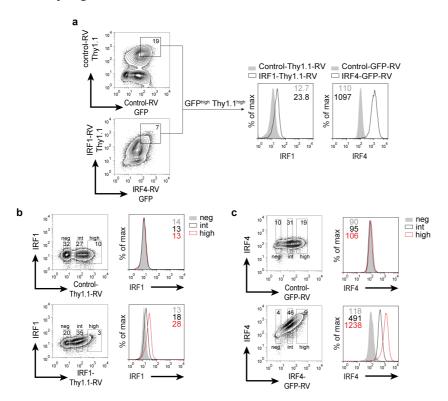
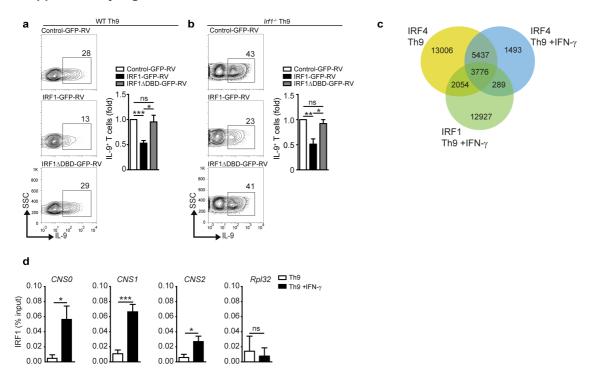


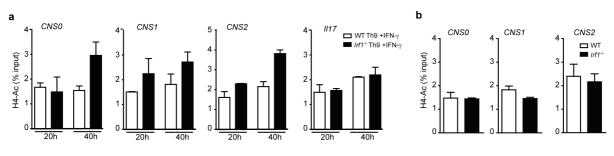
**Supplementary Figure 1. IFN-**//STAT1-induced IRF1 does not affect expression of Th9-related transcription factors. Purified CD4<sup>+</sup> T cells (a,b,e,f) or sorted naïve CD44<sup>+</sup>CD62L<sup>+</sup>CD4<sup>+</sup> T cells (c,d,g,h) were isolated from WT, Irf1<sup>-/-</sup> or Stat1<sup>-/-</sup> mice and then treated under Th9 (TGF-β+IL-4) or Th0 (without skewing cytokines) conditions with/without IFN-γ as indicated. (a) Immunoblot analysis of indicated proteins in CD4<sup>+</sup> T cells untreated or stimulated as indicated for one or two days (d1, d2). (b) ChIP analysis of STAT1 occupancy at the Irf1 5' proximal promoter in WT or Stat1<sup>-/-</sup> T cells stimulated for 13 h under Th9 conditions in the presence orf absence of IFN-γ (n=5, mean ±SD of percent input with substraction of control IgG). Data are combined from two independent experiments. (c,d) Kinetic expression of Spi1 (c) and Batf (d) mRNA (qRT-PCR). mRNA expresion was normalized to Hpr11 and relative expression was calculated by setting of the value for WT naive CD4<sup>+</sup> T cells at the time point 0 to 1. Data represent three individual experiments combined (mean ±SD). (e,f) Immunoblot analysis for the indicated proteins in Th9 cells, harvested after two days, rested in cytokine-free medium for 8 h and then treated for indicated time periods with IFN-γ in combination with rmIL-4 (e) or with rhIL-2 (f). (g,h) Flow cytometric analysis of cells intracellularly stained for T-bet and GATA3 (g) or Foxp3 (h) on day two of culture. (a,e-h) Results are representative of three independent experiments. \*\*\*rp<0.001, 2-tailed Student's t test

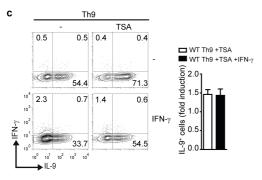


Supplementary Figure 2. The relative intensity of GFP- and Thy1.1-tags represents the relative expression of IRF4 and IRF1 in *Irf4*. Th9 cells upon IRF4-GFP-RV and IRF1-Thy1.1-RV double overexpression. *Irf4*. CD4<sup>+</sup> T cells were activated under Th0 condition overnight and spin-infected with the following retroviruses as indicated: control-GFP-RV, IRF4-GFP-RV, control-Thy1.1-RV and IRF1-Thy1.1-RV. Thereafter, cells were cultured under Th9 conditions for two further days, rested for three days and restimulated under Th9 conditions for additional two days. (a) Highly double positive cells (GFPhiThy1.1hi) were selected for further analysis of IRF1 and IRF4 expression. (b,c) Three subsets (neg, int, high) of cells expressing increasing levels of GFP and Thy1.1 were selected for analysis of IRF1 (b) and IRF4 (c) expression by each subset. (a-c) Numbers in gates represent % of gated cells, whereas numbers in histograms give the mean flurescence intensity (MFI). Data are representative of three independent experiments.

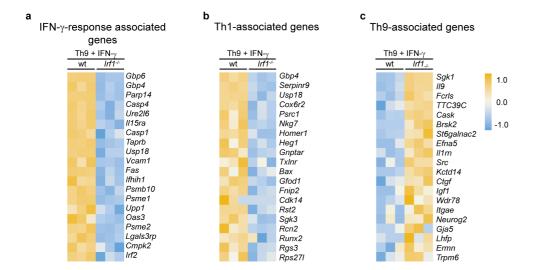


Supplementary Figure 3. IRF1 directly binds to the *II9* locus and DBD-dependently suppresses IL-9 production in Th9 cells. (a) WT or (b)  $Irf1^{-/-}$  CD4<sup>+</sup> T cells were cultured under Th0 condition over night and then spin-infected with the following retroviruses as indicated: control-GFP-RV, IRF1-GFP-RV, IRF1 $\Delta$ DBD. Thereafter, cells were cultured under Th9 conditions for further two days, rested for three days and restimulated under Th9 conditions for additional two days. Highly GFP-expressing cells were selected for further analysis of IL-9 production. Contour-plots show one representative of three independent experiments. Bars to the right give three independent experiments combined and give mean  $\pm$ SD. (c) Venn diagramm of IRF1 and IRF4 peaks in Th9 cells in absence or presence of IFN- $\gamma$  as indicated. (d) ChIP analysis of IRF1 occupancy at the indicated loci (CNS0,1,2 of the *II9* locus) in WT CD4<sup>+</sup> T cells stimulated for 13 h under Th9 conditions with or without IFN- $\gamma$  (three independent experiments combined, mean  $\pm$ SD of percent input with substraction of control IgG). \*p<0.05, \*\*p<0.005, \*\*\*p<0.001 (2-tailed Student's t test).

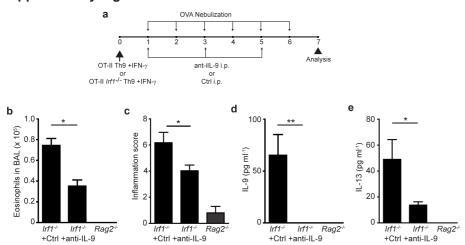




Supplementary Figure 4. IRF1 suppresses the acetylation of histone H4 at II9 CNSs independently of HDAC recruitment. (a,b) ChIP assays of acetylated (Ac) histone H4 at CNSs of the II9 gene. (a) WT and  $Irf1^{-/-}$  CD4+ T cells were (a) cultured for 20 or 40 h under Th9 conditions with or without IFN- $\gamma$  or (b) were directly analyzed. (a,b) The same chromatin was used for control ChIP experiments with control IgG. Precipitated DNA is presented relative to input (% input). Values for nonspecific binding (as determined by using control IgG) were subtracted. (c) WT CD4+ T cells were cultured under Th9 condition with/without IFN- $\gamma$  or TSA (1 nM) for two days and then analyzed for IL-9 and IFN- $\gamma$  levels by flow cytometry. Bars to the right show fold induction of IL-9+ T cells relative to non-TSA-treated cells and are combined from three independent experiments, mean  $\pm$ SD. (a-c) Data and contour-plot are representative for three independent experiments.



Supplementary Figure 5. IRF1 modulates the fate of Th9 cells towards IFN/Th1 gene signature. Naïve CD4 $^{+}$ T cells isolated from WT and  $Irf1^{-/-}$  mice were cultured under Th9 conditions in the presence of IFN- $^{-}$  $\gamma$ . Total RNA was purified from the cells and RNA-Seq was performed from three independent biological samples. Heatmap is color-coded by z-score. Displayed are top 20 core enriched hits of each GSEA of (a) 'Hallmark IFN- $^{-}$  $\gamma$  response', (b) 'Th1-associated' or (c) 'Th9-associated genes'.



Supplementary Figure 6. The IFN- $\gamma$ IRF1 pathway restricts allergic airway inflammation IL-9-dependently. (a) Purified CD4+ T cells from WT and Irf1- $\gamma$  OTII mice cells were polarized under Th9 conditions with IFN- $\gamma$  for two days, then transferred into  $Rag2^{\gamma}$  mice, which were thereafter challenged with nebulized OVA for six days. Some of the recipient mice were treated with IL-9 neutralizing antibodies (anti-IL-9) or control rat IgG (Ctrl) antibodies on the indicated days. Recipient mice were sacrificed 24 h after the last challenge. (b) Cell numbers in the BAL. (c) Tissue inflammation was evaluated with hematoxylin and eosin (H&E) staining. Slides were scored for peribronchial and perivascular inflammation with semiquantitative score from 0 to 10. (d,e) Lung cells were stimulated with 2mM OVA<sub>323-339</sub> for three days. IL-9 and IL-13 production was determined in supernatants by ELISA. (b-e) Data from one representative experiment (n=5 mice per group). The experiments were repeated four times with consistent results. \*p<0.05, \*\*p<0.005 by one-way ANOVA with Tukey's post-test (b,c) or 1-tailed Student's t test (d,e).

# Supplementary Table 1. Primers for ChIP and qPCR analyses

ChIP primers	Sequences	Location (to TSS)
Il9 CNS0	5'-ATGCGGAATGGGTTTTCACT-3'	-6287 to -6093
	5'-AAGCTCCACACACTTAGTTTGT-3'	
Il9 CNS1	5'-CCCTGTAACTCACTGTCTATCAGC-3'	-375 to -270
	5'-GCAGGAATTCTGGTTGTGAG-3'	
Il9 CNS2	5'-TCACCCACTTTAGTCCTTTCAAAA-3'	+4888 to +4983
	5'-AATTACAGAATTTTGCCCCAGGTCCTG-3'	
Il9 gene	5'-TGATTGTACCACACCGTGCT-3'	+1557 to +1657
	5'-TATCCTTTTCACCCGATGGA-3'	
Il17 promoter	5'-GCTCTCCCTGGACTCATGTT-3'	-131 to +74
	5'-TGGTTCTGTGCTGACCTCAT-3'	
RpL32	5'-TCATTTCTCAGGCACATCTT-3'	-116 to +56
promoter	5'-ACTCACCGTAAAACAGATGG-3'	
Irf1 promoter	5'-TACAACAGCCTGATTTCCCC-3'	-104 to +64
	5'-TACCTCGACGAAGGAGTGGT-3'	
		L

qRT-PCR primers	Sequences
Spi1	5'-AAC AGA TGC ACG TCC TCG AT-3'
	5'-GGG CTG GGG ACA AGG TTT GAT AAG-3'
Batf	5'-GAAGAATCGCATCGCTGC-3'
	5'-CGTTCTGTTTCTCCAGGT-3'
Hprt1	5'-CTGGTGAAAAGGACCTCTCG-3'
	5'-TGAAGTACTCATTATAGTCAAGGGCA-3'
IL9	5'-CATGCAAACAAGATACCCACTG-3'
	5'-TTGCCTCTCATCCCTCTCATC-3'
IFNG	5'-TGG GTT CTC TTG GCT GTT ACT G-3'
	5'-ACA CTC TTT TGG ATG CTC TGG TC-3'
IRF1	5'-TGGCTGGGACATCAACAAGG-3'
	5'-CTGCCCTTGTTCCTGCTCTG-3'
IRF4	5'-CTCTTTGACACACAGCAGTTCTTG-3'
	5'-TTCTGGTAAATCGTAGCCCCTC-3'

18S	5'-AGTCCCTGCCCTTTGTACACA-3'
	5'-GATCCGAGGCCTCACTAAAC-3'