

Supplemental Figure 1 Lung cytokine expression in the mild/moderate and severe asthma models. WT mice were subjected to the full 28-day mild/moderate (M/MOD) or severe asthma (SA) models. (A) Cytokine mRNA levels in whole lung homogenates measured by qRT-PCR. (B) Cytokine mRNA levels in an experiment similar to (A) but including comparison to naïve, untreated mice and demonstrating that c-di-GMP alone does not result in induction of cytokine expression in the lung. The data also demonstrate that house dust mite antigen (HDM) alone generates a Type 2 cytokine response whereas HDM + c-di-GMP induces Type 1 and Type 17 cytokine (A, n = 6 mice per group; B, n = 2-3 mice per group). Data are mean  $\pm$  SEM and each experiment was performed at least twice. \*P  $\leq$  0.001 compared to all other conditions using one way ANOVA with Bonferroni's multiple comparison test in (B).



Supplemental Figure 2 Differential BAL cell numbers collected from WT and *IRF5<sup>-/-</sup>* mice subjected to the severe asthma model. WT and *IRF5<sup>-/-</sup>* mice were subjected to the full 28-day severe asthma (SA) model. Differential bronchoalveolar lavage (BAL) cell percentages were determined from at least 300 cells counted per Giemsa-stained cytospin slide. The number of each cell type was then calculated based upon the total BAL cell recovery for each mouse. (Macs, macrophages; Lymphs, lymphocytes, Eos, eosinophils, PMNs, neutrophils). \*\*\*P  $\leq$  0.001 one way ANOVA with Bonferroni's multiple comparison test. n = 6-8 mice per group. Data are mean <u>+</u> SEM and are representative of 3 independent experiments.



Supplemental Figure 3 Significantly higher IRF5 expression in lung DCs versus macrophages in mice treated with HDM + c-di-GMP. Lungs from WT and *IRF5<sup>-/-</sup>* mice were harvested 24 h following immunization with house dust mite (HDM) + c-di-GMP on d 1, 3, and 5. DCs and macrophages (Macs) were enriched from collagenase/DNAse-digested tissue using CD11c magnetic bead separation (Miltenyi Biotec). These two cell populations were then purified by cell sorting using a FACSAria cell sorter (BD Immunocytometry Systems) based upon autofluorescence and MHC Class II staining: lung macrophages are autofluorescence<sup>high</sup>/MHC ClassII<sup>low</sup>, and DCs are autofluorescence<sup>low</sup>/MHC Class II<sup>high</sup>. mRNA was prepared from the purified cells and qRT-PCR was performed for *Irf5* expression relative to *Hprt1*. \*P ≤ 0.05, \*\*P ≤ 0.05 using Mann-Whitney U test. The experiment was performed twice independently.



Supplemental Figure 4 Decreased numbers of IFN- $\gamma$ – and IL-17Aexpressing CD4<sup>+</sup> T cells in the lymph nodes of *IRF5<sup>-/-</sup>* mice in the sensitization phase of the severe asthms model. Lymph nodes were harvested 24 h following immunization with house dust mite (HDM) + c-di-GMP on d 1, 3, and 5. Total lymph node cells were re-stimulated in vitro for 72 h with HDM. For intracellular IFN- $\gamma$  (APC, clone XMG1.2; BD Biosciences catalog 554413; 1:100) and IL-17A (PE, clone TC11-18H10; BD Biosciences catalog 559502; 1:100) staining, in vitro-stimulated cells were treated with PMA (50 ng/ml) and ionomycin (500 ng/ml) for 5 h with brefeldin A (GolgiPlug; BD Biosciences) added during the final 3 h. The cells were fixed in 4% paraformaldehyde for 20 min at RT, followed by permeabilization (CytoPerm; BD Biosciences) for 30 min at 0°C, then optimally diluted antibody also for 30 min at 0°C. Plots shown are gated on CD4<sup>+</sup> T cells.



**Supplemental Figure 5 Lymph node resident DCs express less IRF5 and inflammatory cytokines than migratory DCs.** Lymph nodes were harvested 24 h following immunization with house dust mite (HDM) + c-di-GMP on d 1, 3, and 5. Lymph node resident DCs (rDCs) and migratory DCs (migDCs) were purified by cell sorting using the gating strategy shown in Figure 3A. mRNA was isolated and qRT-PCR was performed on the purified cells for *Irf5, II12p35, II12p40 and II6*. Expression levels for cells pooled from 4-5 mice are shown relative to *Hprt1*.



Supplemental Figure 6 Specificity of HDM-specific ELISAs, and total serum IgGs in the SA model. (A) As described in detail in the Methods, an antigen-specific ELISA was developed incorporating plates coated with house dust mite (HDM), incubation with diluted sera from naïve or mice treated with HDM + c-di-GMP, then detection with labeled antibodies specific for mouse IgG2a or IgG1. Shown is absorbance at 450 nm with correction measured at 570 nm. (B) Total serum IgG2a and IgG1 was measured in sera obtained from WT and *IRF5<sup>-/-</sup>* mice that had been subjected to the full SA model. These values were compared to absorbance values in the HDM-specific assays above in order to generate an arbitrary combined value for individual mice. The data plotted using these arbitrary values are shown in Figure 6C. Total serum and HDM-specific Igs were below the level of detection in serum from naïve, untreated animals. \*P  $\leq$  0.05, \*\*P  $\leq$  0.05using Mann-Whitney U test. n = 5-6 mice per group and the experiment was performed twice independently.