

991 SUPPORTING INFORMATION

992 **Table 1: List of oligonucleotides used in this study.** All oligonucleotides were purchased from
 993 Integrated DNA Technologies. Restriction sites are underlined, InFusion overhangs are
 994 italicized, gRNA sequence is in lowercase.

Primer number	Oligonucleotide sequence
1	<i>CGATTTTTCTCGAGATGTCTTTGAGTATTTATGG</i>
2	<i>ATGTGCTGCACCTGGCCTAGGTGGTTCTTCTCCATCTTACGC</i>
3	<i>CGATTTTTCTCGAGATGAATTCTGTTATGCCTG</i>
4	<i>ATGTGCTGCACCTGGCCTAGGCATTCCTGATCTTTTTCTTGG</i>
5	<i>CGATTTTTCTCGAGATGTTAAAAAATCCAGGTAC</i>
6	<i>ATGTGCTGCACCTGGCCTAGGAAATTCATTTTTGATTG</i>
7	<i>CGATTTTTCTCGAGATGGCAGAAGTAGCAA</i>
8	<i>ATGTGCTGCACCTGGCCTAGGTCTACAAATAACTGTGTCATC</i>
9	<i>CGATTTTTCTCGAGATGAATATTGCTATTGATGAG</i>
10	<i>ATGTGCTGCACCTGGCCTAGGATTGAAGTCGTTTGGAGC</i>
11	<i>CGATTTTTCTCGAGATGTCCATACACCTGTTGAAC</i>
12	<i>ATGTGCTGCACCTGGCCTAGGTTTCTCCTCTCCCATTTGATTTCCC</i>
13	<i>CGATTTTTCTCGAGATGAGTCACTTAATG</i>
14	<i>ATGTGCTGCACCTGGCCTAGGTGAGTAAGGACTTCTC</i>
15	<i>CGATTTTTCTCGAGATGGTAAGCTAAGTTTGG</i>
16	<i>ATGTGCTGCACCTGGCCTAGGGAAGGCTTCGTCATAGTCAG</i>
17	AACGGTAAAAATAATAACACGAATTATCAC
18	TATTCATCTATTTATGGACAATGG
19	CCTATATGCACATATTCATATTAAGTATATAATATTCCTAGGAACTCATCG CTCGGATGCTGCCCGACAGTTTTAGAGCTAGAAATAGCAAGTTAAAATAA GGCTAGTCCGTTATCAACTTGAAAAAGTGGCACCGAGTCGGTGCTTTTTTAT TATTCCTATAAAAATAATATATATA
20	<i>ACGATTTTTCTCGAGATGGACAAGAAGTACAGCATCGGC</i>
21	<i>CCTTGTAGTCCCTAGGCACTTTCCGCTTTTTCTTAGGATCTTCCACCTTGCGT TTTTCTTGGGGTCGCCTCCAGCTGAGACAGGTC</i>
22	CTTTGTTGACTTGTGGTGTGTTGAAGAAAATCCAGGTCCAGACAAGAAGT ACAGCATCG
23	<i>TGCCATATCCCTCGAGT</i> ACTTGTGTCGTCGTCGTCCTTGTAGTCCACTTTCCGC TTTTTCT
24	<i>CGATATGAATTCTAGAC</i> ATTTTGTAATAAAAAAATTAATAATATATTTATATAAT ATTATTTT

25	CAACATCACCACAAGTCAACAAAGAACCTCTACCTTCACCAATGCTGTTCA ACTTCCCAC
26	TAAGTATATAATATTagaaggcttcgcatagtcGTTTTAGAGCTAGAA
27	TTCTAGCTCTAAAACtgactatgacgaagccttctAATATTATATACTTA
28	TAAGTATATAATATTatataattttacatcgaatGTTTTAGAGCTAGAA
29	TTCTAGCTCTAAAACattcgatgtaaaaattatatAATATTATATACTTA
30	GTATACCTTTGAAGACACATCATCAGCC <u>TTAAGG</u> TATGGTCATAGCGAGAG AAGC
31	<i>TTTCTGTTCCGGTAACCGCGAAGGCTTCGTCATAGTCAGGATTTTTGTCACGAG GTTTTGG</i>
32	<i>CCCTTTCGGGGCGCGCCGATATATCAATATATCATTATATCAATATATCAAG ATATC</i>
33	GCTTCTCTCGCTATGACCATAC <u>CTTAAGG</u> CTGATGATGTGTCTTCAAAGGTA TAC
34	CTTATGACGTACCTGATTATGCAC
35	GTAGACCCCATTTGTGAGTACATAAATATATTATATAAACTAGACTAGG

995 **SUPPLEMENTAL FIGURE LEGENDS**

996 **Supplemental Figure 1: Full length blots for Figure 1A- RBC contain TRiC subunits in the**
997 **cytosolic fraction.** As described in Figure 1, lysates from uninfected RBC (U), trophozoite-stage
998 infected RBC (I) or saponin isolated parasites (P) were extracted with RIPA buffer and probed
999 with antibodies against human TRiC subunits A) α , B) β , C) δ , D) ζ and E) θ . PfEF-1 α is a
1000 cytosolic parasite protein. Equivalent fractions were loaded in all blots. Immunoblots are
1001 representative of two-four independent experiments. Marker bands are in kDa.

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1003 **Supplemental Figure 2: Full length blot for GFP-tagged PfTRiC subunits.** As described in
1004 Figure 3, ectopic expression of GFP-tagged PfTRiC subunits was under the control of the strong,
1005 constitutive HSP86 promoter. Dd2 and 3D7 are parent parasite lines. No ~27 kDa GFP 'core' is
1006 observed. Equivalent fractions were loaded in all blots. Marker bands are in kDa.

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1008 **Supplemental Figure 3: Release of hemoglobin into the supernatant fraction upon TetO**

1009 **treatment.** The activity of TetO was confirmed by visually observing release of hemoglobin into

1010 the supernatant fraction. This is also clear in immunoblots as the abundant hemoglobin monomer

1011 cross-reacts (independent of the antibodies used) during the Western blotting procedure. Infected

1012 RBC samples (T = total sample) were fractionated with 1 HU of TetO (20 min, 37 °C) or 0.05 %

1013 saponin (Sap; 20 s, room temperature). PfHAD1 is a soluble parasite protein, remaining in the

1014 saponin- and TetO-pellet (P) fractions, whereas hemoglobin is detected in the saponin- and

1015 TetO-supernatant (SN) fractions. Equivalent fractions were loaded in all blots. Marker bands are

1016 in kDa. This representative blot relates to data presented in Figure 3B panel i).





