

Supporting Information

1,3-Oxazin-6-one Derivatives and Bohemamine-Type Pyrrolizidine

Alkaloids from a Marine-Derived *Streptomyces spinoverrucosus*

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Bioassay Protocols

Antibiotic assays. The antibiotic activities against *Pseudomonas aeruginosa* and *Bacillus subtilis* were evaluated by an agar dilution method. The tested strains were cultivated in LB agar plates at 37 °C. Compounds **1–9**, and positive control (erythromycin) were dissolved in MeOH at different concentrations from 100 to 0.1 μg/mL by the continuous 10-fold dilution methods. A 10 μL quantity of test solution was absorbed by a paper disk (5 mm diameter) and placed on the assay plates. After 24 h incubation, zones of inhibition (mm in diameter) were recorded.

Cytotoxicity assays. Cell lines were cultivated in 10 cm dishes (Corning, Inc.) in NSCLC cell-culture medium: RPMI/L-glutamine medium (Invitrogen, Inc.), 1000 U/mL penicillin (Invitrogen, Inc.), 1 mg/mL streptomycin (Invitrogen, Inc.), and 5% fetal bovine serum (Atlanta Biologicals, Inc.). Cell lines were grown in a humidified environment in the presence of 5% CO₂ at 37 °C. For cell viability assays, HCC366, A549, HCC44 and HCC1171 cells (60 μL) were plated individually at a density of 1200, 750 and 500 cells/well, respectively, in 384-well microtiter assay plates (Bio-one; Greiner, Inc.). After incubating the assay plates overnight under the growth conditions described above, purified compounds were dissolved and diluted in DMSO and subsequently added to each plate with final compound concentrations ranging from 50 μM to 1 nM and a final DMSO concentration of 0.5%. After an incubation of 96 h under growth conditions, Cell Titer Glo reagent (Promega, Inc.) was added to each well (10 mL of a 1:2 dilution in NSCLC culture medium) and mixed. Plates were incubated for 10 min at room temperature, and luminescence was determined for each well using an Envision multimodal plate reader (Perkin-Elmer, Inc.). Relative luminescence units were normalized to the untreated control wells (cells plus DMSO only). Data were analyzed using the Assay Analyzer and Condoseo modules of the Screener Software Suite (GeneData, Inc.) as described previously.^{S1}

Theory and Calculation Details. The calculations were performed by using the density functional theory (DFT) as carried out in the Gaussian 03.^{S2} The preliminary conformational distributions search was performed by HyperChem 7.5 software. All ground-state geometries were optimized at the B3LYP/6-31G(d) level. Solvent effects of methanol solution were evaluated at the same DFT level by using the SCRF/PCM method.^{S3} TDDFT^{S4} at B3LYP/6-31G(d) was employed to calculate the electronic excitation energies and rotational strengths in methanol. The stable conformations obtained at the B3LYP/6-31G(d) level were further used in magnetic shielding constants at the B3LYP/6-311++G(2d,p) level.

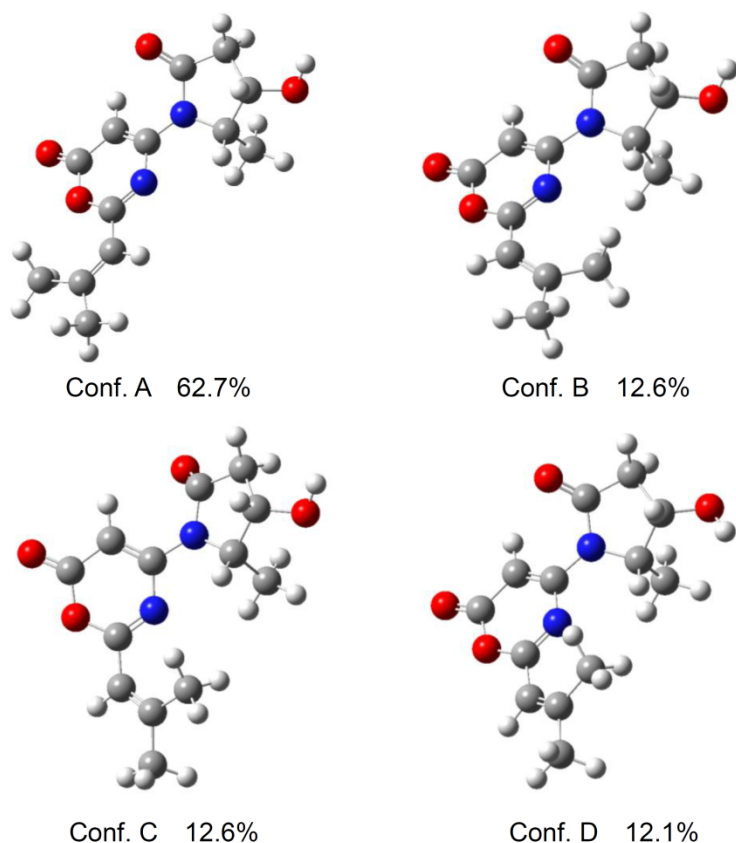


Figure S1. DFT optimized geometries of the four lowest-energy conformers of (4*S*, 5*S*)-1

(S1) Kim, H. S.; Mendiratta, S.; Kim, J.; Pecot, C. V.; Larsen, J. E.; Zubovych, I.; Seo, B. Y.; Kim, J.; Eskiocak, B.; Chung, H.; McMillan, E.; Wu, S.; De Brabander J.; Komurov, K.; Toombs, J. E.; Wei, S.; Peyton, M.; Williams, N.; Gazdar, A. F.; Posner, B. A.; Brekken, R. A.; Sood, A. K.; Deberardinis, R. J.; Roth, M. G.; Minna, J. D.; White, M. A. *Cell* **2013**, *155*, 552–566.

(S2) Gaussian 03, Revision E.01, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, J. A. Montgomery, Jr., T. Vreven, K. N. Kudin, J. C. Burant, J. M. Millam, S. S. Iyengar, J. Tomasi, V. Barone, B. Mennucci, M. Cossi, G. Scalmani, N. Rega, G. A. Petersson, H. Nakatsuji, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, M. Klene, X. Li, J. E. Knox, H. P. Hratchian, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, P. Y. Ayala, K. Morokuma, G. A. Voth, P. Salvador, J. J. Dannenberg, V. G. Zakrzewski, S. Dapprich, A. D. Daniels, M. C. Strain, O. Farkas, D. K. Malick, A. D. Rabuck, K. Raghavachari, J. B. Foresman, J. V. Ortiz, Q. Cui, A. G. Baboul, S. Clifford, J. Cioslowski, B. B. Stefanov, G. Liu, A. Liashenko, P. Piskorz, I. Komaromi, R. L. Martin, D. J. Fox, T. Keith, M. A. Al-Laham, C. Y. Peng, A. Nanayakkara, M. Challacombe, P. M. W. Gill, B. Johnson, W. Chen, M. W. Wong, C. Gonzalez, and J. A. Pople, Gaussian, Inc., Wallingford CT, 2004.

(S3) (a) Miertus, S.; Tomasi, J. *Chem. Phys.* **1982**, *65*, 239–245. (b) Tomasi, J.; Persico, M. *Chem. Rev.* **1994**, *94*, 2027–2094. (c) Cammi, R.; Tomasi, J. *J. Comp. Chem.* **1995**, *16*, 1449–1458.

(S4) (a) Casida, M. E. In *Recent Advances in Density Functional Methods*, part I; Chong, D. P., Eds.; World Scientific: Singapore, 1995; pp 155–192. (b) Gross, E. K. U.; Dobson, J. F.; Petersilka, M. *Top. Curr. Chem.* **1996**, *181*, 81–172. (c) Gross, E. K. U.; Kohn, W. *Adv. Quantum Chem.* **1990**, *21*, 255–291. (d) Runge, E.; Gross, E. K. U. *Phys. Rev. Lett.* **1984**, *52*, 997–1000.

Table S1. The calculated ^{13}C NMR Data for **1** and **1a**

position	measured 1	Calcd. 1	Corr. 1	Error 1	Calcd. 1a	Corr. 1a	Error 1a
1	159.8	163.32	155.7235	4.076468	173.07	163.8759	-4.07586
2	89	95.78	90.54068	-1.54068	94.14	88.71082	0.289178
3	155.3	162.2	154.6426	0.65738	174.23	164.9805	-9.68053
4	57.7	62.76	58.67308	-0.97308	66.85	62.72256	-5.02256
5	64.1	72.98	68.5364	-4.4364	73.08	68.65538	-4.55538
6	39.3	45.32	41.84173	-2.54173	40.57	37.69611	1.603889
7	173.4	177.12	169.0419	4.358088	178.47	169.0183	4.381719
8	13.3	11.91	9.597741	3.702259	14.81	13.16486	0.135137
1'	163.7	173.55	165.5965	-1.89651	173.9	164.6663	-0.96627
2'	116.2	124.56	118.3163	-2.11626	122.83	116.0323	0.167691
3'	158.7	169	161.2053	-2.5053	170.71	161.6284	-2.92843
4'	28	30.57	27.60651	0.393493	30.02	27.64935	0.350654
5'	21	20.84	18.21608	2.783916	20.5	18.58345	2.41655

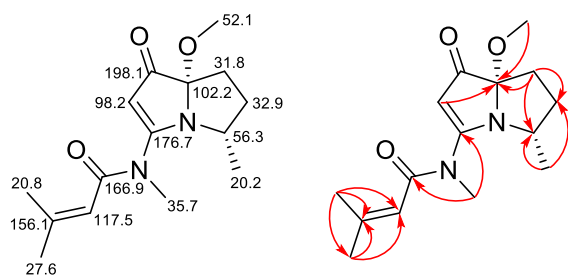
**Figure S2.** ^{13}C and 2D NMR of **3a** (These data were from HMBC and HSQC spectra).

Figure S3. HRESIMS spectrum of spinoxazine A (1)

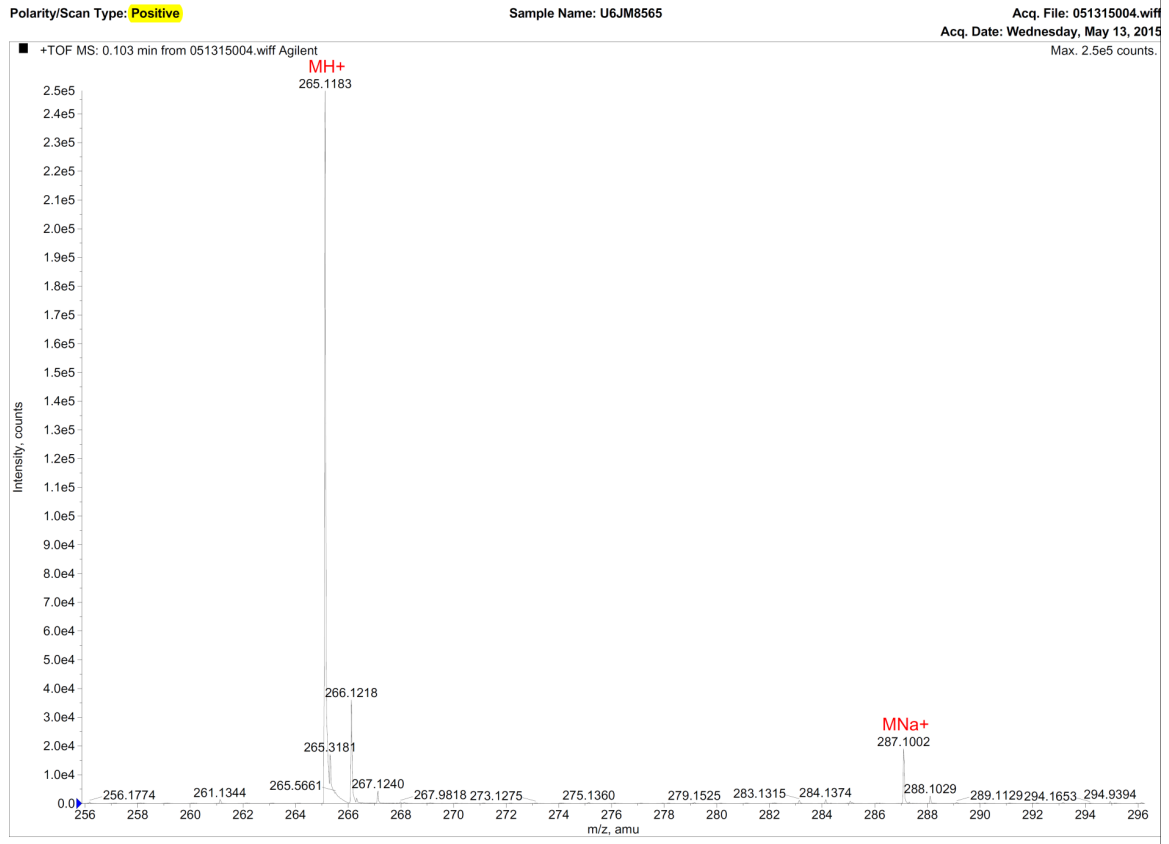


Figure S4. ¹H-NMR spectrum of spinoxazine A (1) in DMSO-d₆

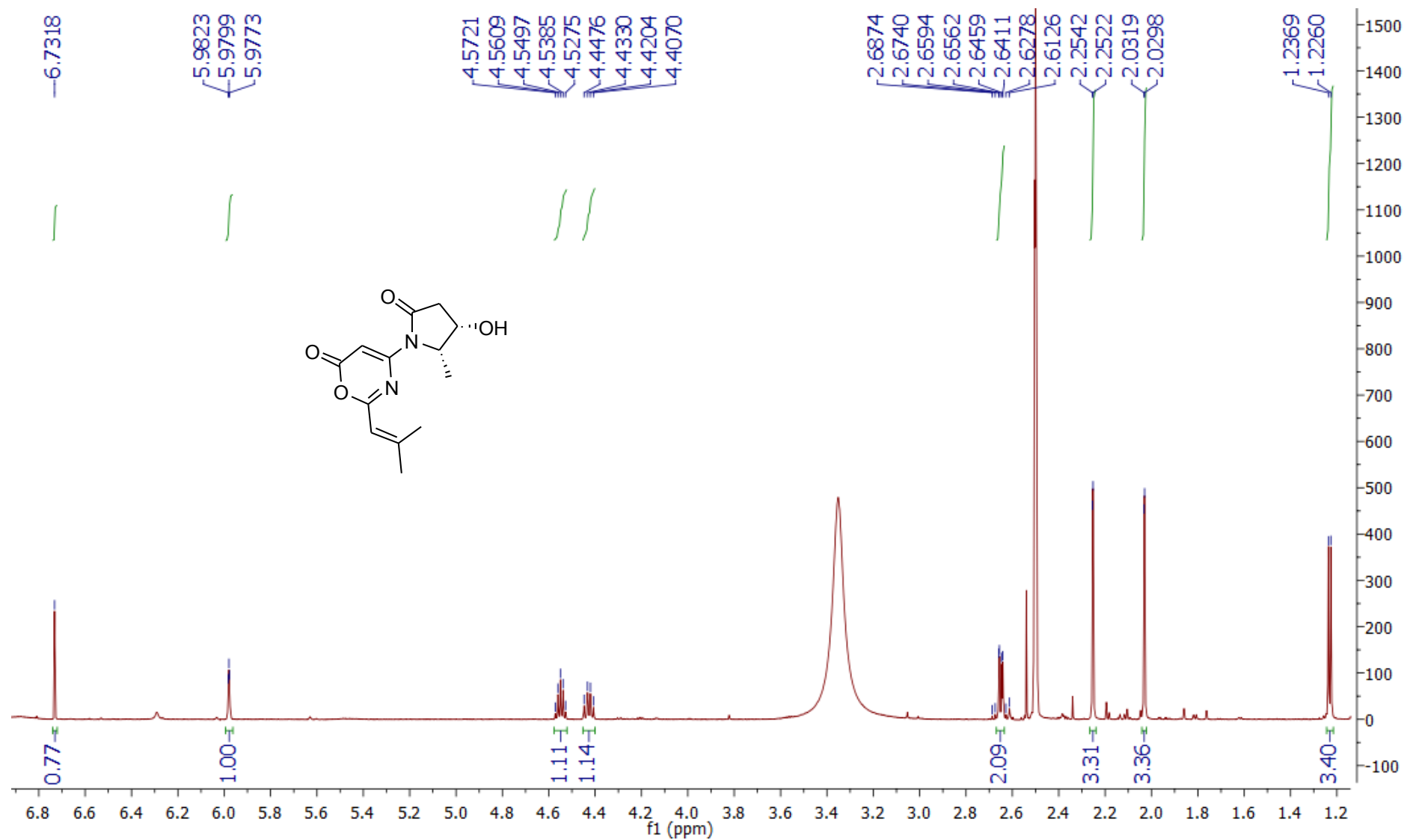


Figure S5. ^{13}C -NMR spectrum of spinoxazine A (1) in $\text{DMSO-}d_6$

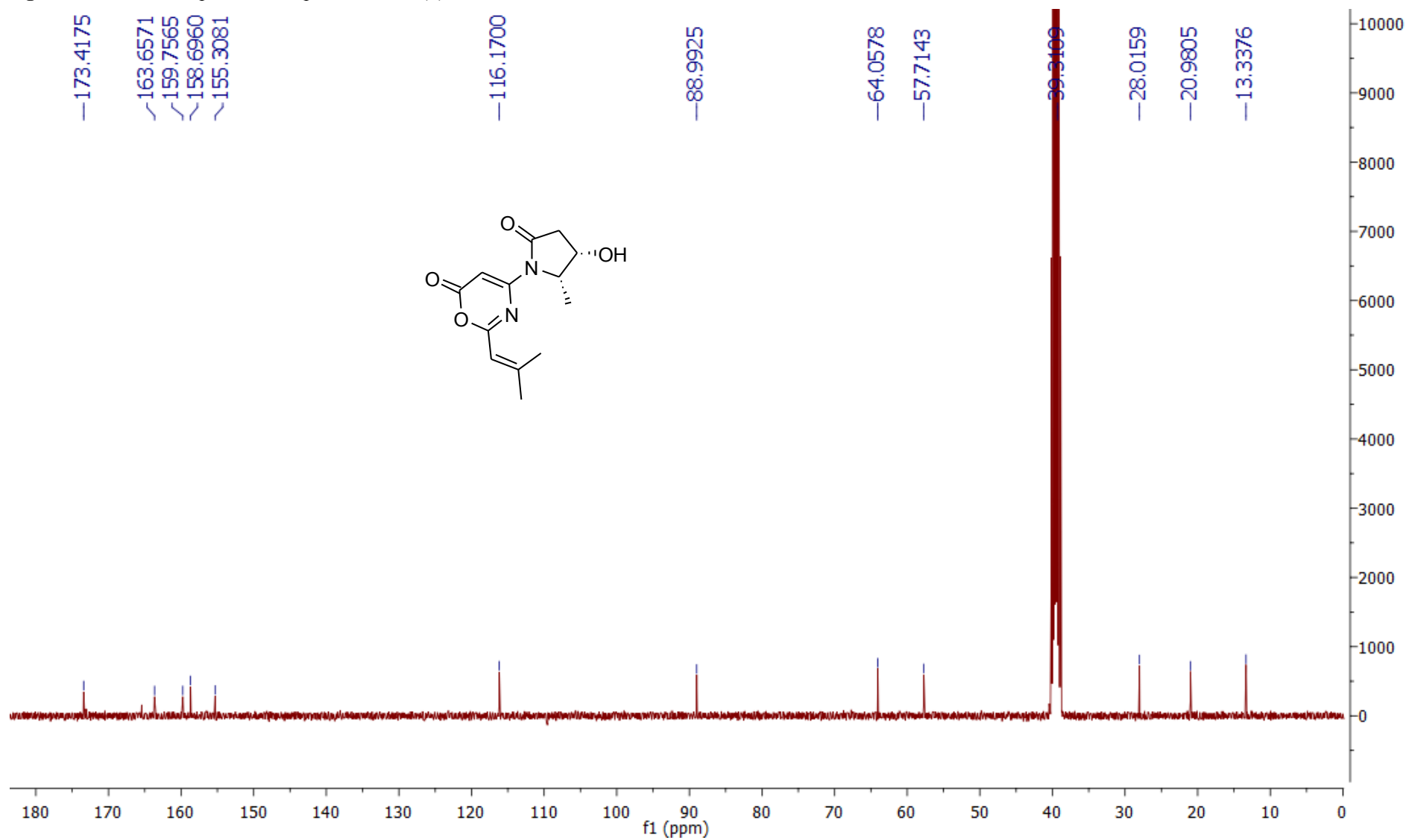


Figure S6. HSQC spectrum of spinoxazine A (1) in DMSO- d_6

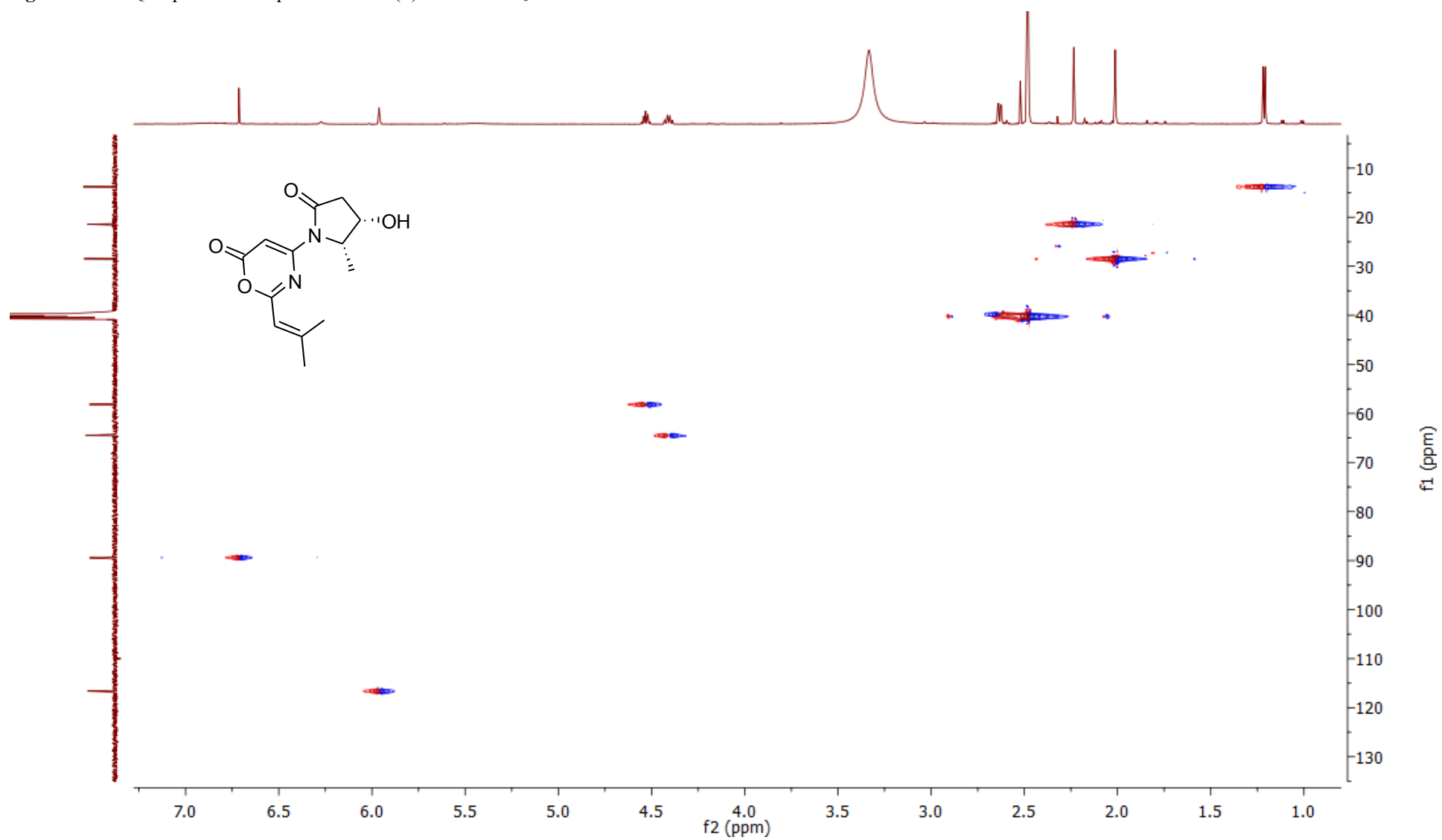


Figure S7. ^1H - ^1H COSY spectrum of spinoxazine A (1) in $\text{DMSO-}d_6$

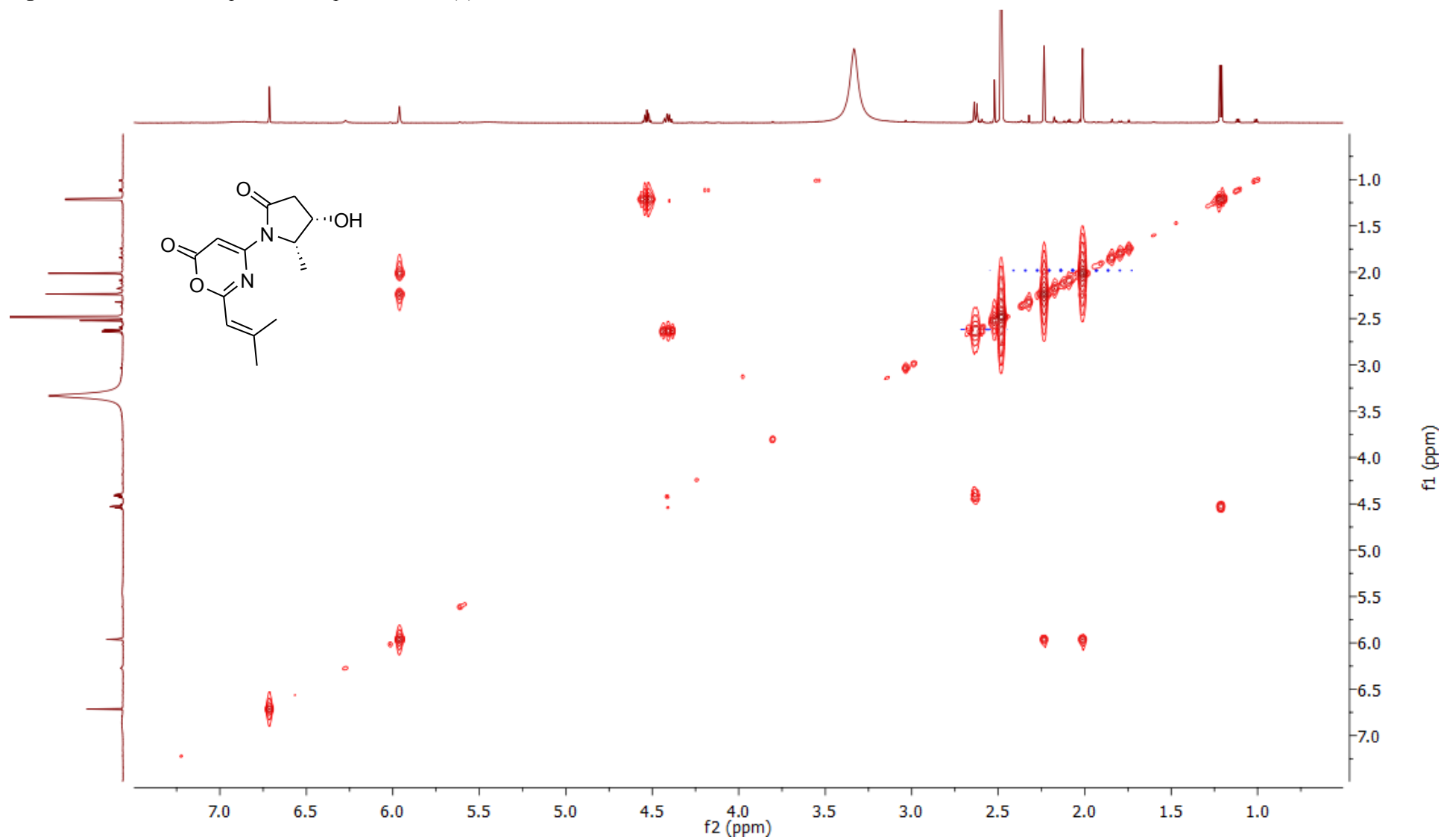


Figure S8. HMBC spectrum of spinoxazine A (1) in DMSO- d_6

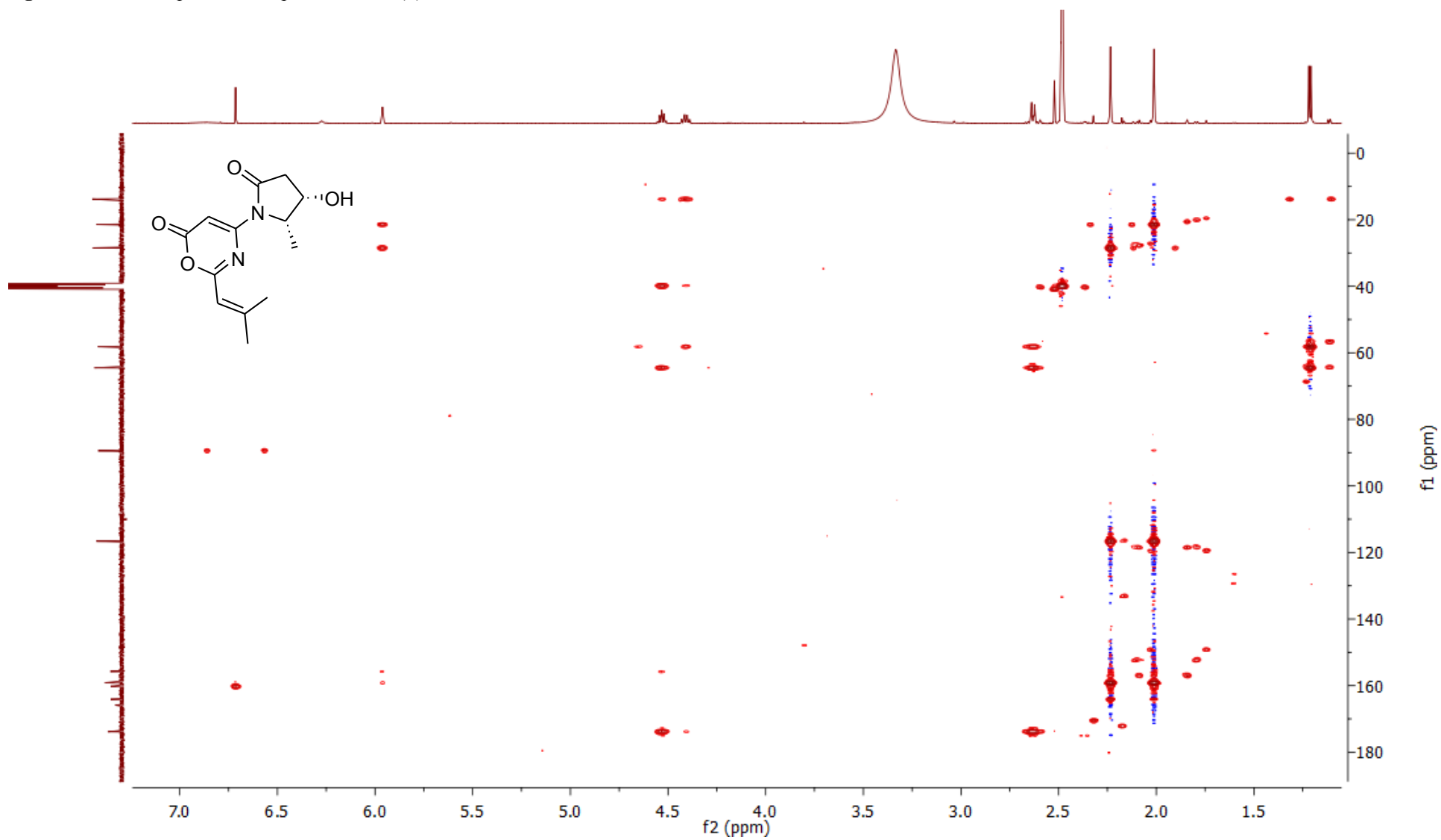


Figure S9. NOESY spectrum of spinoxazine A (1) in DMSO- d_6

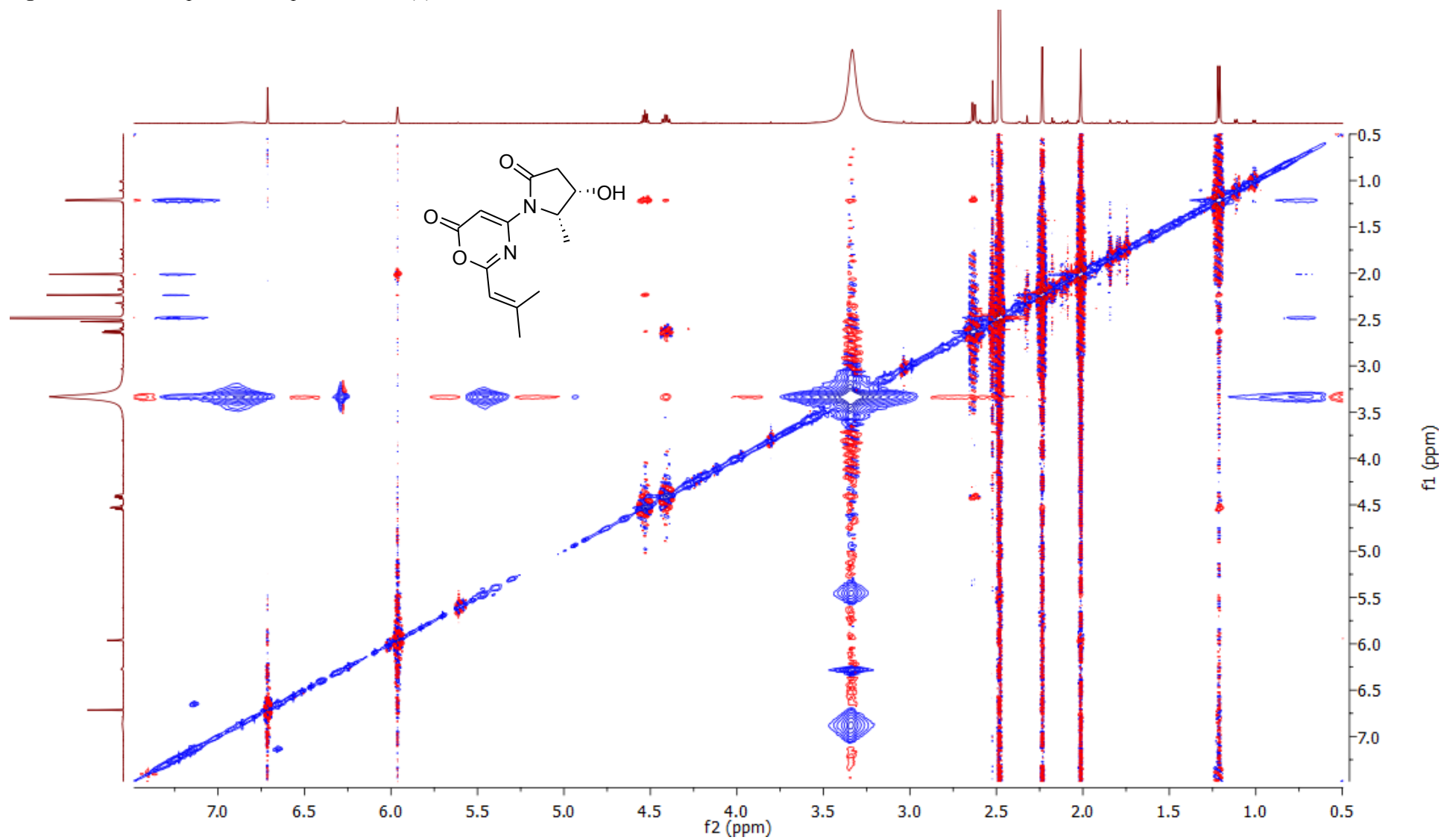


Figure S10. HRESIMS spectrum of spinoxazine B (2)

Polarity/Scan Type: **Positive**

Sample Name: U6JM8648

Acq. File: 052215_A107.wiff
Acq. Date: Friday, May 22, 2015

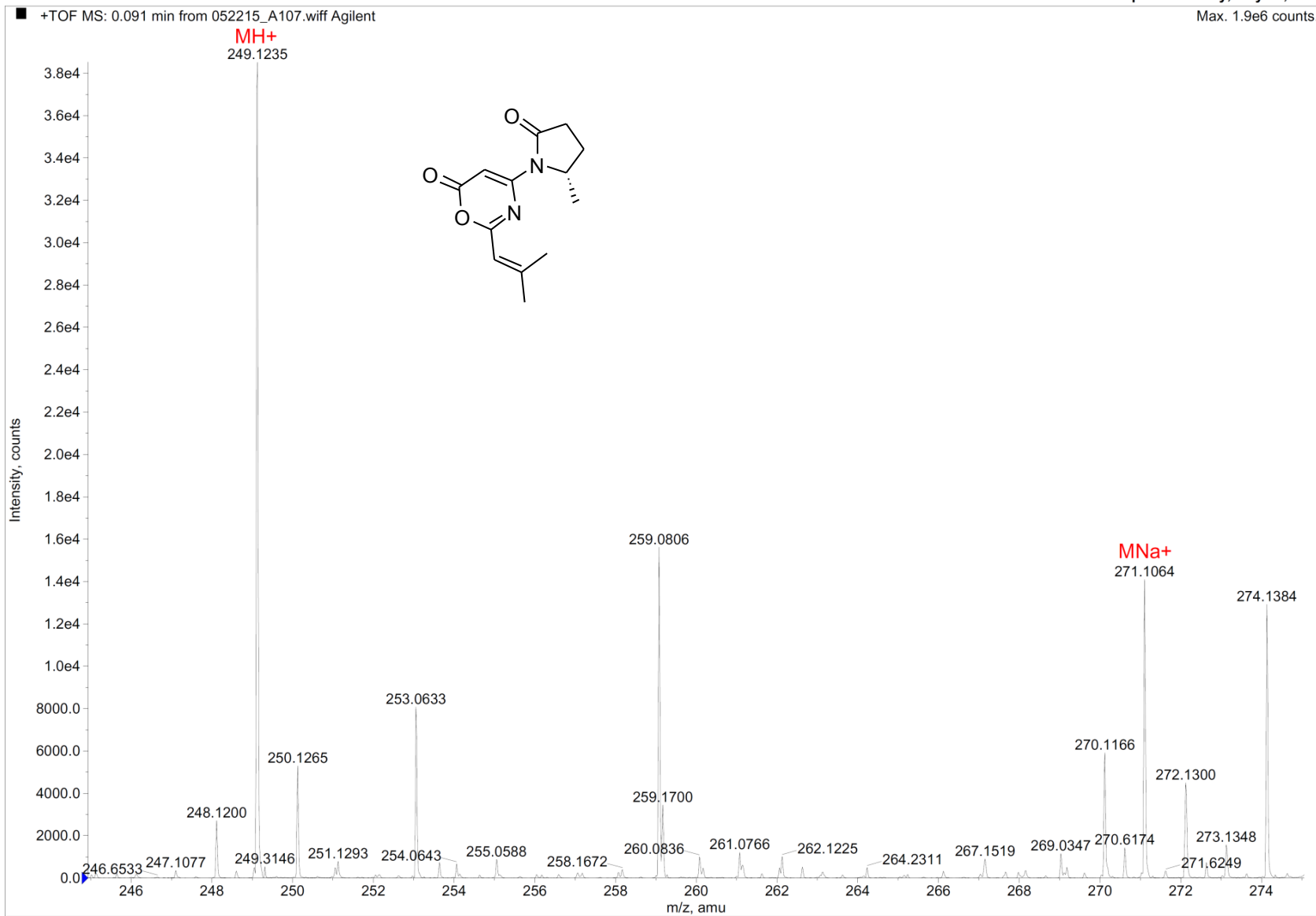


Figure S11. ¹H-NMR spectrum of spinoxazine B (2) in DMSO-*d*₆

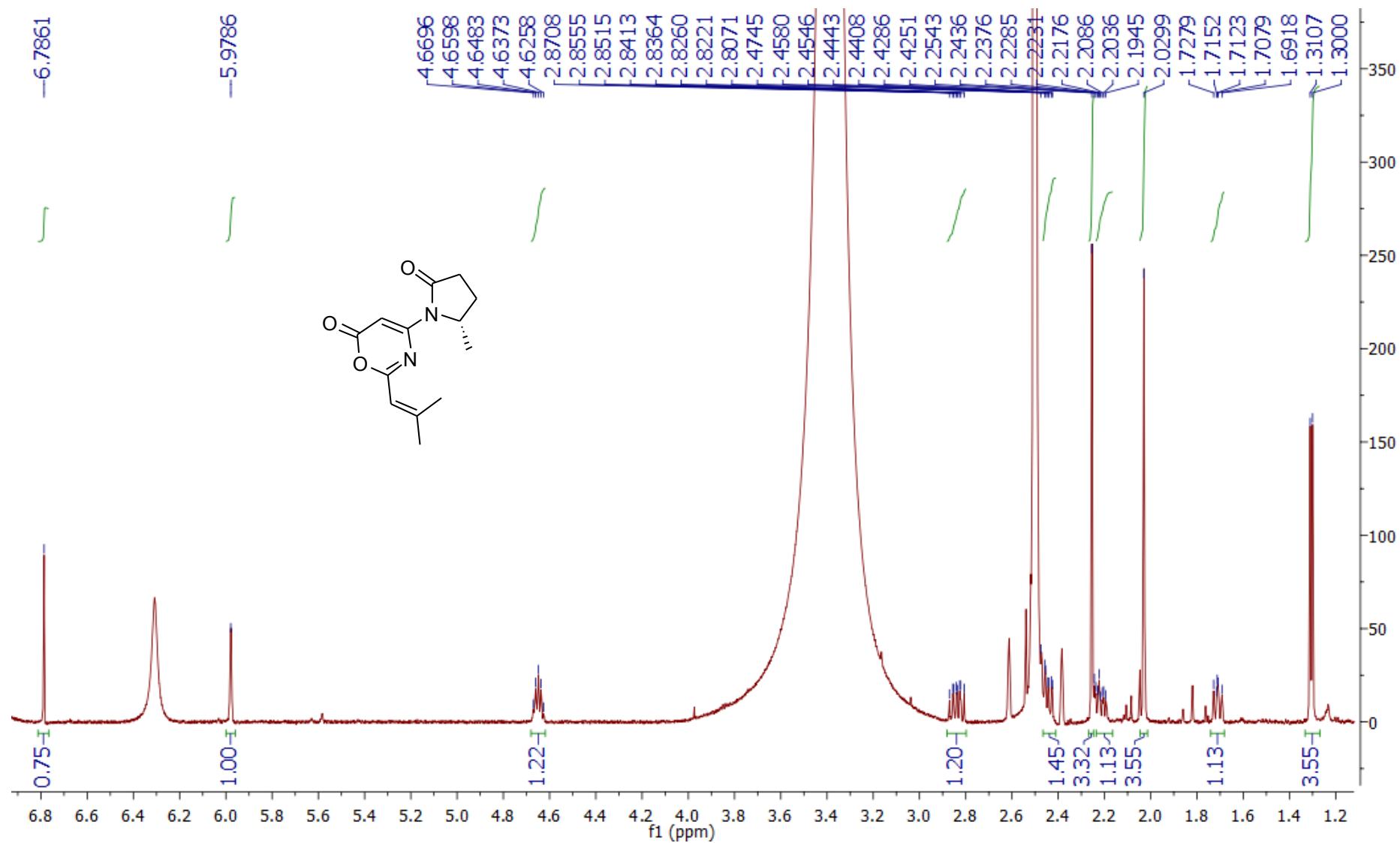


Figure S12. ^{13}C -NMR spectrum of spinoxazine B (2) in $\text{DMSO-}d_6$

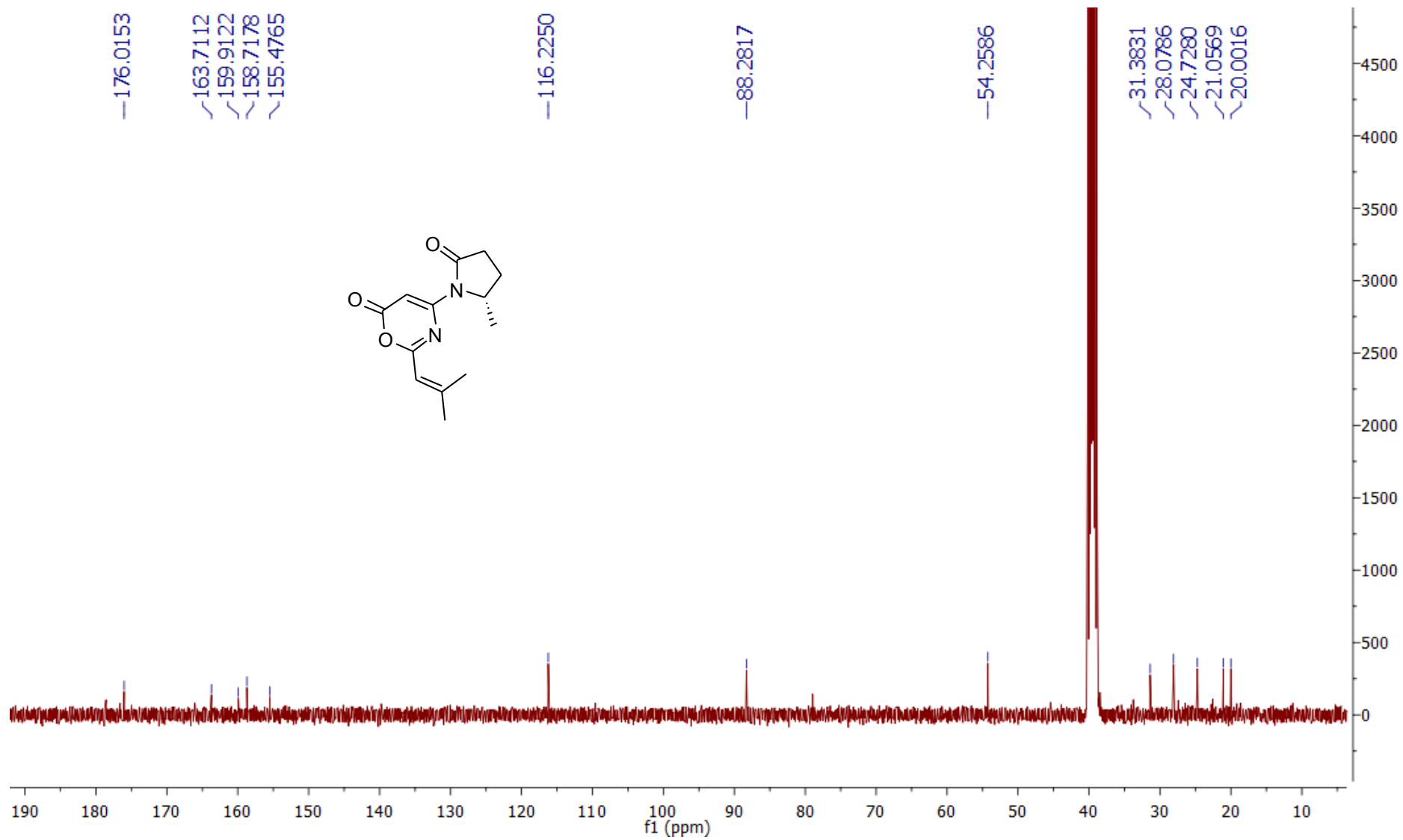


Figure S13. HSQC spectrum of spinoxazine B (2) in DMSO- d_6

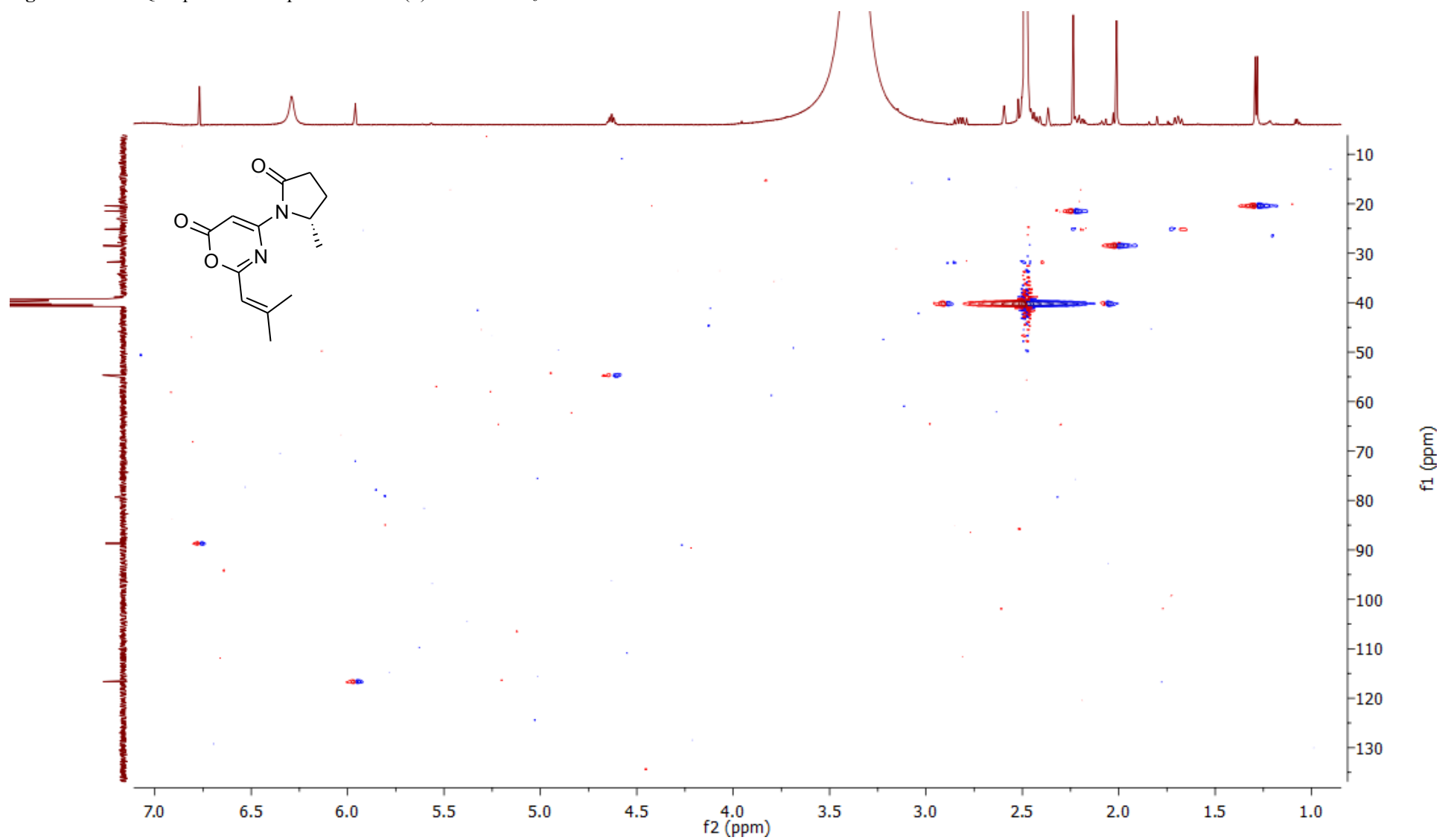


Figure S14. ^1H - ^1H COSY spectrum of spinoxazine B (2) in $\text{DMSO-}d_6$

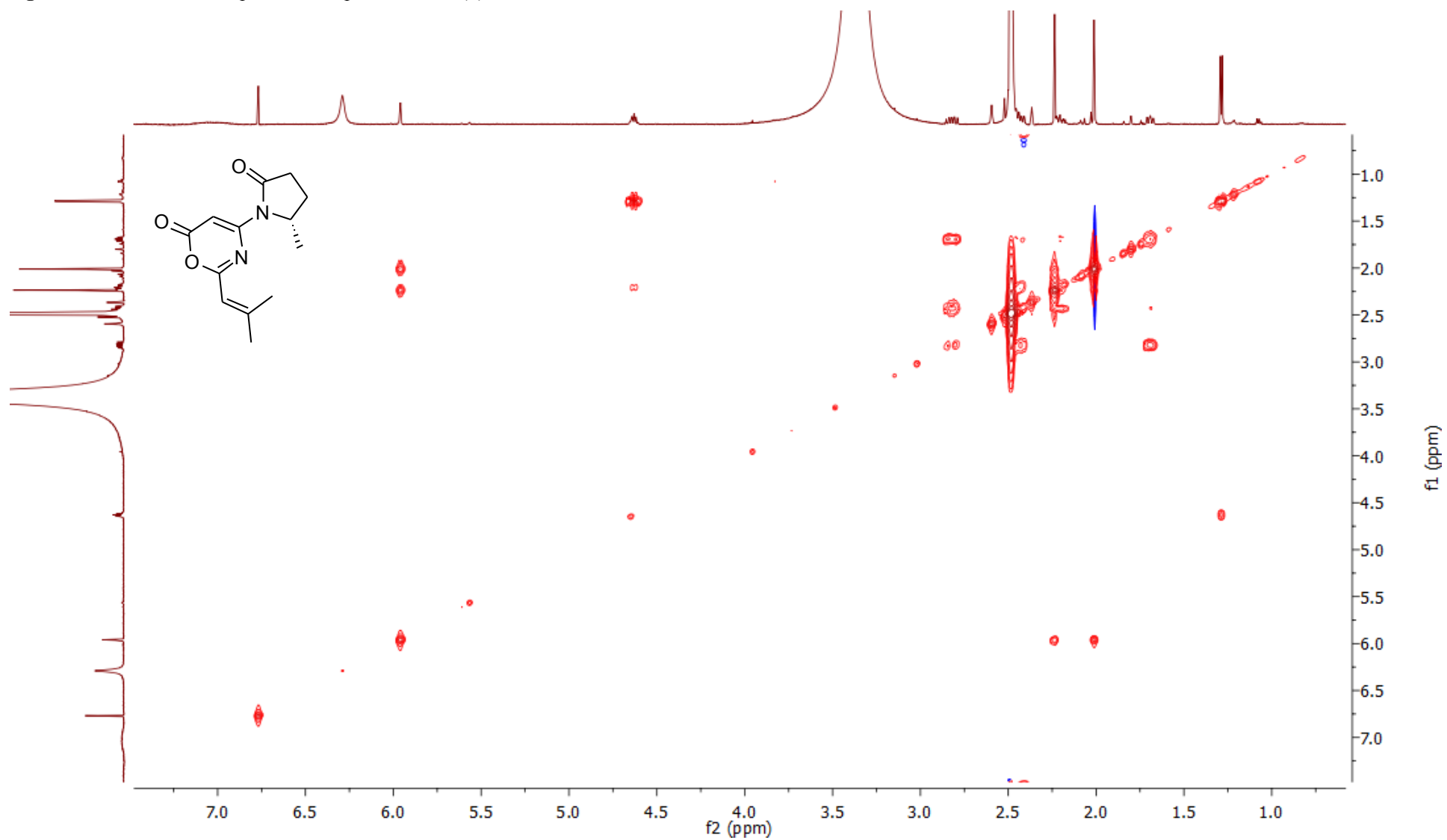


Figure S15. HMBC spectrum of spinoxazine B (2) in DMSO- d_6

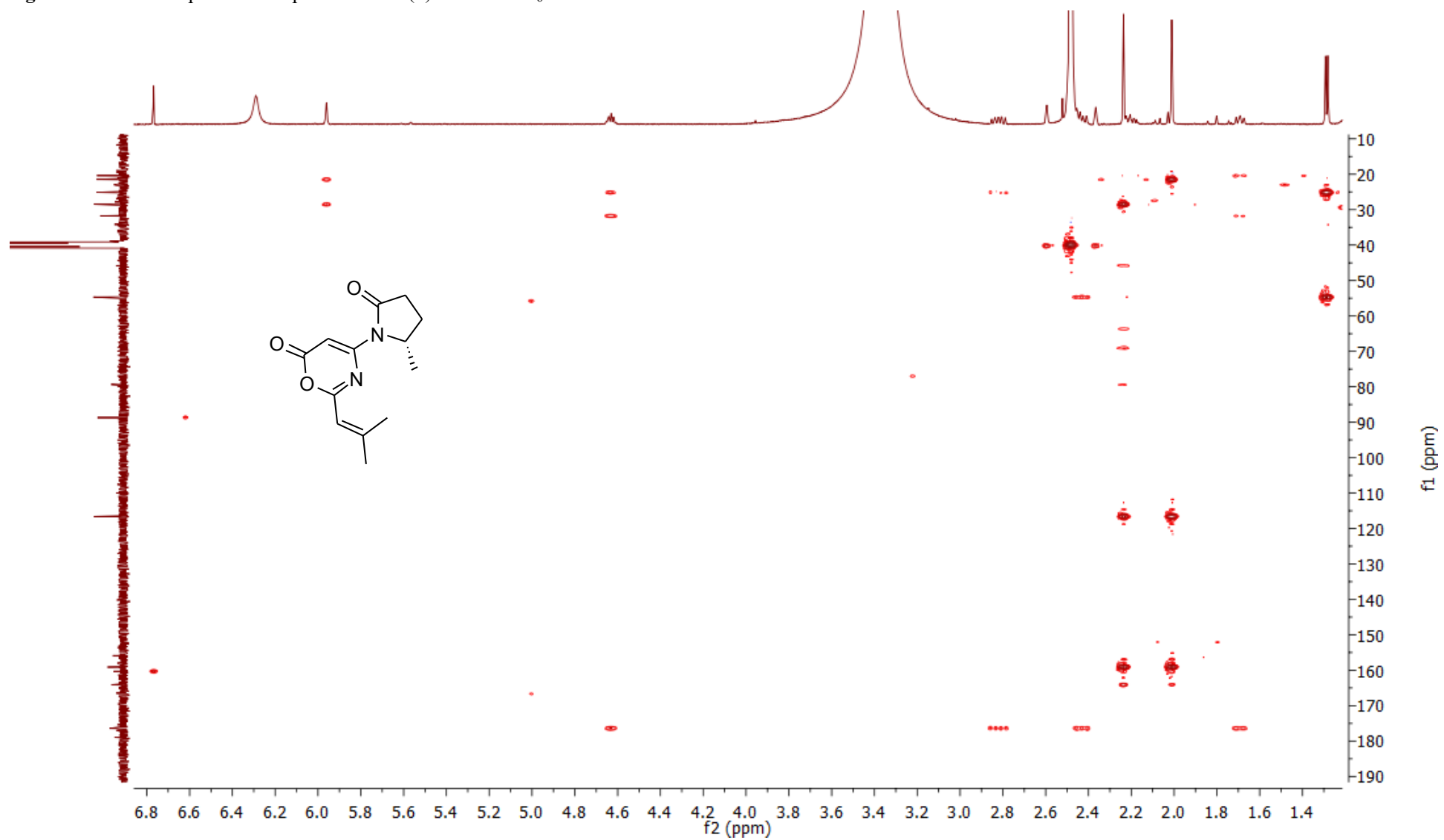


Figure S16. HRESIMS spectrum of bohemamine D (3)

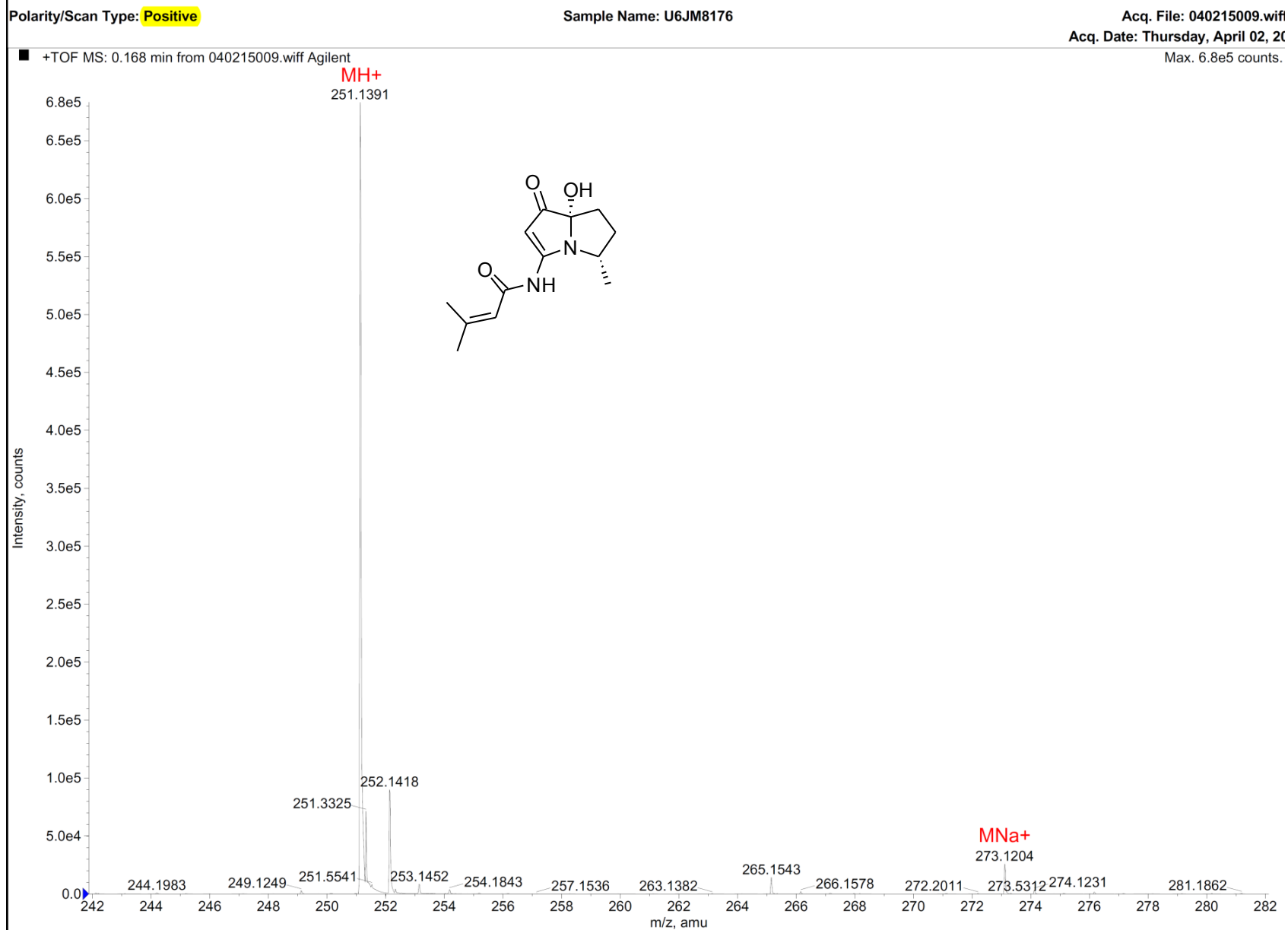


Figure S17. ¹H-NMR spectrum of bohemamine D (3) in DMSO-*d*₆

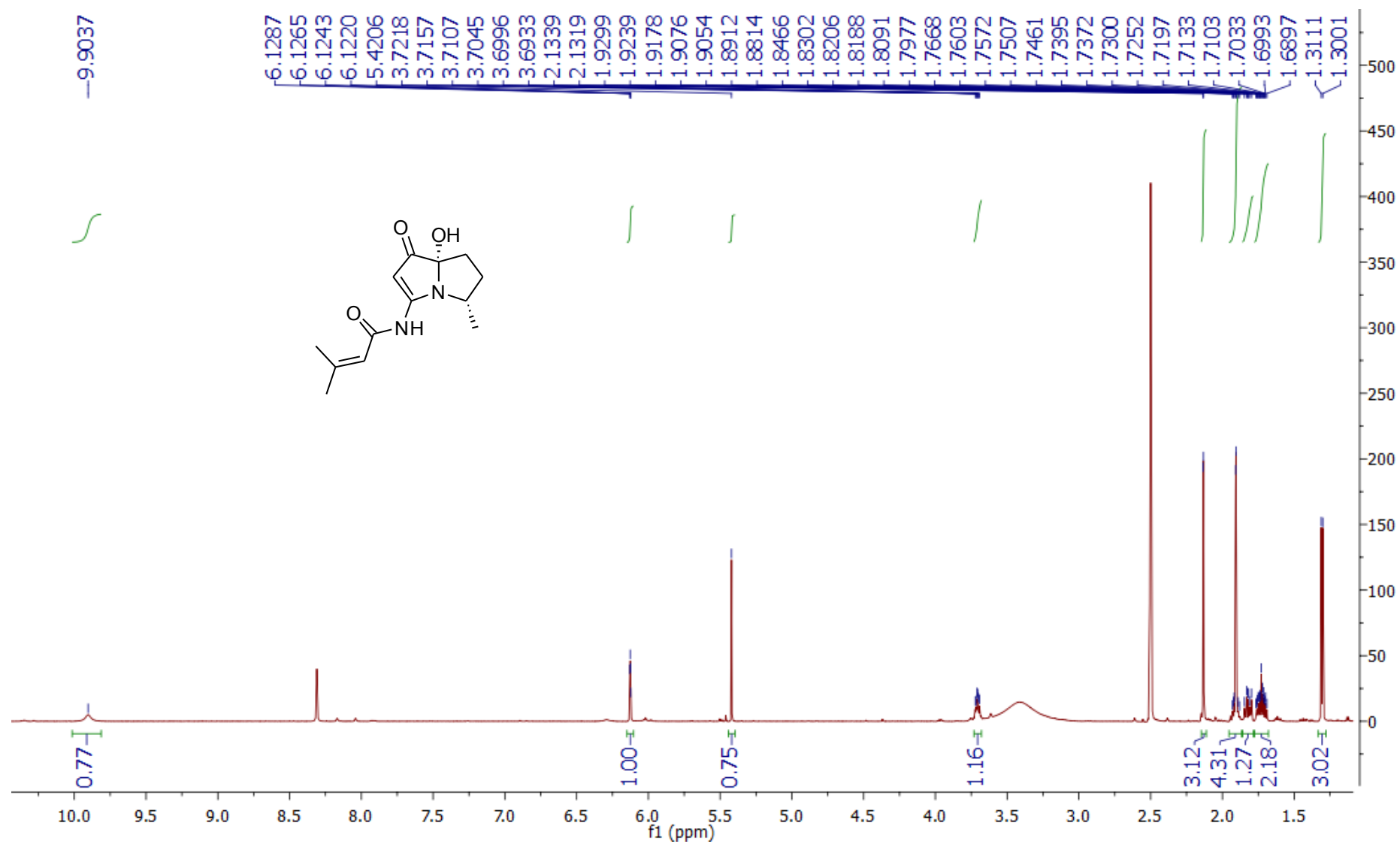


Figure S18. ^{13}C -NMR spectrum of bohemamine D (**3**) in $\text{DMSO-}d_6$

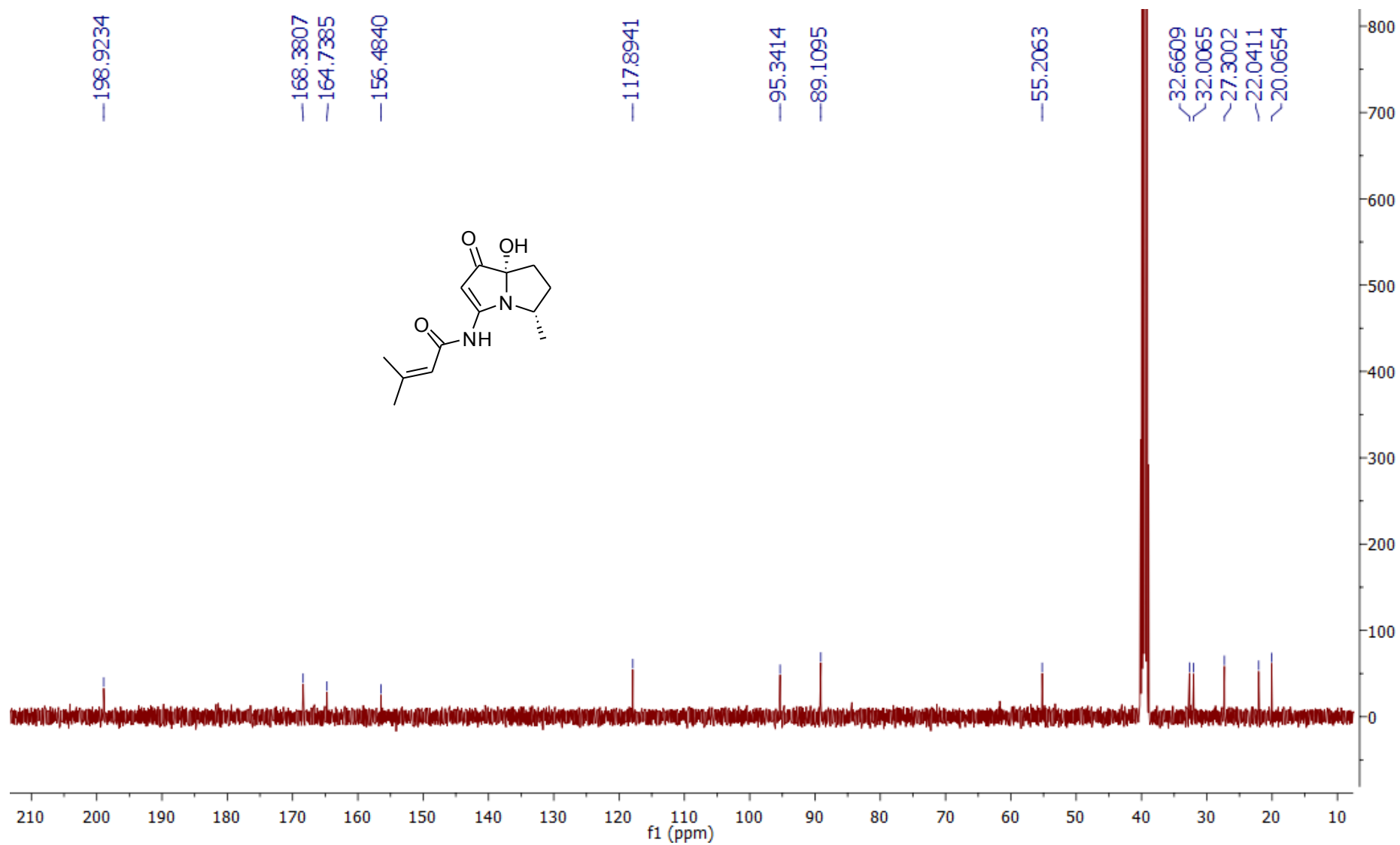


Figure S19. HSQC spectrum of bohemamine D (3) in DMSO-*d*₆

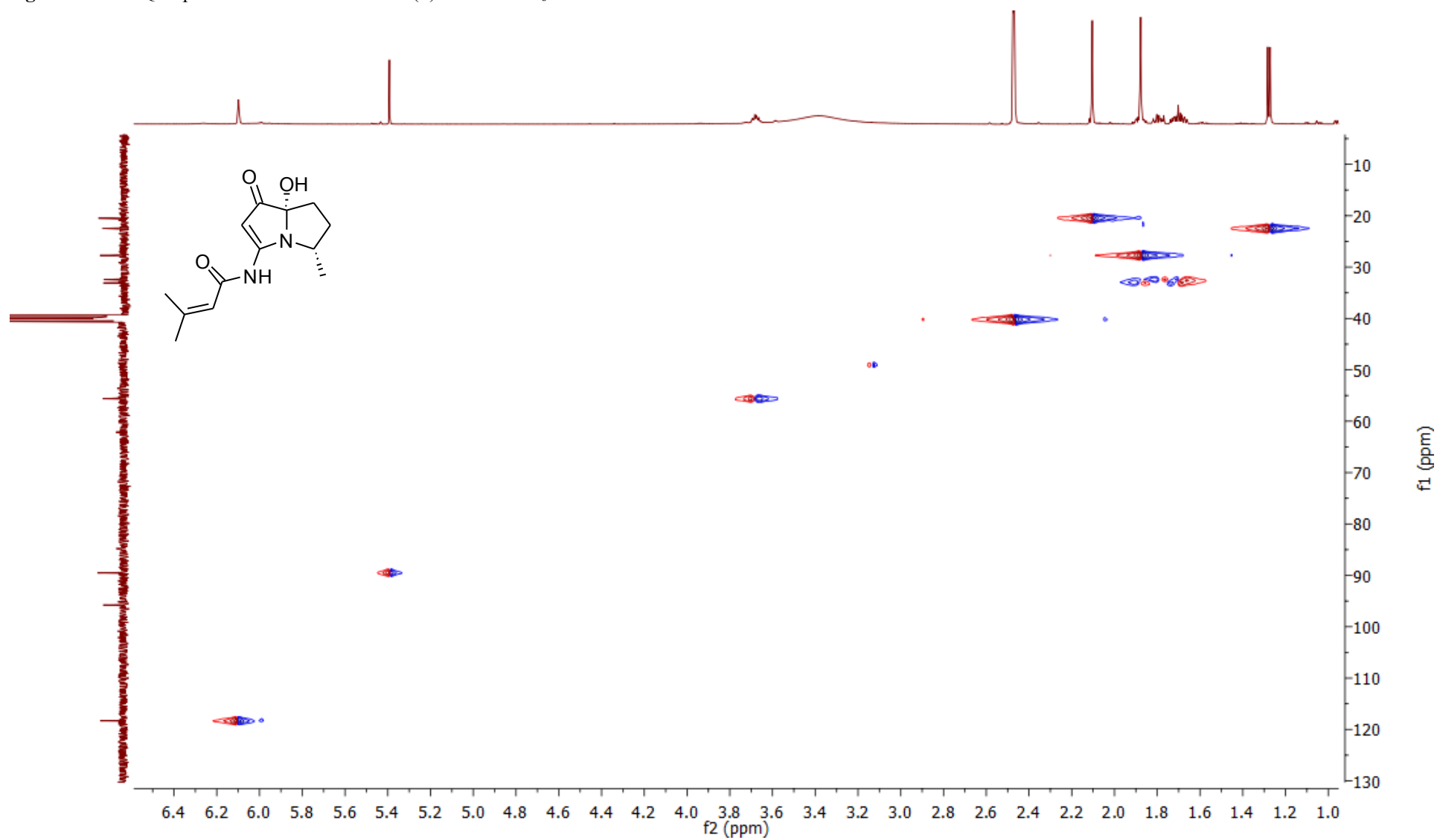


Figure S20. ^1H - ^1H COSY spectrum of bohemamine D (**3**) in $\text{DMSO-}d_6$

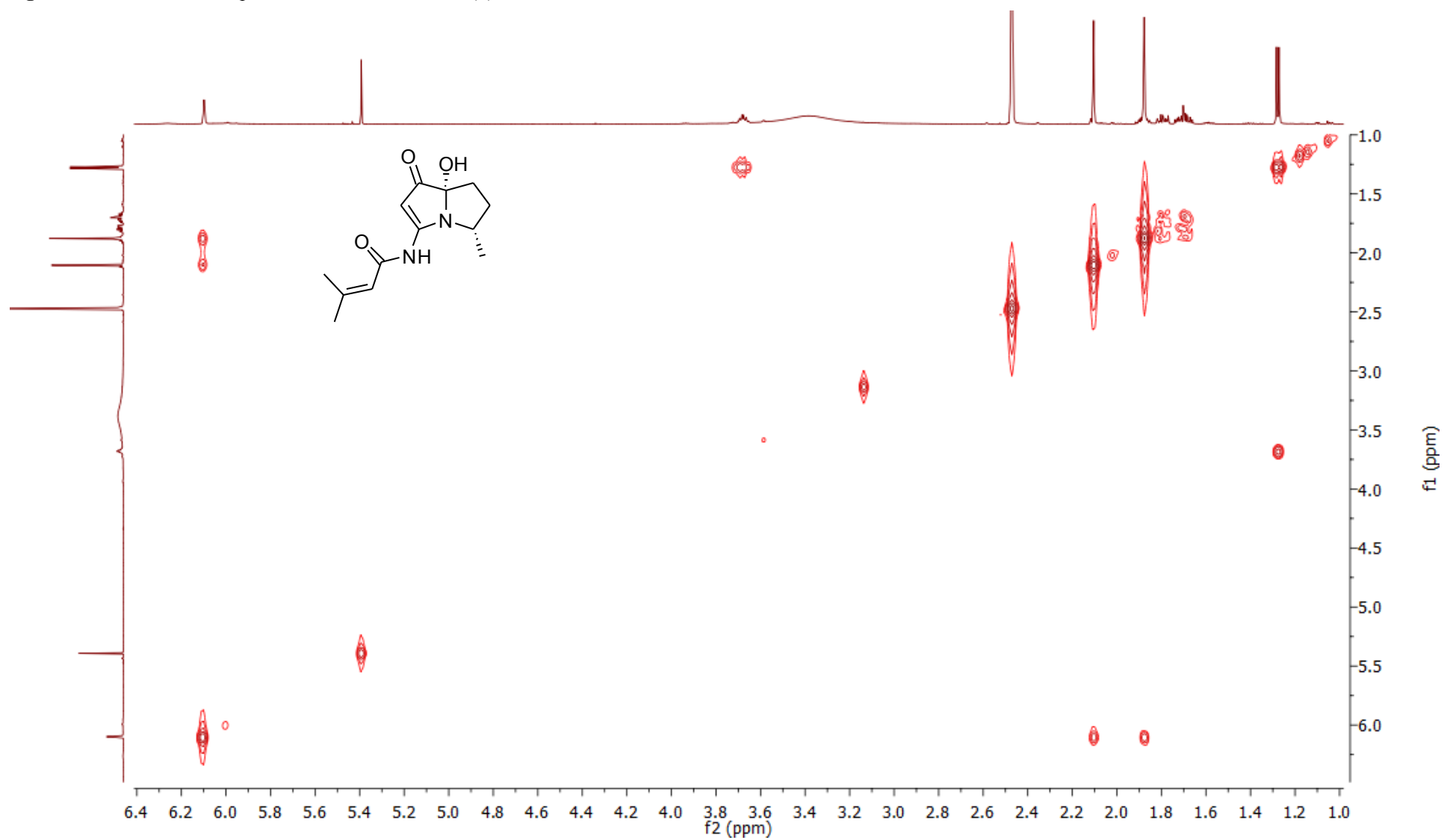


Figure S21. HMBC spectrum of bohemamine D (3) in DMSO- d_6

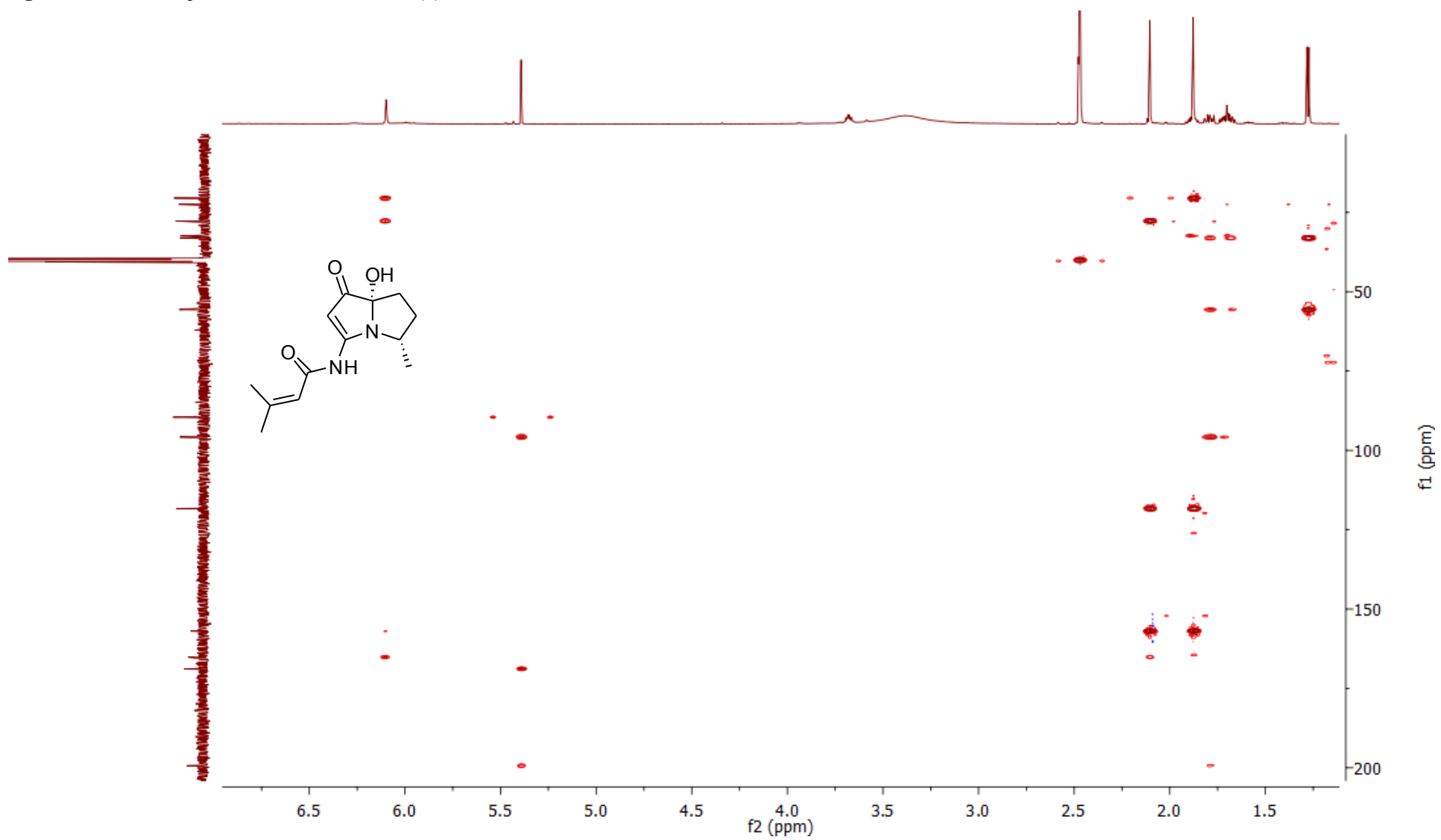


Figure S22. HRESIMS spectrum of compound 3a

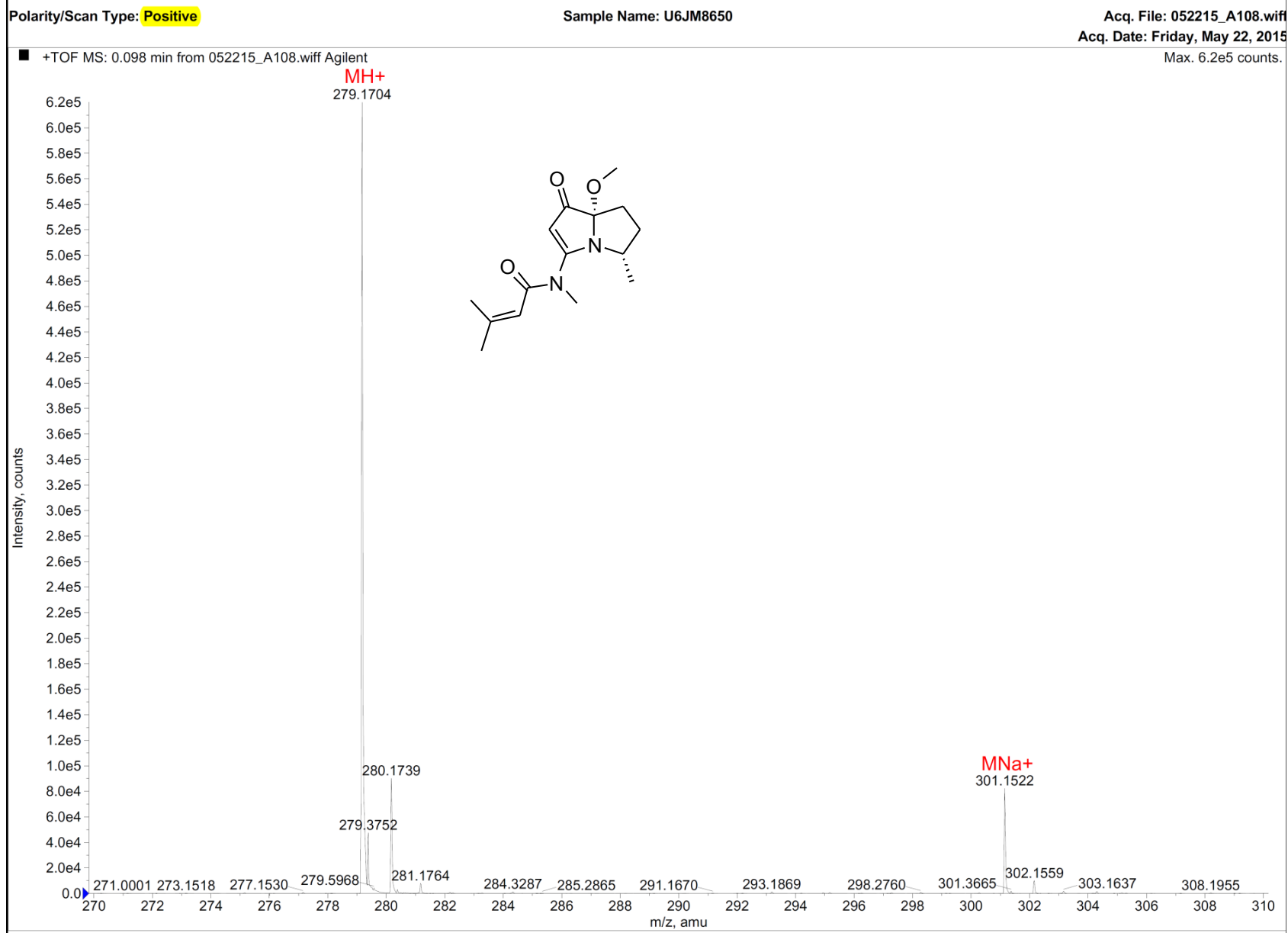


Figure S23. ¹H-NMR spectrum of compound **3a** in CDCl₃

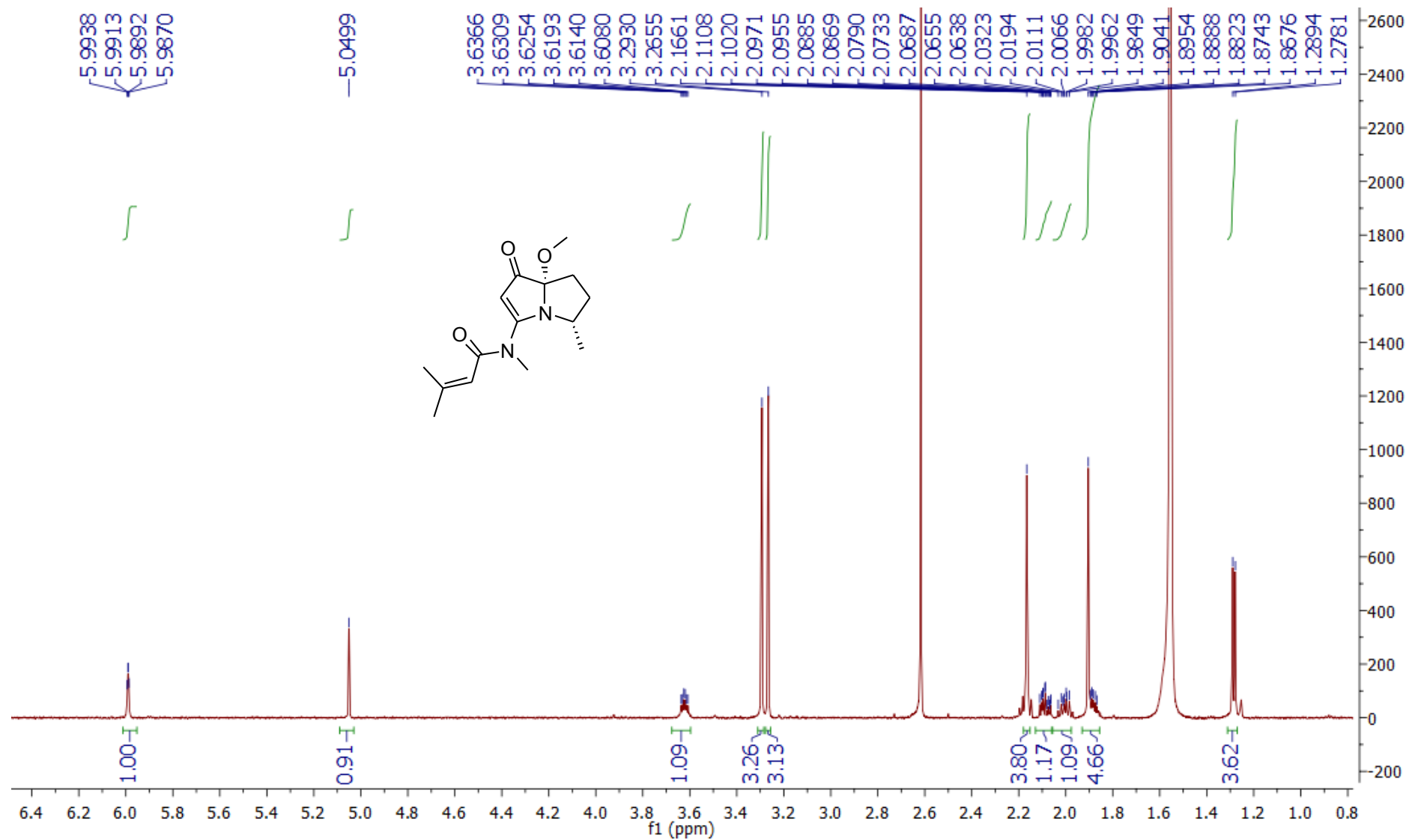


Figure S24. HSQC spectrum of compound **3a** in CDCl₃

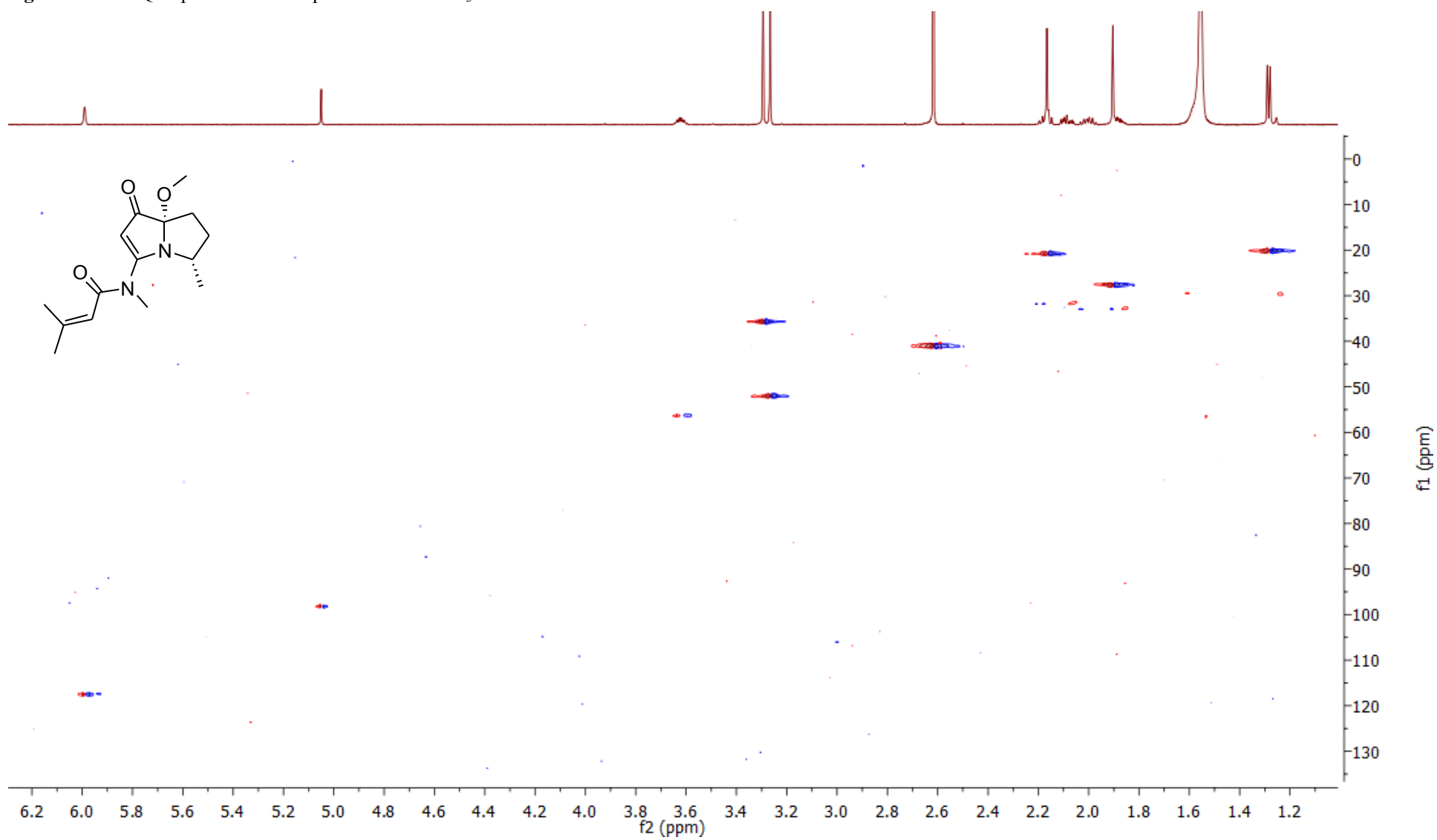


Figure S25. HMBC spectrum of compound **3a** in CDCl₃

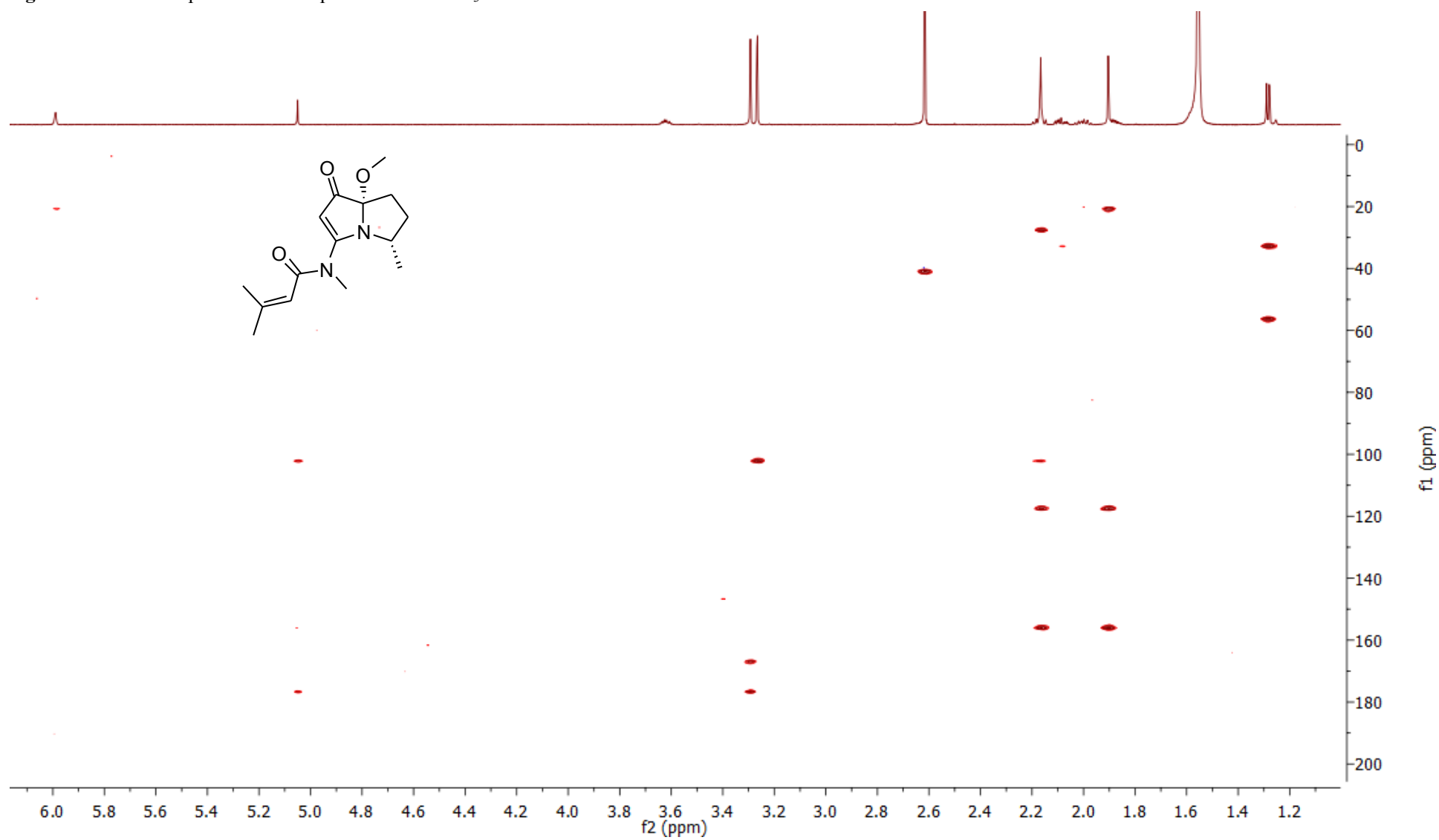


Figure S26. NOESY spectrum of compound **3a** in CDCl₃

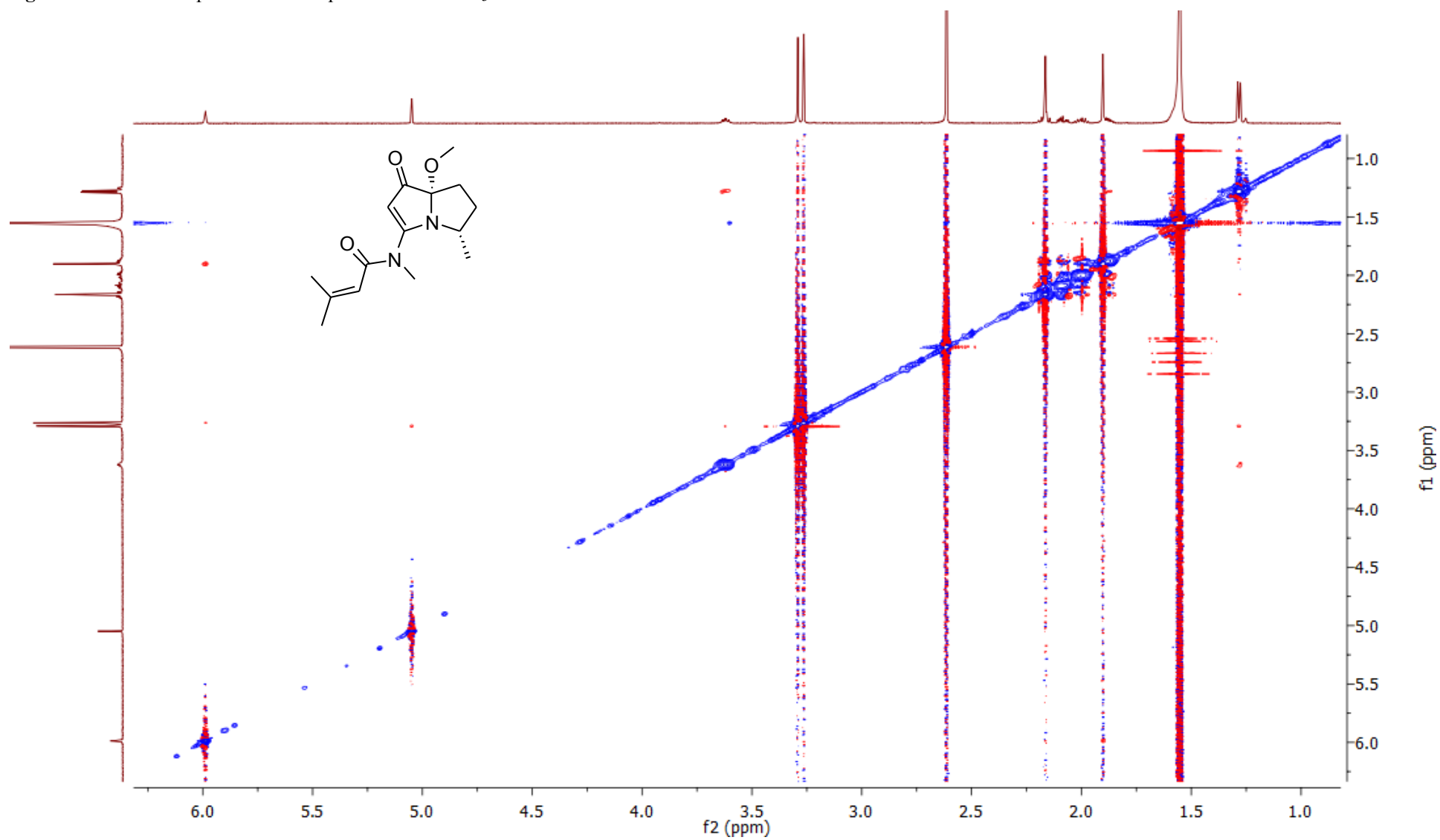


Figure S27. HRESIMS spectrum of bohemamine E (4)

Polarity/Scan Type: **Positive**

Sample Name: U6JM8175

Acq. File: 040215008.wiff
Acq. Date: Thursday, April 02, 20

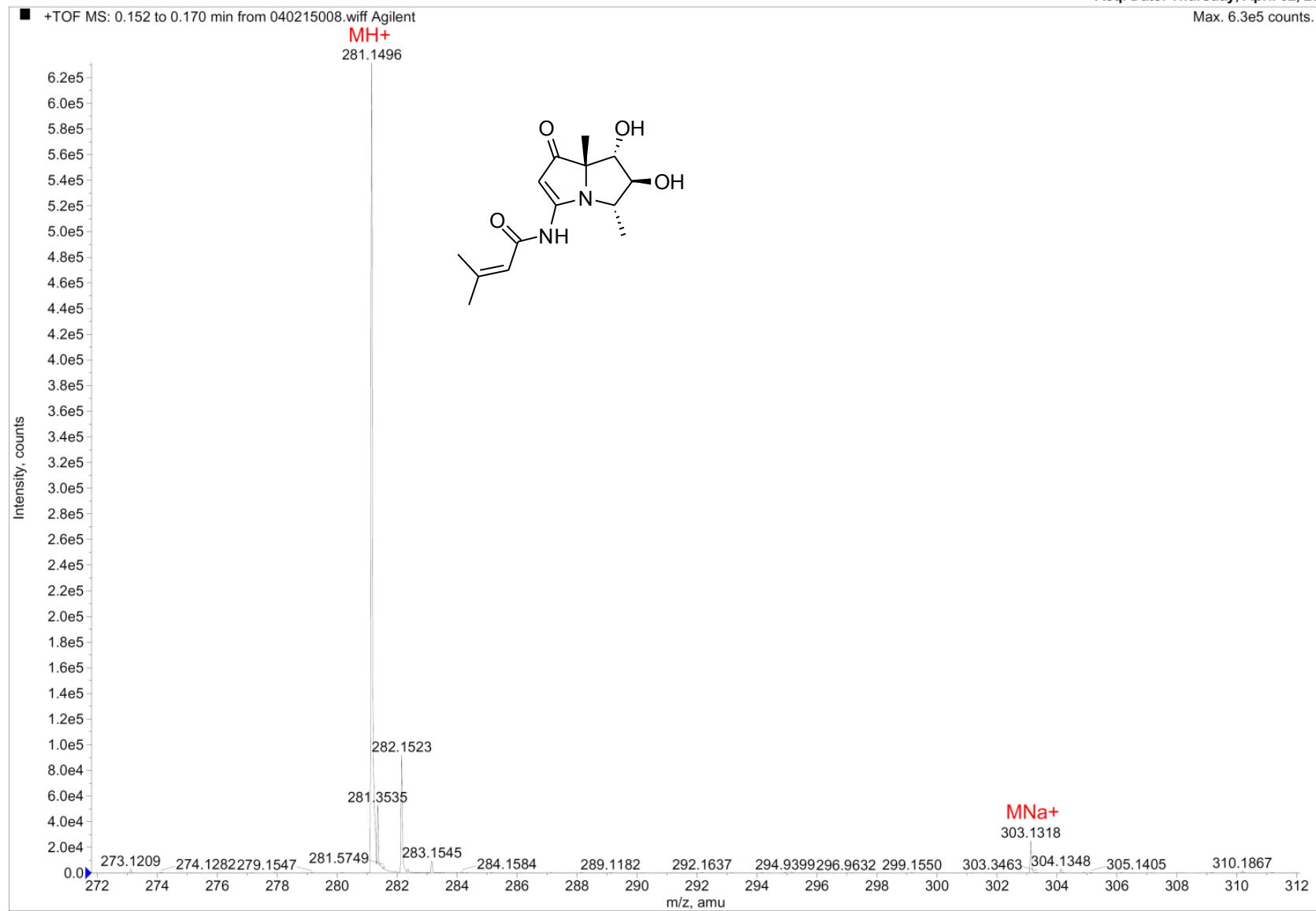


Figure S28. ¹H-NMR spectrum of bohemamine E (4) in DMSO-*d*₆

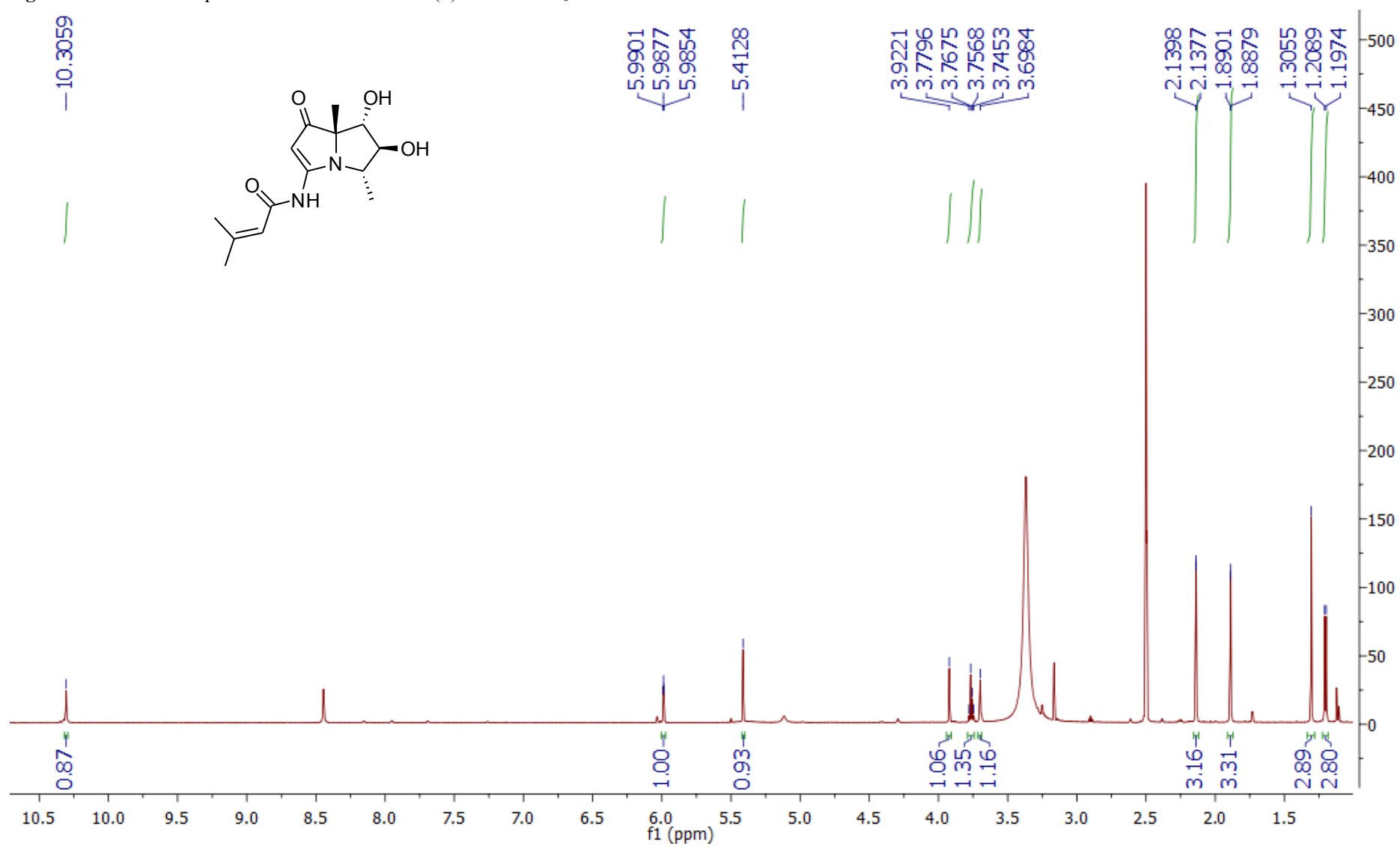


Figure S29. ^{13}C -NMR spectrum of bohemamine E (**4**) in $\text{DMSO-}d_6$

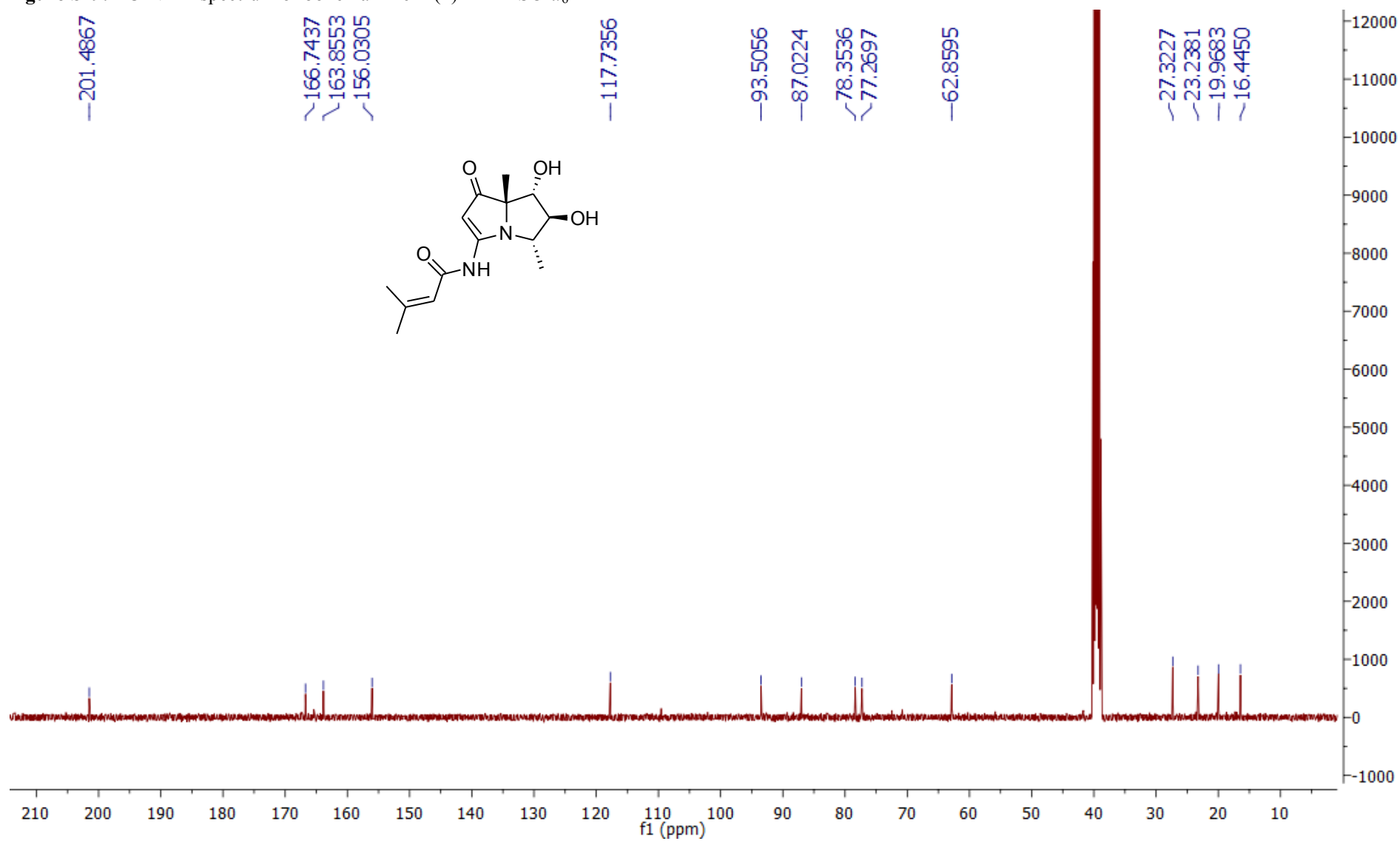


Figure S30. ^1H - ^1H COSY spectrum of bohemamine E (**4**) in $\text{DMSO-}d_6$

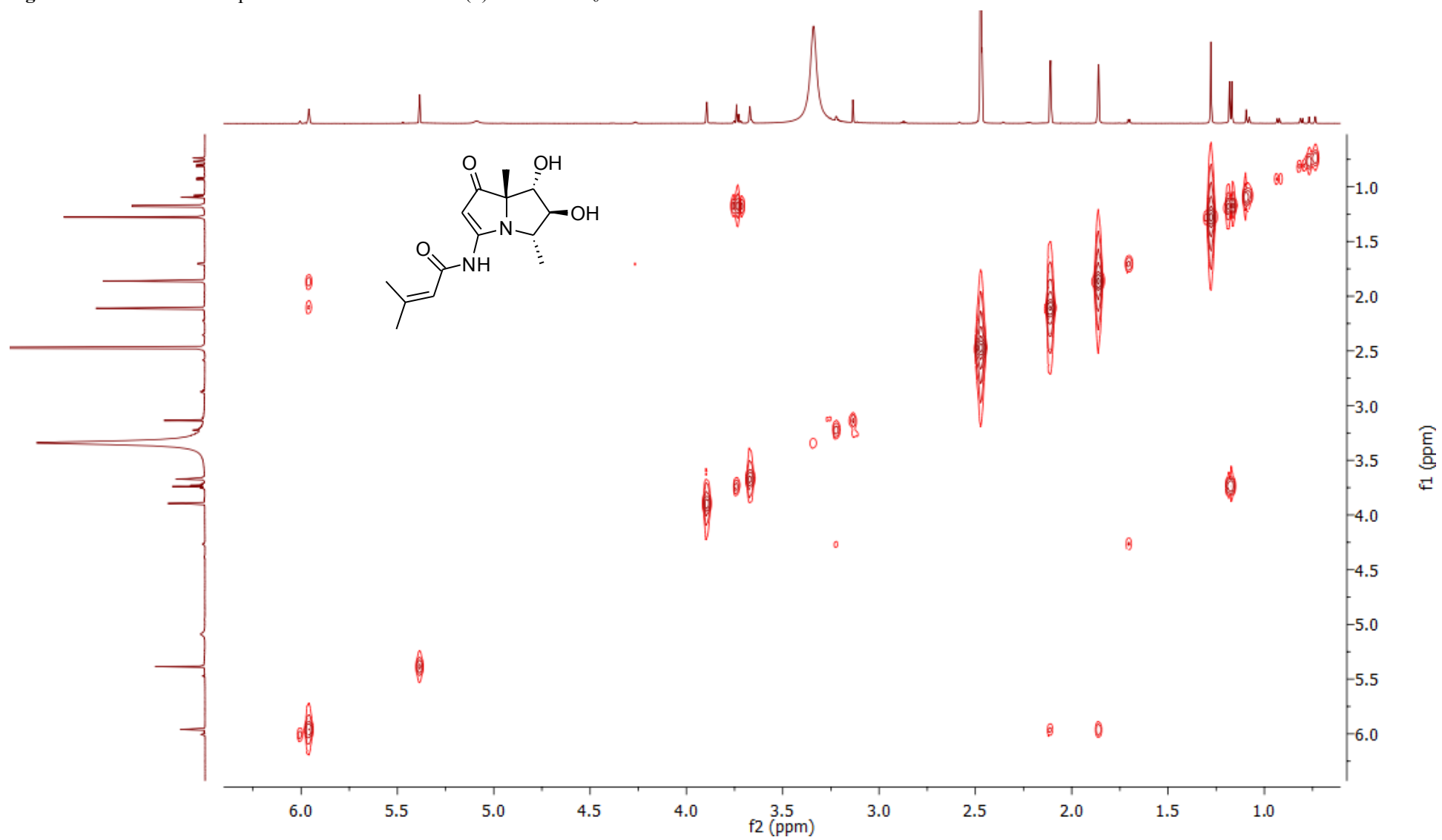


Figure S31. HSQC spectrum of bohemamine E (4) in DMSO- d_6

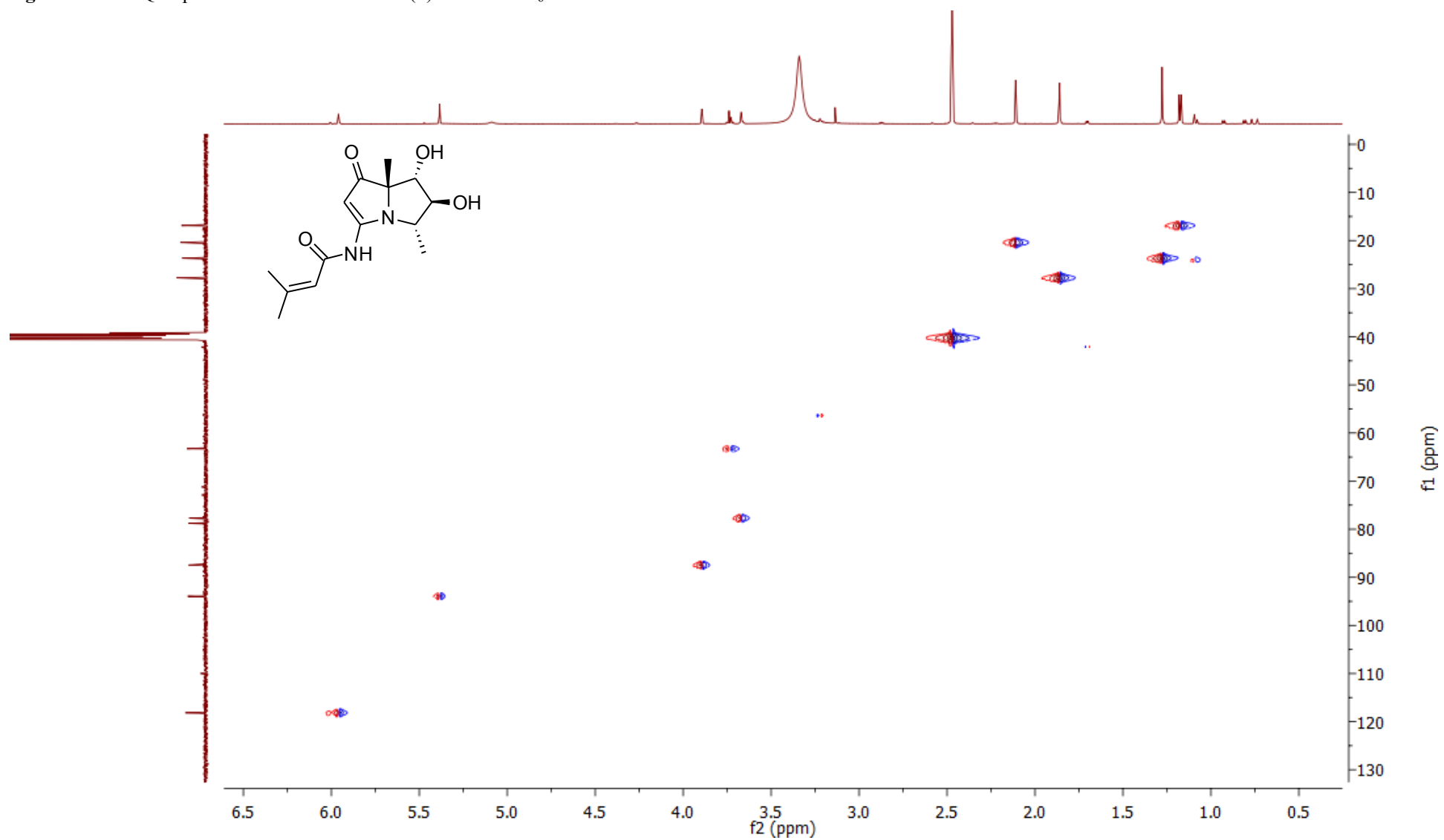


Figure S32. HMBC spectrum of bohemamine E (4) in DMSO- d_6

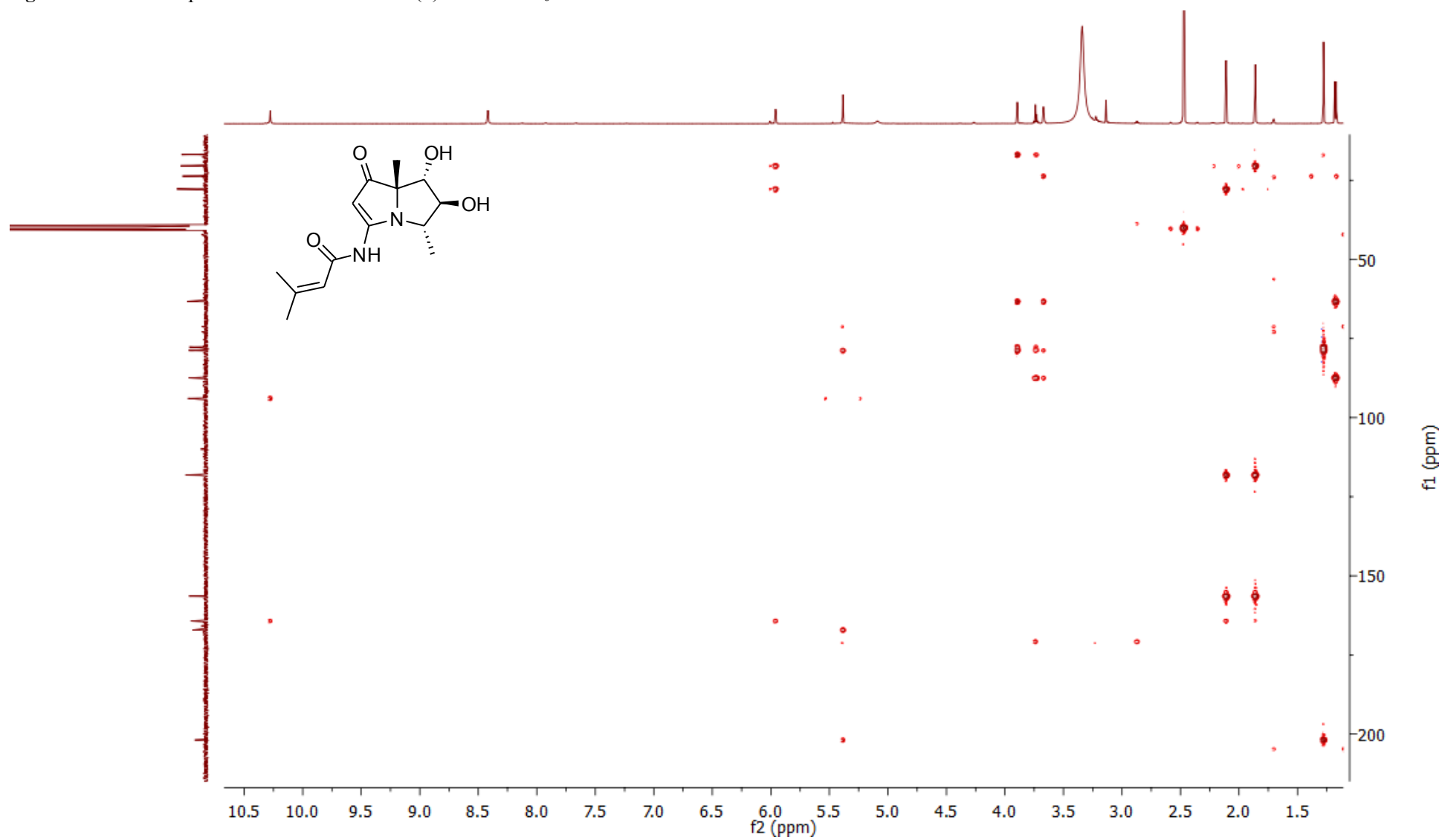


Figure S33. NOESY spectrum of bohemamine E (4) in DMSO-*d*₆

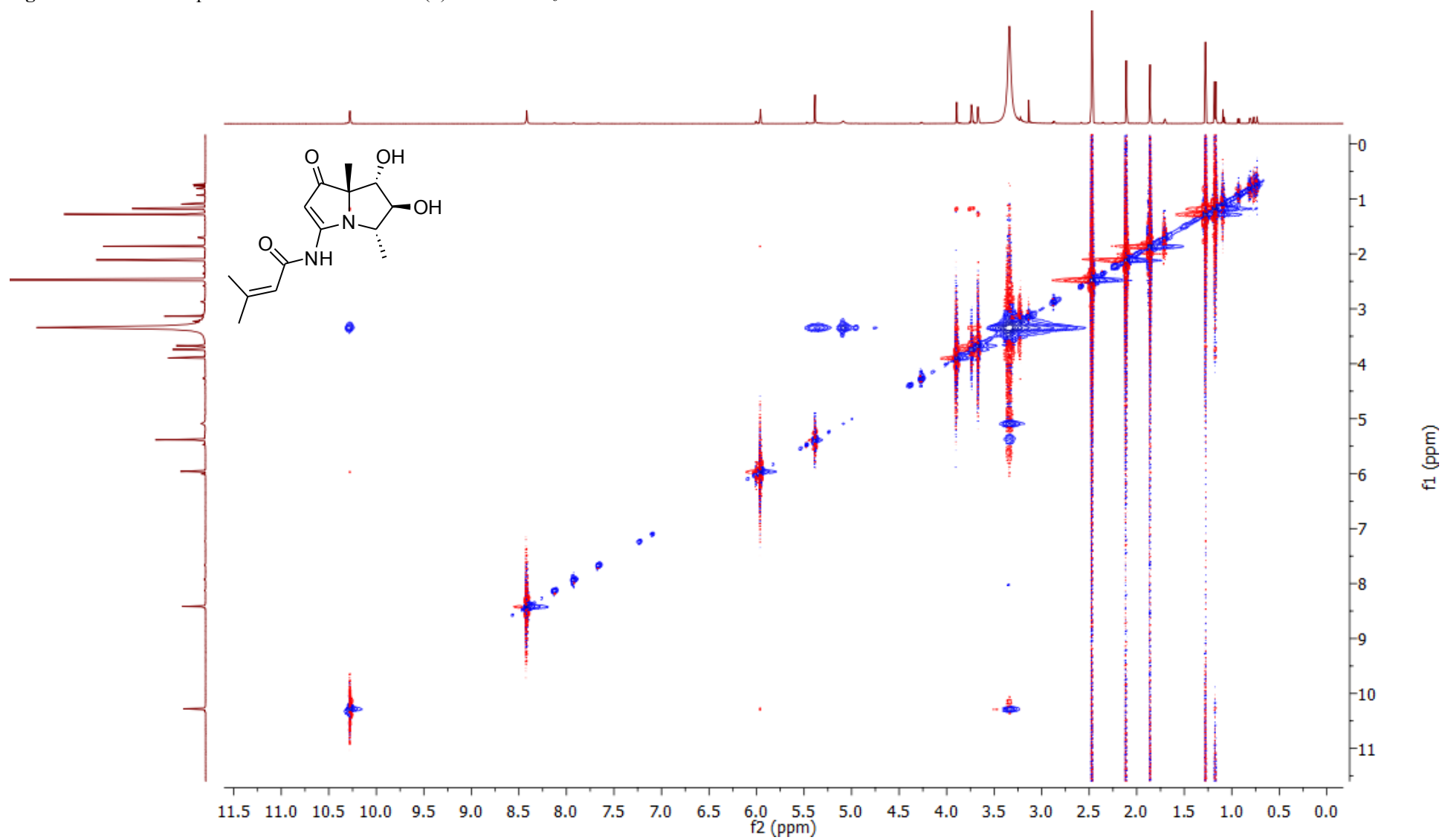


Figure S34. HRESIMS spectrum of bohemamine F (5)

Polarity/Scan Type: **Positive**

Sample Name: U6JM8179

Acq. File: 040215012.wif
Acq. Date: Thursday, April 02, 21

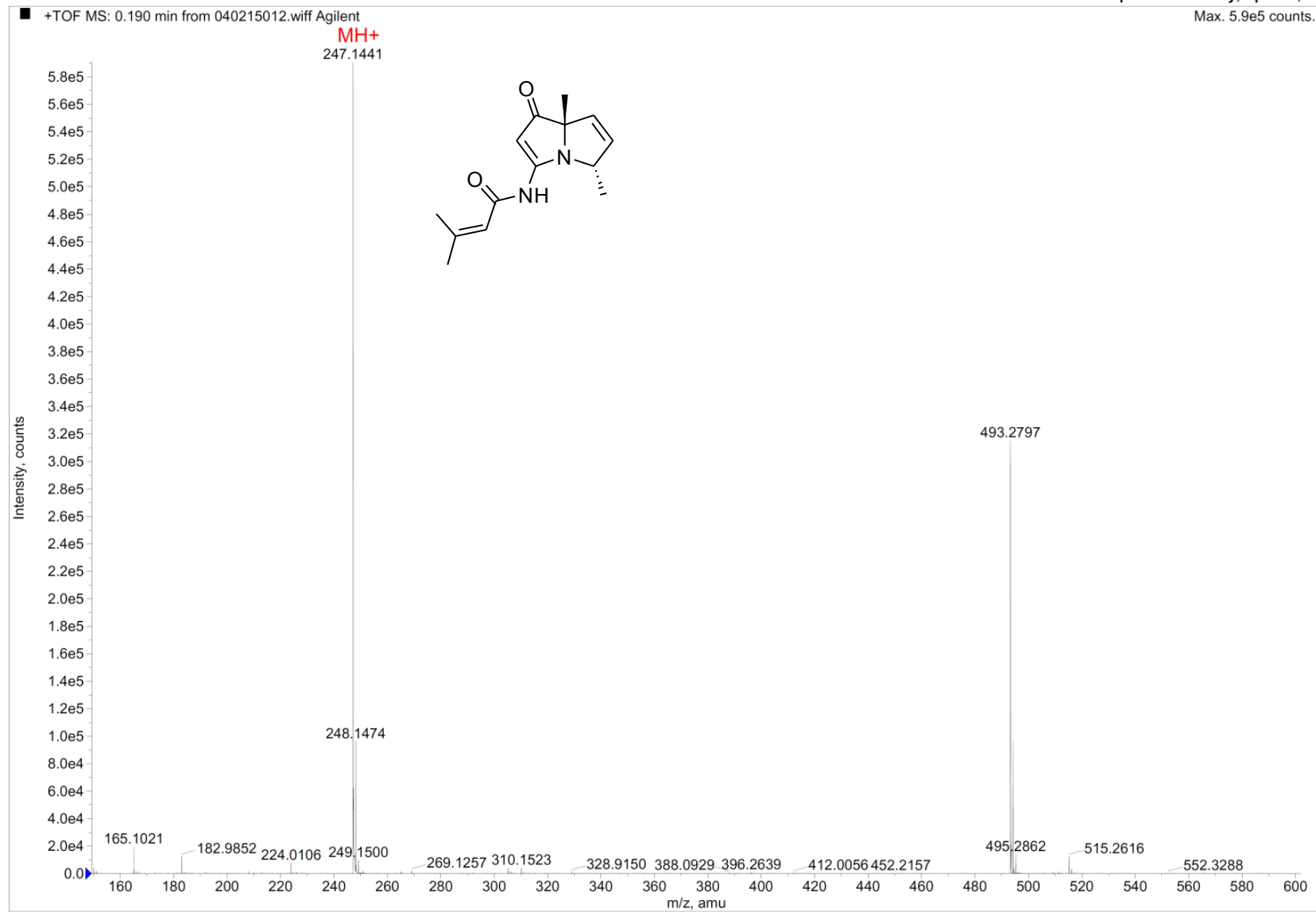


Figure S35. ¹H-NMR spectrum of bohemamine F (5) in DMSO-*d*₆

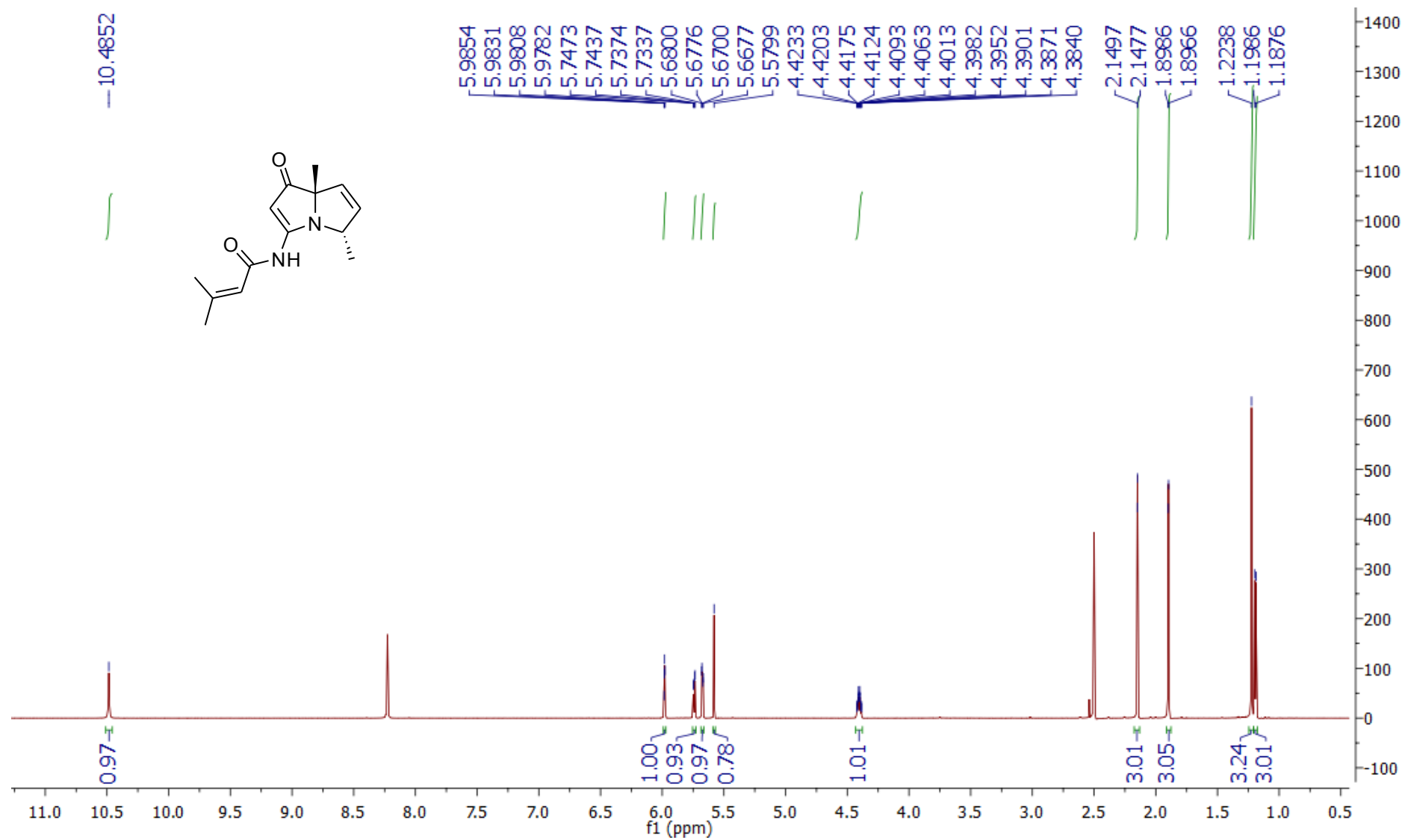


Figure S36. ^{13}C -NMR spectrum of bohemamine F (**5**) in $\text{DMSO-}d_6$

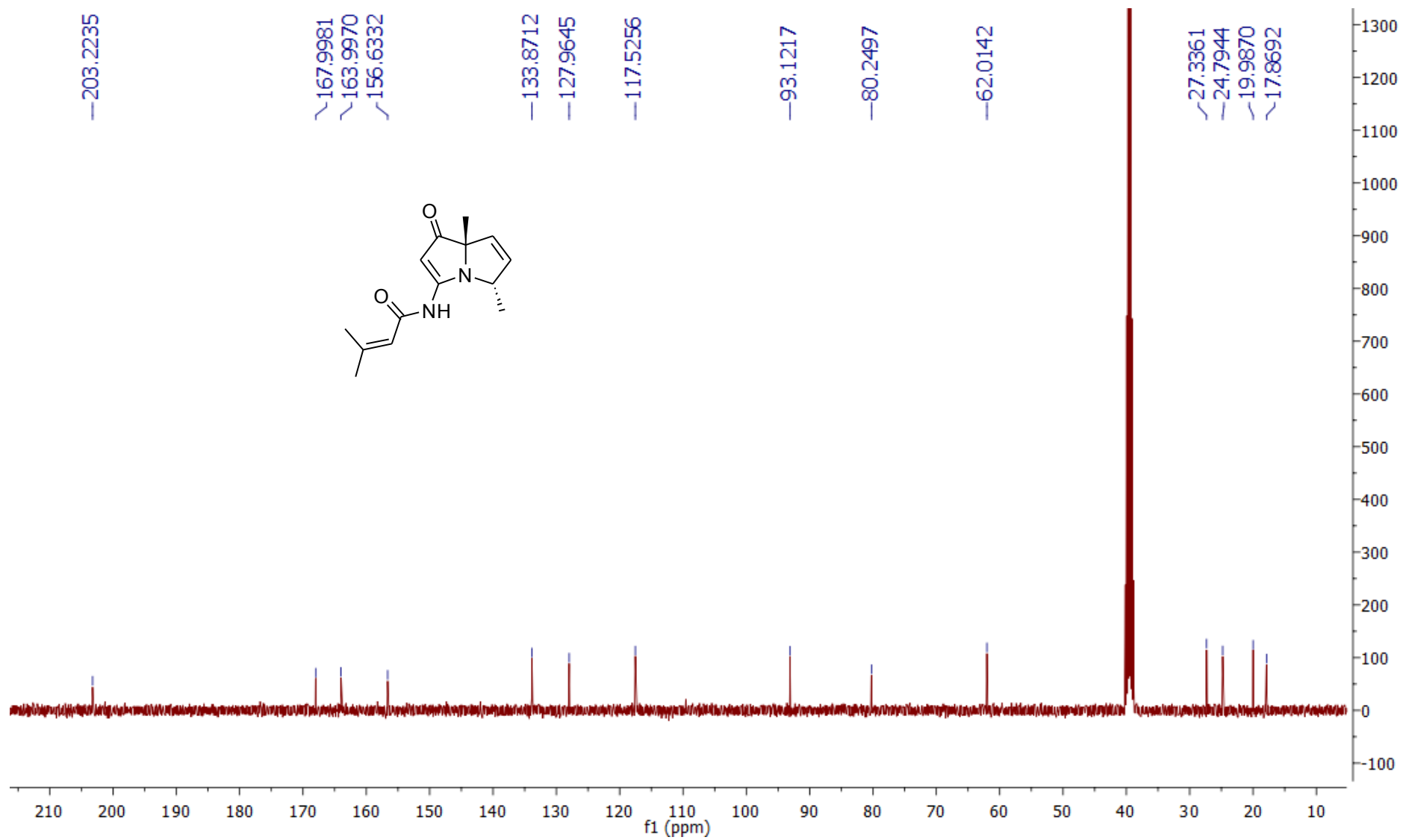


Figure S37. ^1H - ^1H COSY spectrum of bohemamine F (5) in $\text{DMSO-}d_6$

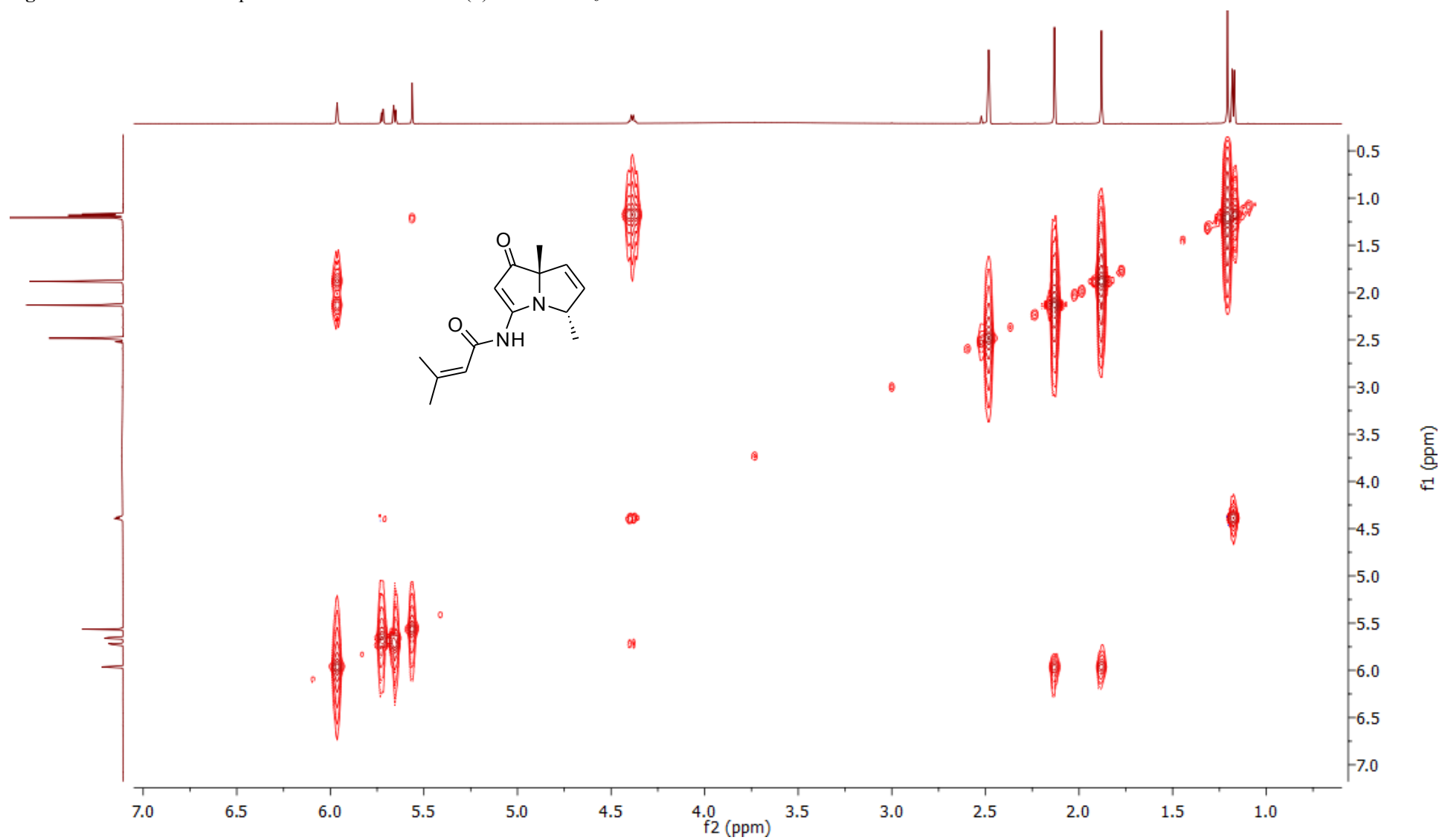


Figure S38. HSQC spectrum of bohemamine F (5) in DMSO- d_6

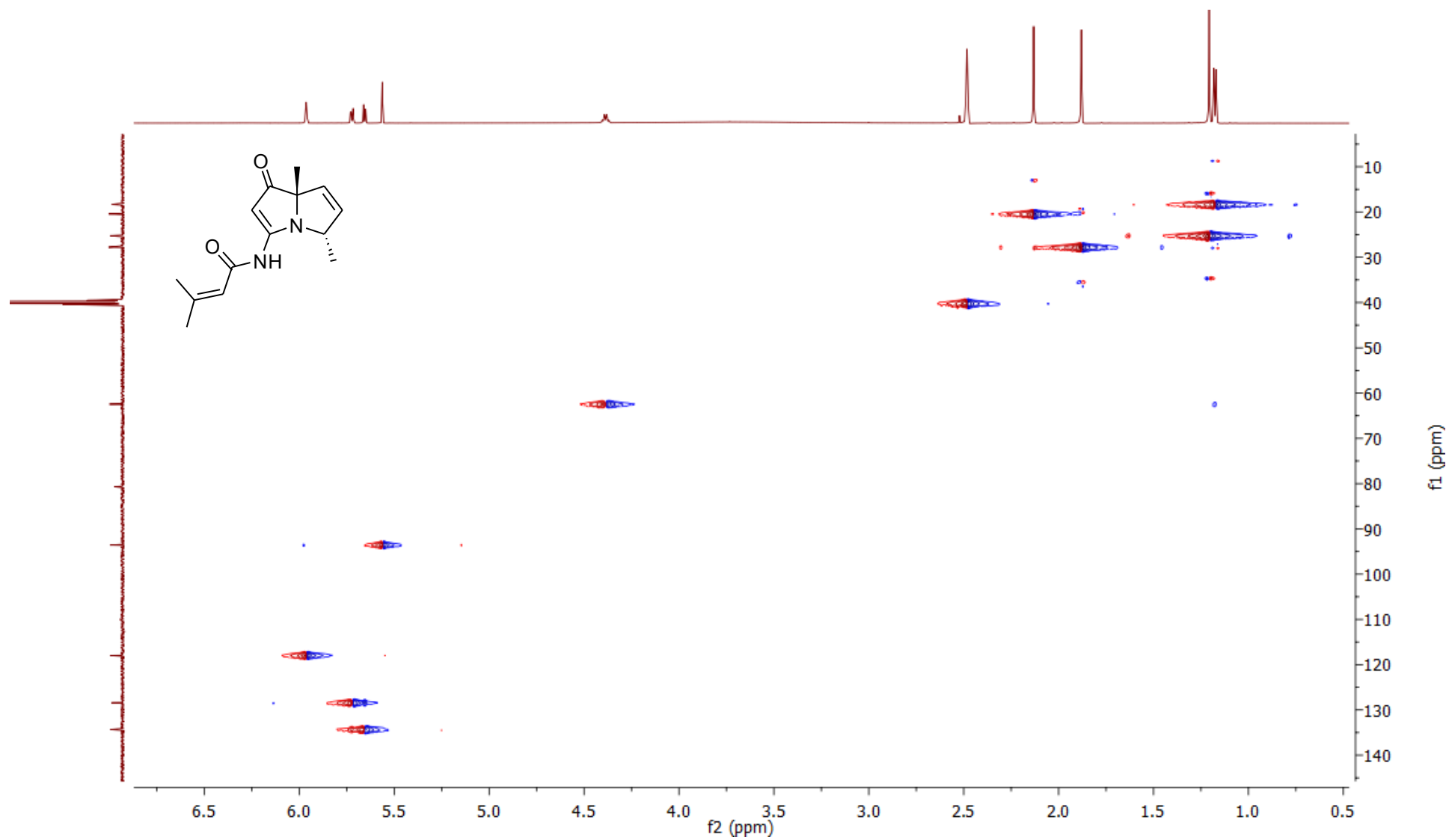


Figure S39. HMBC spectrum of bohemamine F (5) in DMSO-*d*₆

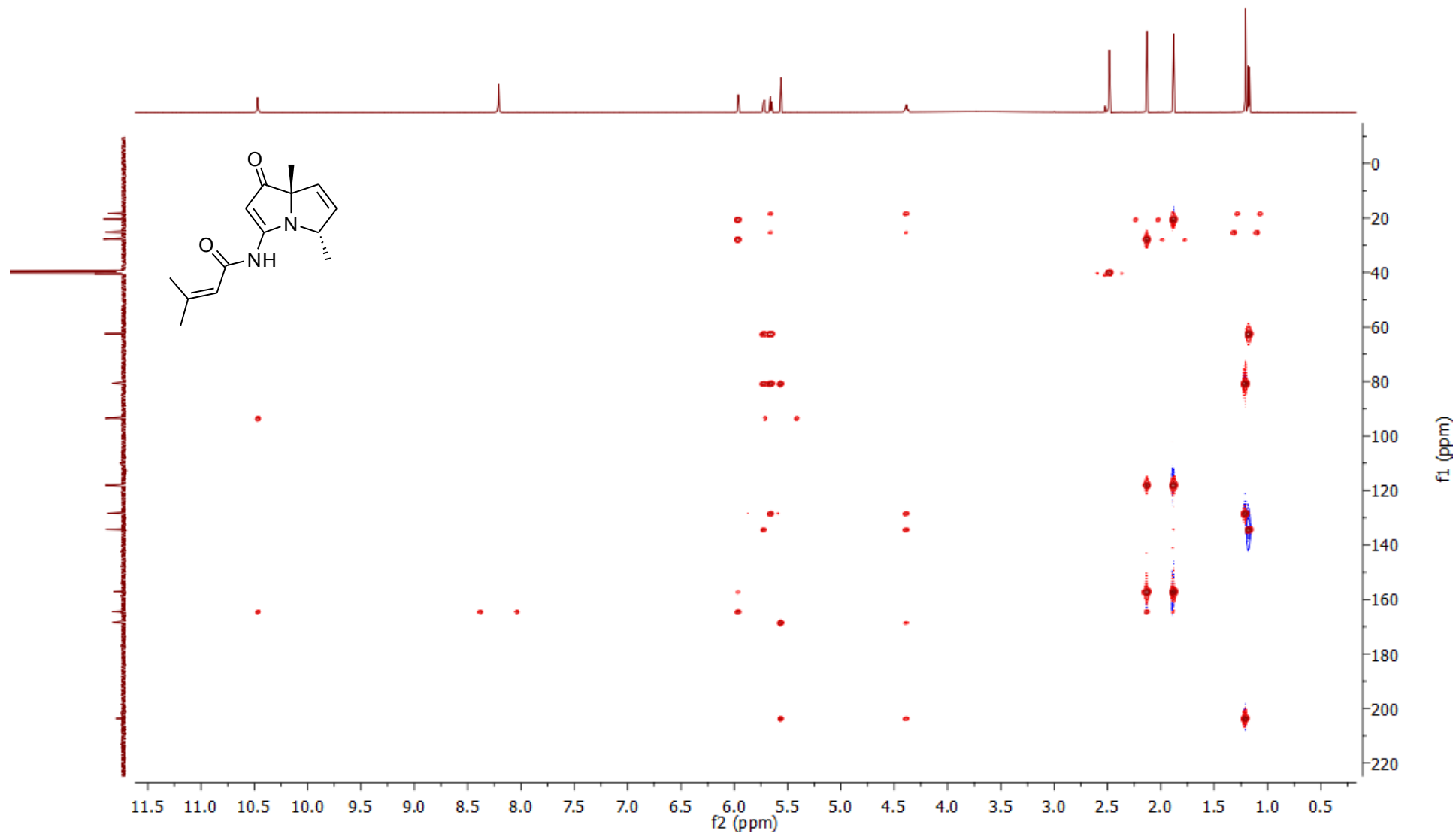


Figure S40. NOESY spectrum of bohemamine F (**5**) in DMSO- d_6

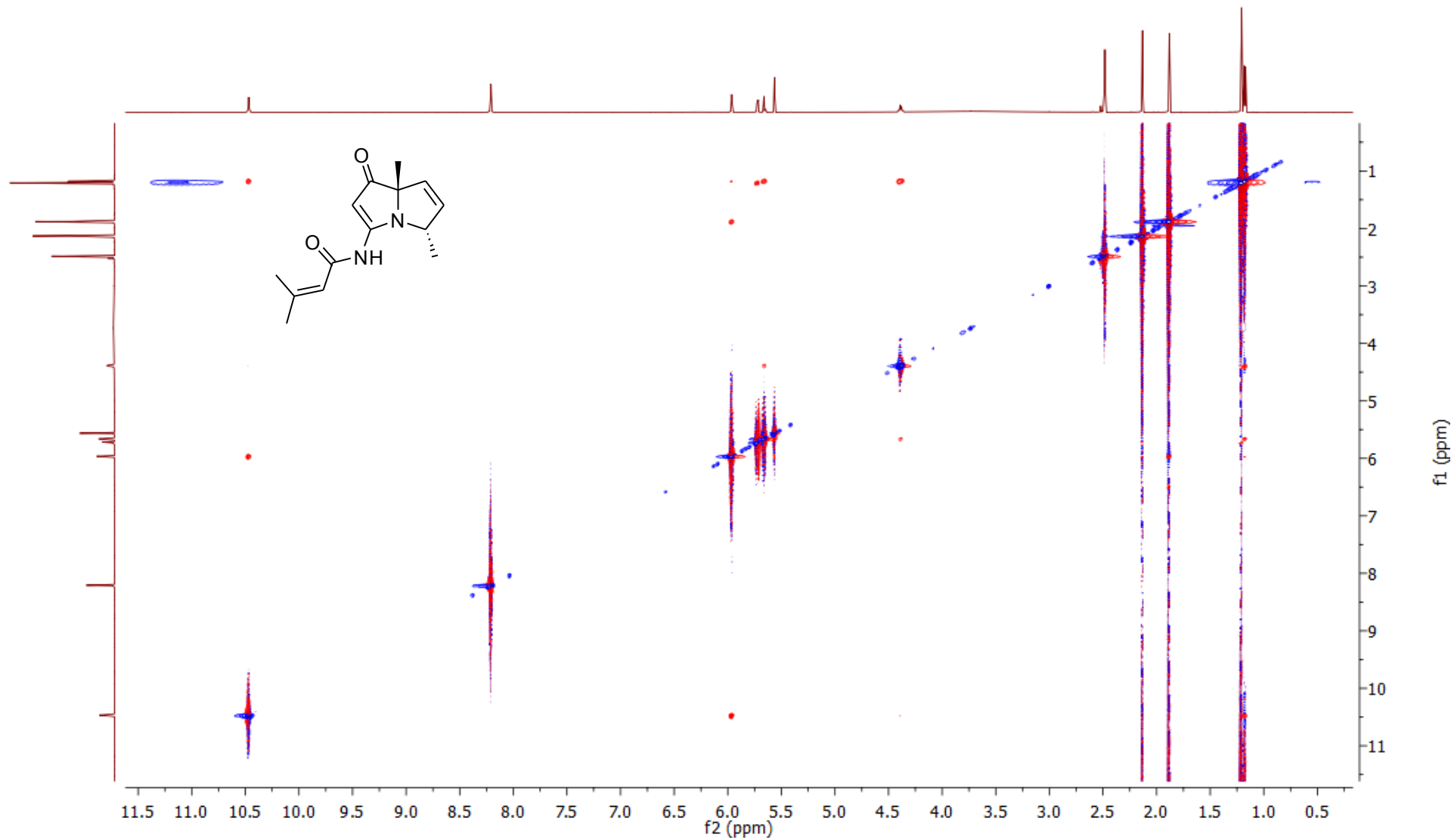


Figure S41. HRESIMS spectrum of bohemamine G (6)

Polarity/Scan Type: **Positive**

Sample Name: U6JM8177

Acq. File: 040215010.wiff

Acq. Date: Thursday, April 02, 20

Max. 4.4e5 counts.

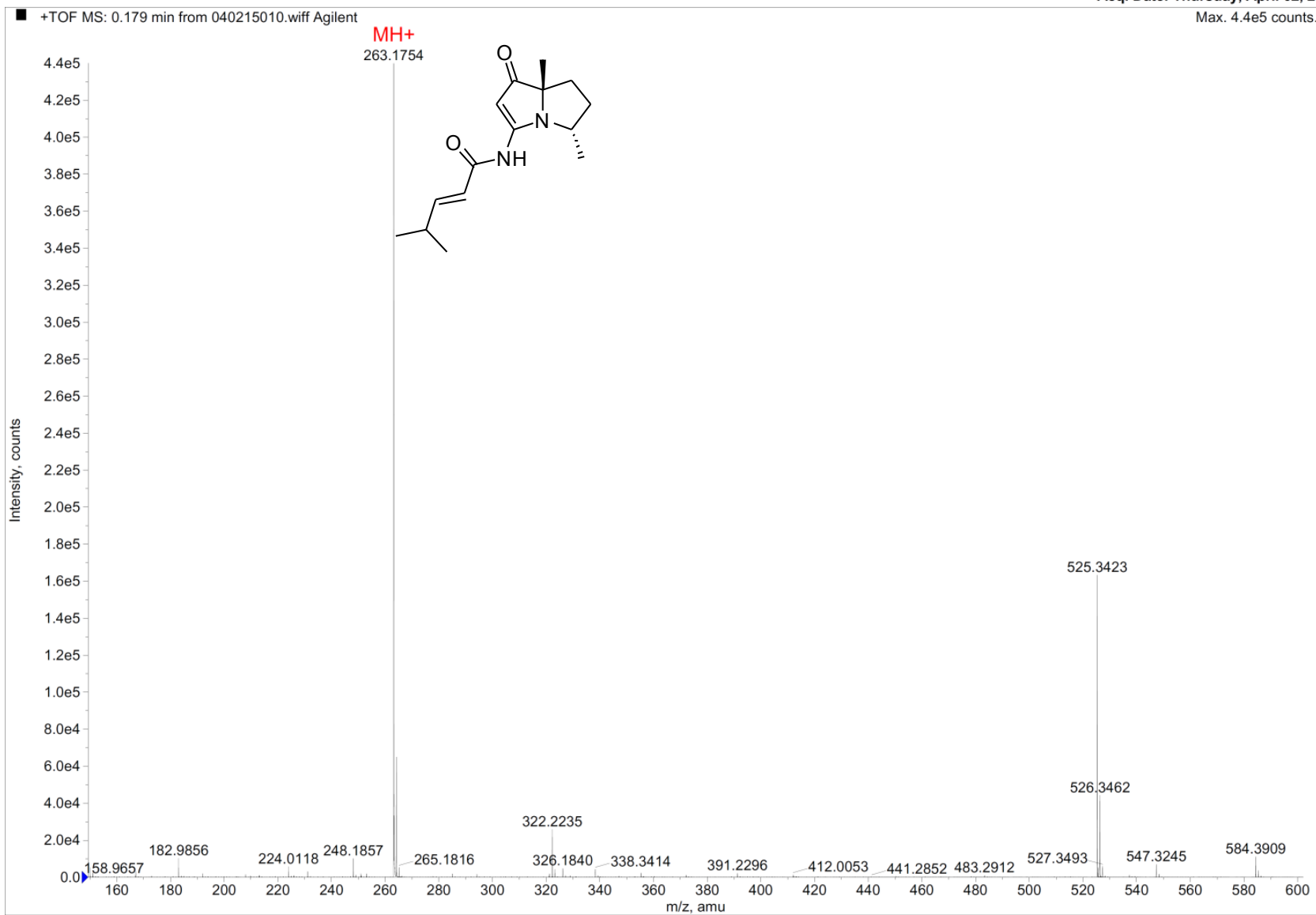


Figure S42. ¹H-NMR spectrum of bohemamine G (6) in DMSO-*d*₆

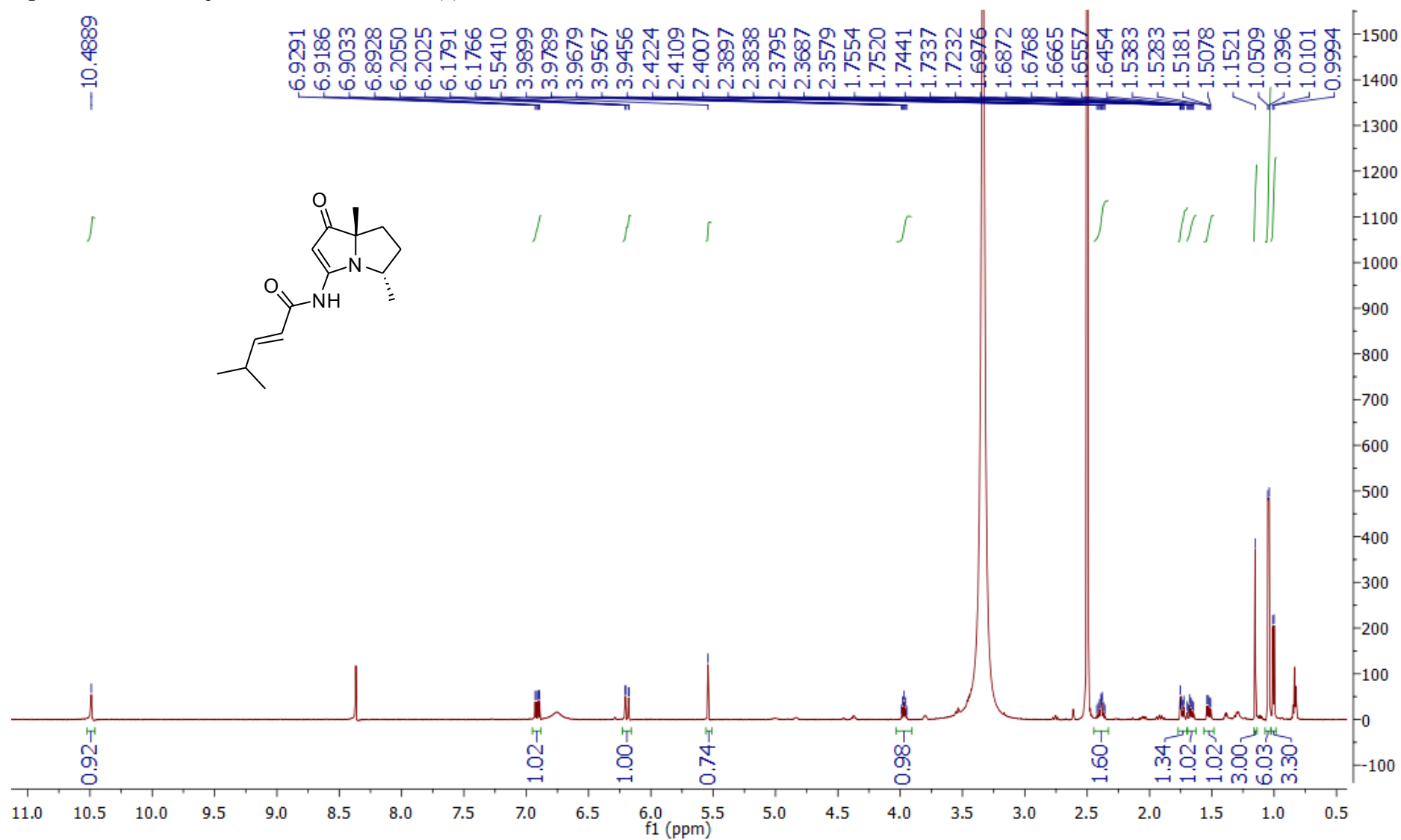


Figure S43. ^{13}C -NMR spectrum of bohemamine G (**6**) in $\text{DMSO-}d_6$

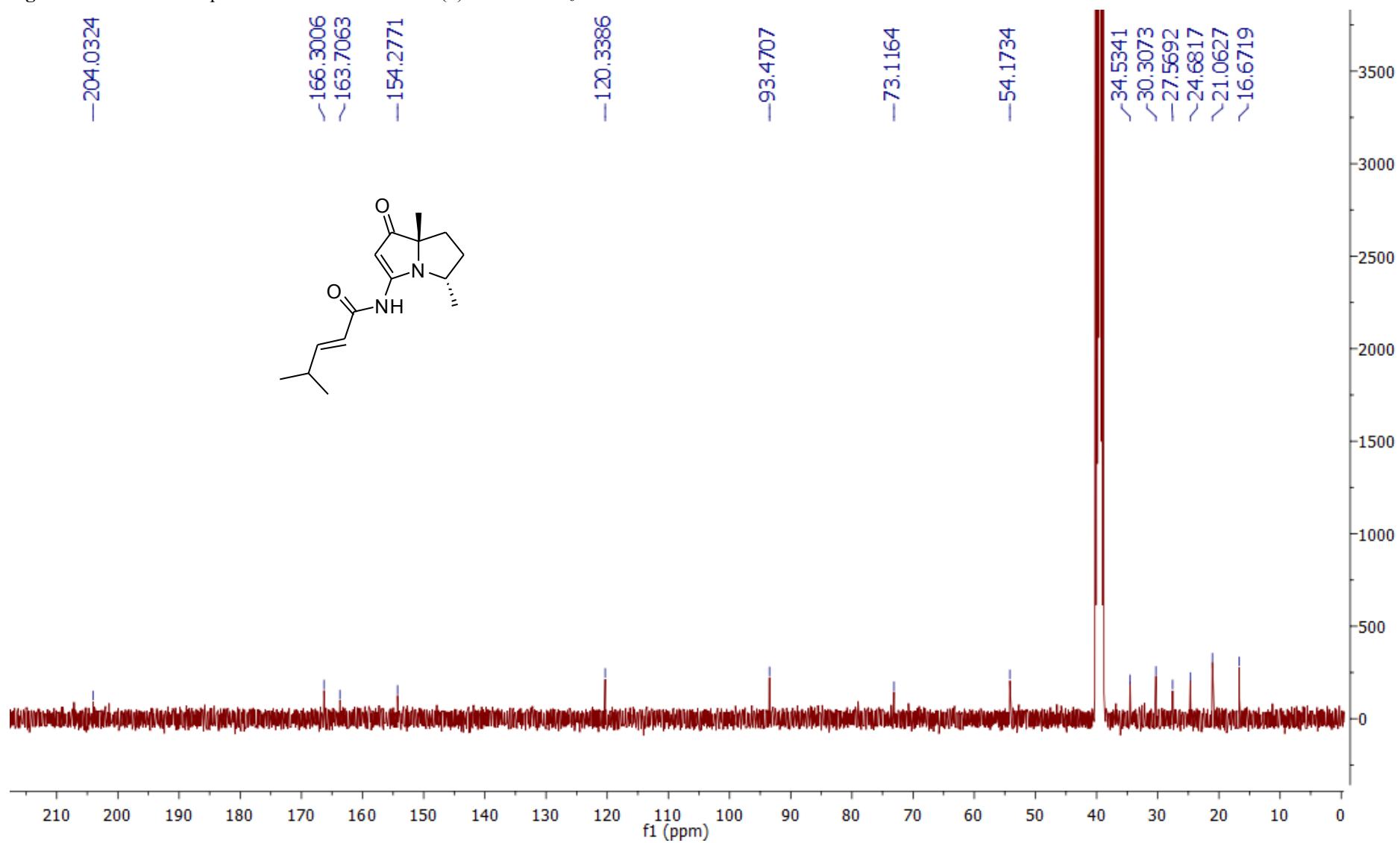


Figure S44. ^1H - ^1H COSY spectrum of bohemamine G (**6**) in $\text{DMSO-}d_6$

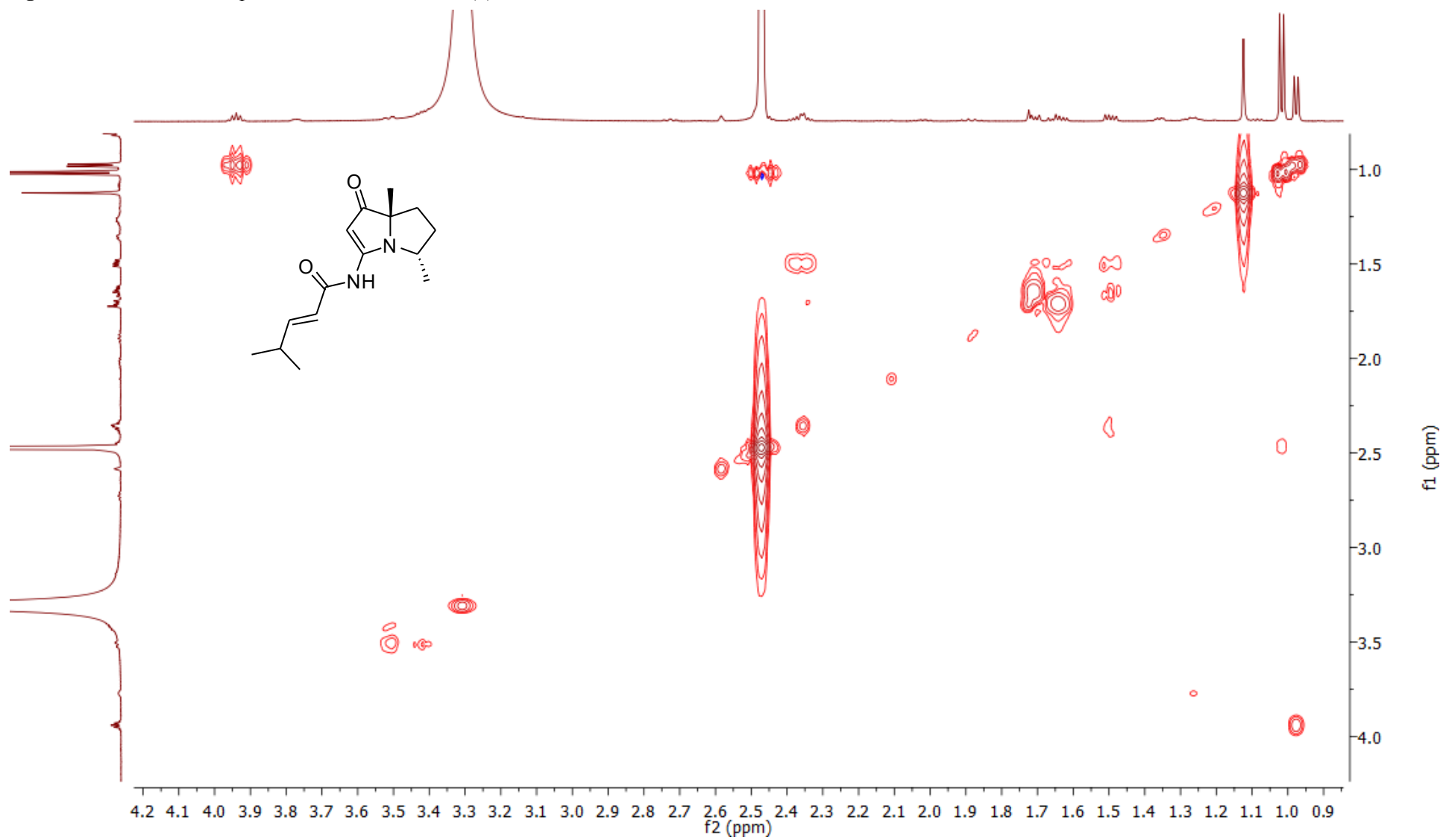


Figure S45. HSQC spectrum of bohemamine G (6) in DMSO- d_6

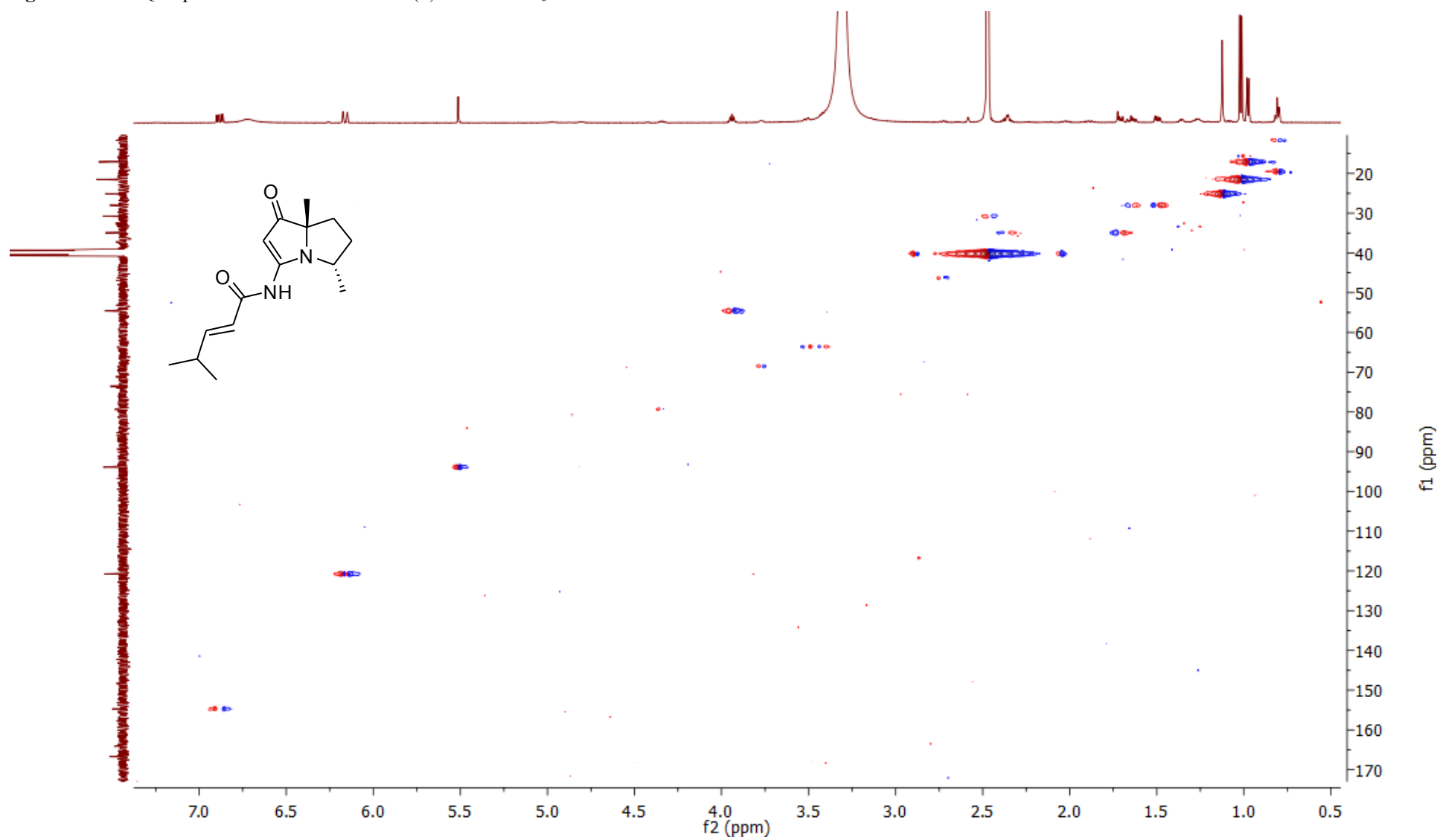


Figure S46. HMBC spectrum of boheminamine G (6) in DMSO- d_6

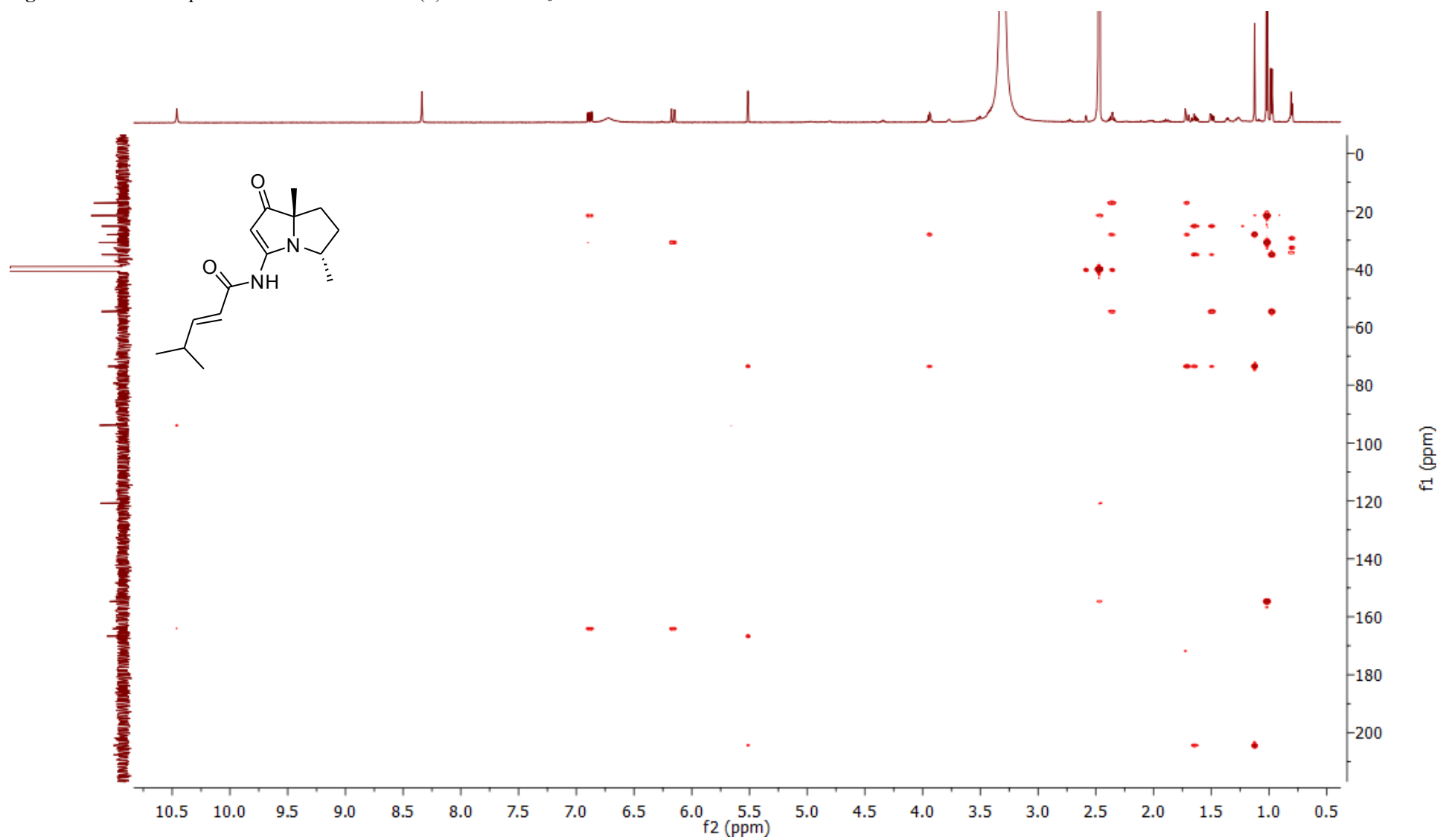


Figure S47. NOESY spectrum of bohemamine G (**6**) in DMSO- d_6

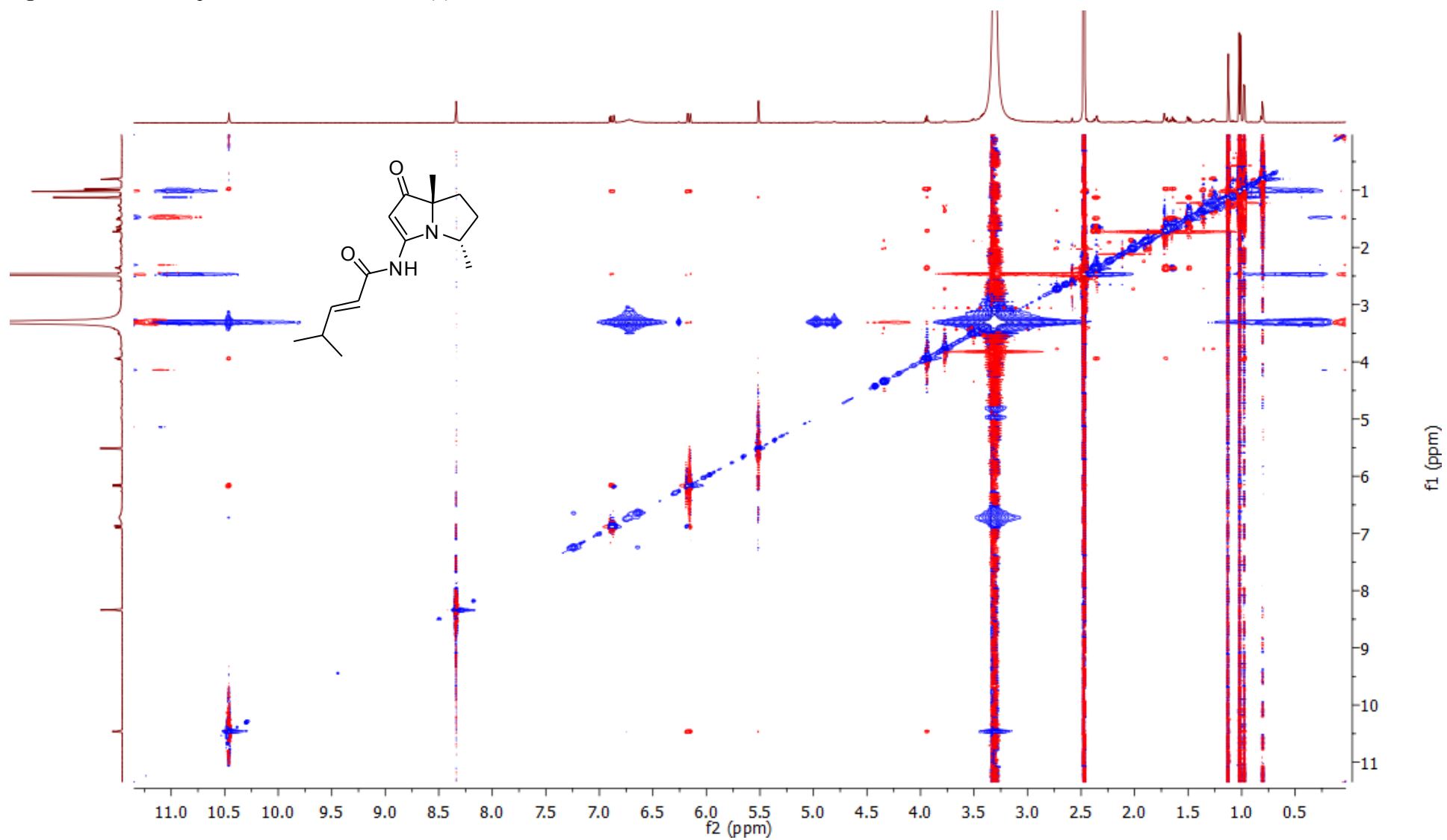


Figure S48. HRESIMS spectrum of bohemamine H (7)

Polarity/Scan Type: **Positive**

Sample Name: U6JM8178

Acq. File: 040215011.wiff
Acq. Date: Thursday, April 02, 20

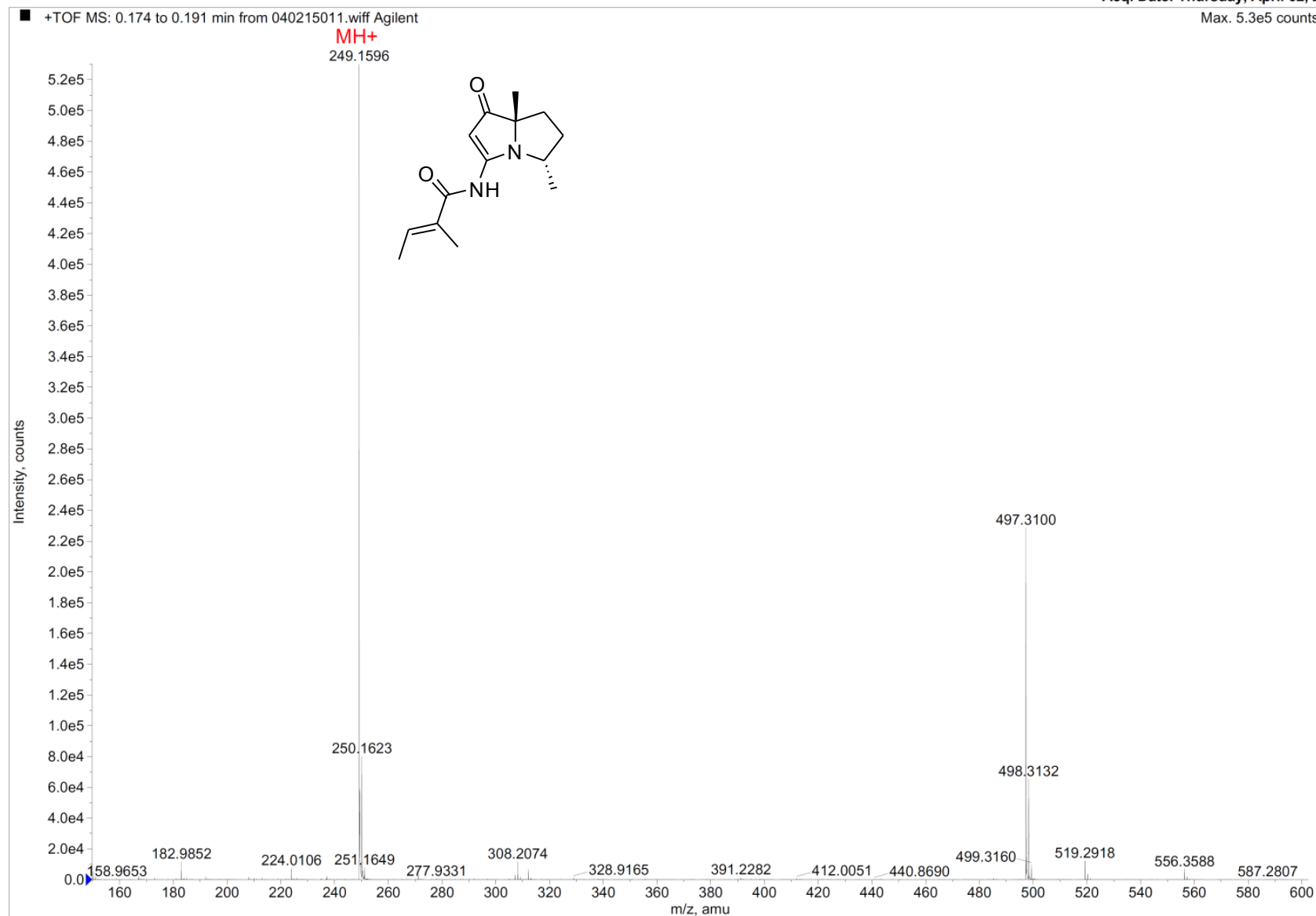


Figure S49. ¹H-NMR spectrum of bohemamine H (7) in DMSO-d₆

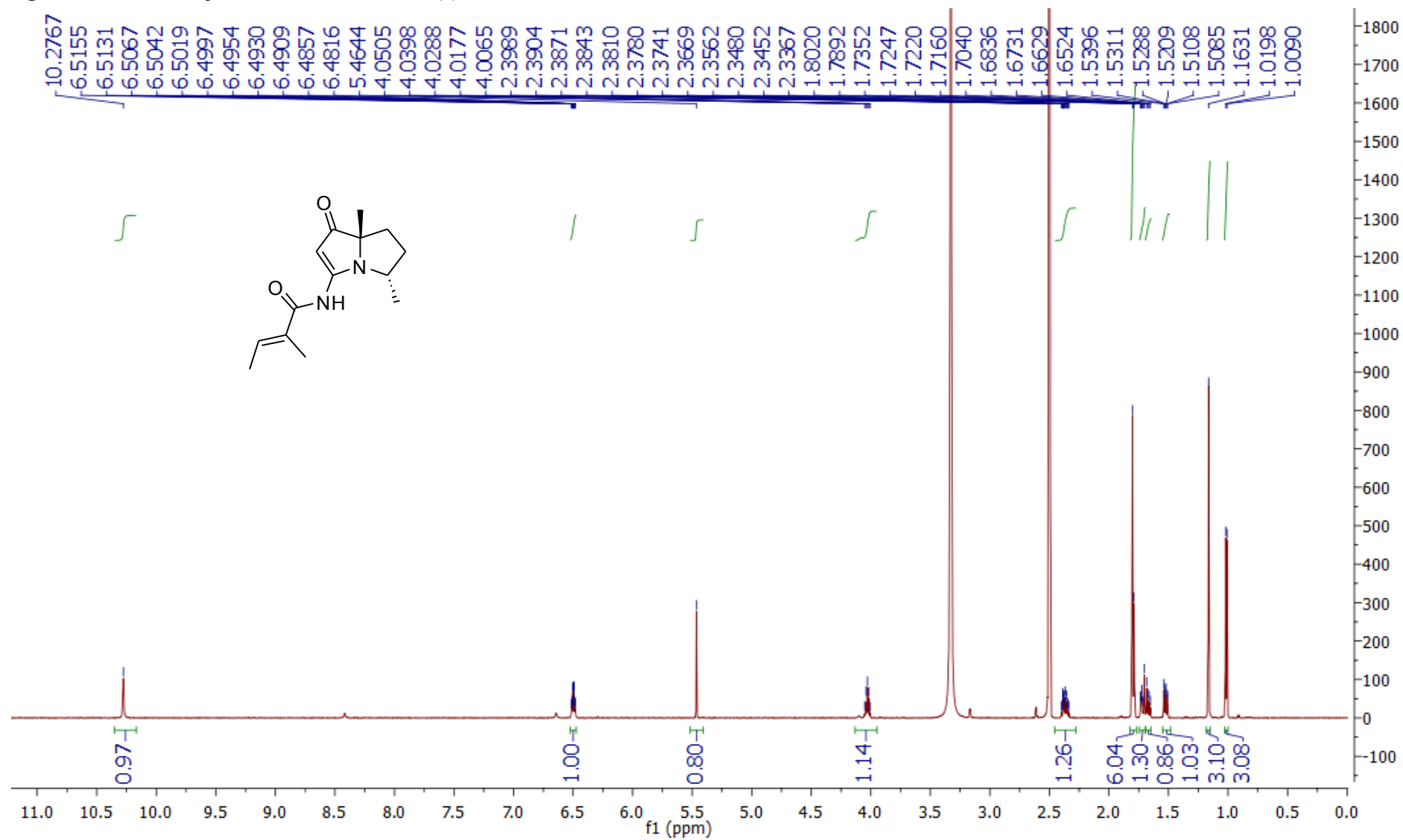


Figure S50. ^{13}C -NMR spectrum of bohemamine H (7) in $\text{DMSO-}d_6$

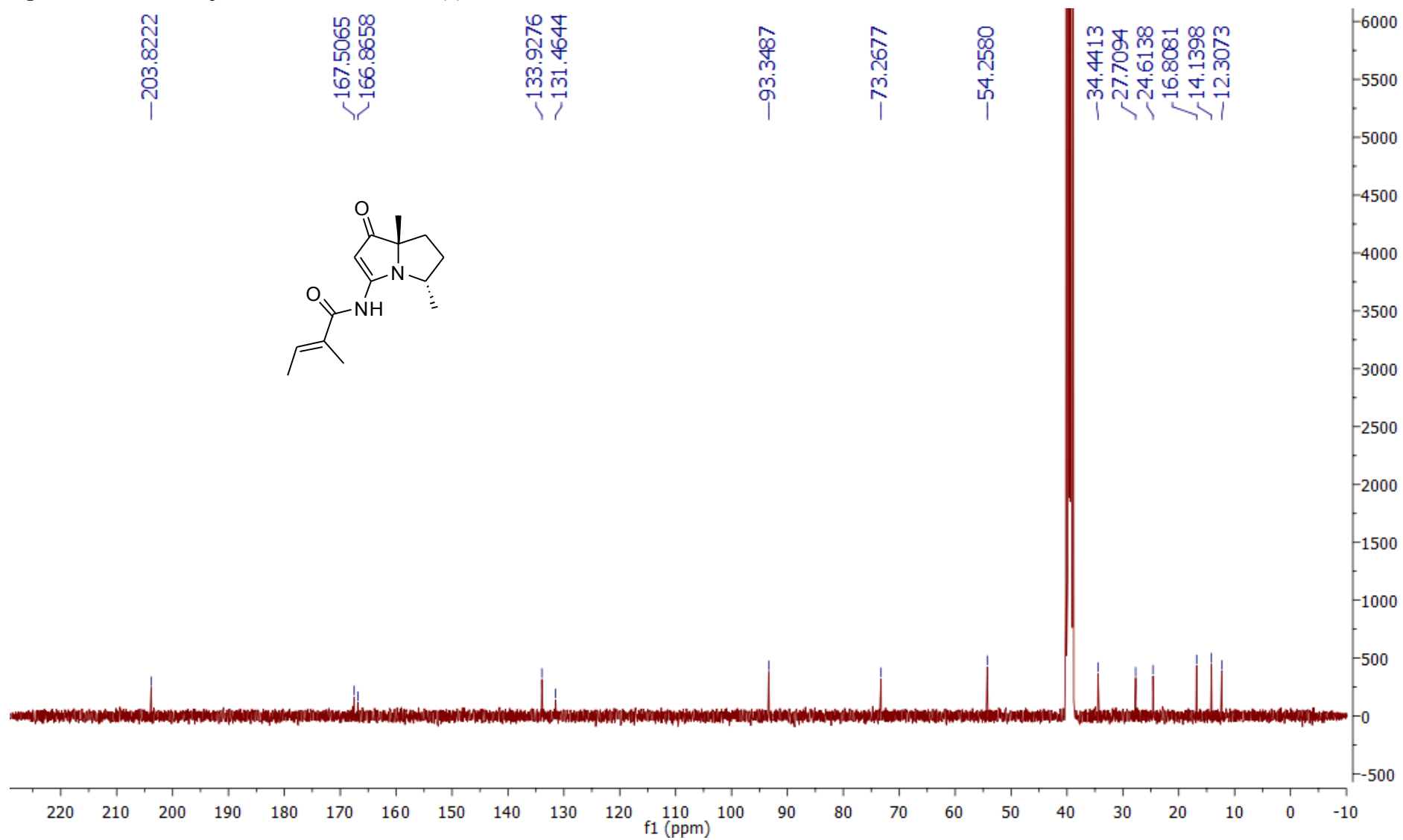


Figure S51. ^1H - ^1H COSY spectrum of bohemamine H (7) in $\text{DMSO-}d_6$

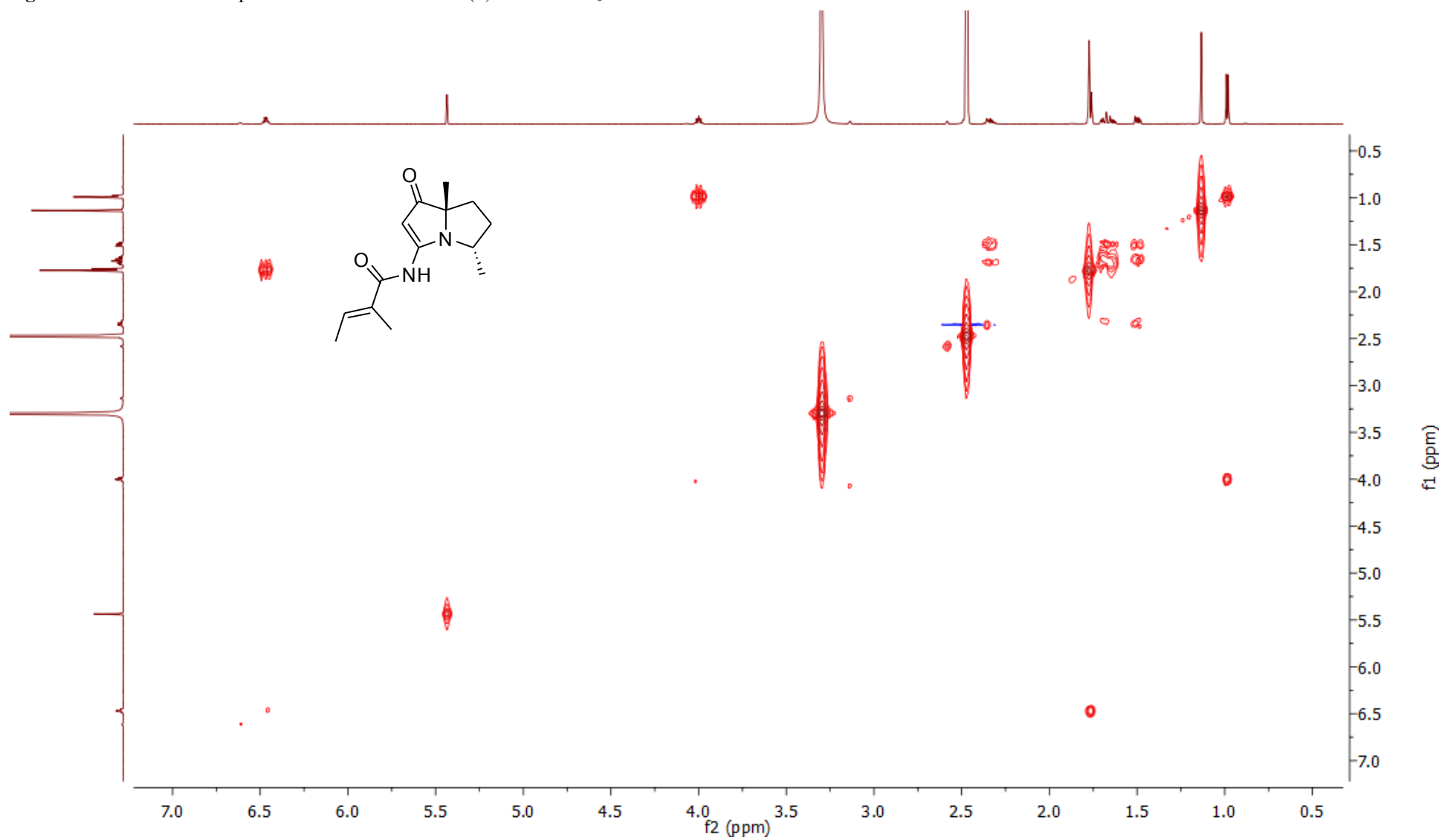


Figure S52. HSQC spectrum of bohemamine H (7) in DMSO- d_6

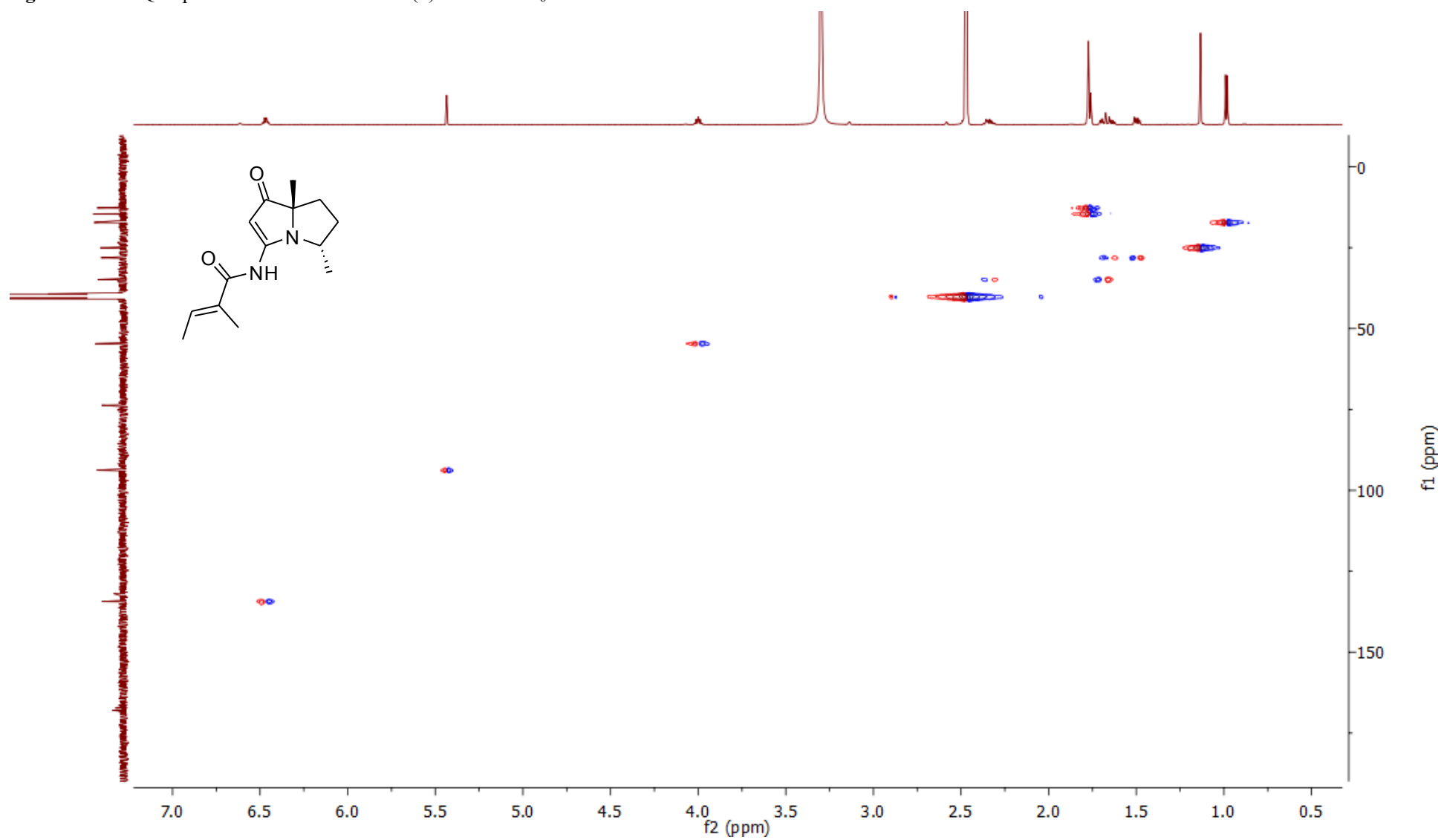


Figure S53. HMBC spectrum of bohemamine H (7) in DMSO- d_6

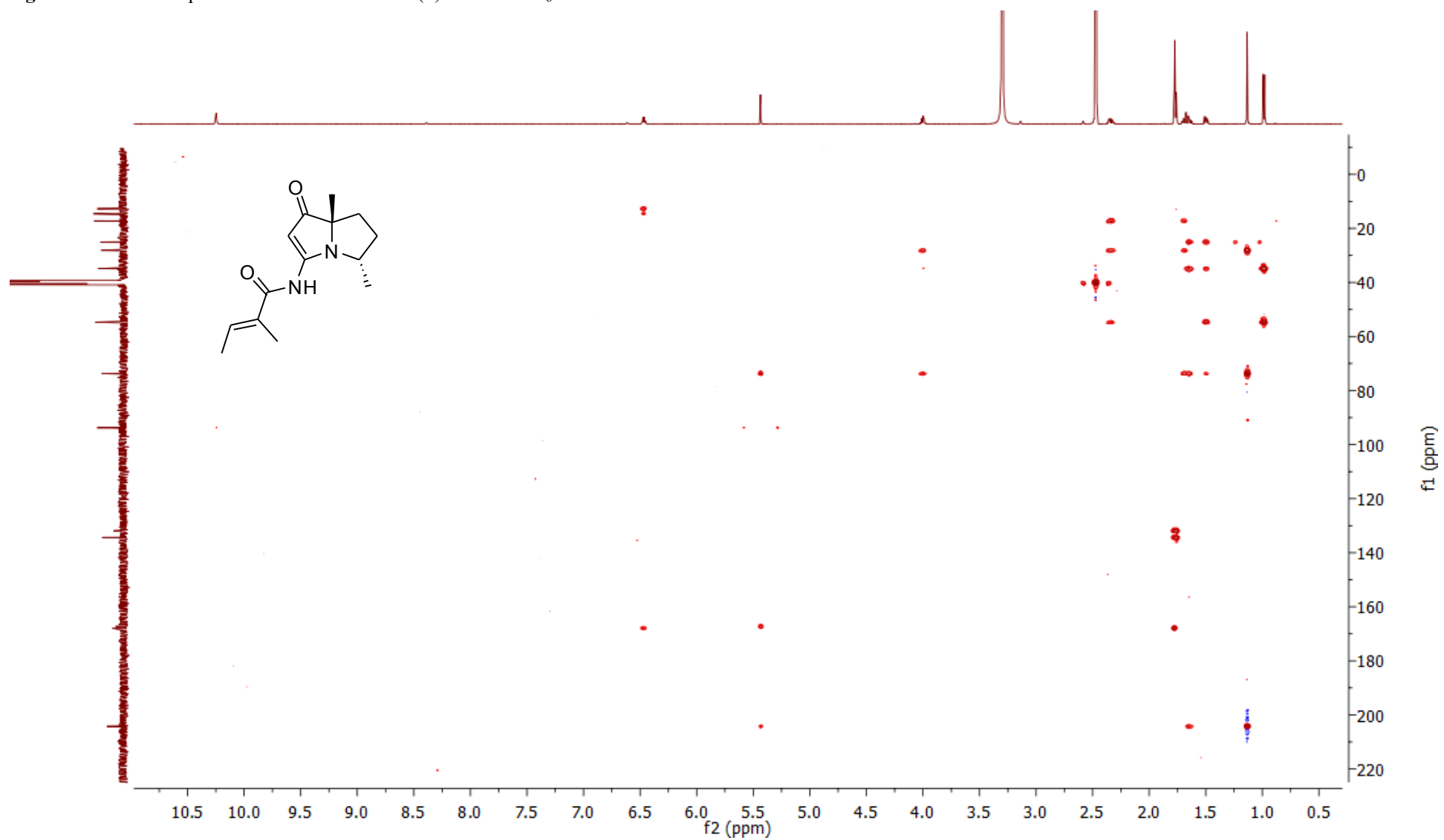


Figure S54. NOESY spectrum of bohemamine H (7) in DMSO- d_6

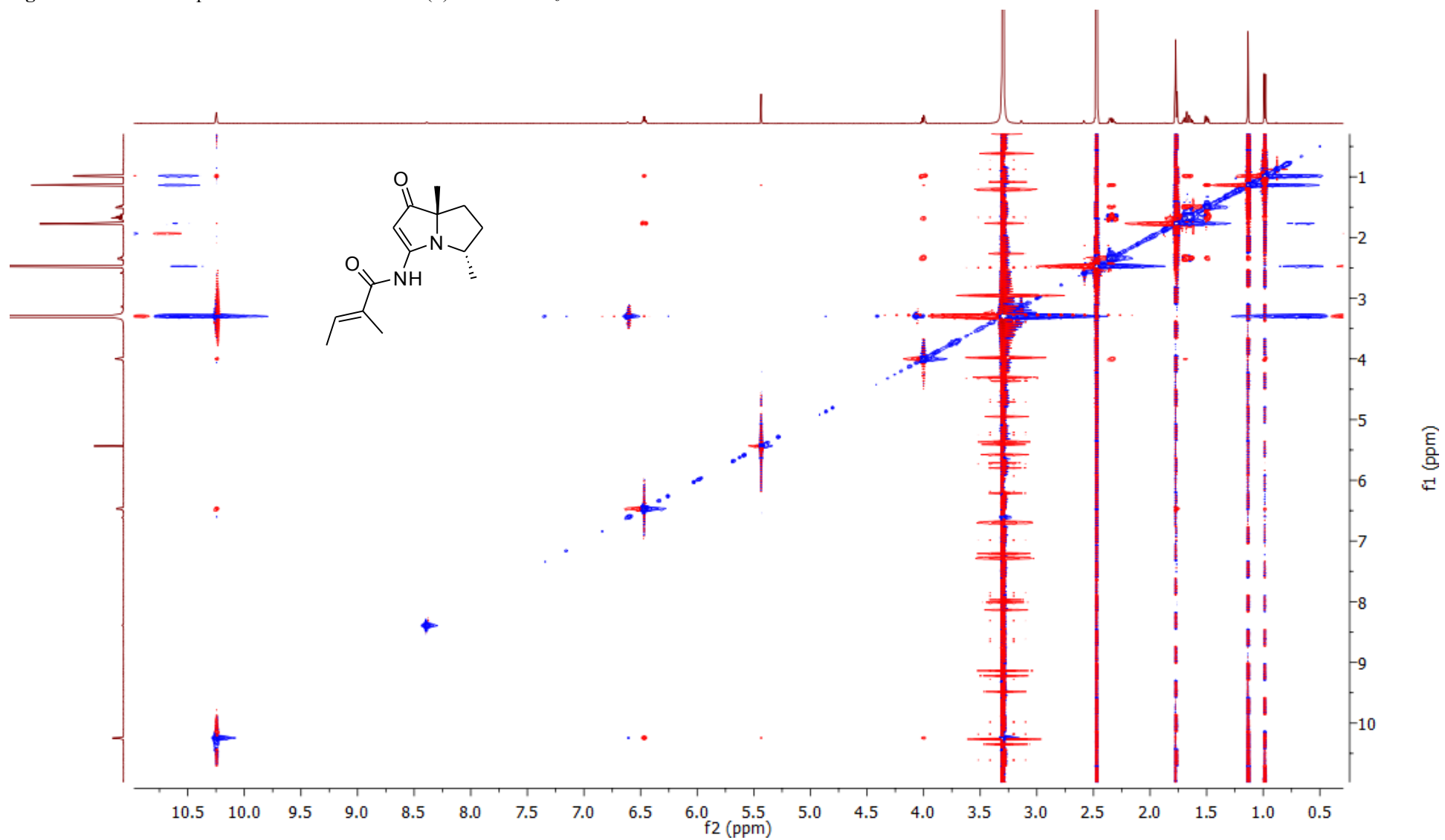


Figure S55. HRESIMS spectrum of bohemamine I (8)

Polarity/Scan Type: **Positive**

Sample Name: U6JM8180

Acq. File: 040215013.wiff

Acq. Date: Thursday, April 02, 20

Max. 3.5e5 counts.

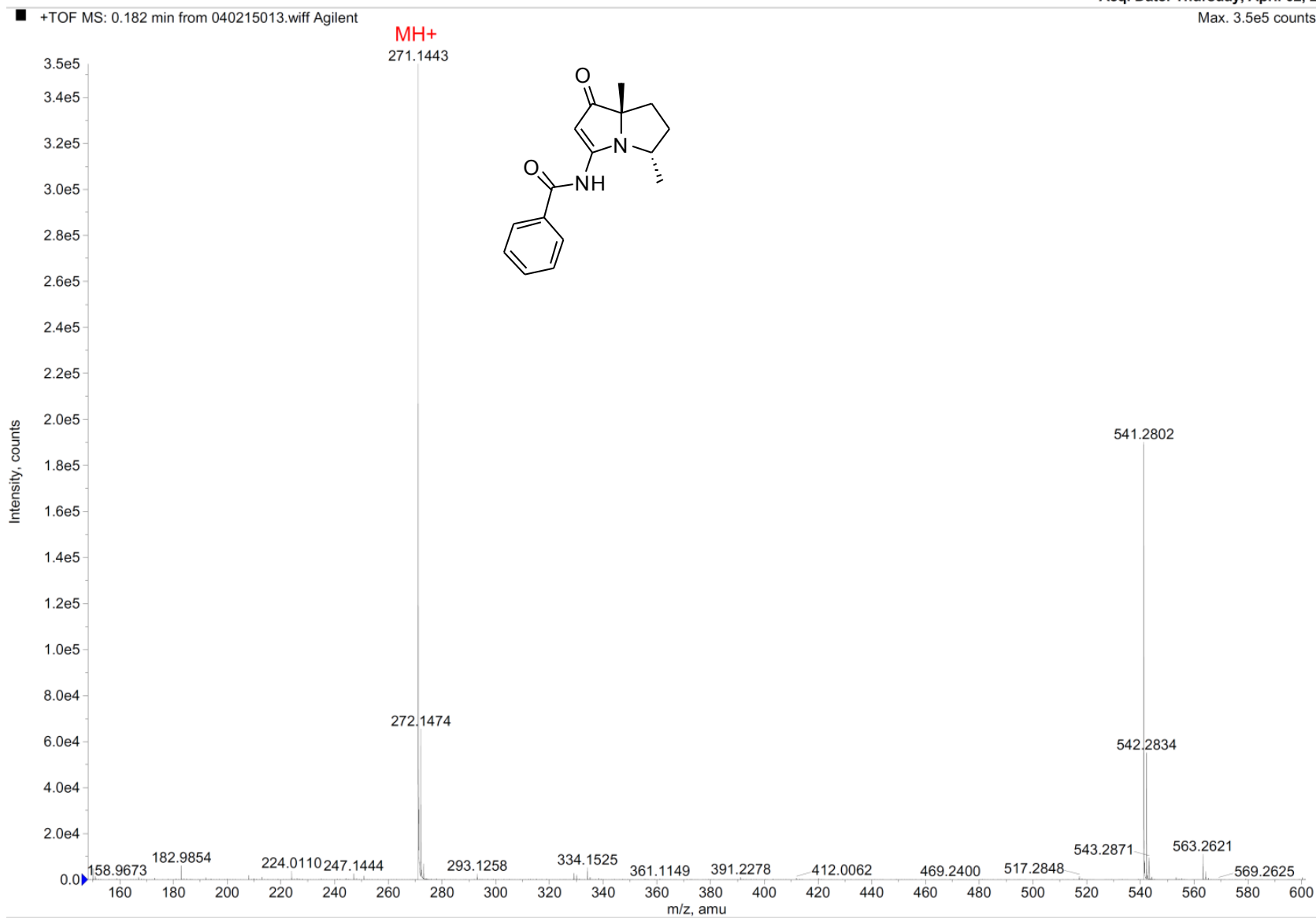


Figure S56. ¹H-NMR spectrum of bohemamine I (**8**) in DMSO-*d*₆

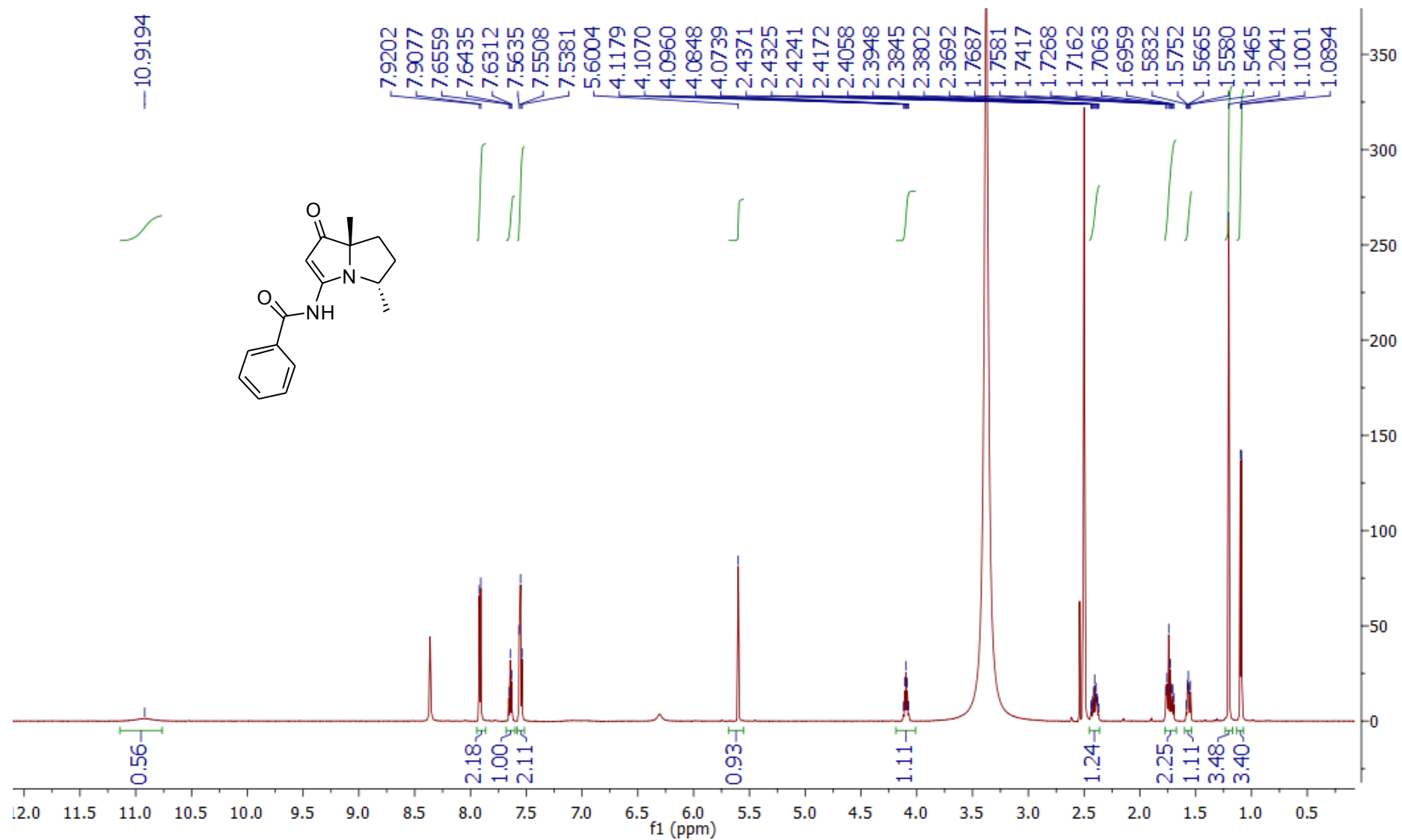


Figure S57. ^{13}C -NMR spectrum of bohemamine I (**8**) in $\text{DMSO-}d_6$

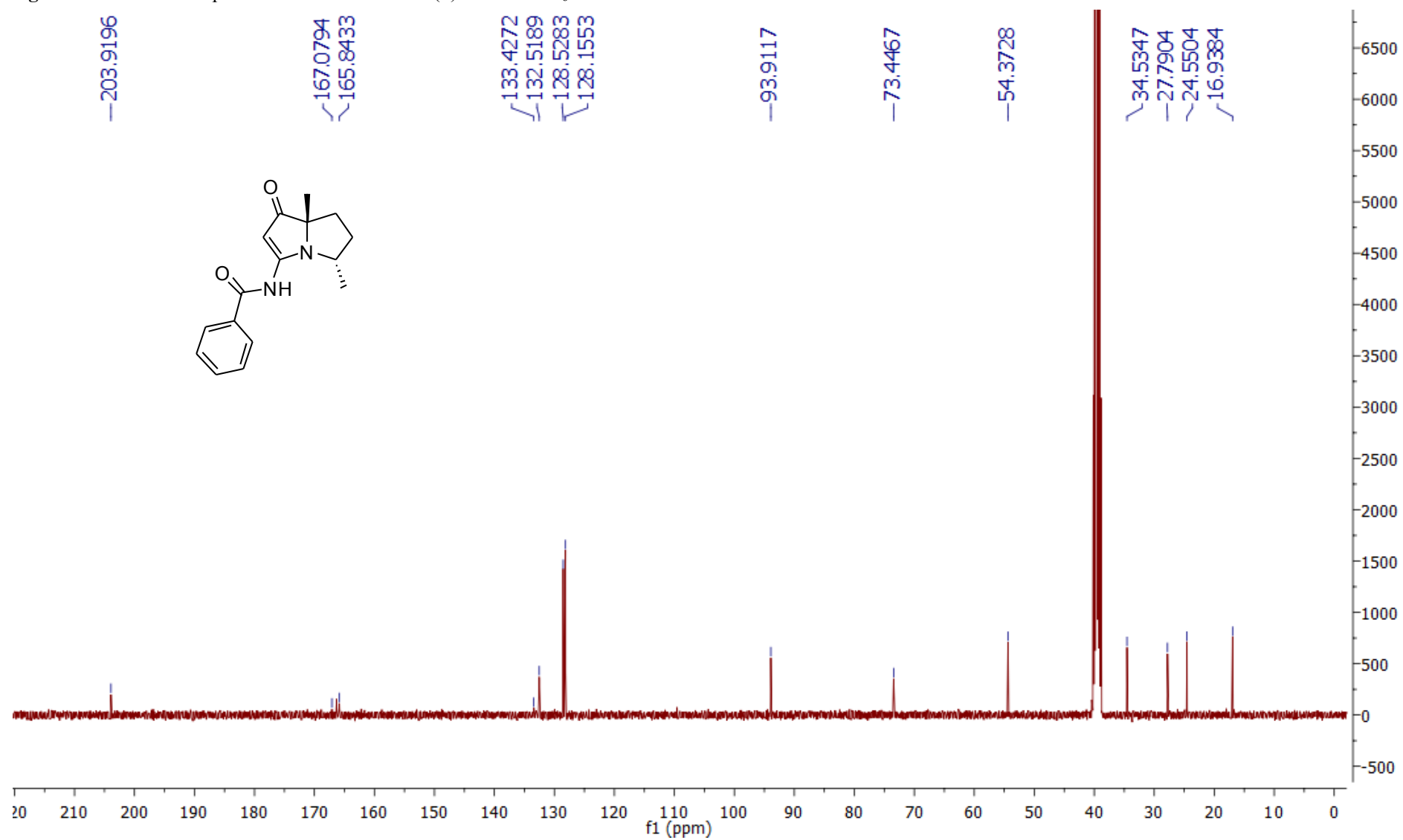


Figure S58. ^1H - ^1H COSY spectrum of bohemamine I (**8**) in $\text{DMSO-}d_6$

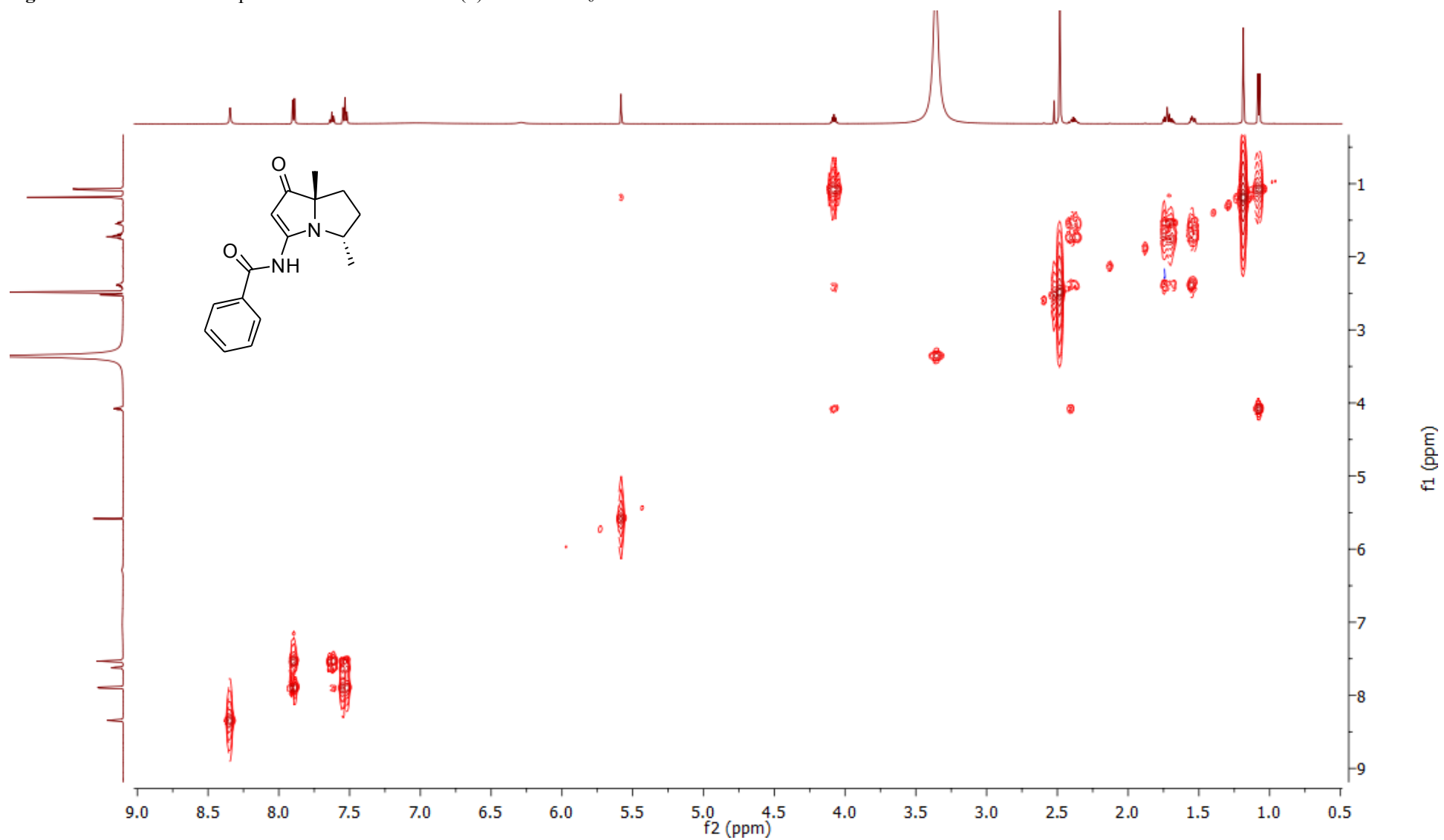


Figure S59. HSQC spectrum of bohemamine I (8) in DMSO- d_6

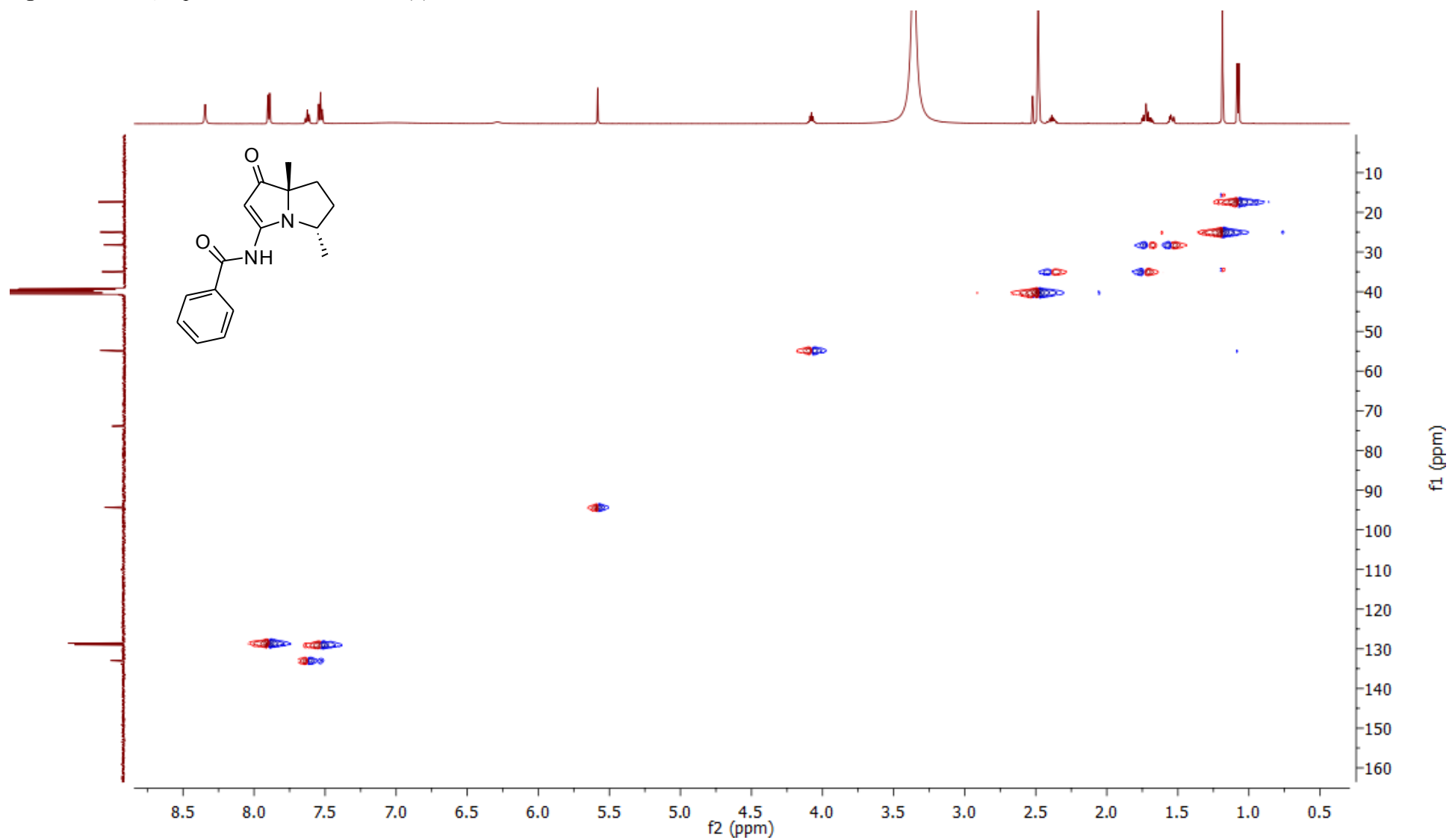


Figure S60. HMBC spectrum of bohemamine I (8) in DMSO- d_6

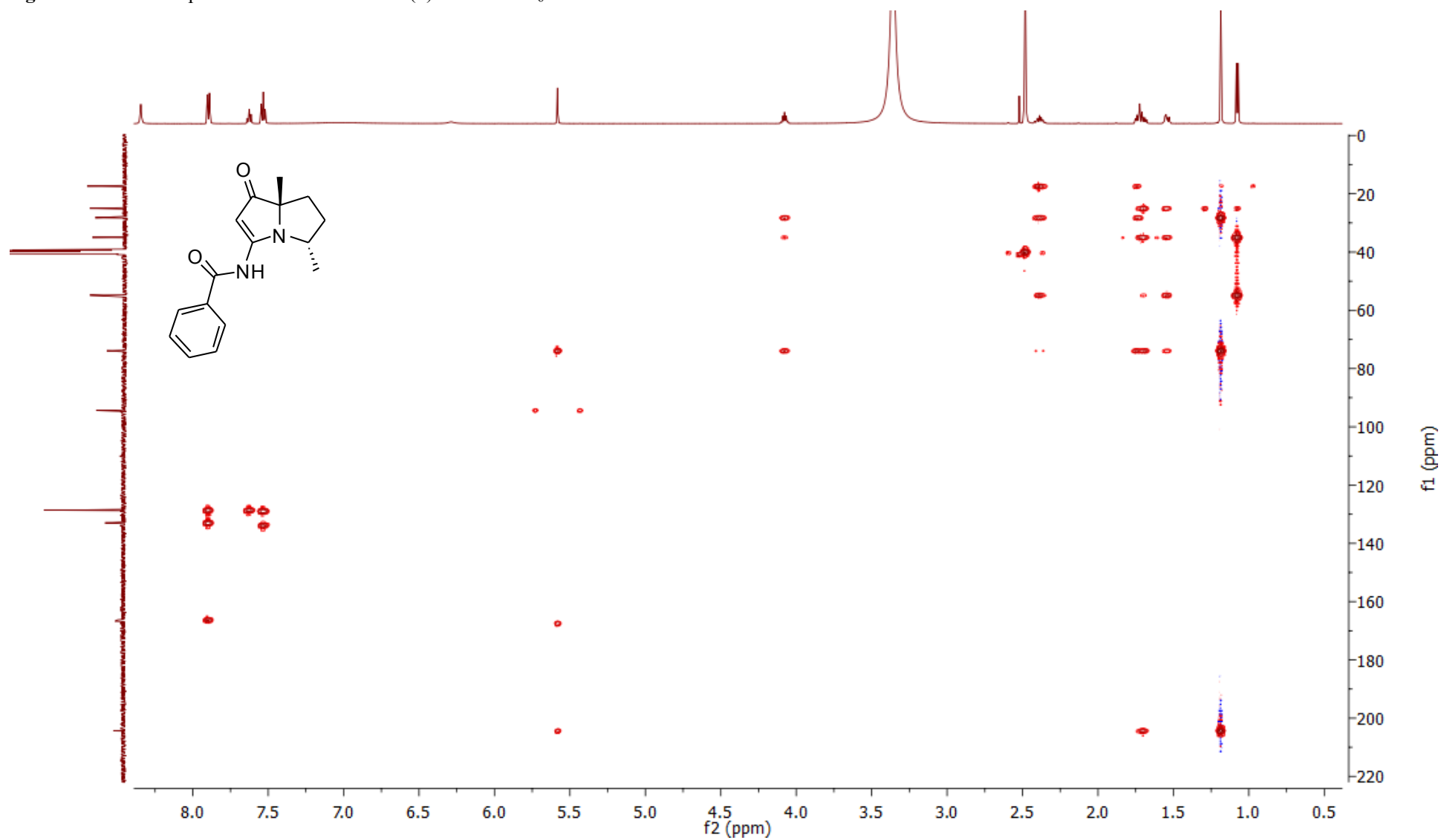


Figure S61. NOESY spectrum of bohemamine I (8) in DMSO- d_6

