Construction of a highly efficient CRISPR/Cas9-mediated duck enteritis virus-based vaccine against H5N1 avian influenza virus and duck Tembusu virus infection

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Supplementary materials

Supplementary Figure 1. Verification of HA and PrM-E insertion in C-KCE by

PCR.



Supplementary Figure 2. Detection of HA, PrM, and E proteins expressions in C-KCE-HA/PrM-E-infected CEF cells by western blot. The precursor PrM-E, PrM, and E indicated by the red arrowhead. A and B are the different exposures.



Supplementary Figure 3. Viral load in organs infected with C-KCE-HA/PrM-E or C-KCE. Ducks were vaccinated with 10<sup>5</sup> PFU of C-KCE or C-KCE-HA/PrM-E (n = 3 per group) subcutaneously. All the ducks in the two groups were humanely euthanized on day 2 post-challenge (pc), and their organs were obtained to determine virus titers using a one-step real-time TaqMan RT-PCR assay.



Supplementary Figure 4. Humoral immune response against virulent DEV in ducks vaccinated with C-KCE-HA/PrM-E or its parental strain C-KCE. Groups of 5 ducks were inoculated subcutaneously with  $10^5$  PFU of C-KCE-HA/PrM-E, C-KCE or with PBS as control. Sera were collected from 0 to 4 weeks to detect NT antibodies against virulent DEV in DEF cells. NT antibody titers for ducks are shown in log2 scale. Dotted lines indicate the thresholds for a positive response. Data are shown as the means  $\pm$  SD. \*\* P < 0.01, \*\*\* P < 0.001.



Supplementary Figure 5. HA responses were assessed against the homologous XN/07 virus. Ducks were vaccinated with PBS,  $10^5$  PFU of C-KCE or C-KCE-HA/PrM-E (n = 10 per group) subcutaneously. Sera were obtained at the indicated time points to detect HA antibodies against XN/07. Dotted lines indicate the thresholds for a positive response.



Challenge	vaccine	Virus replication in the organs in the ducks on 3 days pv (mean $\pm$ SD, lg10EID <sub>50</sub> /g)						
time pv	_	Heart	Liver	Spleen	Lung	Kidney	Brain	
2 weeks	C-KCE-HA-PrM-E	/	/	/	/	/	/	
	C-KCE	$6.3 \pm 1.1$	7.1±0.5	6.8±0.7	5.8±1.1	6.5±0.9	7.2±0.6	
	PBS	$6.7 \pm 0.8$	6.4±0.6	7.0±0.5	6.8±1.4	6.2±0.8	7.5±0.8	
4 weeks	C-KCE-HA-PrM-E	/	/	/	/	/	/	
	C-KCE	6.4±0.7	5.6±1.3	$6.3 \pm 1.0$	5.4±0.9	7.6±0.4	7.3±0.4	
	PBS	6.2±0.5	7.0±0.4	6.8±0.7	6.6±1.2	7.1±0.5	6.8±0.7	

Subplementary rapie 1. Replication of chantenge virus Alv/07 in uu	<b>Supplementary</b>	Table 1. R	eplication of	challenge virus	XN/07 in duck
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Homologous XN/07 replication in the organs of ducks that were vaccinated with C-KCE-HA/PrM-E. Groups of three ducks were inoculated

subcutaneously with  $10^5$  plague-forming units (PFU) of C-KCE-HA, C-KCE, or PBS as a control. Subsequently, the ducks were challenged with homologous (XN/07) intramuscularly at 2 weeks or 4 weeks pv. At day 3 after challenge, the ducks were humanely sacrificed and their organs were collected for virus titration in eggs. Data represent mean titers  $\pm$  standard errors. The backslash indicates that the challenge virus was not recovered at the corresponding time point.

Challenge	vaccine	Viral copy load in the organs in the ducks on 3 days pv (mean $\pm$ SD, log <sub>10</sub> per µg of total RNA)						
time pv		Heart	Liver	Spleen	Lung	Kidney	Brain	
2 weeks	C-KCE-HA-PrM-E	/	/	/	/	/	/	
	C-KCE	$5.91 \pm 0.54$	$5.73 \pm 0.28$	$6.18 \pm 0.42$	$4.29 \pm 0.47$	$5.16 \pm 0.33$	$2.69\pm0.43$	
	PBS	$6.13\pm0.22$	$5.42 \pm 0.37$	$5.78 \pm 0.33$	$5.14 \pm 0.26$	$5.46 \pm 0.26$	$3.17 \pm 0.21$	
4 weeks	C-KCE-HA-PrM-E	/	/	/	/	/	/	
	C-KCE	$5.27\pm0.34$	$4.82 \pm 0.33$	$5.16 \pm 0.27$	$4.52 \pm 0.26$	$4.27 \pm 0.26$	$2.26\pm0.52$	
	PBS	$4.66\pm0.29$	$4.43 \pm 0.47$	$5.45 \pm 0.18$	$4.19 \pm 0.33$	$4.66 \pm 0.42$	$2.53\pm0.33$	

Supplementary Table 2. Replication of challenge virus df2 in ducks.

DTMUV replication in the organs of the ducks vaccinated with C-KCE-HA/PrM-E. Groups of three ducks were inoculated subcutaneously with

10<sup>5</sup> PFU of C-KCE-HA/PrM-E, C-KCE, or with PBS as the control. Subsequently, the ducks were challenged with df2 intramuscularly at

2 weeks or 4 weeks pv. Ducks were euthanized on day 3 post-challenge, and their organs were harvested for virus titration utilizing the

quantitative RT-PCR assay. Data are expressed as means  $\pm$  standard deviations of  $log_{10}$ . The backslash indicates that the challenge virus was not recovered at the corresponding time point.