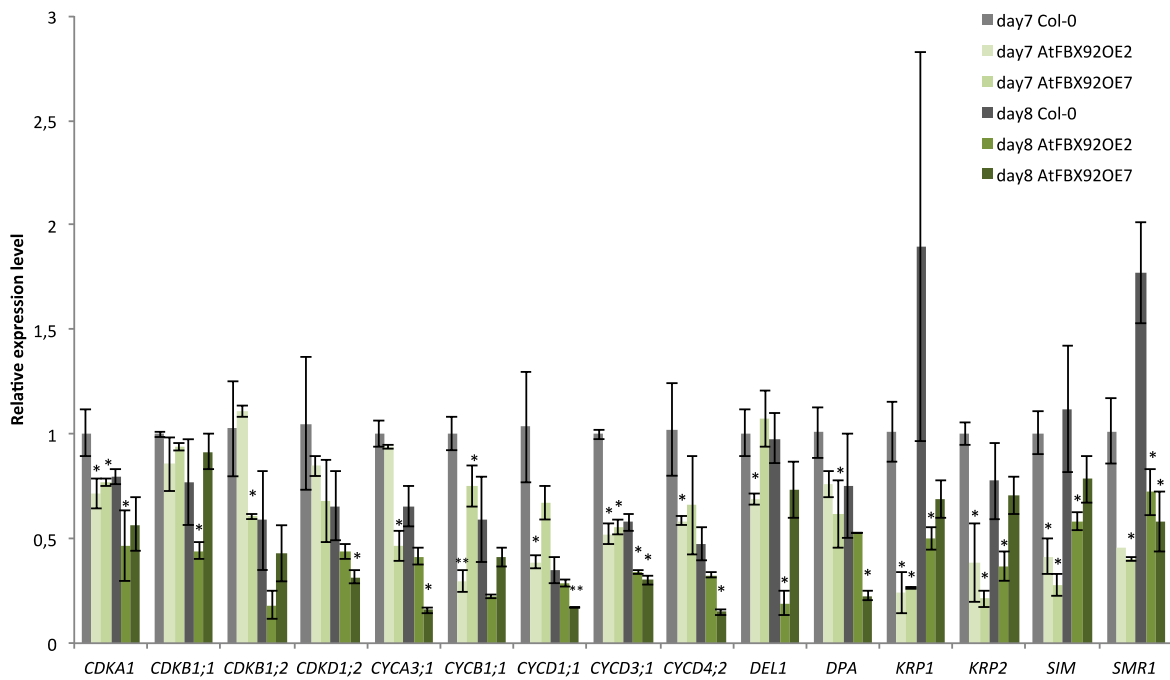


Supplementary Fig. S8. Effect of *AtFBX92* perturbation on rosette growth under mild drought conditions in soil. (A) PRA of *AtFBX92^{OE}* and WT plants grown in soil from 6 until 20 DAS. Plants were grown under mild drought conditions from 10 DAS onwards. Inset: PRA in log scale. Values represent the mean \pm SE ($n = 16$). (B) PRA of *amiFBX92* and WT plants grown in soil from 6 until 20 DAS. Plants were grown under mild drought conditions from 10 DAS onwards. Inset: PRA in log scale. Values represent the mean \pm SE ($n = 16$). (C) PRA of *AtFBX92^{del}* and WT plants grown in soil from 6 until 20 DAS. Plants were grown under mild drought conditions from 10 DAS onwards. Inset: PRA in log scale. Values represent the mean \pm SE ($n = 16$).

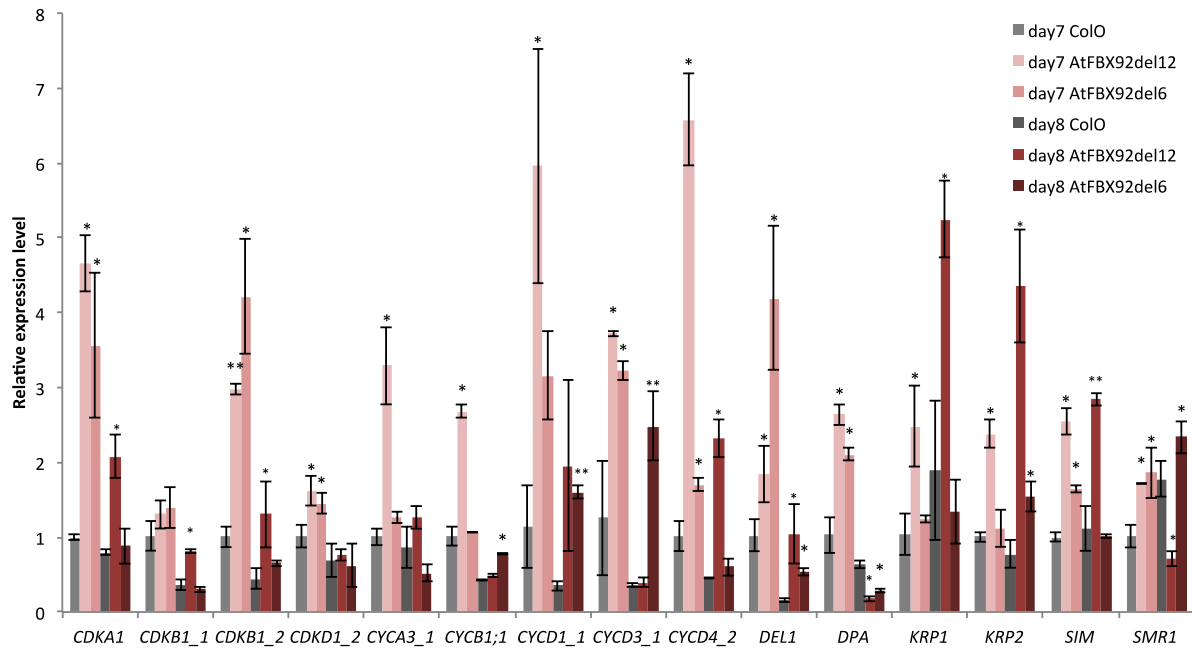
A.



B.



C.



Supplementary Fig. S9. Relative expression levels of cell cycle genes in the first leaf pair of *amiFBX92-13* (A), *AtFBX92^{OE2}* and *AtFBX92^{OE7}* (B) and *AtFBX92^{del12}* and *AtFBX92^{del6}* (C) compared to WT at 7 and 8 DAS as determined by qRT-PCR. Values were normalized against the expression level of the housekeeping gene and represent mean expression levels of indicated transcripts of three technical repeats \pm SE. Significant differences (Student's *t*-test): *, $P < 0.10$; **, $P < 0.01$ relative to WT.

Supplementary Table S1. Primer Sequences.

Construct	Application	Forward primer	Reverse primer
<i>ZmFBX92^{OE}</i>	cloning	AAAAAGCAGGCTTCATCTCTCCAGATCCG ATC	GAAAGCTGGGTCCCCTGTTTCATCTGAAACAG
<i>ZmFBX92</i>	qRT-PCR	GATGCCGATGATGTTGTG	CTGTTGCTGGAAGGACTC
18S rRNA	qRT-PCR	ACCTTACCAGCCCTTGACATATG	GACTTGACCAAACATCTCACGAC
<i>AtFBX92^{OE}</i>	cloning	AAAAAGCAGGCTATGGCTTCAGAG	GAAAGCTGGGTCTGAAGATGTACCC
<i>AtFBX92^{del}</i>	cloning	AAAAAGCAGGCTATGGCTTCAGAG	GAAAGCTGGGTCTTAGACAAGTTCTTGGTCCG
<i>amiFBX92</i>	cloning	gaTACTAGTCTATGGTTGGGCCGtctctcttttg tattcc	gaCGGCCCAACCATAGACTAGTAtcaaagagaatc aatga
<i>amiFBX92</i>	cloning	gaCGACCCAACCATACACTAGTTtcacaggtcg tgatatg	gaAACTAGTGTATGGTTGGGTCgtctacatatatt cct
<i>pAtFBX92</i>	cloning	AAAAAGCAGGCTAAAATGTCTTGACAGCTG	GAAAGCTGGGTCTAGCTCCTCGAATGAAGAG AATCAGAAG
<i>AtFBX92</i>	qRT-PCR	GCGCGGTCGTTTATGGTAAC	GCTTTAGCCCTTTGGCAAG
<i>AtFBX92^{del}</i>	qRT-PCR	GCAACGGTCTTCACTTCCTC	TGGTCGTGGTACTTGTGGA
<i>CDKA1</i>	qRT-PCR	TGATCAGCCAAAAGCCCTTA	ACGGAGTTCCTGATTCTG
<i>ACTIN1</i>	qRT-PCR	GTTGACTACGAGCAGGAGATGG	ACAAACGAGGGCTGGAACAAG
<i>CDKB1;1</i>	qRT-PCR	GCATTGCTTCATATCTTCAGGTT	CAGTCACGCAGTGTGGAAC
<i>CDKB1;2</i>	qRT-PCR	CCAATGAAGAAGTATACCCATGAG	AATGGGTGGCACCAAGAAG
<i>CDKD1;2</i>	qRT-PCR	CAGATTCCAGCTTCAAAGG	TGAATCCTTCAGGACCCATC
<i>CYCA3;1</i>	qRT-PCR	GCCGACGAAAAAGAGAAGT	CTTTGTCTATTGCCGTTCC
<i>CYCB1;1</i>	qRT-PCR	GCTTCTGCAATCTACGCAGC	CCAACAGCTTTGCACAGTCC
<i>CYCD1;1</i>	qRT-PCR	GCGAACGAGTTACCTTCTCTATCC	GCTCAATCCGTACACCAAGTC
<i>CYCD3;1</i>	qRT-PCR	CGAAGAATTCGTGAGGCTCT	ACTTCCACAACCGGCATATC
<i>CYCFBX92;2</i>	qRT-PCR	CCTAGTGGAAGCTTGACTG	TTAGCCTCAAACACGAACATGG
<i>DEL1</i>	qRT-PCR	TGATGACGCTGCAAAATTAAGT	CATAAAGCCGCTCACTTTAGTT
<i>DPA</i>	qRT-PCR	GCAAGAACAGAAGCAAGAACAGAAC	CGGTCAGCAGTATCAATGGATC
<i>KRP1</i>	qRT-PCR	GTGGAAGCAATGAATATAAGAAGAAA	CGACGTTTCAGTGTACCAT
<i>KRP2</i>	qRT-PCR	CGTGGATTTACGATGATTTGAA	GCGGCGAGACTCTACATCTT
<i>SIM</i>	qRT-PCR	CCCACTTCTCCGACCACAA	GCCGAGAGACGACGGTGT
<i>SMR1</i>	qRT-PCR	GCTACGCCGGCTCTGATG	CGGAGGAGAAGAAACGGTCA