# RNA-Seq based genome-wide analysis reveals loss of inter-chromosomal regulation in breast cancer

Jesús Espinal-Enríquez<sup>1,2,+</sup>, Cristóbal Fresno<sup>3+</sup>, Guillermo de Anda-Jauregui<sup>1</sup>, and Enrique Hernandez-Lemus<sup>1,2,\*</sup>

1Computational Genomics Division, National Institute of Genomic Medicine (INMEGEN), 14610, México

2Centro de Ciencias de la Complejidad, Universidad Nacional Autónoma de México (UNAM), 04510 México

2UA AREA CS. AGR. ING. BIO Y S, CONICET - Universidad Católica de Córdoba, Argentina

\*ehernandez@inmegen.gob.mx

+these authors contributed equally to this work

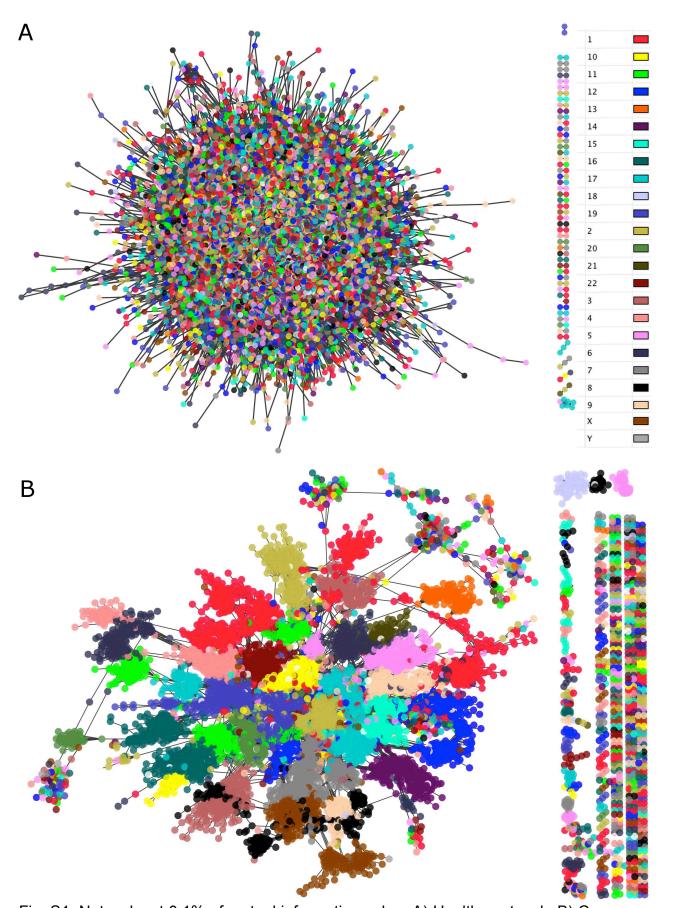


Fig. S1. Networks at 0.1% of mutual information value. A) Healthy network. B) Cancerous network (CN). Both networks are depicted with the same layout as in Fig 6. The colour code is according to the chromosome location in which each gene is located. Despite the fact that both networks have a giant component, a simple visual inspection shows that the CN's gene are more connected to other genes of the same chromosome than to other chromosomes.

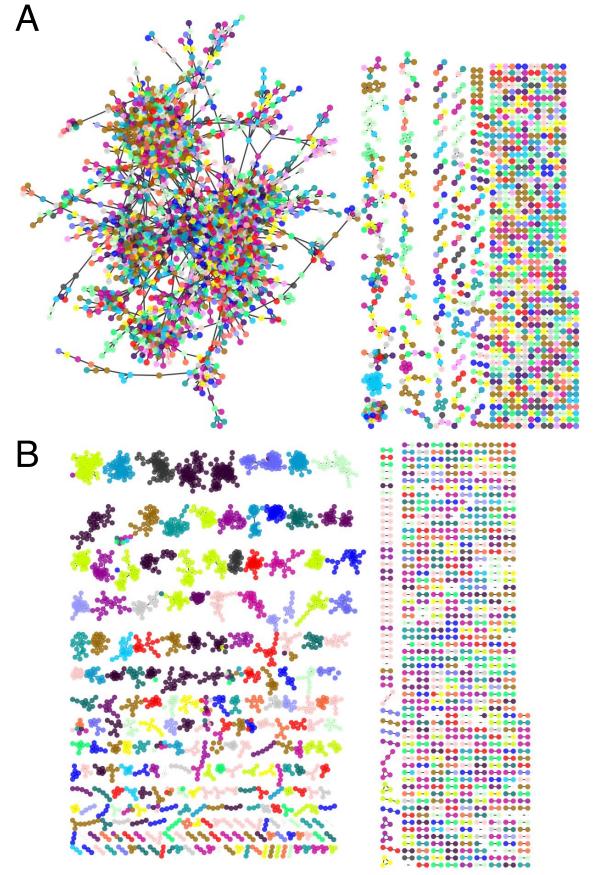


Fig. S2. Networks at 0.001% of mutual information value. Healthy network (HN) B) Cancerous network (CN). Both networks are depicted with the same layout as in Fig 6. The colour code is according to the chromosome location in which each gene is located. Notice that gene connections are similar to the ones showed in Fig. 6.

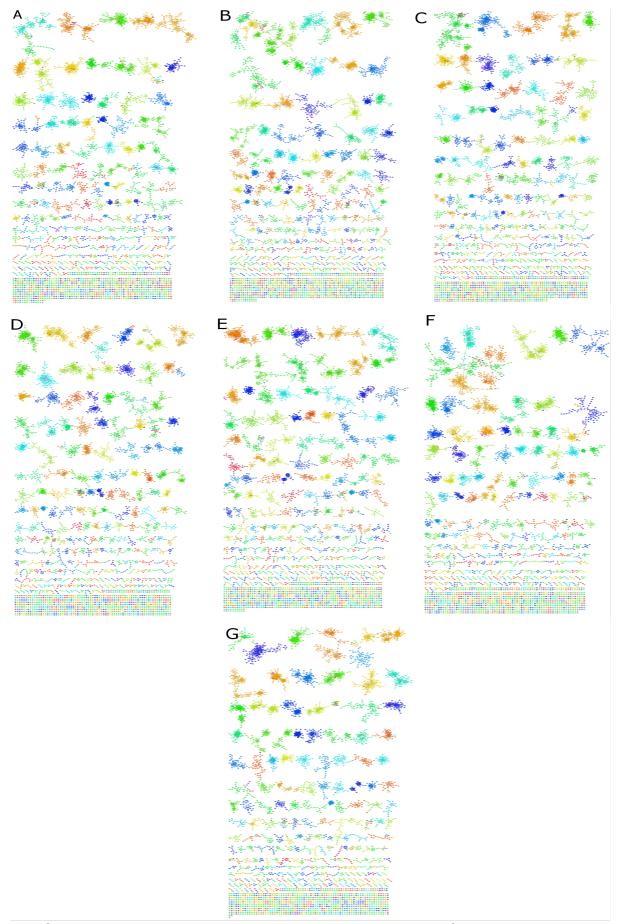


Fig. S3. Network structure does not depend on the number of samples. Networks were constructed with 110 random samplings. A) to G). Genes are coloured according to the chromosomal location of each gene. Notice that for each network, the structure and connectivity are maintained: loss of inter-chromosomal regulation and small connected components.



Fig. S4. Validation cancerous networks with larger number of samples. To corroborate the number of samples does not determine the network structure. Networks were constructed with A) 220 random samplings. B) 330 random samplings C) 440 random samplings D) 550 random samplings and E) 660 random samplings. Genes are again coloured according to the chromosomal location of each gene.

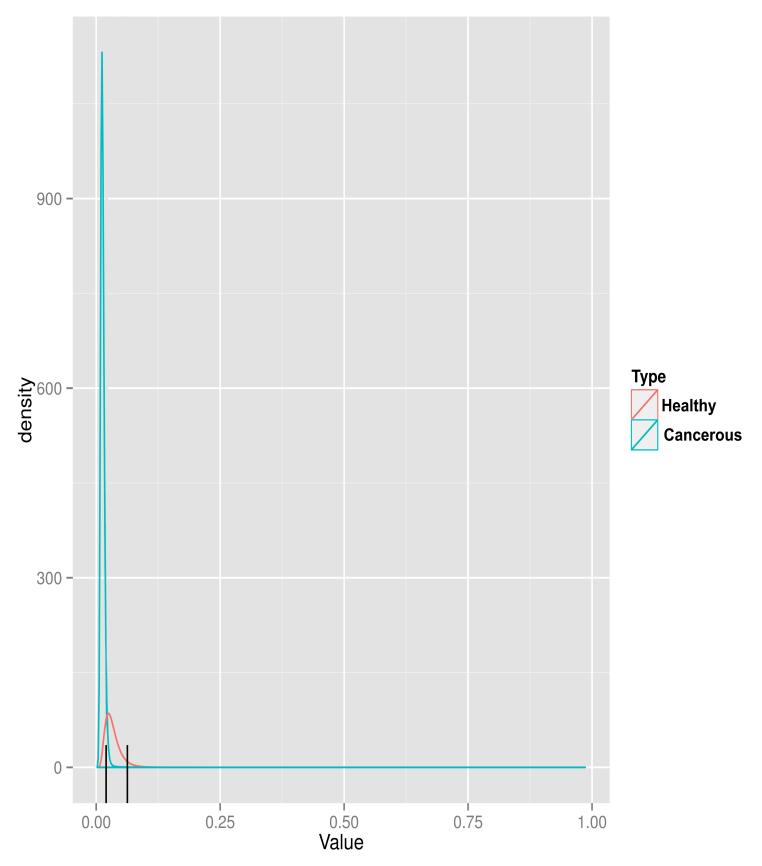


Fig. S5. Mutual information density plots. Healthy and cancerous density plots are shown in red and cyan colour respectively. Vertical black lines represent the top 0.01% cut-off for each experimental condition. Note the density shift to lower mutual information values for cancerous condition compared to the healthy case.

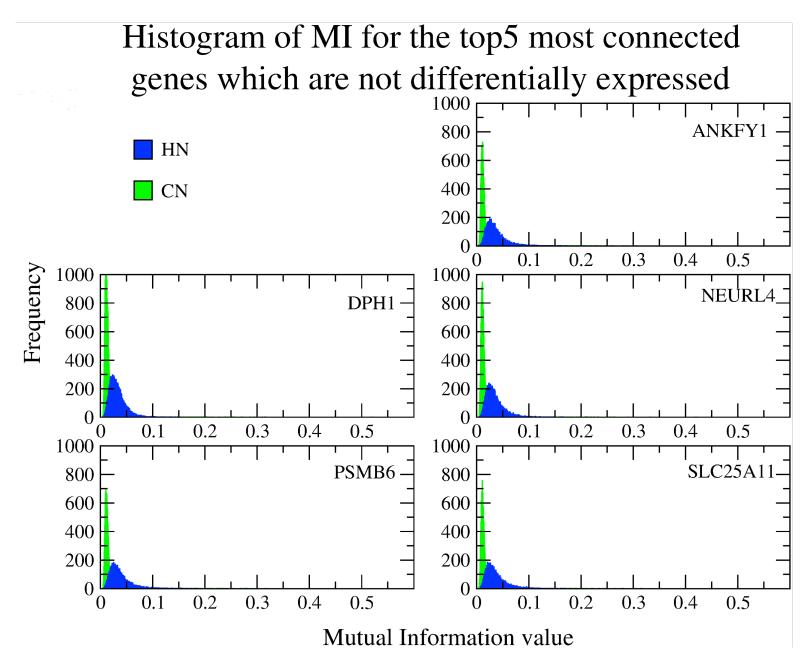


Fig. S6. Histogram of Mutual Information for the 5 most connected genes in the CN and they are not differentially expressed. Each plot represents the frequency of MI values between the mentioned genes and the rest of the genome. Each histogram contains 15,642 values of MI. In blue, the distribution of Healthy samples are depicted, meanwhile for cancerous network, the distributions are in green. Notice that blue values tend to be higher than the green ones, as it can be expected from the visual inspection of Fig. S5.

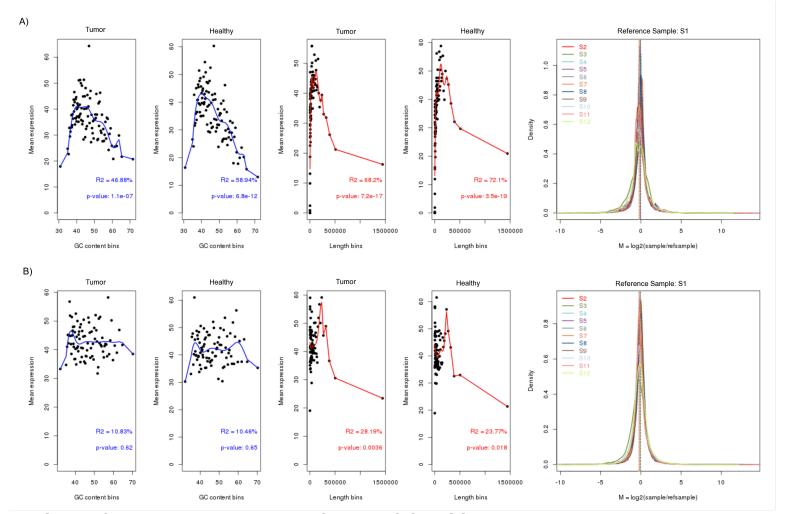


Fig. S7. Artefact removal assessment. A) Original NOISEq GC content (blue), gene length (red) and RNA content bias detection plots for cancerous and healthy experimental conditions. For the first four scatter bias plots the X axis represents the centre value of 200 features bins for the respective artefact against its 5% trimmed gene mean counts, which is then fitted with a cubic spline regression model (coloured line). The RNA content panel shows the density of the M=log2(sample/reference simple) for each sample. In this case sample one (S1) was taken as reference by default. B) Similar plots obtained after EDASeq within and between normalisation for bias removal.

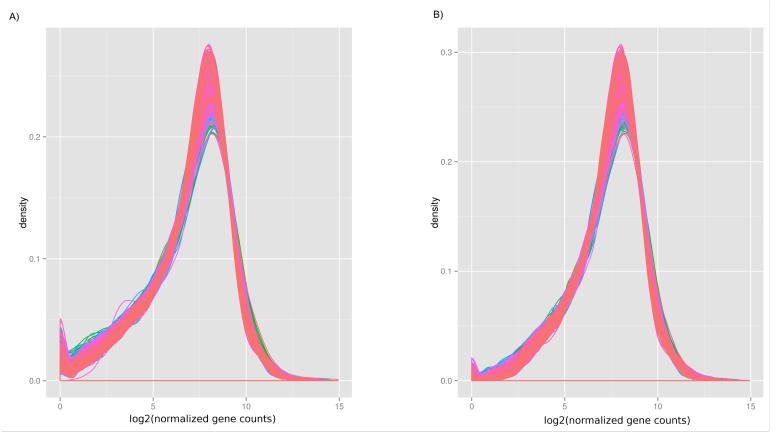


Fig. S8. Normalized samples expression density plot. Values are expressed in log2(normalized genes counts) scale. A) Original bimodal plot. B) Lower expressed genes artefact were removed after CPM < 10 filter was applied.

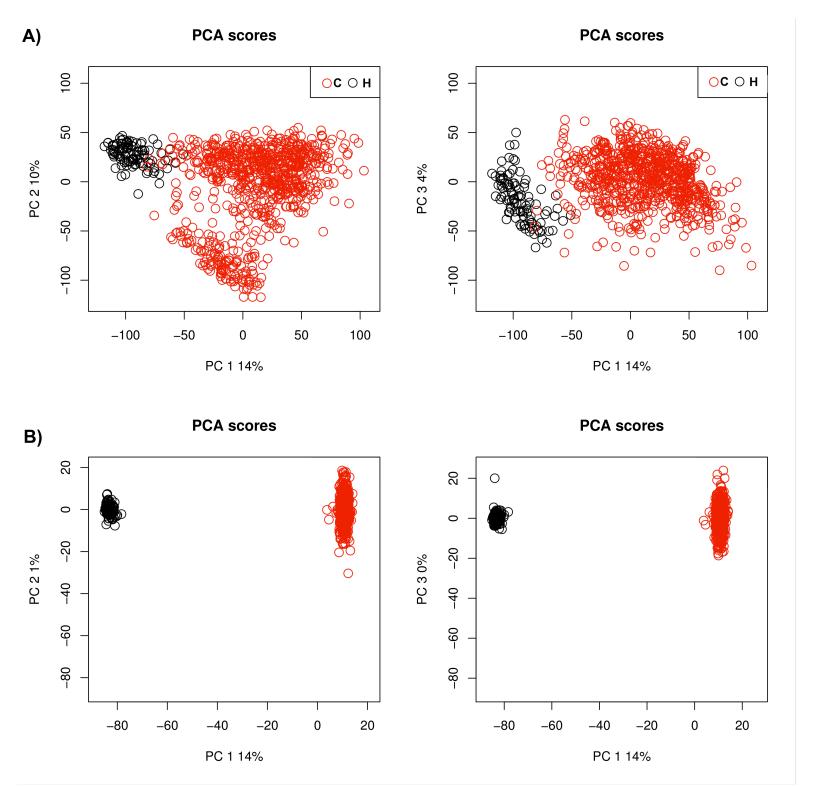


Fig. S9. Multidimensional noise reduction. A) Original scatter principal component (PC) analysis score plot for the first-second/third components for healthy (black) and cancerous (red) samples. B) Similar plot after ARSyN noise reduction

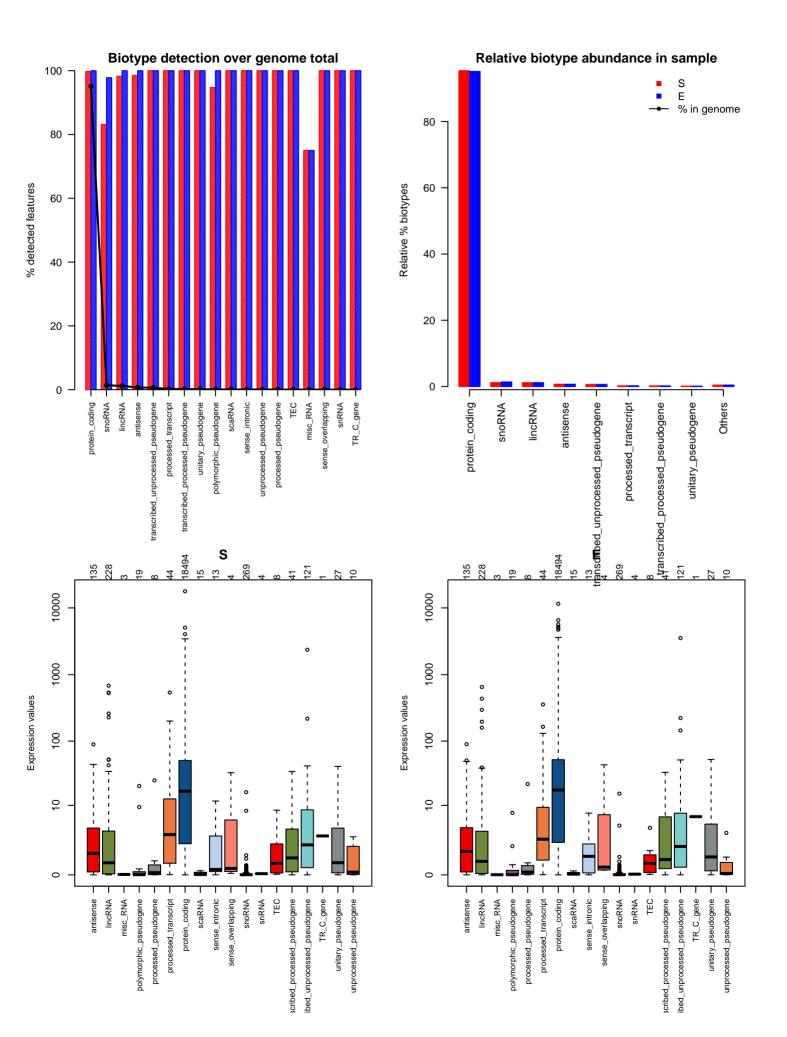
# **Quality Control of Expression Data**

Generated by NOISeq on 08 Jul 2015, 12:42:05

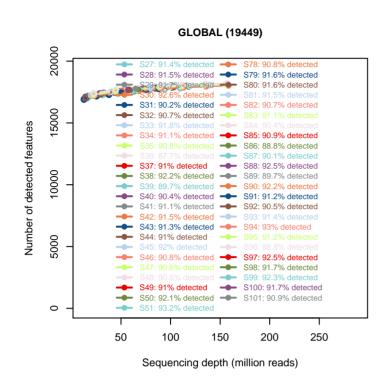
# **Content**

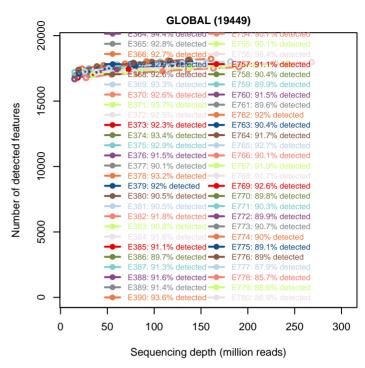
Plot	Description
Biotype detection	Biotype abundance in the genome with %genes detected (counts > 0) in the sample/condition. Biotype abundance within the sample/condition.
Biotype expression	Distribution of gene counts per million per biotype in sample/condition (only genes with counts > 0).
Saturation	Number of detected genes (counts > 0) per sample across different sequencing depths
Expression boxplot	Distribution of gene counts per million (all biotypes) in each sample/condition
Expression barplot	Percentage of genes with >0, >1, >2, >5 or >10 counts per million in each sample/condition.
Length bias	Mean gene expression per each length bin. Fitted curve and diagnostic test.
GC content bias	Mean gene expression per each GC content bin. Fitted curve and diagnostic test.
RNA composition bias	Density plots of log fold changes (M) between pairs of samples.  Confidence intervals for the median of M values.

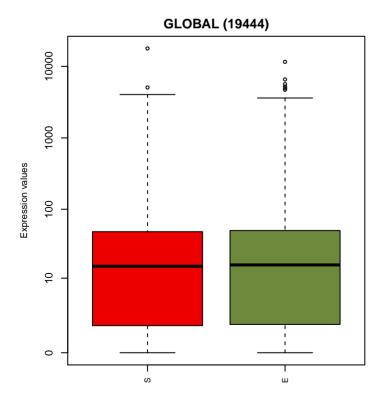
# **Biotype detection**

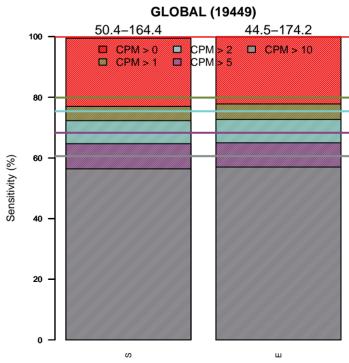


# Sequencing depth & Expression quantification







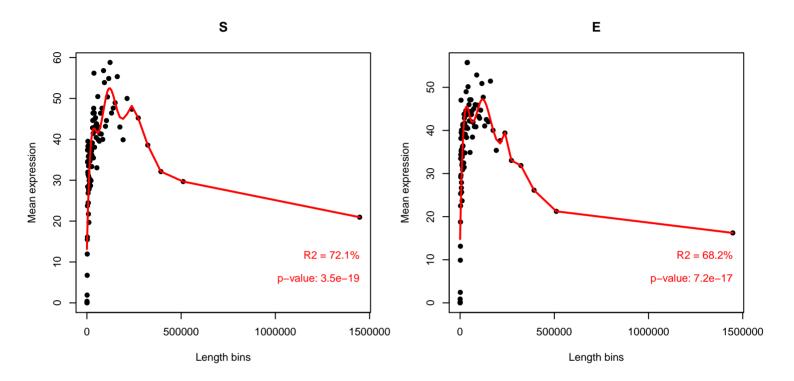


# Sequencing bias detection

#### Diagnostic plot for feature length bias

FAILED. At least one of the model p-values was lower than 0.05 and R2 > 70%.

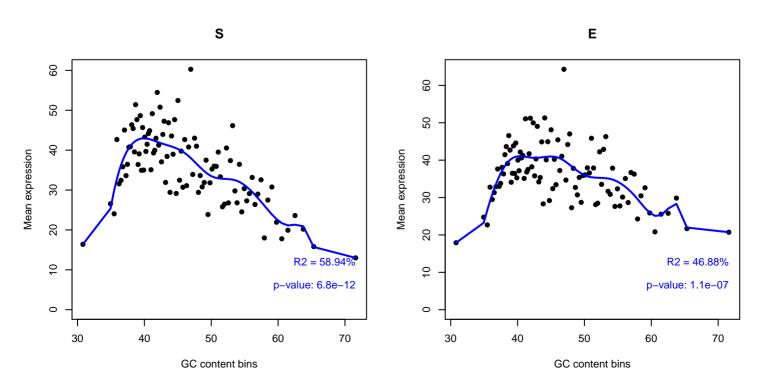
Normalization for correcting length bias is recommended.



#### Diagnostic plot for GC content bias

WARNING. At least one of the model p-values was lower than 0.05, but R2 < 70% for at least one condition.

Normalization for correcting GC content bias could be advisable. Plese check in the plots below the strength of the relationship between GC content and expression.



#### Diagnostic plot for differences in RNA composition

FAILED. There is a pair of samples with significantly different RNA composition Normalization for correcting this bias is required.

#### Reference sample: S1

#### Confidence intervals for median of M values

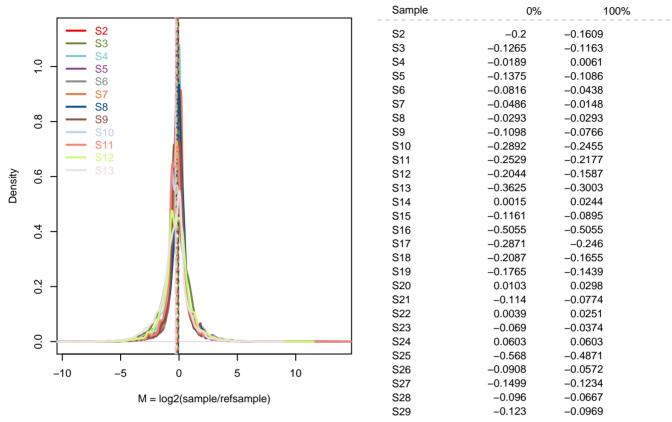
Diagnostic Test

**FAILED** 

**FAILED** 

**PASSED** 

**FAILED** 



# **Exploratory PCA**

Use this plot to see if samples are clustered according to the experimental design.

Use ARSyNseq function to correct potential batch effects.

