SUPPLEMENTARY INFORMATION

Network-assisted analysis of GWAS data identifies a functionally-relevant gene module for childhood-onset asthma

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Supplementary Figure S1. Double Manhattan plot of gene-level *P***-values in META1 and META2.** Gene-level *P*-values were computed from SNP-level *P*-values using fastCGP. META1 and META2 correspond to the results of meta-analysis of 9 COA GWAS each. The GWAS are part of the GABRIEL asthma consortium (ALSPAC, BAMSE, ECRHS, MAS/MAGICS, SLSJ, TOMSK, UFA, CAPPS studies for META1; B58C, BUSSELTON, EGEA, GABRIEL Advanced Surveys, KSMU, MRCA-UKC, PIAMA, SAPALDIA, SAGE studies for META2; see Moffatt et al¹ for details on these studies)

Supplementary Figure S2. Distribution of pairwise module similarities before and after merging the raw modules generated by the Dense Module Search algorithm. The pairwise module similarities (indicating overlaps between modules) were remarkably reduced in both META1 (left panel) and META2 (right panel) after hierarchically merging similar raw modules

Supplementary Figure S3. Distribution of pairwise module similarities between modules in META1 and modules in META2. A total of 1,072,904 module pairs were constructed. The bins represent histograms of pairwise module similarities. The ticks represent rug plot of the similarities. Each tick represents one pairwise module similarity. The red ticks highlight the 10 highest pairwise module similarities

Supplementary Figure S4. Consistent gene modules between META1 and META2. We selected top 10 module pairs showing highest pairwise module similarities among all module pairs between META1 and META2. The involved modules were merged within each dataset, resulting in a subnetwork of 171 genes in META1 (a) and a subnetwork of 201 genes in META2 (b). The intersection of the two subnetworks was retrieved to construct the final module, resulting in a module of 91 genes (nodes in red). The node sizes in this plot are proportional to the gene z-scores

Supplementary Figure S5. An illustrative example of fastCGP. An artificial GWAS result consisting of L=100 SNP *P*-values were created. All *P*-values were set as 0.1 except $P_{11} = 0.05$, $P_{12} = 0.02$, $P_{14} = 0.04$, and $P_{18} = 0.07$. These *P*-values are ordered on a circle according to the chromosomal position of corresponding SNPs. A gene *g* has three SNPs mapped to its genomic region. Its uncorrected *P*-value P_g is set as the minimum SNP *P*-value among the three mapped SNPs ($P_g = P₁₈ = 0.07$). There are four extreme SNP *P*-values on the circle P_{11} , P_{12} , P_{14} and P_{18} ($\leq P_g$). The consecutive extreme *P*-value pairs are $P_{11} \sim P_{12}$, $P_{12} \sim P_{14}$, $P_{14} \sim P_{18}$, $P_{18} \sim P_{11}$

Supplementary Figure S6. Comparison of gene-level *P***-values obtained by fastCGP and VEGAS2**² **.** Gene-level *P*-values were computed from asthma META2 dataset using fastCGP and VEGAS2 (*-bestsnp* sub-model). Their results (-log10 (*P*-value)) were compared for each chromosome from chromosome 1 (Chr1) to chromosome 22 (Chr22). The red diagonal lines indicate perfect match (identical) of the two results. ρ represents the Pearson correlation coefficient between the two results

Supplementary Figure S7. Comparison of gene-level *P***-values obtained by fastCGP and MAGMA**³ **.** Gene-level *P*-values were computed from asthma META2 dataset using fastCGP and MAGMA (*-snpwise=top,1* sub-model). Their results (-log10 (*P*-value)) were compared for each chromosome from chromosome 1 (Chr1) to chromosome 22 (Chr22). The red diagonal lines indicate perfect match (identical) of the two results. ρ represents the Pearson correlation coefficient between the two results

Supplementary Table S1. Gene-level *P***-values in META1 and META2 datasets for the 91 genes in the final childhood-onset asthma module**

Start and end positions of each gene are in accordance with Build 37.1.

The genes at known asthma loci are in bold.

Supplementary Methods 1: computing gene-level *P***-values via fastCGP.** We take advantage of the Circular Genomic Permutation (CGP) strategy⁴ and propose an efficient and exact method, named fastCGP, to compute gene-level *P-*values from SNP-level *P-*values of a GWAS. CGP is a randomization method that permutes SNP-level statistics in a genomic manner to preserve the genomic structure such as regional linkage disequilibrium (LD), thereby to keep similar patterns of correlation in the permutated data as in the original data. Briefly, it considers the genome to be circular and ordered from chromosome 1 to chromosome 22. SNP-level *P*values of a GWAS are ordered according to the position of the SNPs on the circle. A CGP sample is generated by rotating the ordered statistics for a random position and reassigning them to each SNP. This randomization strategy has been successfully applied to several studies⁵⁻⁷, and was shown to have similar performances compared to the gold standard of phenotype permutation in the context of pathway analysis⁸.

Our method starts by mapping SNPs to genes (between the start site and 3'-untranslated region of each gene) using dbSNP Build 132 and human Genome Build 37.1 (a user can choose another mapping strategy). The gene-level *P*-value of a gene g , denoted as P_g , is represented by the best SNP *P-*value among all SNPs mapped to the gene. This *P-*value is biased by gene length (amount of mapped SNPs) as genes with more SNPs mapped tend to have a lower best SNP *P*value by chance. We correct for such bias using a permutation test framework. We define the corrected *P*-value as $P_{\text{corrected}} = 1 - l / (L + 1)$, where *L* is the total number of CGP samples; *l* is the number of samples with $P_{\pi,g} > P_g$, which we call as *normal CGP samples* (nCGP). Particularly, we include all non-repeating CGP samples so that to obtain the best obtainable *P*value within this permutation test framework. In this case, *L* becomes the total amount of SNPs placed on the circle (hence is the number of SNPs in a GWAS), while *l* can be calculated analytically without generating any CGP sample. For illustration convenience, we call a SNP *P*value as an extreme *P*-values if it is less than or equal to P_{g} . We say two extreme *P*-values are consecutive if there is no other extreme *P-*value placed between them on the circle. Note that the SNPs mapped to a gene are consecutive on the circle, hence by rotating the SNP *P-*values, it generates a nCGP only if all *P-*values reassigned to gene *g* are located between some pair of consecutive extreme *P*-values, say, $P_i \sim P_j$ ($1 \le i, j \le L$ are the SNP positions on the circle). Denote d_{ij} as the number of positions between P_i and P_j on the circle, m as the number of SNPs assigned to the *gene*, I_{α} as the indicator function. Then the number of unique rotations with all reassigned *P*-values located within $P_i \sim P_j$ is equal to $\gamma_{ij} = (d_{ij} - m + 1)I_{(d_i \ge m)}$. Since the total amount of non-repeating nCGPs is the summation of γ_{ij} for all pairs of consecutive extreme *P*-

values, this leads to the formula $P_{\text{corrected}} = 1 - \sum_{i \sim j} \gamma_{ij} / (L+1)$. The complete algorithm is summarized below

Algorithm Computation of corrected gene-level *P*-values via fastCGP

- **Step 1.** Order GWAS SNP *P-*values on a circle according to the genomic positions of SNPs
- **Step 2.** Map SNPs to genes according to their genomic positions
- **Step 3.** For a gene g , set P_g as the minimum *P*-value of all SNPs mapped to it
- **Step 4.** Find all extreme SNP *P*-values on the circle: $\{P_{SNP} | P_{SNP} \leq P_{\rho}\}\$
- **Step 5.** Compute $\gamma_{ii} = (d_{ii} m + 1)I_{(d_{ii} \ge m)}$ for all pairs of consecutive extreme *P*-value $P_i \sim P_j$
- **Step 6.** Compute the corrected gene-level *P*-value: $P_{\text{corrected}} = 1 \sum_{i \sim j} \gamma_{ij} / (L+1)$

In the following we present an illustrative example of fastCGP. We constructed an artificial GWAS result consisting of *L* = 100 SNP *P-*values (Supplementary Fig. S5). We set all *P-*values as 0.1 except $P_{11} = 0.05, P_{12} = 0.02, P_{14} = 0.04$, and $P_{18} = 0.07$. These *P*-values are ordered on a circle according to the chromosomal position of corresponding SNPs. A gene *g* has *m* = 3 SNPs mapped to its genomic region. Its uncorrected *P*-value P_g is set as the minimum SNP *P*-value among the three mapped SNPs ($P_q = P_{18} = 0.07$). There are four extreme SNP *P*-values on the circle : P_{11} , P_{12} , P_{14} and P_{18} . The consecutive extreme *P*-value pairs are $P_{11} \sim P_{12}$, $P_{12} \sim P_{14}$, $P_{14} \sim P_{18}$, $P_{18} \sim P_{11}$. For each pair, the amount of unique rotations with all SNP *P-*values reassigned to gene *g* falling into this pair is 0, 0, 1 and 92 respectively. Thereby $P_{corrected} = 1 - (0 + 0 + 1 + 92)/101 = 0.079.$

We compared the performance of fastCGP with two other popular methods $VEGAS2²$ and $MAGMA³$. The META2 asthma dataset in our study was used for the comparison. For both VEGAS2 and MAGMA we implemented their best-SNP sub-model (use -*bestsnp* option for VEGAS2 and *snp-wise=top,1* option for MAGMA). Both methods apply Monte-Carlo simulations to correct the best-SNP *P*-value for the gene length bias. During simulation, the LD patterns between SNPs within a gene are estimated on the basis of the LD structure of a set of reference individuals. As we could not use the original genotypes of our asthma dataset, we used the 1,000 Genomes European population as external reference. We observed that the results obtained by fastCGP were concordant with those obtained using VEGAS2 or MAGMA. At the chromosome level, the Pearson correlation coefficients between the gene-level *P*-values (-log10 transformed) of fastCGP and VEGAS2 range from 0.93 to 0.97, with an average value of 0.96 (Supplementary Fig. S6). The correlation coefficients of gene-level *P*-values between fastCGP and MAGMA range from 0.95 to 0.98, with an average value of 0.97 (Supplementary Fig. S7). As

for computational efficiency, both fastCGP and MAGMA took ~30 minutes on a PC (Intel Core i7 3.40GHz CPU, 8GB RAM); while VEGAS2 took 13 hours to perform the analysis.

Supplementary Methods 2: generating random modules via Metropolis-Hasting Random Walk algorithm. We inherited the Metropolis-Hasting Random Walk (MHRW) algorithm⁹ to generate random modules. It has the property that the stationary probability of each node to be sampled follows the uniform distribution, and has been demonstrated to work well in practice⁹. In the beginning, a seed gene V is chosen from the scored-PPI. The next gene W is selected at random from all neighbours of *V* . *W* is added to the module and set as the next seed if the degree ratio of V and W is smaller than a random number drawn from uniform distribution $U(0,1)$. Otherwise stay at *V* and repeat the step. The procedure iterates until the module has the same number of genes as the module under test. To ensure sufficient coverage of the whole scored-PPI, we set each gene in the network as a seed to generate a module. Then, a total of *N* (equals to the number of genes in the scored-PPI) random modules are generated.

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