SUPPLEMENTARY INFORMATION

Network-assisted analysis of GWAS data identifies a functionally-relevant gene module for childhood-onset asthma

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Supplementary Figure S1. Double Manhattan plot of gene-level *P***-values in META1 and META2.** Gene-level *P*-values were computed from SNP-level *P*-values using fastCGP. META1 and META2 correspond to the results of meta-analysis of 9 COA GWAS each. The GWAS are part of the GABRIEL asthma consortium (ALSPAC, BAMSE, ECRHS, MAS/MAGICS, SLSJ, TOMSK, UFA, CAPPS studies for META1; B58C, BUSSELTON, EGEA, GABRIEL Advanced Surveys, KSMU, MRCA-UKC, PIAMA, SAPALDIA, SAGE studies for META2; see Moffatt et al¹ for details on these studies)



Supplementary Figure S2. Distribution of pairwise module similarities before and after merging the raw modules generated by the Dense Module Search algorithm. The pairwise module similarities (indicating overlaps between modules) were remarkably reduced in both META1 (left panel) and META2 (right panel) after hierarchically merging similar raw modules



Supplementary Figure S3. Distribution of pairwise module similarities between modules in META1 and modules in META2. A total of 1,072,904 module pairs were constructed. The bins represent histograms of pairwise module similarities. The ticks represent rug plot of the similarities. Each tick represents one pairwise module similarity. The red ticks highlight the 10 highest pairwise module similarities



Supplementary Figure S4. Consistent gene modules between META1 and META2. We selected top 10 module pairs showing highest pairwise module similarities among all module pairs between META1 and META2. The involved modules were merged within each dataset, resulting in a subnetwork of 171 genes in META1 (a) and a subnetwork of 201 genes in META2 (b). The intersection of the two subnetworks was retrieved to construct the final module, resulting in a module of 91 genes (nodes in red). The node sizes in this plot are proportional to the gene z-scores



Supplementary Figure S5. An illustrative example of fastCGP. An artificial GWAS result consisting of L=100 SNP *P*-values were created. All *P*-values were set as 0.1 except $P_{11} = 0.05$, $P_{12} = 0.02$, $P_{14} = 0.04$, and $P_{18} = 0.07$. These *P*-values are ordered on a circle according to the chromosomal position of corresponding SNPs. A gene *g* has three SNPs mapped to its genomic region. Its uncorrected *P*-value P_g is set as the minimum SNP *P*-value among the three mapped SNPs ($P_g = P_{18} = 0.07$). There are four extreme SNP *P*-values on the circle P_{11} , P_{12} , P_{14} and P_{18} ($\leq P_g$). The consecutive extreme *P*-value pairs are $P_{11} \sim P_{12}$, $P_{12} \sim P_{14}$, $P_{18} \sim P_{11}$



Supplementary Figure S6. Comparison of gene-level *P*-values obtained by fastCGP and VEGAS2². Gene-level *P*-values were computed from asthma META2 dataset using fastCGP and VEGAS2 (*-bestsnp* sub-model). Their results (-log10 (*P*-value)) were compared for each chromosome from chromosome 1 (Chr1) to chromosome 22 (Chr22). The red diagonal lines indicate perfect match (identical) of the two results. ρ represents the Pearson correlation coefficient between the two results



Supplementary Figure S7. Comparison of gene-level *P*-values obtained by fastCGP and MAGMA³. Gene-level *P*-values were computed from asthma META2 dataset using fastCGP and MAGMA (*-snp-wise=top,1* sub-model). Their results (-log10 (*P*-value)) were compared for each chromosome from chromosome 1 (Chr1) to chromosome 22 (Chr22). The red diagonal lines indicate perfect match (identical) of the two results. ρ represents the Pearson correlation coefficient between the two results



Supplementary Table S1. Gene-level *P*-values in META1 and META2 datasets for the 91 genes in the final childhood-onset asthma module

		Start	End	# SNPs		META1		META2			
Chr	Gene				Best SNP	Best SNP <i>P</i> -value	Corrected Best SNP <i>P</i> -value (by fastCGP)	Best SNP	Best SNP <i>P</i> -value	Corrected Best SNP <i>P</i> -value (by fastCGP)	
Gene	Genes nominally significant in both META1 and META2										
2	IL1RL1	102 927 962	102 968 497	80	rs4988957	2.9E-05	1.5E-03	rs10192157	2.2E-05	5.6E-04	
2	IL18R1	102 979 097	103 015 218	48	rs3771166	1.9E-05	8.2E-04	rs1974675	2.4E-05	4.1E-04	
2	IL18RAP	103 035 254	103 069 025	71	rs6543135	2.6E-03	4.5E-02	rs2310300	2.6E-05	6.9E-04	
4	CRMP1	5 822 491	5 894 785	60	rs13144677	2.7E-03	4.1E-02	rs10011385	1.3E-03	1.7E-02	
5	RAD50	131 892 630	131 979 599	35	rs2240032	1.2E-03	1.5E-02	rs2897443	3.8E-04	3.9E-03	
6	ZNF192	28 109 716	28 125 236	21	rs13205911	6.4E-05	1.2E-03	rs2622321	7.4E-03	4.1E-02	
6	RAETIE	150 209 601	150 212 097	10	rs9371533	1.1E-02	4.5E-02	rs9371533	1.0E-02	3.6E-02	
9	IL33	6 241 678	6 257 982	20	rs7019575	4.8E-03	3.2E-02	rs7019575	8.7E-05	6.7E-04	
9	CTSL1	90 340 974	90 346 384	4	rs2378757	1.8E-03	5.6E-03	rs2378757	6.9E-03	1.4E-02	
12	C12orf43	121 440 848	121 454 300	10	rs3751150	4.4E-04	3.1E-03	rs3751151	1.2E-02	4.3E-02	
17	PNMT	37 824 507	37 826 728	1	rs876493	1.3E-07	3.1E-05	rs876493	4.0E-05	7.3E-05	
17	ERBB2	37 844 393	37 884 915	7	rs1058808	3.5E-06	1.6E-04	rs1058808	1.6E-05	8.2E-05	
17	IKZF3	37 921 198	38 020 441	48	rs907091	2.5E-15	4.6E-05	rs9909593	1.6E-07	7.1E-05	
17	GSDMB	38 060 848	38 074 903	12	rs9303281	2.6E-16	5.5E-06	rs2305480	7.5E-08	1.9E-05	
17	ORMDL3	38 077 296	38 083 854	4	rs12603332	4.6E-15	9.7E-06	rs8076131	7.3E-08	7.2E-06	
17	GSDMA	38 119 226	38 134 019	6	rs7212938	2.4E-13	2.7E-05	rs3902025	3.2E-07	3.3E-05	
17	PSMD3	38 137 060	38 154 212	21	rs11655264	1.0E-07	9.6E-05	rs12453334	6.9E-05	5.8E-04	
17	MED24	38 175 350	38 210 889	21	rs12309	3.0E-06	2.5E-04	rs12451897	1.2E-04	9.2E-04	
19	JAK3	17 935 591	17 958 841	12	rs2110586	5.8E-03	2.8E-02	rs3212701	1.2E-02	4.5E-02	
Gene	s nominally s	ignificant in e	ither META1	or MET	42						
2	MARCH4	217 122 585	217 236 750	113	rs1477235	3.0E-02	4.0E-01	rs1510836	1.3E-03	2.8E-02	
3	CCRL1	132 316 094	132 321 382	2	rs7626622	3.0E-04	8.5E-04	rs7626622	1.7E-01	2.2E-01	
6	BTN3A3	26 440 763	26 453 643	15	rs13220495	1.8E-04	1.9E-03	rs3846845	1.3E-01	4.3E-01	
6	ZNF165	28 046 572	28 057 341	9	rs1321505	3.7E-03	1.6E-02	rs203878	2.8E-02	8.8E-02	
6	MICA	31 371 371	31 383 090	41	rs2844518	3.1E-08	1.0E-04	rs12213831	6.4E-02	4.0E-01	
6	MICB	31 465 855	31 478 901	26	rs3130614	2.9E-05	7.6E-04	rs2855814	3.5E-01	9.0E-01	
6	BAI3	69 345 632	70 099 403	770	rs3757043	4.6E-05	1.3E-02	rs17502590	3.7E-02	9.5E-01	
6	ULBP1	150 285 143	150 294 846	6	rs9478311	1.2E-01	2.8E-01	rs9478311	1.2E-03	3.2E-03	
9	CCL19	34 689 567	34 691 274	2	rs3176813	4.1E-02	5.5E-02	rs3136658	2.6E-02	3.3E-02	
9	SHC3	91 628 046	91 793 682	134	rs1331180	2.0E-05	1.6E-03	rs2316280	3.6E-02	4.9E-01	

10	CNNM2	104 678 114	104 838 241	129	rs943036	2.3E-05	1.8E-03	rs2296569	6.5E-02	6.7E-01	
12	STAT6	57 489 191	57 505 161	10	rs324015	8.7E-04	5.3E-03	rs1059513	2.0E-02	6.8E-02	
17	CCL8	32 646 066	32 648 421	2	rs3138036	9.2E-01	9.7E-01	rs3138036	2.2E-02	2.8E-02	
18	ZNF24	32 912 178	32 924 426	12	rs7239712	4.2E-03	2.2E-02	rs1064753	1.9E-02	7.2E-02	
19	JSRP1	2 252 252	2 255 344	3	rs7250822	1.1E-02	2.2E-02	rs7250822	2.0E-01	3.2E-01	
19	FCER2	7 753 643	7 767 032	22	rs2287866	1.4E-03	1.3E-02	rs2303112	1.4E-02	7.8E-02	
Othe	Other genes										
1	FBXO6	11 724 150	11 734 411	2	rs747863	6.1E-01	7.5E-01	rs747863	6.1E-02	7.9E-02	
1	CD48	160 648 536	160 681 585	14	rs3796502	1.6E-02	7.2E-02	rs10489636	8.2E-02	2.9E-01	
1	CD247	167 399 877	167 487 847	116	rs864537	2.7E-03	6.7E-02	rs864537	3.0E-03	6.0E-02	
1	CR2	207 627 670	207 663 240	41	rs17258982	4.6E-02	3.2E-01	rs7543913	9.1E-02	5.1E-01	
2	CEBPZ	37 428 772	37 458 740	12	rs2239650	3.4E-02	1.3E-01	rs11689186	1.0E-01	3.3E-01	
2	IL1RL2	102 803 433	102 855 811	134	rs17026782	6.6E-02	6.9E-01	rs12987222	1.4E-02	2.4E-01	
2	DNER	230 222 345	230 579 286	353	rs10192168	1.6E-03	1.1E-01	rs6726280	3.2E-03	1.6E-01	
3	CCR8	39 371 197	39 375 171	2	rs2853699	5.4E-02	7.2E-02	rs4676633	2.3E-01	3.0E-01	
3	CCBP2	42 850 964	42 908 775	36	rs13093968	8.3E-03	7.4E-02	rs4396867	4.4E-01	9.7E-01	
3	CCR9	45 928 019	45 944 667	12	rs17714101	4.5E-02	1.6E-01	rs6441931	2.6E-01	6.7E-01	
3	RYK	133 875 978	133 969 586	52	rs4280635	8.4E-02	5.4E-01	rs10935104	7.5E-02	4.9E-01	
3	SEC62	169 684 580	169 716 161	23	rs9813592	1.8E-02	1.1E-01	rs16854694	3.2E-01	8.5E-01	
4	LNX1	54 326 437	54 457 724	156	rs2117600	2.6E-02	4.4E-01	rs9312642	1.5E-02	2.9E-01	
4	LPHN3	62 362 839	62 938 168	336	rs17082520	1.3E-02	4.6E-01	rs1497906	8.9E-03	3.4E-01	
4	CXCL13	78 432 907	78 532 988	27	rs17406477	2.1E-01	7.3E-01	rs355687	2.8E-02	1.6E-01	
5	PCDHGA11	140 800 537	140 892 546	57	rs11958830	4.5E-02	3.7E-01	rs1423149	1.8E-01	8.1E-01	
6	RAETIG	150 238 014	150 244 214	4	rs6927913	2.3E-01	4.2E-01	rs9397070	5.0E-02	9.7E-02	
6	ULBP3	150 385 743	150 390 202	7	rs12202737	3.0E-01	6.3E-01	rs2010212	2.7E-02	7.3E-02	
6	PARK2	161 768 590	163 148 834	1776	rs4623220	1.6E-03	3.9E-01	rs11966738	1.9E-03	4.0E-01	
7	TPST1	65 670 259	65 825 438	68	rs778732	3.8E-01	9.8E-01	rs4149463	1.9E-01	8.6E-01	
8	TERF1	73 921 097	73 959 987	26	rs12334686	2.4E-01	7.7E-01	rs10107605	5.8E-01	9.9E-01	
9	CCL27	34 661 893	34 662 689	1	rs11575584	6.3E-01	6.1E-01	rs11575584	3.4E-01	3.1E-01	
9	SPTLC1	94 793 427	94 877 690	74	rs16908106	9.5E-02	6.6E-01	rs12235495	4.1E-03	5.8E-02	
10	PCDH21	85 954 517	85 977 122	34	rs12781048	2.1E-01	7.6E-01	rs12781048	4.1E-02	2.5E-01	
11	DCHS1	6 642 558	6 677 080	28	rs11607376	7.1E-02	3.6E-01	rs997263	2.0E-01	7.1E-01	
11	NCR3LG1	17 373 279	17 398 868	13	rs6486364	4.9E-02	1.9E-01	rs12791318	1.1E-01	3.7E-01	
11	IL18	112 013 976	112 034 840	17	rs1834481	1.0E-01	3.9E-01	rs2043055	1.5E-01	5.1E-01	
11	CXCR5	118 754 541	118 766 971	14	rs12363277	1.0E-01	3.6E-01	rs12363277	5.1E-02	1.9E-01	
12	KLRK1	10 524 952	10 542 640	20	rs2617149	1.5E-02	8.2E-02	rs12826560	9.0E-02	3.7E-01	

12	KLRC4	10 559 983	10 562 356	5	rs2734565	1.4E-01	2.9E-01	rs2617170	4.2E-01	7.2E-01
12	KIF21A	39 687 030	39 836 918	71	rs11171691	2.4E-02	2.6E-01	rs11172108	4.9E-02	4.3E-01
12	BIN2	51 674 822	51 717 938	20	rs4761998	6.4E-02	2.9E-01	rs4761995	5.9E-02	2.6E-01
12	IFNG	68 548 550	68 553 521	4	rs1861494	6.6E-01	8.9E-01	rs1861493	3.4E-01	5.8E-01
12	CMKLR1	108 681 821	108 733 094	48	rs11113818	8.2E-02	5.1E-01	rs10861889	5.1E-02	3.6E-01
13	FREM2	39 261 173	39 461 268	220	rs11618650	4.1E-03	1.5E-01	rs2218722	1.3E-02	3.3E-01
13	PCDH20	61 983 818	61 989 655	6	rs3829388	1.8E-01	3.9E-01	rs3812872	3.0E-01	6.1E-01
15	CD276	73 976 622	74 006 859	35	rs12591553	3.1E-01	8.9E-01	rs12594595	9.0E-02	4.7E-01
16	ZNF434	3 432 085	3 451 025	14	rs28603	8.1E-02	2.9E-01	rs17136367	2.3E-01	6.4E-01
16	ZNF174	3 451 190	3 459 364	5	rs39728	3.7E-01	6.6E-01	rs37811	2.1E-01	4.3E-01
16	ITGAX	31 366 509	31 394 318	16	rs2929	1.9E-01	6.0E-01	rs8052139	4.7E-02	1.9E-01
17	CCL13	32 683 471	32 685 629	2	rs159313	7.2E-01	8.5E-01	rs2072069	8.5E-01	9.4E-01
17	CCLI	32 687 399	32 690 252	5	rs3136682	4.4E-02	1.0E-01	rs3136682	2.3E-01	4.5E-01
17	CCL16	34 303 535	34 308 523	4	rs2063979	8.2E-01	9.7E-01	rs11080369	3.4E-02	6.6E-02
17	CCL4	34 431 220	34 433 014	2	rs1719147	4.4E-02	6.0E-02	rs1634517	5.0E-01	6.3E-01
17	ZNHIT3	34 842 473	34 851 662	4	rs2306589	8.1E-02	1.6E-01	rs2277662	2.4E-01	4.3E-01
18	SERPINB3	61 322 431	61 329 197	3	rs1065205	4.5E-01	6.7E-01	rs7228687	2.0E-01	3.3E-01
19	ELAVLI	8 023 457	8 070 529	24	rs7251814	3.9E-01	9.1E-01	rs12977189	8.3E-02	3.8E-01
19	CCL25	8 117 934	8 127 547	8	rs2287936	3.9E-01	7.7E-01	rs2032887	3.5E-01	7.2E-01
19	C19orf66	10 196 806	10 203 928	2	rs2232066	3.9E-02	5.3E-02	rs2232066	9.5E-02	1.2E-01
19	CDKN2D	10 677 138	10 679 655	1	rs1465701	6.3E-01	6.1E-01	rs1465701	2.7E-01	2.4E-01
19	ZNF20	12 242 803	12 251 140	2	rs155955	2.1E-01	2.8E-01	rs12608894	1.9E-01	2.5E-01
19	RCN3	50 030 875	50 046 890	5	rs10419198	3.3E-01	6.1E-01	rs8108243	1.9E-01	3.9E-01
20	SIRPD	1 514 897	1 538 343	26	rs16995146	5.1E-02	2.7E-01	rs2249673	2.3E-01	7.5E-01
21	APP	27 252 861	27 543 446	306	rs7281055	8.2E-03	3.2E-01	rs2830076	3.1E-02	6.8E-01
21	COL18A1	46 825 097	46 933 634	82	rs2236483	1.3E-01	7.8E-01	rs17004785	2.0E-01	9.1E-01
22	TPST2	26 921 714	26 986 089	94	rs5752349	3.4E-02	4.0E-01	rs4149484	4.7E-02	4.8E-01

Start and end positions of each gene are in accordance with Build 37.1.

The genes at known asthma loci are in bold.

Supplementary Methods 1: computing gene-level *P***-values via fastCGP.** We take advantage of the Circular Genomic Permutation (CGP) strategy⁴ and propose an efficient and exact method, named fastCGP, to compute gene-level *P*-values from SNP-level *P*-values of a GWAS. CGP is a randomization method that permutes SNP-level statistics in a genomic manner to preserve the genomic structure such as regional linkage disequilibrium (LD), thereby to keep similar patterns of correlation in the permutated data as in the original data. Briefly, it considers the genome to be circular and ordered from chromosome 1 to chromosome 22. SNP-level *P*-values of a GWAS are ordered according to the position of the SNPs on the circle. A CGP sample is generated by rotating the ordered statistics for a random position and reassigning them to each SNP. This randomization strategy has been successfully applied to several studies^{5–7}, and was shown to have similar performances compared to the gold standard of phenotype permutation in the context of pathway analysis⁸.

Our method starts by mapping SNPs to genes (between the start site and 3'-untranslated region of each gene) using dbSNP Build 132 and human Genome Build 37.1 (a user can choose another mapping strategy). The gene-level P-value of a gene g , denoted as P_{g} , is represented by the best SNP P-value among all SNPs mapped to the gene. This P-value is biased by gene length (amount of mapped SNPs) as genes with more SNPs mapped tend to have a lower best SNP Pvalue by chance. We correct for such bias using a permutation test framework. We define the corrected *P*-value as $P_{\text{corrected}} = 1 - l / (L + 1)$, where *L* is the total number of CGP samples; *l* is the number of samples with $P_{\pi,g} > P_g$, which we call as normal CGP samples (nCGP). Particularly, we include all non-repeating CGP samples so that to obtain the best obtainable Pvalue within this permutation test framework. In this case, L becomes the total amount of SNPs placed on the circle (hence is the number of SNPs in a GWAS), while l can be calculated analytically without generating any CGP sample. For illustration convenience, we call a SNP Pvalue as an extreme P-values if it is less than or equal to P_g . We say two extreme P-values are consecutive if there is no other extreme P-value placed between them on the circle. Note that the SNPs mapped to a gene are consecutive on the circle, hence by rotating the SNP P-values, it generates a nCGP only if all P-values reassigned to gene g are located between some pair of consecutive extreme P-values, say, $P_i \sim P_j$ ($1 \le i, j \le L$ are the SNP positions on the circle). Denote d_{ij} as the number of positions between P_i and P_j on the circle, *m* as the number of SNPs assigned to the gene, $I_{(\cdot)}$ as the indicator function. Then the number of unique rotations with all reassigned *P*-values located within $P_i \sim P_j$ is equal to $\gamma_{ij} = (d_{ij} - m + 1)I_{(d_{ij} \ge m)}$. Since the total amount of non-repeating nCGPs is the summation of γ_{ii} for all pairs of consecutive extreme P-

values, this leads to the formula $P_{\text{corrected}} = 1 - \sum_{i \sim j} \gamma_{ij} / (L+1)$. The complete algorithm is summarized below

Algorithm Computation of corrected gene-level P-values via fastCGP

Step 1.	Order GWAS S	SNP P-values	on a circle ac	cording to the	genomic	positions	of SNPs
				1.7			

- Step 2. Map SNPs to genes according to their genomic positions
- **Step 3.** For a gene g, set P_g as the minimum *P*-value of all SNPs mapped to it
- **Step 4.** Find all extreme SNP *P*-values on the circle: $\{P_{SNP} | P_{SNP} \leq P_g\}$
- **Step 5.** Compute $\gamma_{ij} = (d_{ij} m + 1)I_{(d_{ij} \ge m)}$ for all pairs of consecutive extreme *P*-value $P_i \sim P_j$
- **Step 6.** Compute the corrected gene-level *P*-value: $P_{\text{corrected}} = 1 \sum_{i \sim j} \gamma_{ij} / (L+1)$

In the following we present an illustrative example of fastCGP. We constructed an artificial GWAS result consisting of L = 100 SNP *P*-values (Supplementary Fig. S5). We set all *P*-values as 0.1 except $P_{11} = 0.05$, $P_{12} = 0.02$, $P_{14} = 0.04$, and $P_{18} = 0.07$. These *P*-values are ordered on a circle according to the chromosomal position of corresponding SNPs. A gene *g* has m = 3 SNPs mapped to its genomic region. Its uncorrected *P*-value P_g is set as the minimum SNP *P*-value among the three mapped SNPs ($P_g = P_{18} = 0.07$). There are four extreme SNP *P*-values on the circle : P_{11}, P_{12}, P_{14} and P_{18} . The consecutive extreme *P*-value pairs are $P_{11} \sim P_{12}, P_{12} \sim P_{14}, P_{14} \sim P_{18}, P_{18} \sim P_{11}$. For each pair, the amount of unique rotations with all SNP *P*-values reassigned to gene *g* falling into this pair is 0, 0, 1 and 92 respectively. Thereby $P_{corrected} = 1 - (0 + 0 + 1 + 92)/101 = 0.079$.

We compared the performance of fastCGP with two other popular methods VEGAS2² and MAGMA³. The META2 asthma dataset in our study was used for the comparison. For both VEGAS2 and MAGMA we implemented their best-SNP sub-model (use *-bestsnp* option for VEGAS2 and *snp-wise=top,1* option for MAGMA). Both methods apply Monte-Carlo simulations to correct the best-SNP *P*-value for the gene length bias. During simulation, the LD patterns between SNPs within a gene are estimated on the basis of the LD structure of a set of reference individuals. As we could not use the original genotypes of our asthma dataset, we used the 1,000 Genomes European population as external reference. We observed that the results obtained by fastCGP were concordant with those obtained using VEGAS2 or MAGMA. At the chromosome level, the Pearson correlation coefficients between the gene-level *P*-values (-log10 transformed) of fastCGP and VEGAS2 range from 0.93 to 0.97, with an average value of 0.96 (Supplementary Fig. S6). The correlation coefficients of gene-level *P*-values between fastCGP and MAGMA range from 0.95 to 0.98, with an average value of 0.97 (Supplementary Fig. S7). As

for computational efficiency, both fastCGP and MAGMA took ~30 minutes on a PC (Intel Core i7 3.40GHz CPU, 8GB RAM); while VEGAS2 took 13 hours to perform the analysis.

Supplementary Methods 2: generating random modules via Metropolis-Hasting Random Walk algorithm. We inherited the Metropolis-Hasting Random Walk (MHRW) algorithm⁹ to generate random modules. It has the property that the stationary probability of each node to be sampled follows the uniform distribution, and has been demonstrated to work well in practice⁹. In the beginning, a seed gene V is chosen from the scored-PPI. The next gene W is selected at random from all neighbours of V. W is added to the module and set as the next seed if the degree ratio of V and W is smaller than a random number drawn from uniform distribution U(0,1). Otherwise stay at V and repeat the step. The procedure iterates until the module has the same number of genes as the module under test. To ensure sufficient coverage of the whole scored-PPI, we set each gene in the network as a seed to generate a module. Then, a total of N (equals to the number of genes in the scored-PPI) random modules are generated.

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