## SCIENTIFIC REPORTS

## Supplementary information

Molecular diagnostic assays based on cpn60 UT sequences reveal the geographic

distribution of subgroup 16SrXIII-(A/I)I phytoplasma in Mexico

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0.01

**Figure S1.** Phylogenetic tree reconstructed through the maximum likelihood method of the F2nR2 sequences obtained from samples S07-P-JC, S41-L-JB, and S10-L-JC, along with previously characterized phytoplasma strains. Accession numbers of the F2nR2 sequences obtained for the samples mentioned above are in the main text. Phylogenetic analysis was conducted using the maximum likelihood method with MEGA v6.0<sup>1</sup> and bootstrapping 1000 times to estimate stability. *Acholeplasma laidlawii* strain PG-8 A (U14905) was used as outgroup to root the tree. Bar, 1 substitution in 100 positions.





**Figure S2.** Symptoms detected in the berry plants analyzed in this study. (**A**) Strawberry plants showing symptoms previously described for SbGP disease featuring inedible fruit associated with the presence of green leaf-like structures. The symptoms depicted were associated with samples S12-P-JC, S10-P-JC, and S31-L-MB. (**B**) Blackberry showing inedible, yellowing fruit associated with the precense of green leaf-like structures (sample Bk01-L-JC). (**C**) Blueberry plants showing small leaves along with yellowing and redness in leaves (samples Bl01-L-JA and Bl02-L-JB). (**D**) Raspberry showing white leaf mosaic (R06-L-MC), little leaves and green structures in the fruits, a symptom referred to as 'fruit witch's broom' (R01to 03-L-MA).



**Fig. S3.** LAMP assay linearity and detection limits. Assays were performed using calcein (closed circles) or isothermal (open circles) detection chemistry with known copy numbers of SbGP/MPV *cpn60* plasmids in a background of uninfected strawberry DNA.



Fig. S4. Analytical specificity of the assays. a. and c. Quantitative PCR assay. b. and d. LAMP assay. a. and b. show a lack of assay signal (\*) using no template as well as genomic DNA templates isolated from plants affected by the following strains: AY-Ruta (<u>16SrI-A</u>, '*Ca*. P. asteris- related strain), SF1 (<u>16SrI-B</u>, '*Ca*. P. asteris'-related strain), CVB, AY-Col (<u>16SrI-C</u>, '*Ca*. P. asteris'-related strain), RS (<u>16SrV-A</u>, '*Ca*. P. ulmi'-related strain), AshY (<u>16SrVII-A</u>, '*Ca*. P. fraxini'-related strain), Cr (<u>16SrIX-H</u>, '*Ca*. P. phoenicium'-related strain), AP (<u>16SrX-A</u>, '*Ca*. P. mali'-related strain), PYLR (<u>16SrX-C</u>, '*Ca*. P. pyri'-related strain), ESFY (<u>16SrX-F</u>, '*Ca*. P. prunorum'-related strain), and BN44948 (<u>16SrXII-A</u>, '*Ca*. P. solani'-related strain). c. and d. show a lack of signal (\*) using no template control and using genomic DNA extracted from uninfected strawberry, blueberry, blackberry, raspberry, and periwinkle plants.



**Fig. S5.** Effect of dilution on the sensitivity of the LAMP, qRT- PCR, and ddPCR assays. **a.** Amplification curves obtained using LAMP assays where red is undiluted, orange 1:5, yellow 1:10, light green 1:20, dark green 1:50, light blue 1:10, dark blue no template control, and pink SbGP/MPV *cpn60* (+ control) of the corresponding sample. **b.** qRT-PCR dilution effect on the calculated number of genomes/g tissue. **c.** ddPCR dilution effect on the proportional abundance of SbGP genomes normalized to strawberry rubisco genes.



**Fig. S6.** F2nR2 gene fragment amplification of the samples collected in 2015 through nested PCR with P1/P7 primers in the first reaction, and R16F2n/R16R2 in the nested reaction. **a.** Dilution optimization of P1/P7 PCR product for use in the subsequent nested reaction. **b.** Electrophoresis of the nested PCR products of samples collected in 2015 with P1/P7 PCR product diluted 1:30. 1 to 55 is S01-L-MA to S43-L-JB (Table 1), 56 to 63 is R01-L-MA to R08-L-MC (Table 1), 64 to 65 is Bl01-L-JA to Bl02-L-JB (Table 1), 66 to 69 is Bk01-L-JC to Bk04-L-ME (Table 1).

Sample	F2nR2 PCR	cpn60 PCR	<i>cpn60</i> LAMP	Fluorescent microsphere hybridization	<i>cpn60</i> sequencing	<i>cpn60</i> qRT-PCR <sup>a</sup>	<i>cpn60</i> ddPCR <sup>b</sup>
S11-L-JC	+	+	+	SbGP/MPV	NT <sup>c</sup>	3.86x10 <sup>7</sup>	0.5650 (fractional
S33-L-MB	+	+	+	-	SbGP/MPV 100%	2.02x10 <sup>6</sup>	abundance) 19.35 (fractional abundance)
S43-L-JB	+	+	+	-	$ND^d$	1.51x10 <sup>5</sup>	0.0325 (fractional abundance)
R01-L-MA	+	+	+	NT	SbGP/MPV 100%	1.87x10 <sup>5</sup>	$2.66 \times 10^{6}$ (genomes/g)
R01-P-MA	+	+	+	NT	NT	DNQ	$9.75 \times 10^5$ (genomes/g)
R02-L-MA	-	+	+	NT	NT	DNQ	1.31x10 <sup>5</sup> (genomes/g)
R03-L-MA	-	+	+	NT	ND	DNQ	$1.31 \times 10^5$ (genomes/g)
R06-L-MC	-	+	+	NT	SbGP/MPV 100%	1.66x10 <sup>5</sup>	$3.32 \times 10^6$ (genomes/g)
R08-L-MC	+	+	+	NT	ND	$1.03 \times 10^5$	$3.15 \times 10^6$ (genomes/g)
Bk01-L-JC	-	+	+	NT	ND	$8.87 \times 10^4$	$3.13 \times 10^{6}$ (genomes/g)
Bk03-L- MD	-	+	+	NT	SbGP/MPV 100%	$3.49 \times 10^4$	$9.00 \times 10^5$ (genomes/g)
Bl01-L-JA	+	+	+	NT	SbGP/MPV 100%	1.34x10 <sup>5</sup>	$2.79 \times 10^{6}$ (genomes/g)
S22-L-MB	+	-	+	SbGP/MPV	ND	$1.40 \times 10^{6}$	0.008 (fractional abundance)
S06-P-MB	+	-	+	SbGP/MPV	NT	1.89x10 <sup>7</sup>	1.24 (fractional abundance)

**Table S1.** F2nR2 direct and nested and *cpn*60 direct PCR discordant samples confirmed by sequencing of *cpn*60 UT PCR product or the *cpn*60 UT-based molecular diagnostics developed in this study.

<sup>a</sup>qRT-PCR data expressed as genomes/g tissue

<sup>b</sup>ddPCR data expressed as fractional abundance for S (Strawberry) samples or genomes/g tissue for R (raspberry), Bk (blackberry), and Bl (blueberry) samples.

<sup>c</sup> NT, not tested.

<sup>d</sup>ND, not determined. Samples from raspberry, blueberry, and blackberry yielded low amounts of PCR product that provided poor quality sequence in many cases.

GenBank accession number of the *cpn60* UT sequenced from the samples: S33-L-MB (KY061181), R01-L-MA (KY061169), R06-L-MC (KY061172), Bk03-L-MD (KY061185), and Bl01-L-JA (KY061168).

Primer name	Sequence (5'-3')			Amplificatio	on conditions	
		Product	Detection			
		size (bp)	assay	qRT-PCR	ddPCR	
D0414	AGGGGCTAATCCTATTTTC			1x 95°C, 3		
D0415	CCTGAAGAAATAGCTGCTA			min; 40 x	1x 94°C 10	
SbGP	FAM-ATTAGCGTC/ZEN/TCAAGTGGTTTCTCAAA-IB	132	qRT-PCR and ddPCR	95°C, 10 sec; 56°C, 10 sec; 72°C, 30 sec (data collection)	min; 50x 94°C, 30 sec; 52°C, 1:00 (ramp 2°C/sec);	
D0490	ACTACCTTGGCTCATTAT				- 1X 98°C, 10	
D0491	CCATGATTCTTCTGTCTATC	92	ddPCR		111111	
Frag-rubisco <sup>a</sup>	HEX-CAACGGTCT/ZEN/ACTTCTTCACATC-IB					
MS31_2-F3	AAAGGATTTGAAACCGAGTTA					
MS31_2-B3	CCCTTAATTTATTAGCAACCAA					
MS31 2-FIP	ATTCAACCGCCATAGATTCTCT-					
10001 <u>2</u> 11	GAAGTAGTTCAAGGTTTACAAT		LAMP	63°C 60 minutes (calcein detection) or 30 minutes (isothermal detection)		
MS31 2-BIP	AACCATTAGTTTTAGTGACCGA-	204				
	TTCAACTGATTCAGCTACAATT					
MS31_2-					,	
loopF	AATACGGAGAAGCATATCCTT					
MS31_2-						
ТоорВ	AGAAGGIGIIGIIAAAGAAICI					
		(0.1	Fluorescent			
		604	microsphere			
	ACCACITGAGACGCIAAI	1 1 1	nybridization			
Primers and pr	obe designed using <i>Fragaria</i> x ananassa ribulose-bisphosphate c	carboxylase I	arge subunit ( <i>rbc</i>	L), GenBank F	$^{\circ}AU06805$	
D0490 D0491 Frag-rubisco <sup>a</sup> MS31_2-F3 MS31_2-B3 MS31_2-FIP MS31_2-FIP MS31_2-BIP MS31_2- loopF MS31_2- loopB <u>SbGP<sup>b</sup></u> <sup>a</sup> Primers and pr <sup>b</sup> hybridization of	ACTACCTTGGCTCATTAT CCATGATTCTTCTGTCTATC HEX-CAACGGTCT/ZEN/ACTTCTTCACATC-IB AAAGGATTTGAAACCGAGTTA CCCTTAATTTATTAGCAACCAA ATTCAACCGCCATAGATTCTCT- GAAGTAGTTCAAGGTTTACAAT AACCATTAGTTTAGTGACCGA- TTCAACTGATTCAGCTACAATT AATACGGAGAAGCATATCCTT AGAAGGTGTTGTTAAAGAATCT ACCACTTGAGACGCTAAT obe designed using <i>Fragaria</i> x ananassa ribulose-bisphosphate c capture probe used in combination with <i>cpn60</i> -targeted universal	92 204 604 carboxylase l phytoplasma	ddPCR LAMP Fluorescent microsphere hybridization arge subunit ( <i>rbc</i> a amplification p	63°C 60 min detection) o (isotherma L), GenBank F	nutes (calcein or 30 minutes I detection) FAU06805 ibed <sup>2</sup>	

**Table S2.** Sequences of amplification primers and hydrolysis probes used for detection and quantification of SbGP/MPV phytoplasmas.

												L	AMP		
		DCD 1	DCD2			1 104	CD			LAMP		Time to p	positive	(Tp),	
		PCRI	PCR2	PCR3		ddPo	L.K	aRT-PCR		(calcein)		m	inutes		
	DNA		cpn60	cpn60-				SbGP					Isothe	rmal	
	conc.	F2nR2	-univ <sup>a</sup>	<b>F</b> MA <sup>b</sup>	FMA		Sb-	(genomes/				Calcein	detect	ion	Tanneal,
Sample	ng/µl	(1:50)	(1:50)	(1:50)	type	SbGP <sup>c</sup>	rubisco	g tissue)	host	neat	1:50	1:50	1:50		°C
S26b-GP-MA	$NT^{d}$	pos	pos	$NT^{d}$	NT	$3.85 \times 10^{8}$	NT	$1.12 \times 10^{9}$	strawberry	pos	NT	35.38	NT		
S26b-P-MA	NT	NT <sup>e</sup>	NT	NT	NT	$4.23 \times 10^{8}$	NT	$1.04 \mathrm{X} 10^{9}$	strawberry	pos	NT	25.25	NT		
S26b-F-MA	NT	NT <sup>e</sup>	NT	NT	NT	$1.46 \mathrm{X} 10^{8}$	NT	$3.98 \times 10^{8}$	strawberry	pos	NT	38.38	NT		
S27b-GP-MA	NT	pos	pos	NT	NT	6.92X10 <sup>8</sup>	NT	$4.01 \times 10^{9}$	strawberry	pos	NT	30.88	NT		
S31b-L-MA	NT	pos	pos	NT	NT	$2.41 \times 10^{8}$	NT	$4.82 \mathrm{X} 10^{8}$	strawberry	pos	NT	45.88	NT		
S31b-GP-MA	NT	NT <sup>e</sup>	pos	NT	NT	$4.80 \mathrm{X} 10^{8}$	NT	$2.74 \times 10^{9}$	strawberry	pos	NT	31.88	NT		
S31b-P-MA	NT	NT <sup>e</sup>	NT	NT	NT	$2.11X10^{8}$	NT	$4.34 X 10^8$	strawberry	pos	NT	35.00	NT		
S31b-F-MA	NT	NT <sup>e</sup>	NT	NT	NT	$5.48 \times 10^{7}$	NT	$9.69 \times 10^7$	strawberry	pos	NT	45.00	NT		
S267-GP-MA	NT	NT <sup>e</sup>	neg	NT	NT	neg	NT	$8.21X10^{2}$	strawberry	neg	NT	$ND^{f}$	NT		
S267-P-MA	NT	NT <sup>e</sup>	NT	NT	NT	neg	NT	neg	strawberry	neg	NT	ND	NT		
S267-F-MA	NT	NT <sup>e</sup>	NT	NT	NT	neg	NT	neg	strawberry	neg	NT	ND	NT		
S289-L-MA	NT	pos	NT	NT	NT	$7.94 \times 10^{7}$	NT	$1.34 X 10^{8}$	strawberry	pos	NT	49.63	NT		
S289-GP-MA	NT	NT <sup>e</sup>	NT	NT	NT	neg	NT	neg	strawberry	neg	NT	ND	NT		
S289-P-MA	NT	NT <sup>e</sup>	pos	NT	NT	$7.16 \text{X} 10^7$	NT	$4.24 X 10^{8}$	strawberry	pos	NT	35.38	NT		
S289-F-MA	NT	NT <sup>e</sup>	NT	NT	NT	neg	NT	neg	strawberry	neg	NT	ND	NT		
S01-L-MA	16.9	neg	neg	neg		0.0025	pos	<b>DNQ</b> <sup>g</sup>	strawberry	neg	neg	NT	neg		
S01-P-MA	9.72	pos	pos	pos	SbGP	2.6300	pos	$1.94 X 10^{7}$	strawberry	pos	pos	NT		13.5	81.14
S02-P-JA	6.32	neg	neg	neg		0.0095	pos	DNQ	strawberry	neg	neg	NT	neg		
S03-P-JB	5.28	neg	neg	neg		0.0000	pos	DNQ	strawberry	neg	neg	NT	neg		
S04-L-MB	14.4	pos	pos	pos	SbGP	3.5900	pos	$5.50 \times 10^{7}$	strawberry	pos	pos	NT		15.25	81.09
S05-L-MB	8.08	pos	pos	pos	SbGP	2.7800	pos	$2.21X10^{7}$	strawberry	pos	pos	NT		13.75	81.09
S05-P-MB	56.8	pos	pos	pos	SbGP	5.5300	pos	$1.47 X 10^{8}$	strawberry	pos	pos	NT		13.75	81.14
S06-L-MB	38.2	pos	pos	pos	SbGP	6.5950	pos	$2.23X10^{8}$	strawberry	pos	pos	NT		13.5	81.14
S06-P-MB	37.7	pos	neg	pos	SbGP	1.2400	pos	$1.89 \times 10^{7}$	strawberry	pos	pos	NT		13.75	81.09
S07-P-JC	14.1	pos	pos	pos	SbGP	16.0850	pos	$2.61 \times 10^{8}$	strawberry	pos	pos	NT		12.5	81.24
S08-L-JA	13.4	neg	neg	neg		0.0200	pos	$2.09 \times 10^{5}$	strawberry	neg	neg	NT	neg		
S09-L-MB	10.6	pos	pos	pos	SbGP	3.0100	pos	$1.83 X 10^{7}$	strawberry	pos	pos	NT		13	81.27
S09-P-MB	25.3	pos	pos	pos	SbGP	1.9150	pos	$5.04 \times 10^{7}$	strawberry	pos	pos	NT		13.75	81.09

S10-L-JC	11.5	pos	pos	pos	SbGP	11.6700	pos	$3.75 \times 10^{8}$	strawberry	pos	pos	NT		12.5	81.14
S10-P-JC	12.7	pos	pos	pos	SbGP	17.9500	pos	$2.57 X 10^8$	strawberry	pos	pos	NT		13.5	81.19
S11-L-JC	20.7	pos	pos	pos	SbGP	0.5650	pos	$3.86 \times 10^7$	strawberry	pos	pos	NT		14	81.09
S12-P-JC	7.2	pos	pos	pos	SbGP	1.345	pos	$6.52 \times 10^{6}$	strawberry	pos	pos	NT		15	81.14
S13-L-JC	11.4	pos	pos	pos	SbGP	2.905	pos	$2.35 \times 10^{7}$	strawberry	pos	pos	NT		17.75	81.02
S14-P-JA	17	neg	neg	neg		0.013	pos	$2.37 \times 10^{5}$	strawberry	pos	neg	NT	neg		
S15-L-JA	21.5	neg	neg	neg		0.0025	pos	$6.26 \text{X} 10^4$	strawberry	pos	pos	NT	neg		
S16-P-JA	9.88	neg	neg	neg		0.019	pos	DNQ	strawberry	pos	neg	NT	neg		
S17-P-JA	11	neg	neg	neg		0.007	pos	$6.34 X 10^4$	strawberry	pos	neg	NT	neg		
S18-P-JA	10.6	neg	neg	neg		0.0375	pos	$2.54X10^{5}$	strawberry	pos	neg	NT		22.75	81.09
S19-L-JD	8.48	pos	pos	pos	SbGP	3.305	pos	$7.41 \text{X} 10^7$	strawberry	pos	pos	NT		14.25	81.05
S19-P-JD	7	pos	pos	pos	SbGP	2.415	pos	$2.16 \times 10^7$	strawberry	pos	pos	NT		14	81.15
S20-L-JD	12.8	neg	neg	neg		0.0025	pos	DNQ	strawberry	neg	neg	NT	neg		
S21-L-MB	8.56	pos	pos	pos	SbGP	0.64	pos	$1.12X10^{7}$	strawberry	pos	pos	NT		16.75	81.14
S21-P-MB	7.44	pos	pos	pos	SbGP	0.315	pos	$1.60 \mathrm{X} 10^7$	strawberry	pos	pos	NT		15.5	81.15
S22-L-MB	14.2	pos	neg	pos	SbGP	0.008	pos	$1.40 \mathrm{X} 10^{6}$	strawberry	pos	pos	NT		15.25	81.1
S22-P-MB	13.2	pos	pos	pos	SbGP	0.995	pos	$1.07 X 10^{8}$	strawberry	pos	pos	NT		13.5	81.06
S23-L-MB	15.4	pos	pos	pos	SbGP	14.195	pos	$2.62 \times 10^{8}$	strawberry	pos	pos	NT		12.75	81.05
S24-P-JB	9.24	neg	neg	neg		0.0135	pos	DNQ	strawberry	pos	neg	NT	neg		
S25-L-JC	19.6	pos	pos	pos	SbGP	3.05	pos	$8.51 \times 10^{7}$	strawberry	pos	pos	NT		14	81.07
S25-P-JC	3.97	pos	pos	pos	SbGP	1.365	pos	$2.31X10^{8}$	strawberry	pos	pos	NT		13.25	80.97
S26-L-JA	17.8	pos	pos	pos	SbGP	3.065	pos	$8.38 \times 10^{7}$	strawberry	pos	pos	NT		14.25	80.95
S26-P-JA	15.2	neg	neg	neg		0.011	pos	$4.64 \text{X} 10^4$	strawberry	pos	pos	NT	neg		
S27-P-JA	0.896	neg	neg	neg		0	pos	0.00	strawberry	neg	neg	NT	neg		
S28-L-MB	0.808	pos	pos	pos	SbGP	6.95	pos	$1.14 X 10^{6}$	strawberry	pos	pos	NT		13.25	81.09
S29-L-MB	2.38	neg	neg	neg		0.16	pos	$4.45 X 10^4$	strawberry	pos	neg	NT	neg		
S30-P-MB	0.596	neg	neg	neg		0.07	pos	0.00	strawberry	pos	neg	NT	neg		
S31-L-MB	0.832	pos	pos	neg		2.3	pos	$3.02 \times 10^5$	strawberry	pos	pos	NT		12.5	81.04
S32-L-MB	0.952	pos	pos	neg		2.55	pos	$3.20 \times 10^5$	strawberry	pos	pos	NT		12	81
S33-L-MB	0.876	pos	pos	neg		19.35	pos	$2.02X10^{6}$	strawberry	pos	pos	NT		10.75	80.91
S34-L-MC	1.21	neg	neg	neg		0	pos	$2.18 \times 10^{5}$	strawberry	pos	neg	NT		15.25	80.96
S35-L-MD	1.44	neg	neg	neg		0.055	pos	DNQ	strawberry	pos	pos	NT		10.25	80.78
S36-L-ME	5.56	neg	neg	neg		5.95	pos	0.00	strawberry	neg	pos	NT		11.5	80.81
S37-P-MD	1.2	neg	neg	neg		0	pos	DNQ	strawberry	pos	neg	NT		15.5	81.3

S38-L-MF	2.66	neg	neg	neg		0.04	pos	DNQ	strawberry	pos	pos	NT		12.25	80.8
S39-L-MG	2.03	neg	neg	neg		NT	pos	0.00	strawberry	neg	neg	NT		12.5	81.16
S40-L-MH	1.29	neg	neg	neg		0.04	pos	0.00	strawberry	neg	neg	NT		12.25	80.81
S41-L-JB	8.4	pos	pos	pos	AY- OY-M AY-	NT	pos	6.52X10 <sup>4</sup>	strawberry	pos	pos	NT		10.5	80.92
S41-P-JB	1.8	pos	pos	pos	OY-M	0.0565	pos	$5.02X10^4$	strawberry	pos	pos	NT		11	80.92
S42-L-JB	3.43	pos	pos	neg		0.605	pos	$1.64 X 10^{6}$	strawberry	pos	pos	NT		9.75	80.89
S43-L-JB	6.16	pos	pos	neg		0.0325	pos	$1.51 \times 10^{5}$	strawberry	pos	pos	NT		10	80.92
R01-L-MA	5.6	pos	pos	NT		$2.66 \times 10^{6}$	NT	$1.87 \mathrm{X} 10^{5}$	raspberry	pos	pos	NT		9.75	81.05
R01-P-MA	1.44	pos	pos	NT		9.75X10 <sup>5</sup>	NT	DNQ	raspberry	pos	pos	NT		10.5	81.15
R02-L-MA	12.9	neg	neg	NT		$1.31X10^{5}$	NT	DNQ	raspberry	pos	pos	NT		9.25	81.07
R03-L-MA	3.7	neg	pos	NT		$1.31X10^{5}$	NT	DNQ	raspberry	pos	pos	NT		9.5	81.02
R04-L-MB	7.96	neg	neg	NT		$1.11X10^{6}$	NT	$5.36X10^{4}$	raspberry	pos	pos	NT		9.25	81
R05-L-MC	17.1	pos	pos	NT		$5.44 X 10^{6}$	NT	$2.05 \times 10^{5}$	raspberry	pos	pos	NT		9.5	81
R06-L-MC	4.2	neg	pos	NT		$3.32 \times 10^{6}$	NT	$1.66 \times 10^5$	raspberry	pos	pos	NT		9.5	81.05
R07-L-MC	9	neg	neg	NT		$7.13 \times 10^{5}$	NT	DNQ	raspberry	pos	pos	NT		9.5	81.2
R08-L-MC	4.92	pos	pos	NT		$3.15 \times 10^{6}$	NT	$1.03 \times 10^{5}$	raspberry	pos	pos	NT		11.75	82.1
Bk01-L-JC	1.74	neg	pos	NT		$3.13 \times 10^{6}$	NT	$8.87 X 10^4$	blackberry	pos	pos	NT		11.5	80.86
Bk02-L-JD	3.54	pos	pos	NT		$1.50 \times 10^{6}$	NT	$4.22X10^{4}$	blackberry	pos	pos	NT		9.5	80.99
Bk03-L-MD	19.9	neg	pos	NT		$9.00 \text{X} 10^5$	NT	$3.49X10^4$	blackberry	pos	pos	NT		8.75	81.09
Bk04-L-ME	26.5	neg	neg	NT		0.00	NT	DNQ	blackberry	neg	neg	NT	neg		
Bl01-L-JA	3.56	pos	pos	NT		$2.79 \times 10^{6}$	NT	$1.34 X 10^{5}$	blueberry	pos	pos	NT		9	81.05
Bl02-L-JB	3.01	neg	neg	NT		0.00	NT	DNQ	blueberry	pos	pos	NT		10.25	81.11
P83-L-SLP		pos	pos	NT		2.53X10 <sup>9</sup>	NT	$4.67 \times 10^{9}$	periwinkle	pos	NT	38.42	NT		
P86-L-SLP		pos	pos	NT		$2.82 \times 10^{9}$	NT	6.13X10 <sup>9</sup>	periwinkle	pos	NT	37.08	NT		

<sup>a</sup>cpn60-univ, PCR using cpn60-targeted universal phytoplasma primers<sup>2</sup>

<sup>b</sup>FMA, fluorescent microsphere hybridization assay

<sup>c</sup>ddPCR data expressed as genomes/g tissue extracted where internal control data is NT; otherwise expressed as fractional abundance

<sup>d</sup>NT, Not tested

<sup>e</sup>sample was neg using 16S-targeted pan-phytoplasma primers P1-Tint<sup>3</sup>

<sup>f</sup>ND, Not detected

<sup>g</sup>DNQ, Detectable but not quantifiable

 Table S3. Complete results of all assays on all samples.

Sample	qRT-PCR genomes/g tissue							
	Mean (n)	Standard deviation						
S03-P-JB	DNQ (2)	-						
S24-P-JB	DNQ (2)	-						
S41-L-JB	$7.68 \times 10^{6} (2)$	$1.03 \times 10^{6}$						
S41-P-JB	$8.61 \times 10^{6} (2)$	$5.07 \times 10^{6}$						
S42-L-JB	0 (2)	0						
S43-L-JB	DNQ (2)	-						
Bl02-L-JB	DNQ (2)	-						

**Table S4.** Results obtained using AY-specific qRT-PCR on samples obtained from Jaliscolocation B.

## REFERENCES

- 1 Tamura, K., Nei, M. & Kumar, S. Prospects for inferring very large phylogenies by using the neighbor-joining method. *Proc. Natl. Acad. Sci. U.S.A.* **101**, 11030-11035, doi:10.1073/pnas.0404206101 (2004).
- 2 Dumonceaux, T. J., Green, M., Hammond, C., Perez, E. & Olivier, C. Molecular diagnostic tools for detection and differentiation of Phytoplasmas based on chaperonin-60 reveal differences in host plant infection patterns. *PLoS ONE* **9**, e116039, doi:10.1371/journal.pone.0116039 (2014).
- 3 Smart, C. D. *et al.* Phytoplasma-specific PCR primers based on sequences of the 16S-23S rRNA spacer region. *Appl. Environ. Microbiol.* **62**, 2988-2993 (1996).