Title:

A robust in vitro model for trans-lymphatic endothelial migration

Authors

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Supplementary table 1. Comparison of gene array data for primary LEC and SVEC4-10.

Supplementary Table 2. Significant Probe Set Differences Between Primary LEC and SVEC4-10.

Probe Set ID	p-value	adjusted p- value	Gene Accession	Gene Symbol
17283364	2.37E-09	9.80E-05	NM_001082976	Tc2n
17456934	4.25E-08	0.000879	NM_001252292	Mest
17359344	2.22E-07	0.003062	NM_009848	Entpd1
17304523	3.61E-07	0.003318	NM_138672	Stab1
17279670	4.29E-07	0.003318	NM_175930	Rapgef5
17480880	5.05E-07	0.003318	NM_001008548	Pde2a
17206471	6.27E-07	0.003318		
17334803	6.42E-07	0.003318	NM_028101	Jmjd8
17248426	1.06E-06	0.004887	NM_011412	Slit3
17315133	1.29E-06	0.005351	NM_001277255	Acvrl1
17239113	8.61E-06	0.032359	NM_175155	Sash1
17355565	1.09E-05	0.037647	NM_001171010	Slc14a1
17206479	1.24E-05	0.039357		
17271118	1.35E-05	0.039857	NM_145823	Pitpnc1



Supplementary Figure 1. (A) SVEC on the inserts were stained with DAPI (violet) and phalloidin (gold) and images were captured with z-stack confocal, $\times 40$, scale bar 21 µm. (B) Mouse LEC on the inserts were stained with DAPI (violet) and anti-VE-cadherin (green) and

images were captured with z-stack confocal imaging, ×40, scale bar 21 μ m. (C) SVEC on the inserts were stained with anti-laminin (cyan), anti-collagen (red) and DAPI (blue) and images were captured fluorescence microscopy, ×20, scale bar 42 μ m; ×100, scale bar 8 μ m. Representative of three independent experiments.



Supplementary Figure 2. Diagram for inserts: (A, B) standard orientation of inserts (SVEC or LEC) and inverted orientation insets (iSVEC or iLEC); (C) inserts with various coating on inserts; (D, E) static assay and fluid flow assay; (F) inserts pre-coated with CCL21.



Supplementary Figure 3. Permeability of different monolayers. Evans blue dye added to the upper chamber of the transwell assay with different monolayers coated on the inserts; samples collected at the indicated time points from the lower chamber. Representative of three independent experiments.

Α	VCAM-1	DAPI	Merge
Medium			
S1P			
TNFα			
В	VE-cadherin	DAPI	Merge
			SAR POST
Medium			
Medium S1P			

Supplementary Figure 4. (A) SVEC on the inserts were stained with DAPI (blue) and anti-VCAM-1 (green) and images were captured with fluorescence microscopy, \times 60, scale bar 14 µm. (B) SVEC on the inserts were stained with DAPI (blue) and anti-VE-cadherin (green) and images were captured with fluorescence microscopy, \times 60, scale bar 14 µm. Representative of three independent experiments.



Supplementary Figure 5. Fluorescence staining of LEC layers. (A) VCAM-1 expression on mLEC treated with medium, TNF α or S1P and images were captured with fluorescence microscopy, ×10, scale bar 80 µm. (B) ICAM-1 expression on SVEC4-10 treated with medium, TNF α or S1P and images were captured with fluorescence microscopy, x10, scale bar 80 µm. Representative of three independent experiments. **P* < 0.05, ****P* < 0.0001.



Supplementary Figure 6. (A, B) SVEC on the inserts were stained with DAPI (blue), phalloidin (red) and anti-VCAM-1 (green) and images were captured with fluorescence microscopy, $\times 20$, scale bar 42 μ m; $\times 60$, scale bar 14 μ m. Representative of three independent experiments.



Supplementary Figure 7. (A) SVEC on the inserts were stained with DAPI (blue), anti- β -catenin (red) and anti-VCAM-1 (green) and images were captured with fluorescence microscopy, ×60, scale bar 14 µm. (B) SVEC on the inserts were stained with DAPI (blue), anti-moesin (red) and anti-VCAM-1 (green) and images were captured with fluorescence microscopy, × 60, scale bar 14 µm. Representative of three independent experiments.



Supplementary Figure 8. LEC conditioned medium does not induce CD4 T cell migration. CD4 T cell migration toward LEC conditioned medium or CCL19. Representative of three independent experiments.