

Supplemental information (Figures, and Materials and Methods)

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Title: Integrin beta8 (ITGB8) activates VAV-RAC1 signaling via FAK in the acquisition of endometrial epithelial cell receptivity for blastocyst implantation.

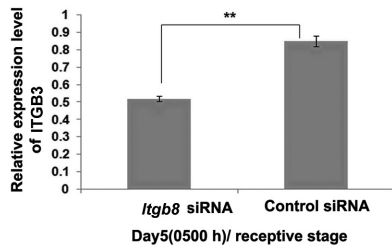
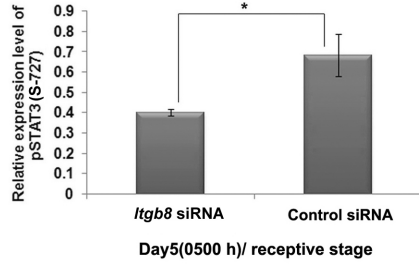
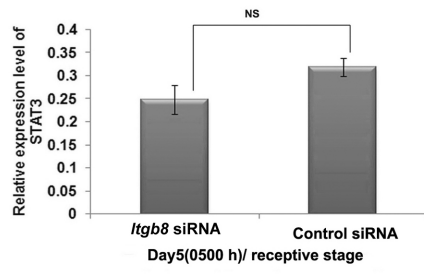
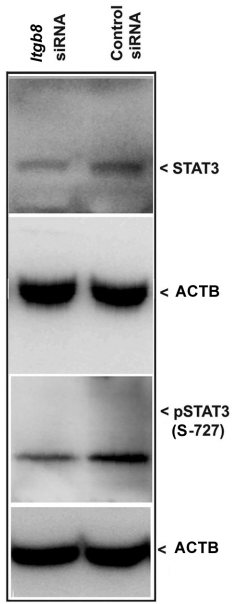
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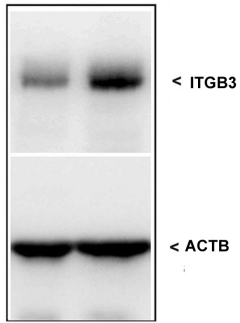
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A.



B.



C.

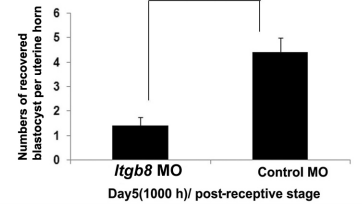
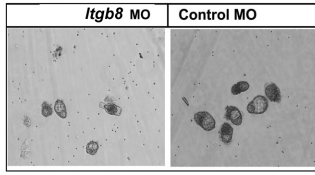
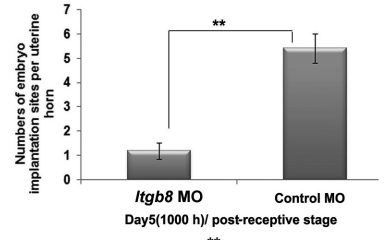
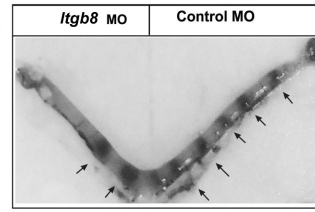


Figure-S1.

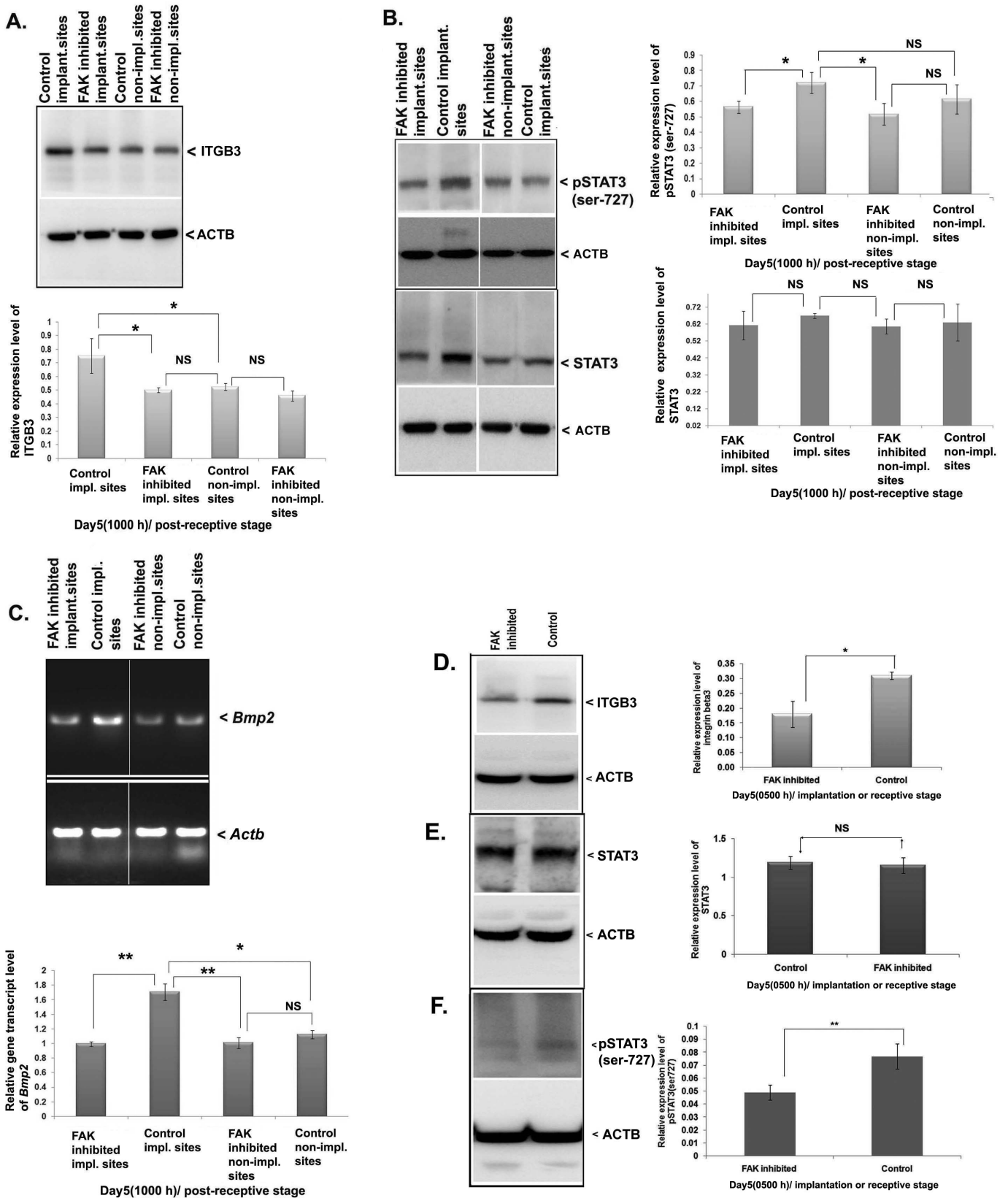


Figure-S2.

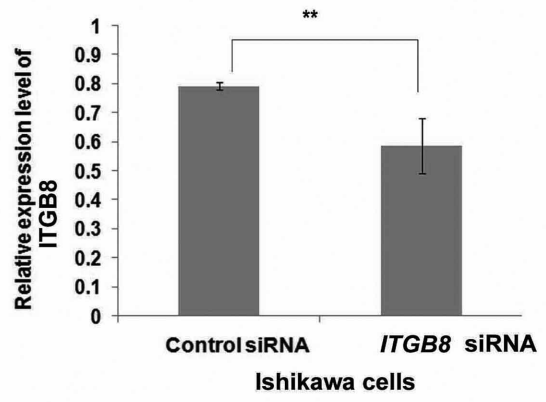
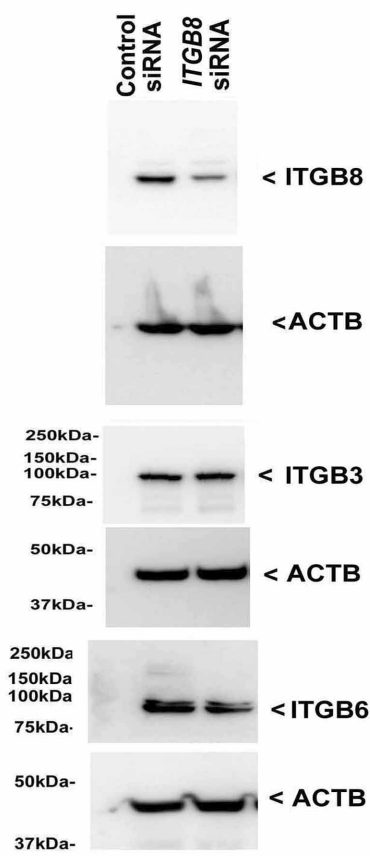


Figure-S3.

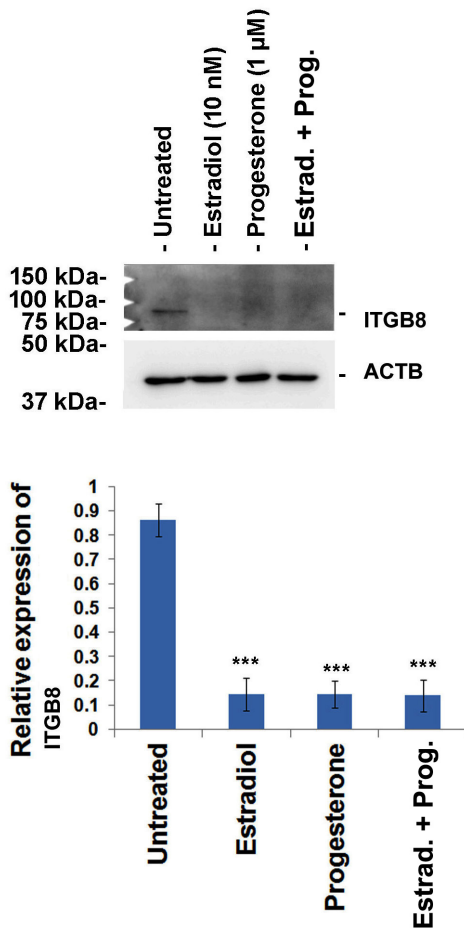
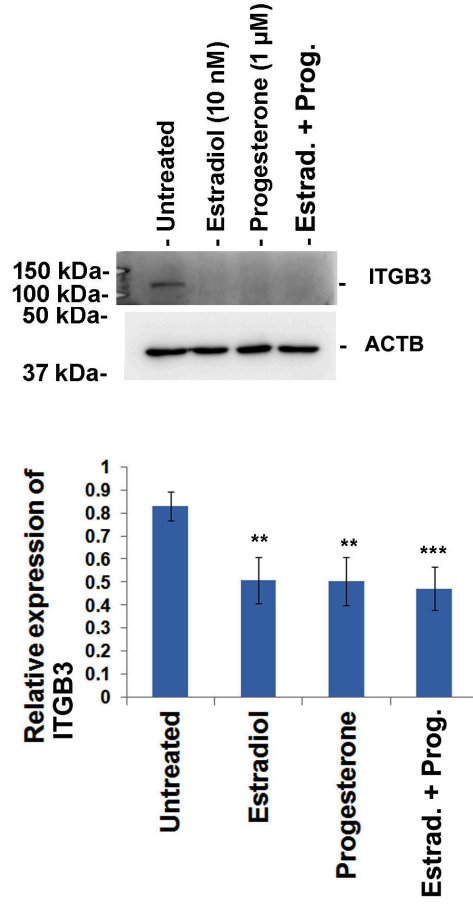
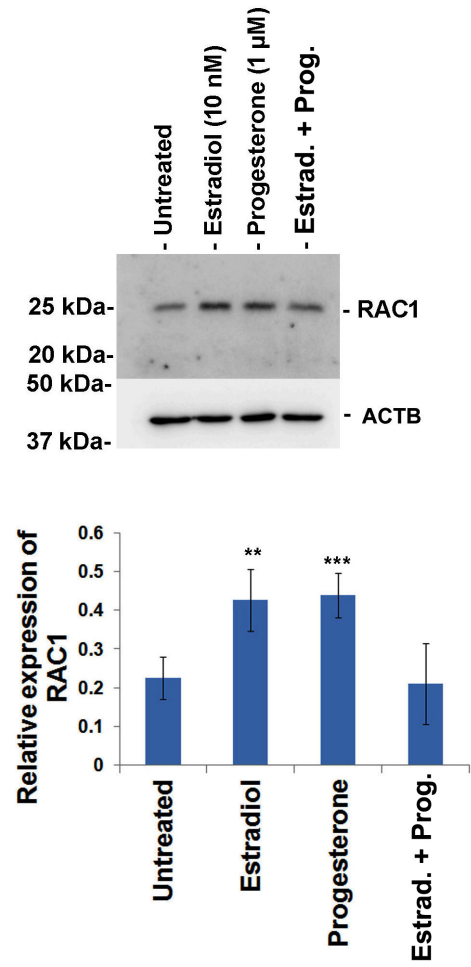
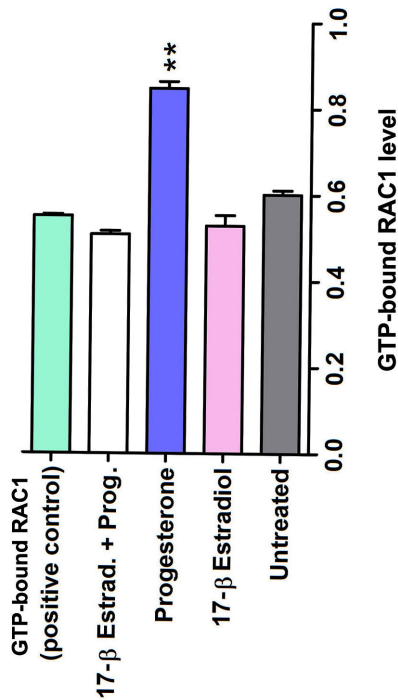
A.**B.****C.****D.****Figure-S4**

Figure-S1: Integrin beta8 regulates embryo implantation associated biochemical markers in the endometrial tissue and endometrial epithelial cells in the mouse model.

(A) Integrin beta8 silencing from mouse endometrial epithelial cells post 24 hrs culture from day5 (0500 h) stage caused poor phosphorylation of STAT3 (serine 727). (B) Transient knockdown of *Itgb8* from the mouse endometrial epithelial cells (day5, 0500 h) led down-regulation of expression of ITGB3. (C) Transient silencing of *Itgb8* from one of the mouse uterine horns at pre-implantation or day4 (1000 h) stage showed the reduction of implantation sites and blastocyst numbers (recovered). Actin-beta was used as loading control to normalize the immunoblot values. (** p<0.01,*p<0.05, NS p>0.05).

Figure-S2: FAK affected the embryo implantation related biomarkers, i.e., ITGB3, STAT3, and *Bmp2* signaling in the process of mouse endometrial epithelial cell receptivity.

(A) The expression level of ITGB3 was determined using immunoblotting in mouse endometrium during day5 (1000 h) after the FAK activity inhibition at day4 (1000 h). (B) Similarly, the expression of total and phosphorylated forms of STAT3 was determined by immunoblotting in the mouse endometrium at day5(1000 h) /post-implantation stage after the FAK activity inhibition. (C) PCR analysis of *Bmp2* was done in the mouse uterus post-FAK activity inhibition during day5(1000 h) of window of endometrial receptivity. (D) Later, the expression of ITGB3 was assayed using immunoblotting in the mouse endometrial epithelial cells after the FAK activity inhibition. (E and F) Similarly, the expression level of total and phosphorylated forms of STAT3 was assayed after the FAK activity inhibition in the endometrial cells from day5, 0500 h stage. Actin beta was used as a loading control to normalized the immunoblot values. (** p<0.01,*p<0.05, NS p>0.05).

Figure-S3: Silencing RNA against *ITGB8* found to be specific for its target and did not have an off-target effect.

Integrin beta8 was silenced in the human endometrial epithelial cells (Ishikawa) and we determined the expression level of ITGB8 along with other beta integrins (ITGB3 and ITGB6). Post-*ITGB8* silencing in the Ishikawa cells, the expression of ITGB3 and ITGB6 was detected by immunoblotting and found unchanged when compared with the scrambled siRNA. (** p<0.001).

Figure-S4. Integrins beta3 and beta8 were seen downregulated in the human origin endometrial cells (Ishikawa cells) in response to either progesterone or 17-β-estradiol alone or in combination with progesterone and 17-β-estradiol supplementation.

However, RAC1 was seen upregulated in progesterone or 17- β -estradiol supplementation, but not in progesterone and 17- β -estradiol combined supplementation. (A, B and C) Using immunoblotting and densitometry, the expression analysis of ITGB3, ITGB8, and RAC1 was done post-progesterone (P4), 17- β -estradiol (E2), and progesterone and 17- β -estradiol combined treatment in the Ishikawa cells during 24 hrs of culture. (D) The level of GTP-bound form of RAC1 was determined biochemically in response to progesterone, 17- β -estradiol, and progesterone and 17- β -estradiol combined supplementation (24 hrs) in the Ishikawa cells. (** $p < 0.01$, NS $p > 0.05$).

Supplementary Materials and Methods

Steroid treatment to Ishikawa cells: Ishikawa (Human endometrial epithelium carcinoma cells) cells were grown in a minimum essential medium as described earlier¹. After reaching cells confluency at 80%, cells were treated either with estradiol (10 nM), or progesterone (1 μ M) or with both estradiol (10 nM) and progesterone (1 μ M) or left untreated as a control for 24 hours at 37°C in 5% CO₂ incubator². After 24 hours, cells were washed with PBS and the total protein lysate was prepared using RIPA buffer (0.5M Tris-HCl, pH 7.4, 1.5M NaCl, 2.5% deoxycholic acid, 10% NP-40, 10mM EDTA) at 4°C.

Reference List

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2. Somkuti,S.G., Yuan,L., Fritz,M.A., & Lessey,B.A. Epidermal growth factor and sex steroids dynamically regulate a marker of endometrial receptivity in Ishikawa cells. *J. Clin. Endocrinol. Metab.* **82**, 2192-2197 (1997).