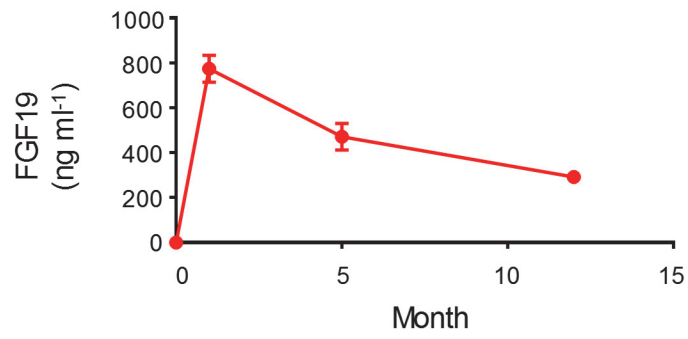
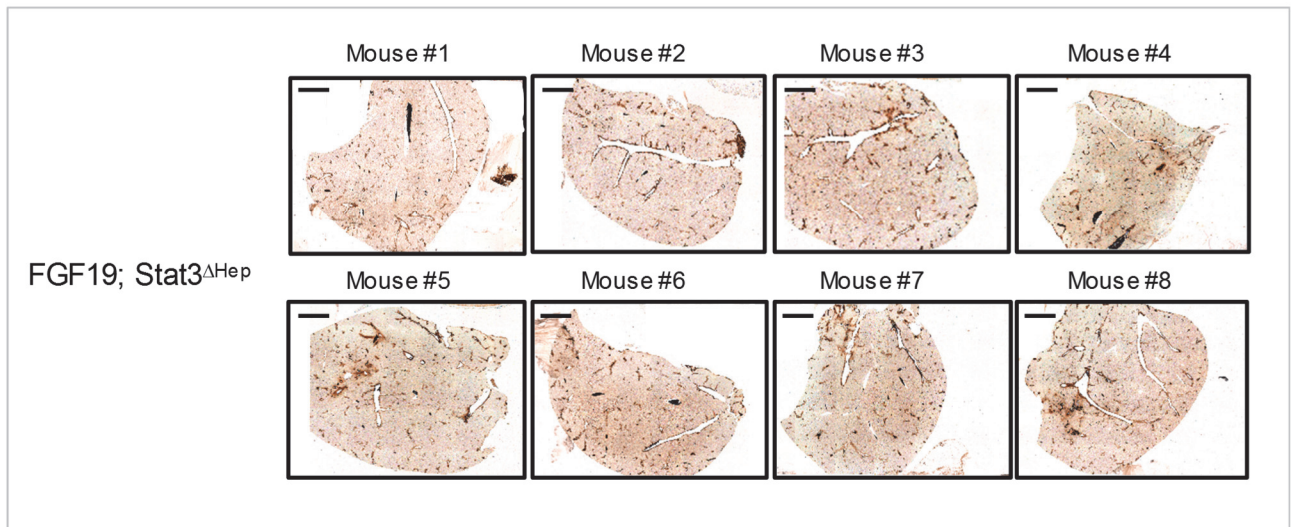
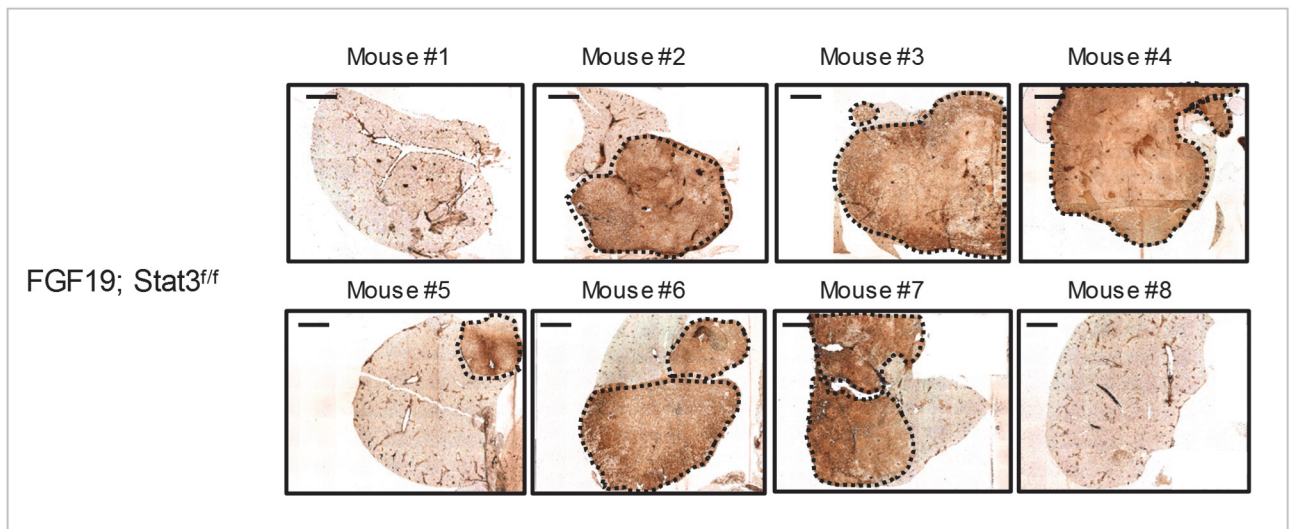
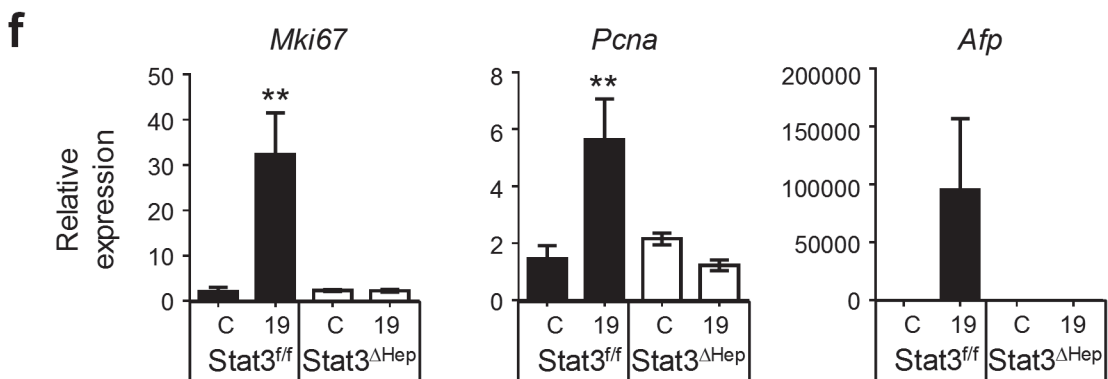
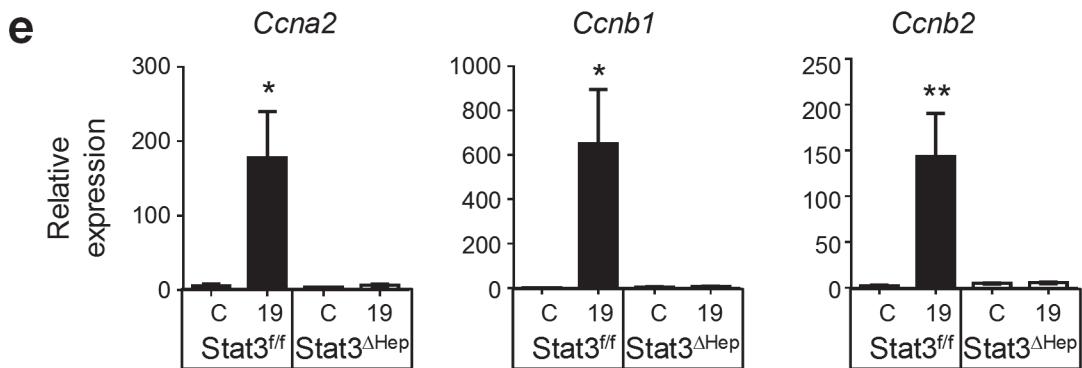
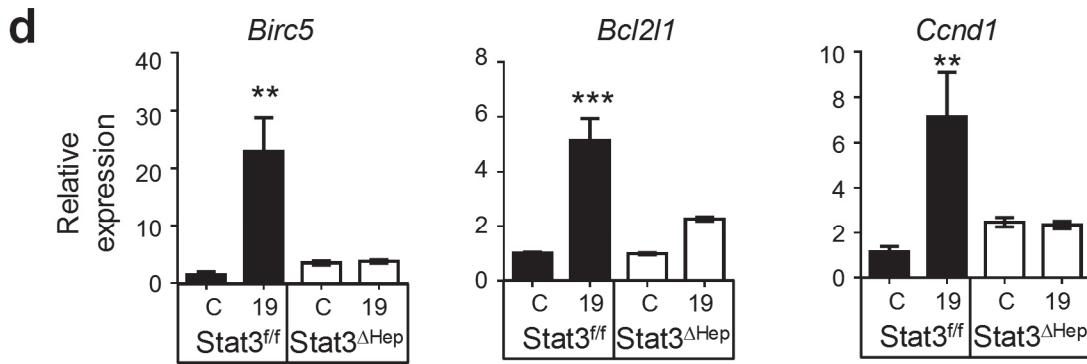
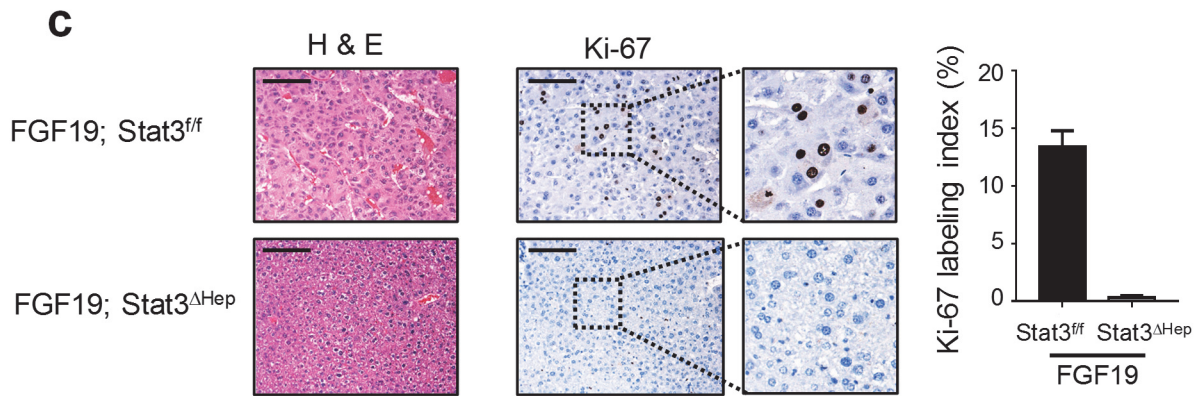
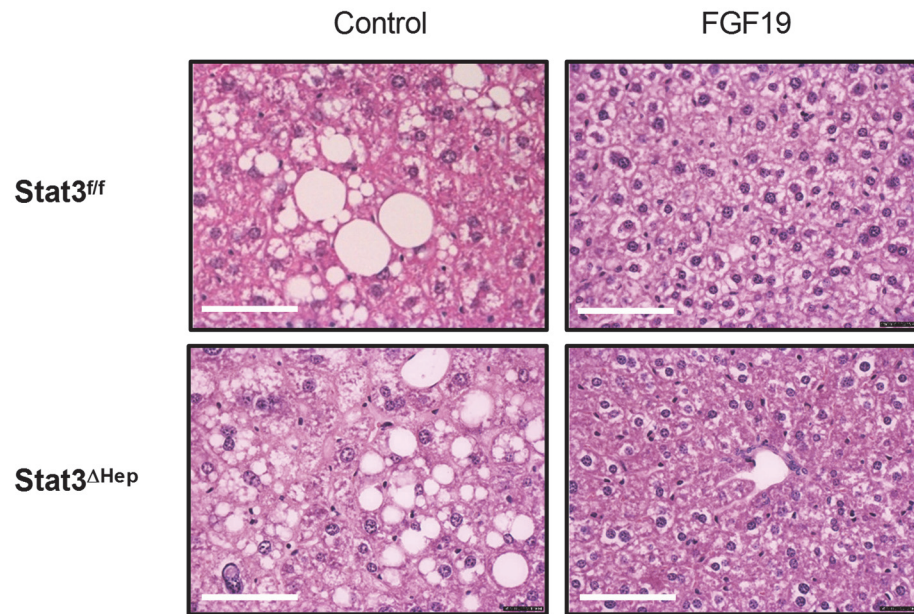


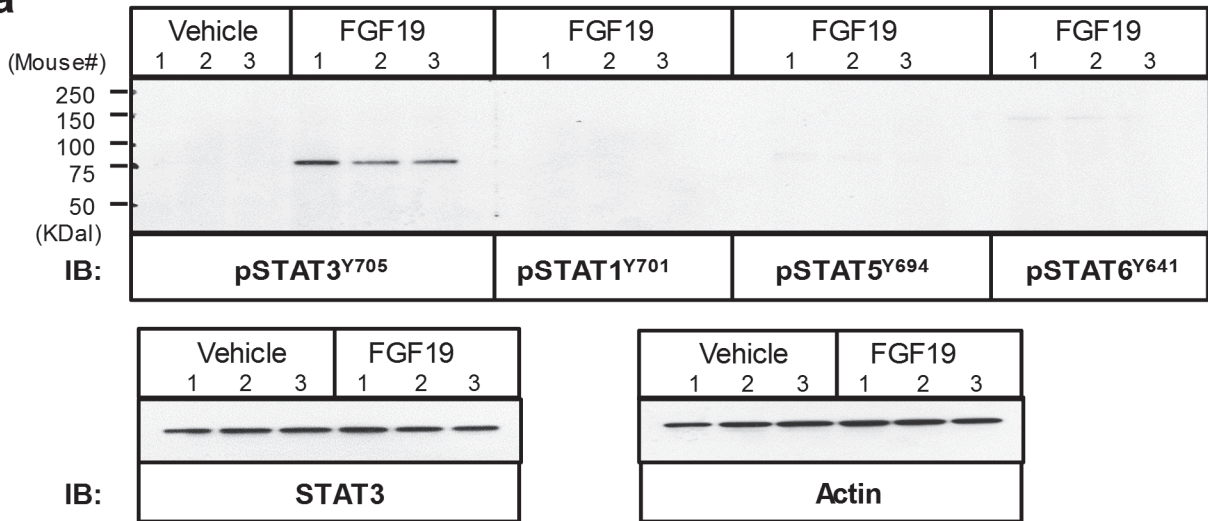
**a****b**



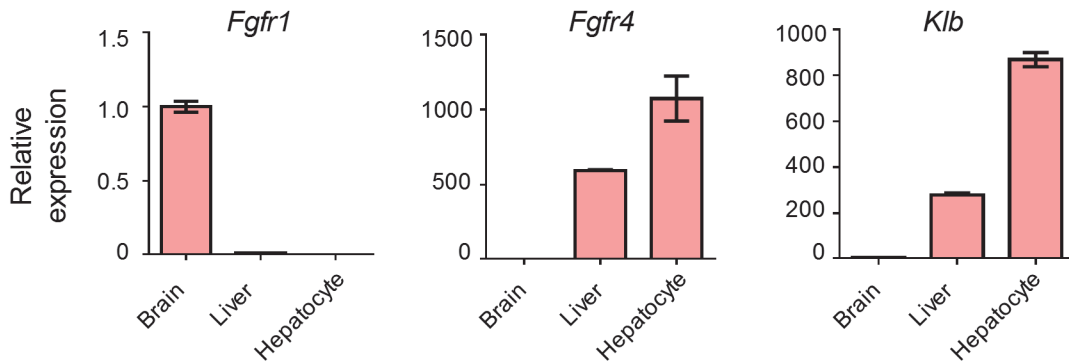
**Supplementary Figure 1. Hepatocyte-specific ablation of STAT3 eliminates FGF19-induced pro-tumorigenic gene expression.** (a) Time course of circulating FGF19 levels in *Stat3<sup>fl/fl</sup>* mice following AAV-FGF19 administration. 14 to 18 week-old *Stat3<sup>fl/fl</sup>* mice ( $n = 8$ ) received a single tail vein injection of AAV-FGF19. Blood samples were taken prior to AAV injection, and at 1, 5, and 12 months after AAV injection. FGF19 levels were determined by ELISA. (b) Tumor area for individual mouse by immunohistochemistry. Liver sections were stained with anti-glutamine synthetase and DAB substrates (brown color). Tumors are indicated by dashed lines; scale bars, 5 mm. (c) Immunohistochemical staining with antibodies against Ki67, a marker of proliferation. Note that FGF19 increases Ki-67-positive cells in *Stat3<sup>fl/fl</sup>* mice, but not *Stat3<sup>ΔHep</sup>* mice. Scale bars, 200  $\mu\text{m}$ .  $n = 8$  mice per group. Quantification of Ki-67 labeling index is shown as a bar graph. (d) STAT3 target genes, *Birc5* (*Survivin*), *Bcl2l1* (*Bcl-xL*), and *Ccnd1* (*Cyclin D1*), were induced by FGF19 (19) in *Stat3<sup>fl/fl</sup>* mice, but not *Stat3<sup>ΔHep</sup>* mice. Mice were euthanized 12 months after AAV-FGF19 administration. qRT-PCR analysis were performed on RNA samples extracted from the liver. C, control virus. (e) Cyclins were induced by FGF19 in *Stat3<sup>fl/fl</sup>* mice, but not *Stat3<sup>ΔHep</sup>* mice. (f) FGF19 induces proliferative and HCC-related genes in *Stat3<sup>fl/fl</sup>* mice, but not *Stat3<sup>ΔHep</sup>* mice. Values are mean  $\pm$  s.e.m. \*\*\* $P < 0.001$ , \*\* $P < 0.01$ , \* $P < 0.05$  versus control virus-injected *Stat3<sup>fl/fl</sup>* mice by one-way ANOVA.



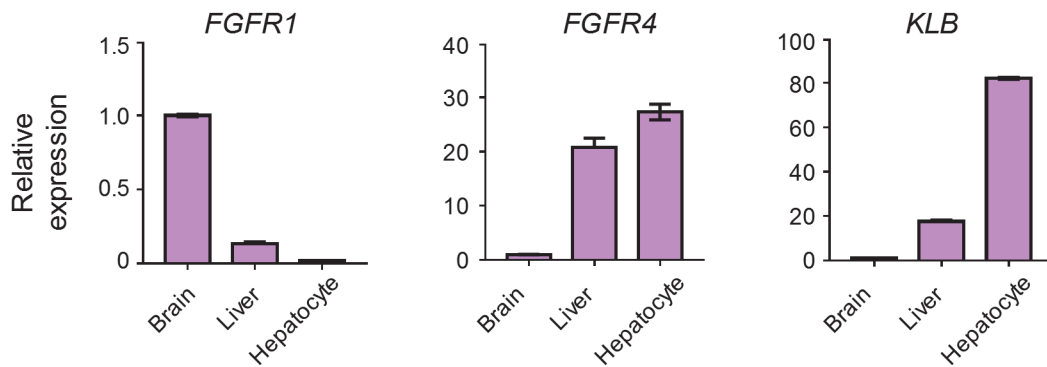
**Supplementary Figure 2. Loss of STAT3 in hepatocytes does not impair FGF19-dependent improvement in steatosis.** *Stat3<sup>f/f</sup>* or *Stat3<sup>ΔHep</sup>* mice were euthanized 12 months after a tail vein injection of AAV-FGF19 ( $n = 8$ ) or a control virus ( $n = 5$ ). Representative H & E images of liver sections are shown. Note vacuoles (steatosis) present in both *Stat3<sup>f/f</sup>* and *Stat3<sup>ΔHep</sup>* mice treated with the control virus, but not in FGF19-treated mice. Scale bars, 100  $\mu$ m.

**a****b**

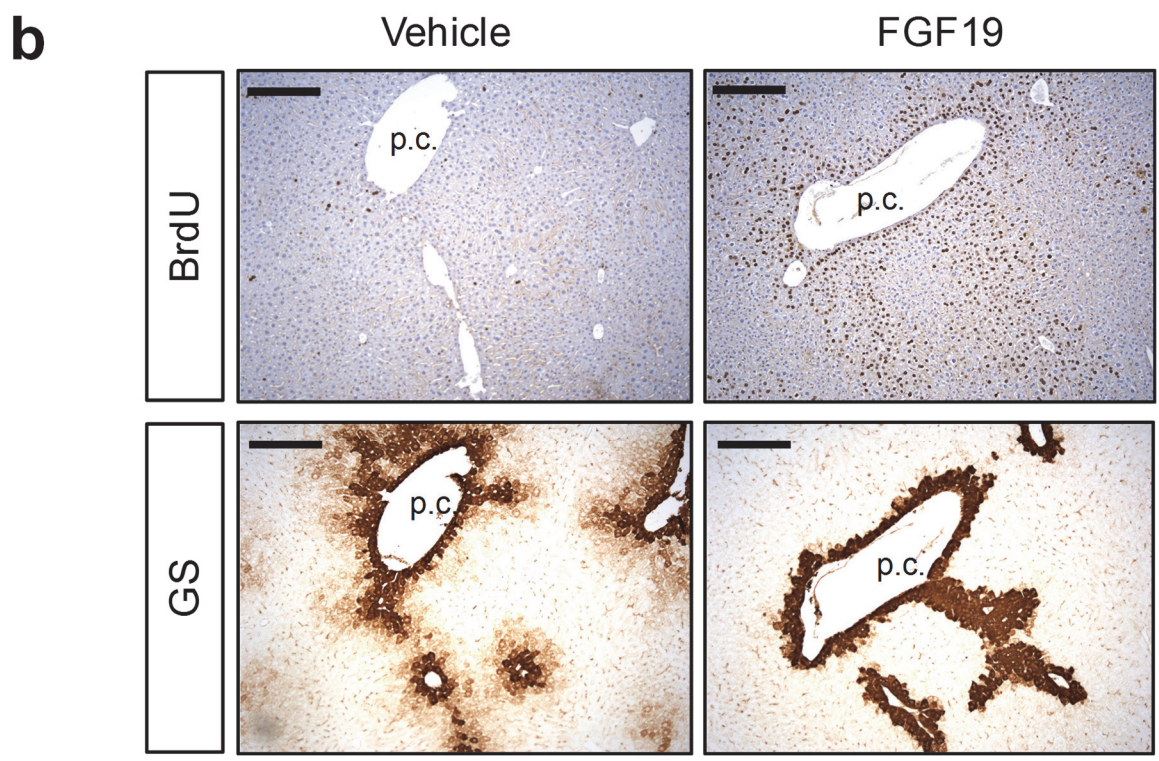
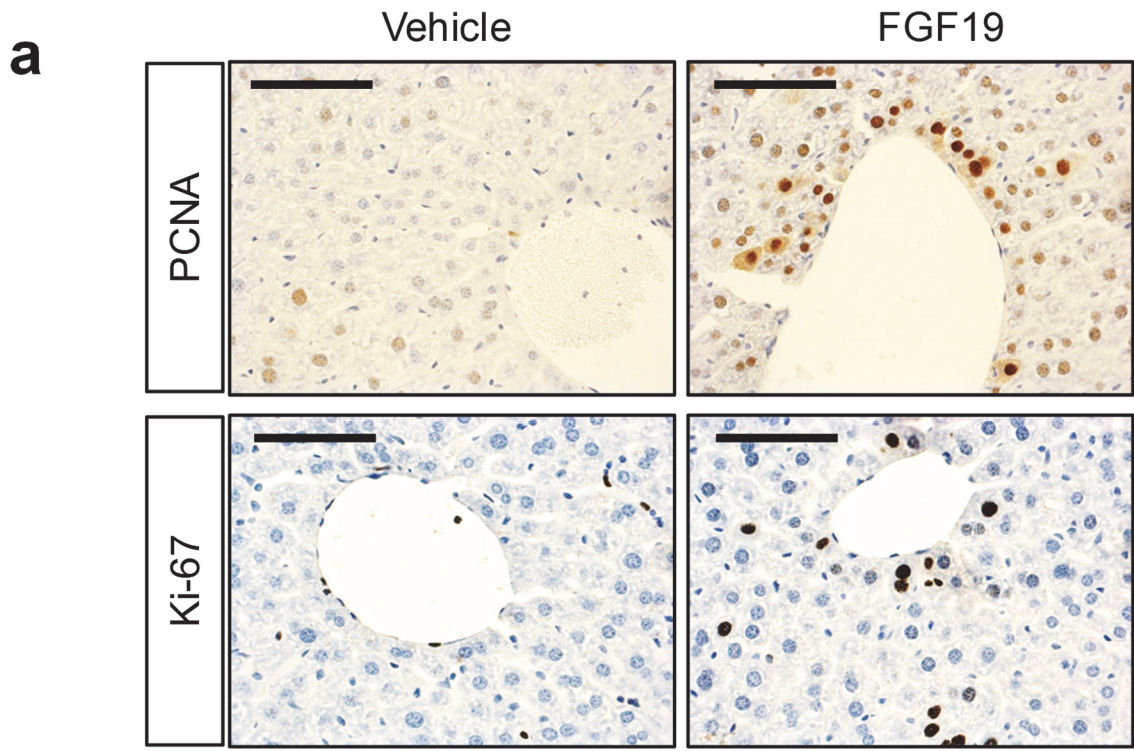
Mouse hepatocytes



Human hepatocytes

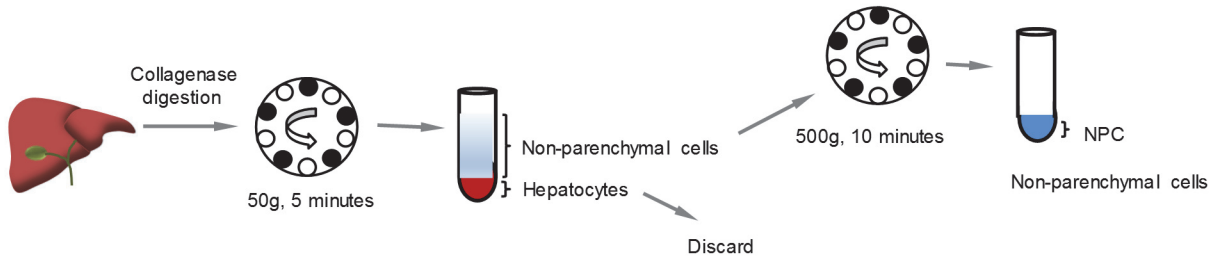
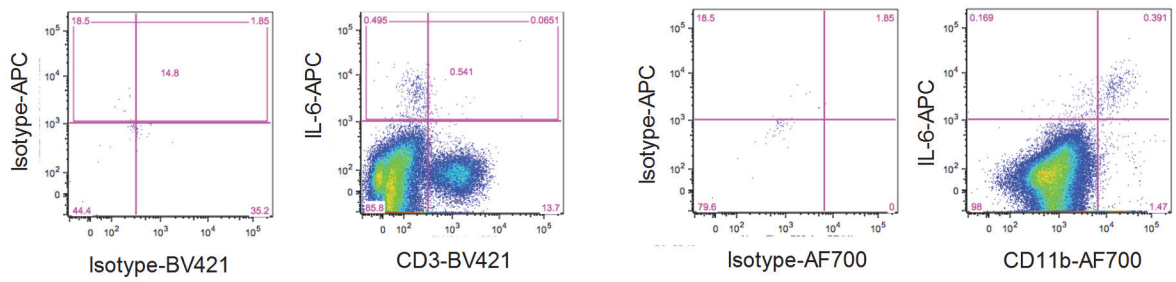
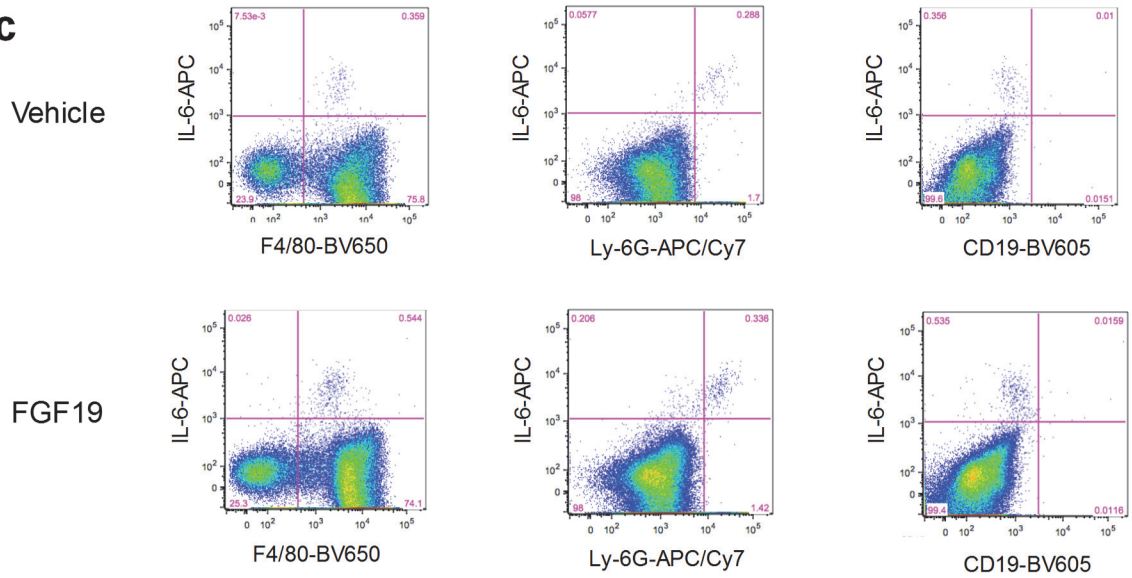


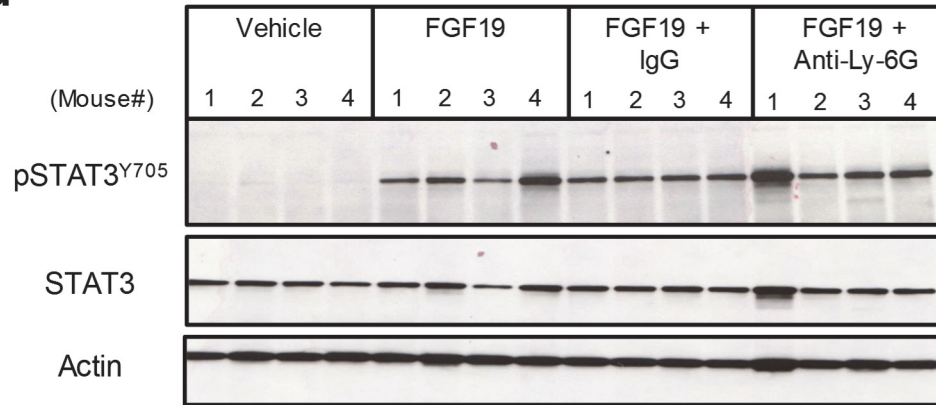
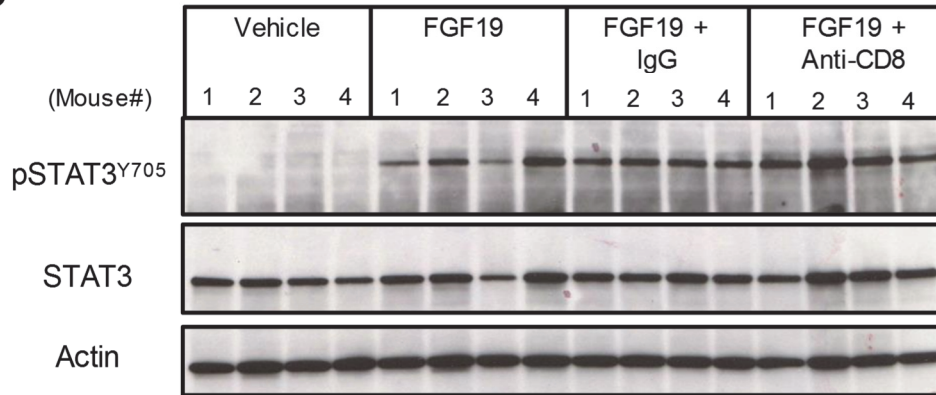
**Supplementary Figure 3. Specificity of FGF19-induced STAT3 phosphorylation in vivo and receptor expression in isolated hepatocytes.** (a) Phosphorylation of STAT3 at tyrosine residue 705 (pSTAT3<sup>Y705</sup>), but not the conserved tyrosine residue in other STAT isoforms, was induced by FGF19. 11~12-week old *db/db* mice ( $n = 3$  per group) received a single intraperitoneal injection of 1 mg/kg FGF19 or vehicle, and livers were harvested 2 hours post dose. Shown are immunoblot analyses of liver lysates with antibodies against pSTAT3<sup>Y705</sup>, pSTAT1<sup>Y701</sup>, pSTAT5<sup>Y694</sup>, pSTAT6<sup>Y641</sup>, STAT3 and anti- $\beta$ -actin. (b) Primary hepatocytes were isolated by collagenase digestion method from mouse or human livers. Real-time quantitative RT-PCR was performed to determine mRNA levels of FGFR1, FGFR4, and KLB receptors. Data are normalized to housekeeping gene GAPDH, and are relative to the expression in the brain.



**Supplementary Figure 4. Non-cell-autonomous promotion of hepatocellular proliferation by FGF19.** (a) FGF19 increases PCNA and Ki-67 staining in the liver. Mice ( $n = 5$  per group) were implanted with osmotic pumps releasing BrdU (8.5 mg/kg/day) and FGF19 protein (0.4 mg/kg/day). Livers were harvested 6 days post implant. Shown are representative images of livers stained with anti-PCNA or anti-Ki-67 antibodies followed by DAB substrates (brown color). Sections were counter-stained with hematoxylin. Scale bars, 100  $\mu\text{m}$ . (b) FGF19 increases proliferation of peri-central hepatocytes. Adjacent liver sections from mice implanted with osmotic pumps releasing BrdU and FGF19 were stained with anti-BrdU (top panels) or anti-glutamine synthetase (marker of peri-central hepatocytes, bottom panels). DAB substrates (brown color) were used for color development. *p.c.*, peri-central. Scale bars, 100  $\mu\text{m}$ .



**a****b****c**

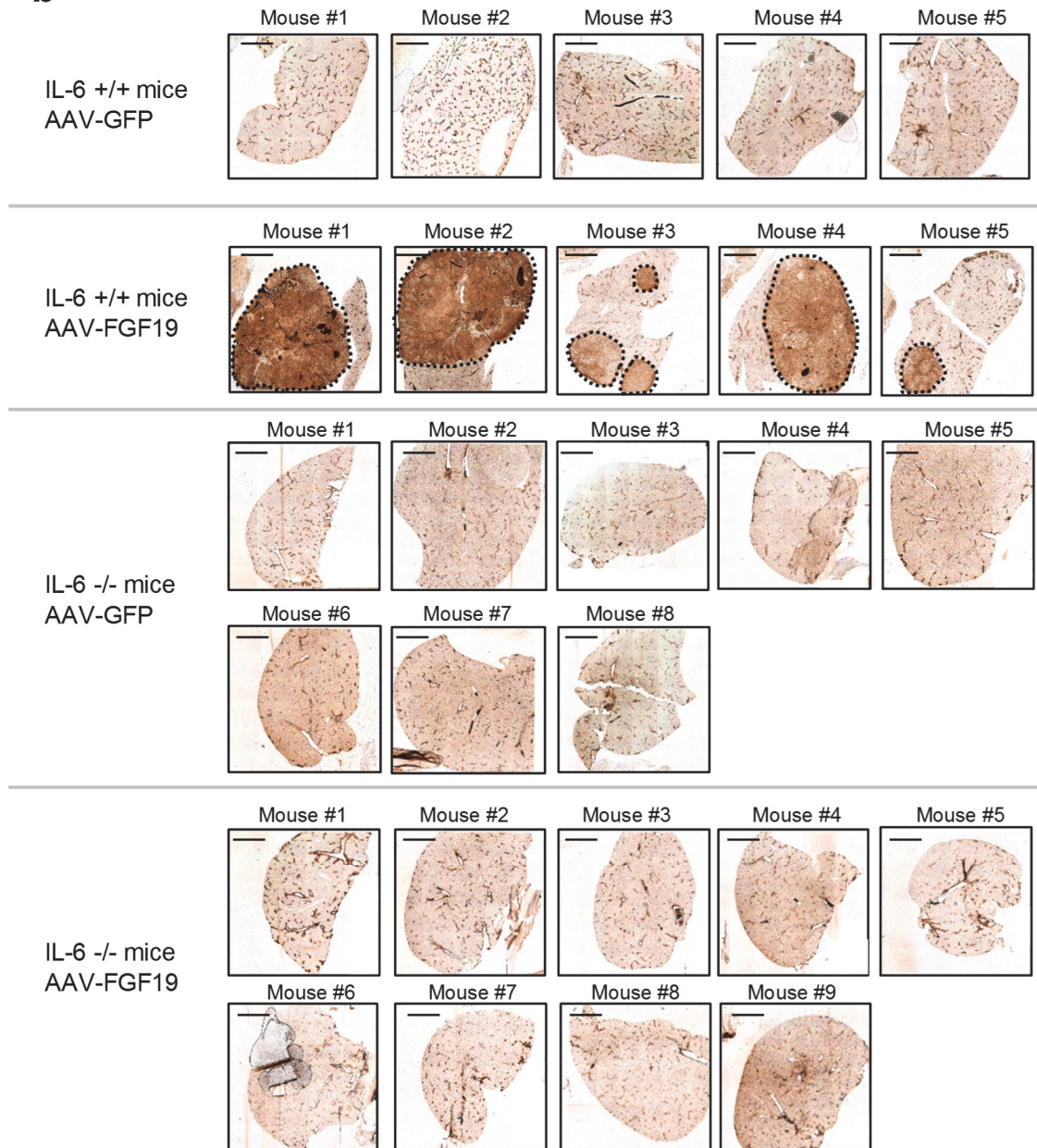
**d****e**

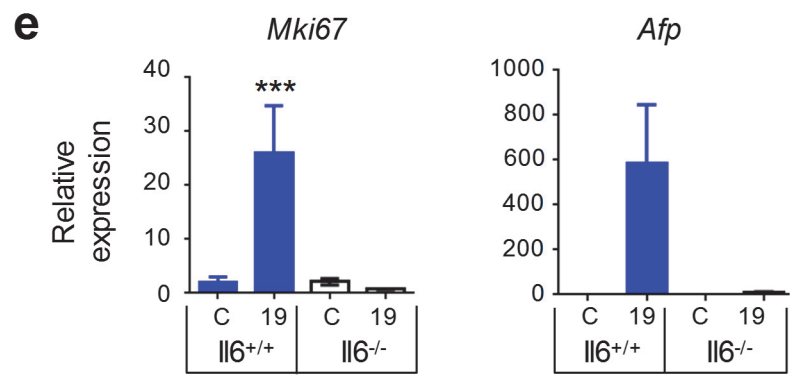
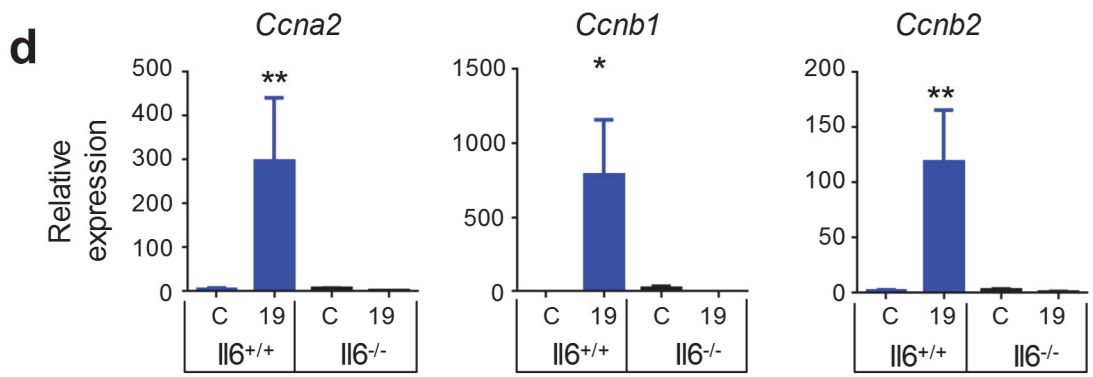
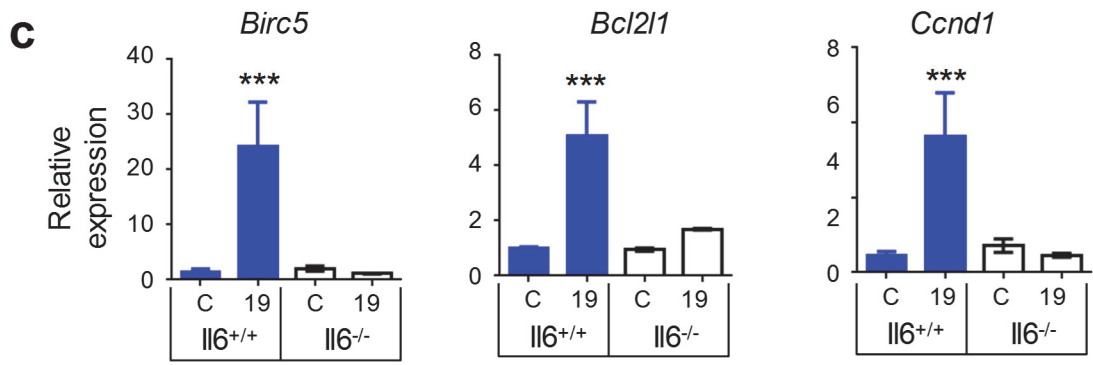
**Supplementary Figure 5. Identification of secreted factor(s) mediating non-cell-autonomous activation of STAT3 by FGF19.** (a) Schematic diagram of isolation of hepatic non-parenchymal (NPC) cells. (b-c) Intracellular IL-6 cytokine staining of non-parenchymal cells analyzed by flow cytometry. Isolated hepatic non-parenchymal cells were incubated in RPMI1640 media containing vehicle or 50 ng/mL FGF19 and protein transport inhibitors (10  $\mu$ M brefeldin A and 2 mM monensin). Cells were subsequently stained with fluorophore-labeled antibodies (CD45, clone 30F-11; CD3, clone 17A2; CD19, clone 6D5; CD11b, clone M1/70; Ly6-G, clone 1A8; NK1.1, clone PK136; F4/80, clone BM8). Surface-stained cells were fixed and permeabilized, and incubated with anti-IL-6-APC (clone MP5-20F3). Staining with isotype control antibodies was included as negative controls in b. (d-e) Effect of immune cell depletion on FGF19-induced pSTAT3 activation. Antibody-mediated depletion of neutrophils (d) or CD8<sup>+</sup> T cells (e) was performed prior to FGF19 injection in *db/db* mice ( $n = 4$  per group). Isotype IgG serves as control. Livers were harvested 2 hours after FGF19 (1 mg/kg) injection. Shown are immunoblots of pSTAT3<sup>Y705</sup>, total-STAT3 and anti- $\beta$ -actin in liver lysates.

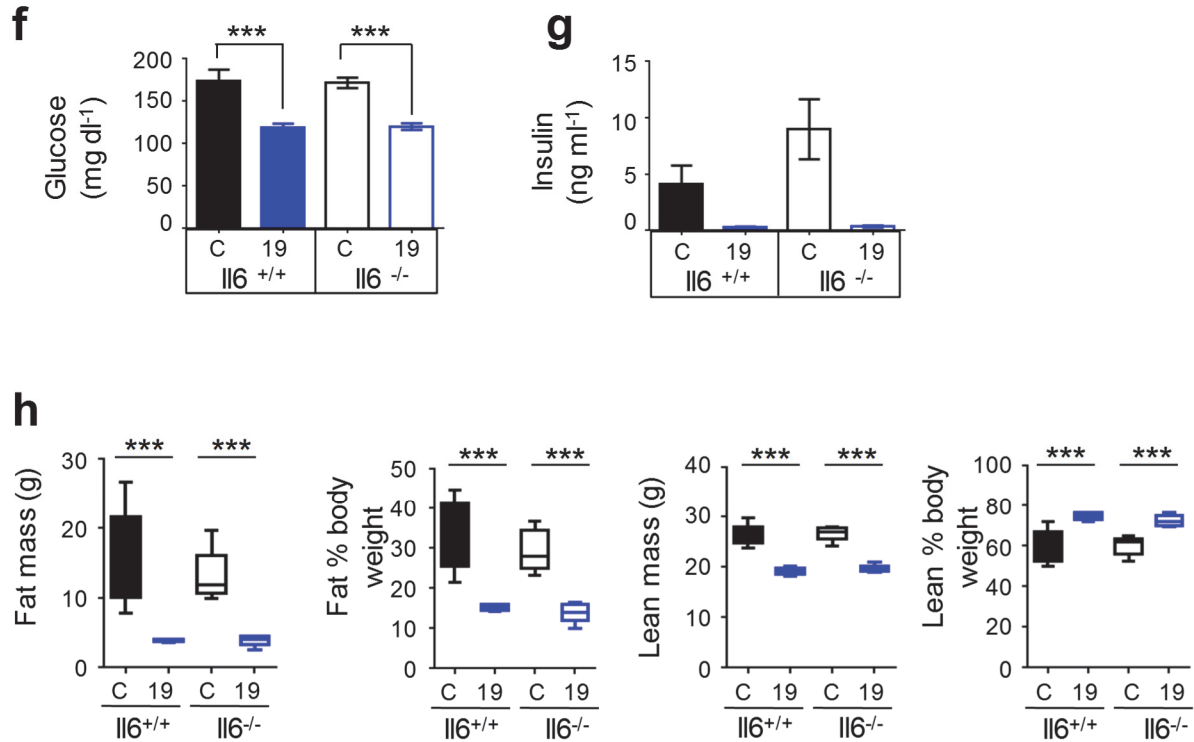
**a**



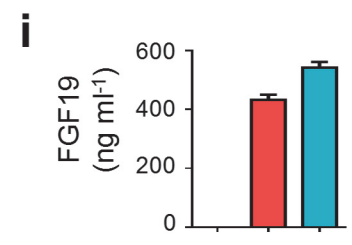
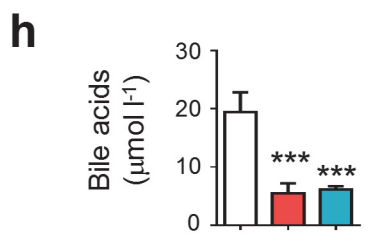
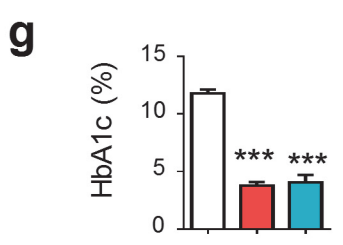
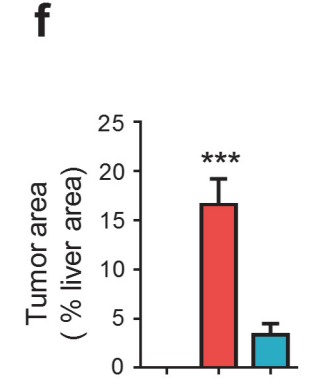
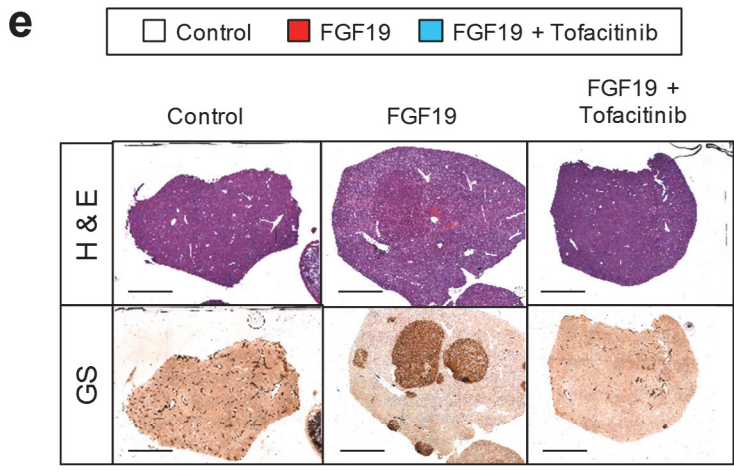
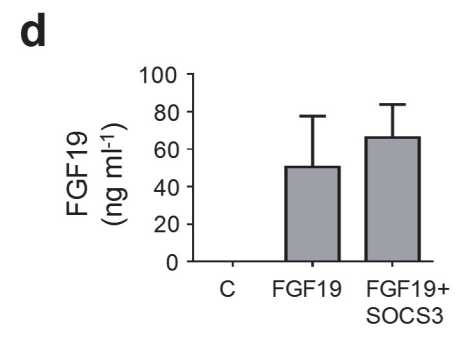
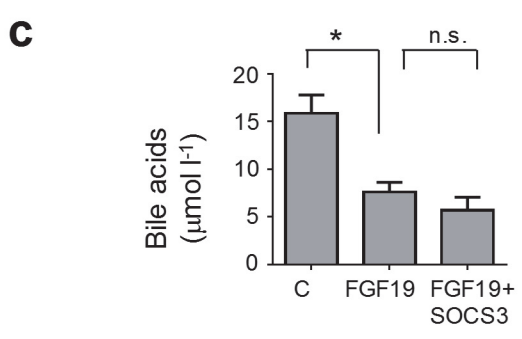
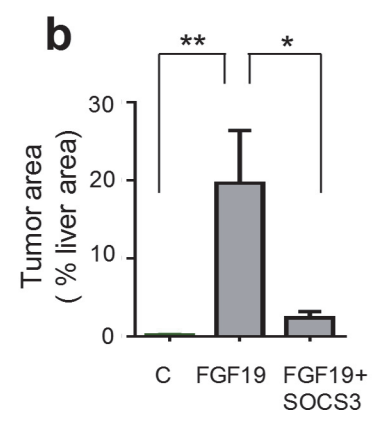
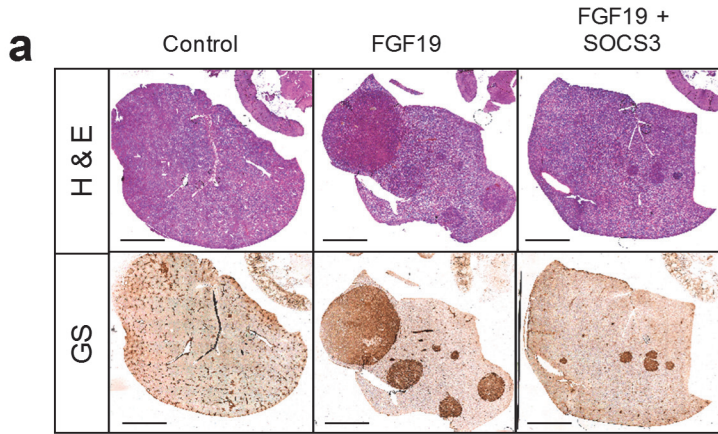
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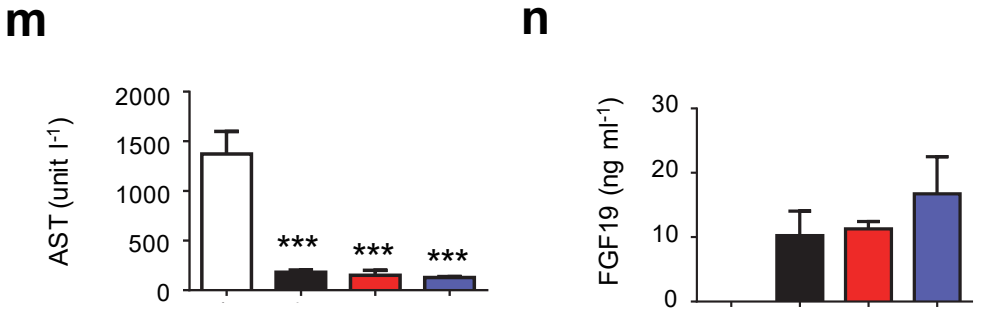
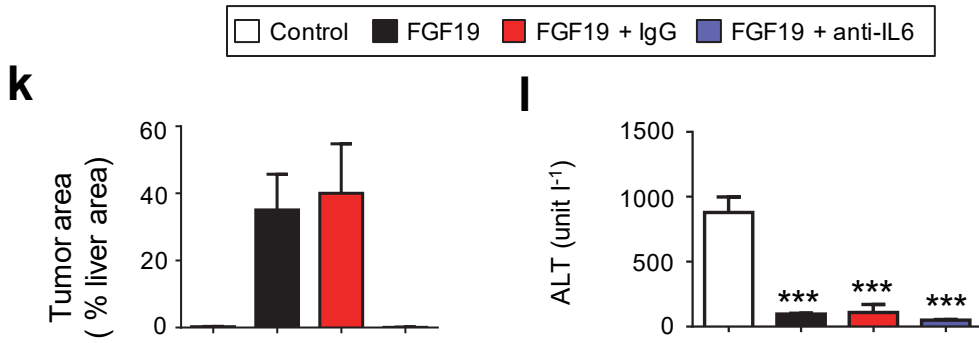
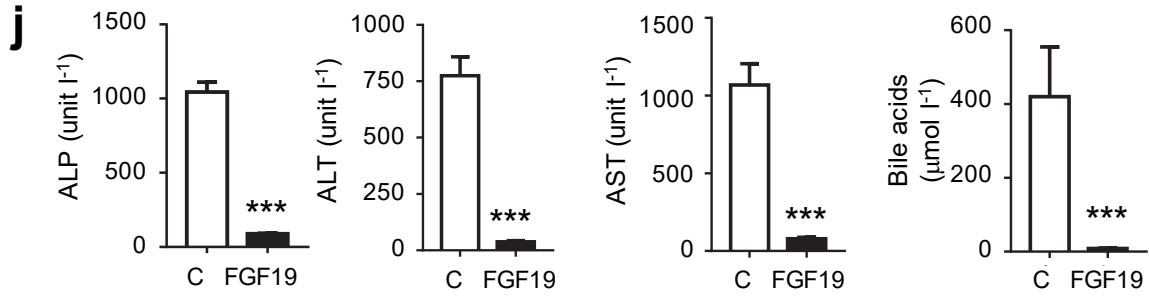






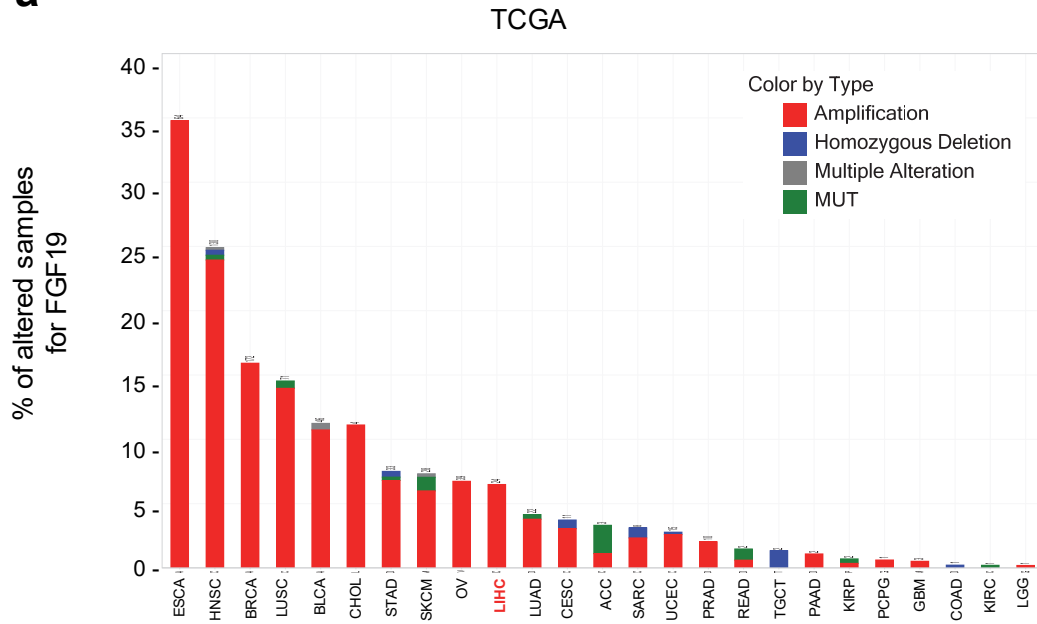
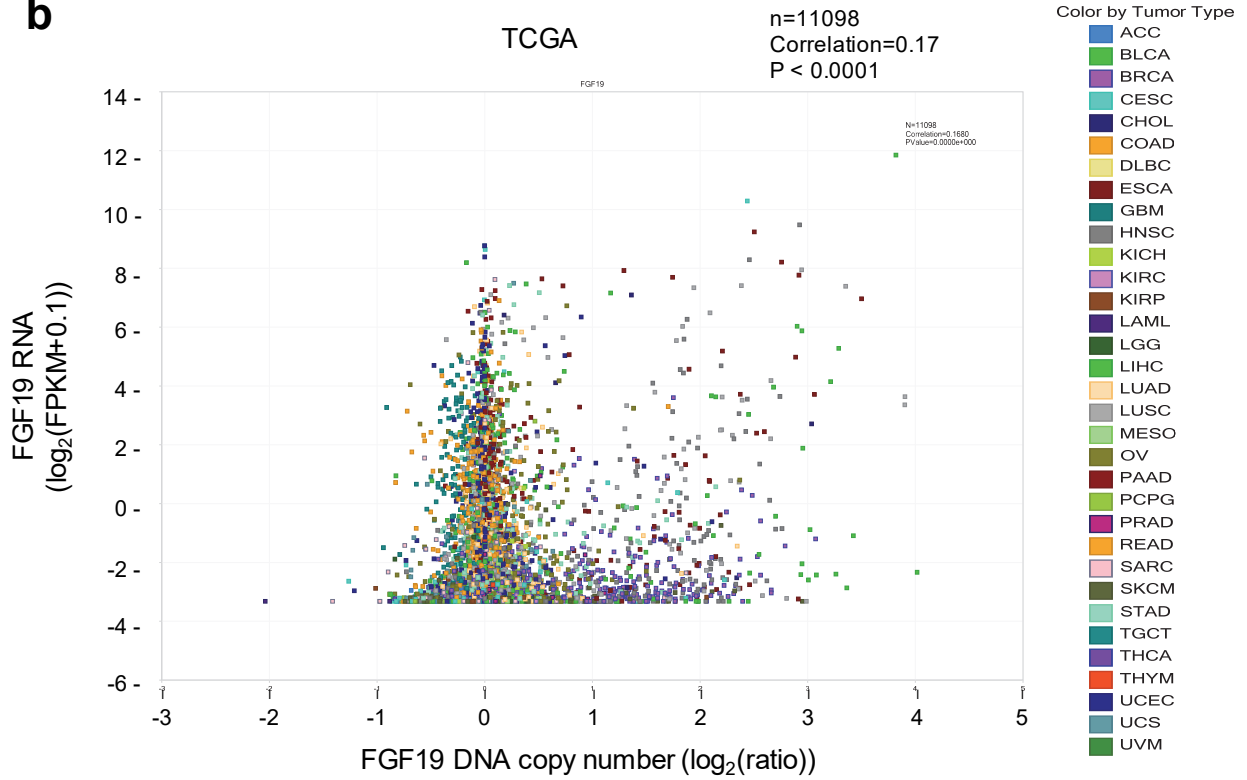
**Supplementary Figure 6. Homozygous deletion of IL-6 prevents FGF19-induced HCC development while sparing FGF19-controlled metabolic improvements.** (a) Study design. *Il6*<sup>+/+</sup> or *Il6*<sup>-/-</sup> mice received a single tail vein injection of AAV-FGF19 (19) or a control (C) virus. Body weight, glucose, insulin, and body composition were determined in live animals 11 months post-dose. Mice were sacrificed 12 months after AAV administration for tumor and gene expression analysis. Animal groups are *Il6*<sup>+/+</sup> control ( $n = 5$ ), *Il6*<sup>+/+</sup>FGF19 ( $n = 5$ ), *Il6*<sup>-/-</sup> control ( $n = 8$ ), *Il6*<sup>-/-</sup>FGF19 ( $n = 9$ ). (b) Tumor area for individual mouse by immunohistochemistry. Liver sections were stained with anti-glutamine synthetase and DAB substrates (brown color). Tumors are indicated by dashed lines; scale bars, 5 mm. (c) STAT3 target genes, *Birc5*, *Bcl2l1*, and *Ccnd1*, were induced by FGF19 in *Il6*<sup>+/+</sup> mice, but not *Il6*<sup>-/-</sup> mice. (d) Cyclins were induced by FGF19 in *Il6*<sup>+/+</sup> mice, but not *Il6*<sup>-/-</sup> mice. (e) FGF19 induces proliferative and HCC-related genes in *Il6*<sup>+/+</sup> mice, but not *Il6*<sup>-/-</sup> mice. (f-h) Glucose (f), insulin (g), and body composition (h) of the animals. Values are mean  $\pm$  s.e.m. \*\*\* $P < 0.001$ , \*\* $P < 0.01$ , \* $P < 0.05$  versus control virus-injected *Il6*<sup>+/+</sup> mice by one-way ANOVA.

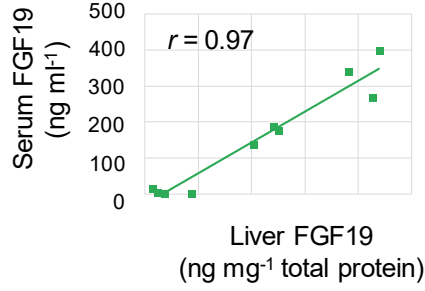
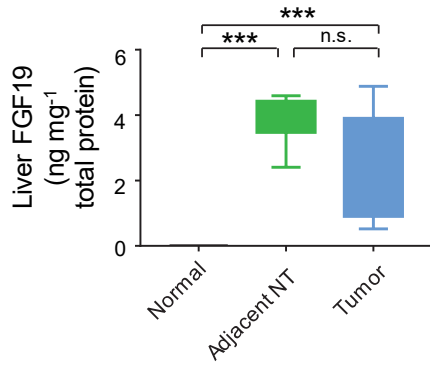
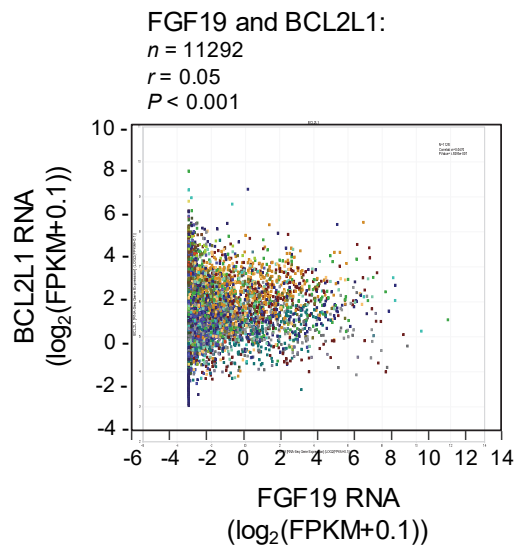
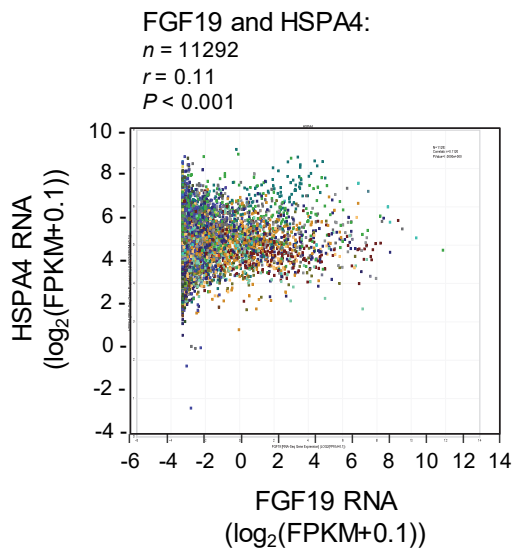
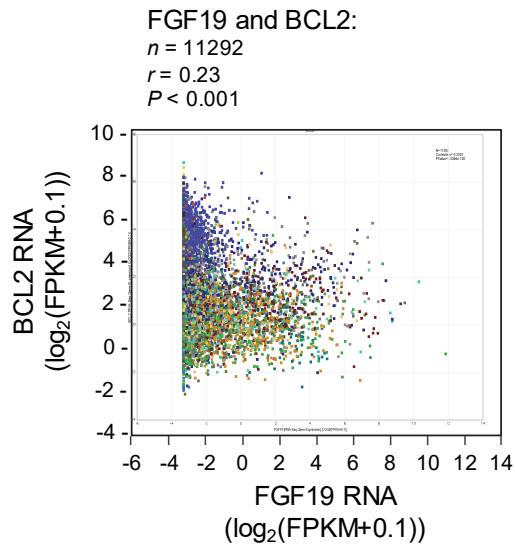
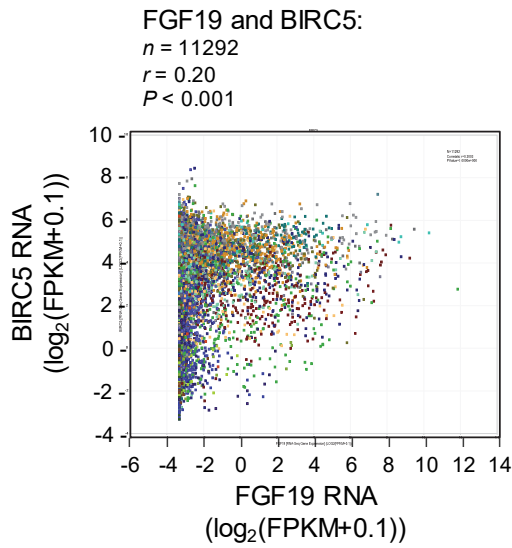




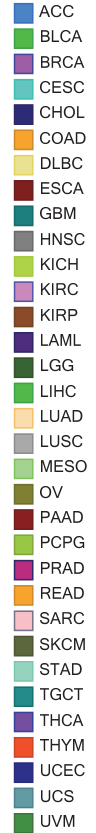


**Supplementary Figure 7. Pharmacological inhibition of the STAT3/IL-6 axis abolishes FGF19-dependent HCC. (a-d) AAV-SOCS3 study in *db/db* mice.** 11~12-week old *db/db* mice ( $n = 5$  per group) were *i.v.* administered with AAV-FGF19 with or without AAV-SOCS3, or a control virus. Mice were euthanized 24 weeks later for liver tumor analysis. Representative images of livers **(a)** (top panels, H&E staining; bottom panels, anti-glutamine synthetase staining with DAB substrates; scale bars, 5 mm), quantification of glutamine synthetase-positive tumor area **(b)**, serum levels of total bile acids **(c)** and FGF19 **(d)** are shown. **(e-i) Tofacitinib study in *db/db* mice.** 11~12-week old *db/db* mice ( $n = 5$  per group) were *i.v.* administered with AAV-FGF19 or a control virus. Tofacitinib treatment was initiated 4 weeks later. Mice were euthanized 24 weeks after AAV injection. Representative images of livers **(e)** (top panels, H&E staining; bottom panels, anti-glutamine synthetase staining with DAB substrates; scale bars, 5 mm), quantification of glutamine synthetase-positive tumor area **(f)**, blood levels of HbA1c **(g)**, total bile acids **(h)** and FGF19 **(i)** are shown. **(j-n) Anti-IL-6 study in *Mdr2*<sup>-/-</sup> mice.** *Mdr2*<sup>-/-</sup> mice received a single tail vein injection of AAV-FGF19 or a control virus. Starting from week 14 after AAV injection, AAV-FGF19-injected mice were dosed intraperitoneally with 10 mg/kg anti-mouse IL-6 ( $n = 10$ ) or an isotype control antibody ( $n = 8$ ) weekly. Blood concentrations of liver enzymes were measured prior to antibody administration at week 14 **(j)**. At the end of the study, glutamine positive tumor areas were quantified **(k)**, and serum levels of ALT **(l)**, AST **(m)**, and FGF19 **(n)** were determined. Values are mean  $\pm$  s.e.m. \*\*\* $P < 0.001$ , \*\* $P < 0.01$ , \* $P < 0.05$  by unpaired two-tailed t-test when comparing two groups, or versus control group by one-way ANOVA when comparing multiple groups.

**a****b**

**c****d**

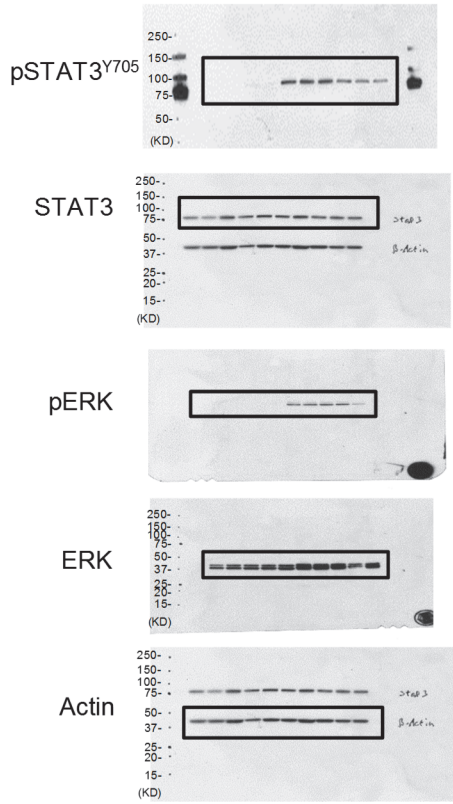
Color by Tumor Type



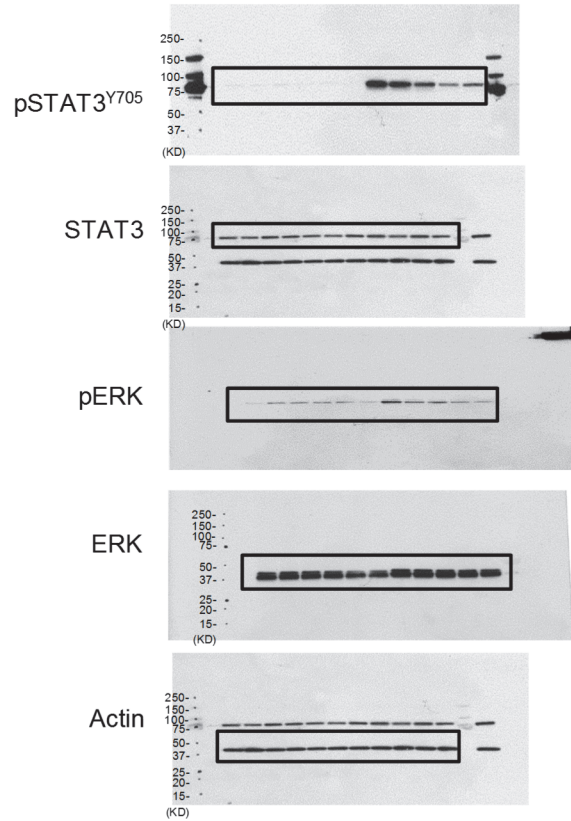
**Supplementary Figure 8. Genetic alterations and correlations in FGF19 in human cancers.**

**(a)** Frequency of FGF19 genetic alterations in human tumors from the Cancer Genome Atlas (TCGA) database. Types of alterations include amplification, homozygous deletion, mutation and multiple alterations. Genetic alterations in genes encoding CTNNB1, PTEN and TP53 are shown for comparison. **(b)** Scatter plot of FGF19 DNA copy number versus mRNA expression in TCGA database. Spearman correlation coefficient ( $r$ ) and  $P$  value are displayed. mRNA levels retrieved from RNA-seq data are expressed as Fragments Per Kilobase of transcript per Million mapped reads (FPKM). **(c)** Quantification of FGF19 levels in liver tumors and adjacent non-tumor (NT) liver tissues in mice injected with AAV-FGF19. Note that FGF19 is undetectable in normal livers from untreated mice. FGF19 concentrations in liver extracts were determined by ELISA, and expressed as ng per mg of total liver protein. Scatter plot shows serum FGF19 concentrations correlate with FGF19 levels in the liver in mice injected with AAV-FGF19. \*\*\* $P < 0.001$ ; n.s., not statistically significant. **(d)** FGF19 mRNA levels correlate with the expression of STAT3 target genes in human cancers in TCGA database. Correlation of RNA FPKM for FGF19 and BIRC5, BCL2, HSPA4, or BCL2L1 is shown as scatter plots, with each tumor type indicated on the right. Spearman correlation coefficient ( $r$ ) and  $P$  value are displayed.

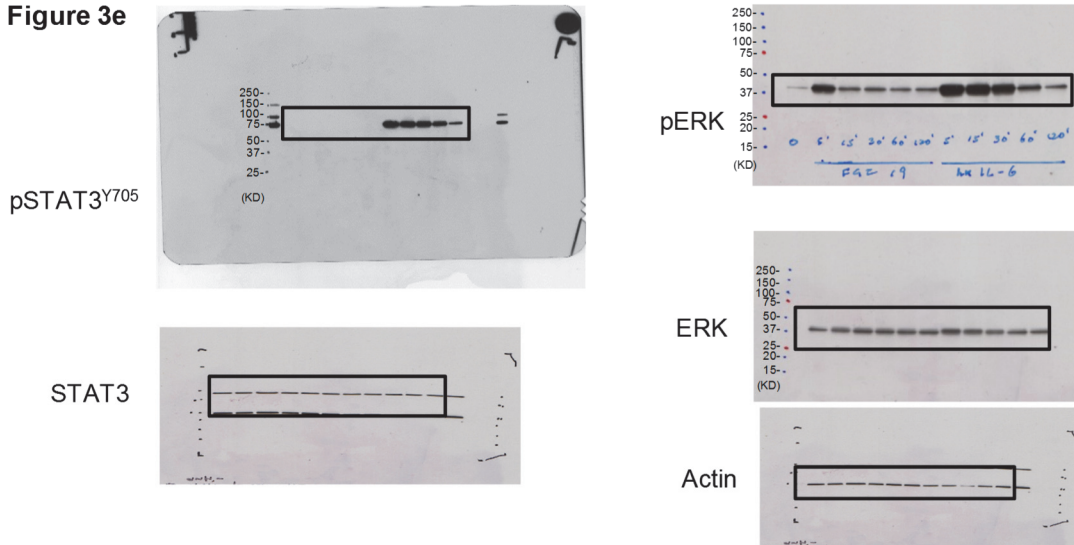
**Figure 3b**

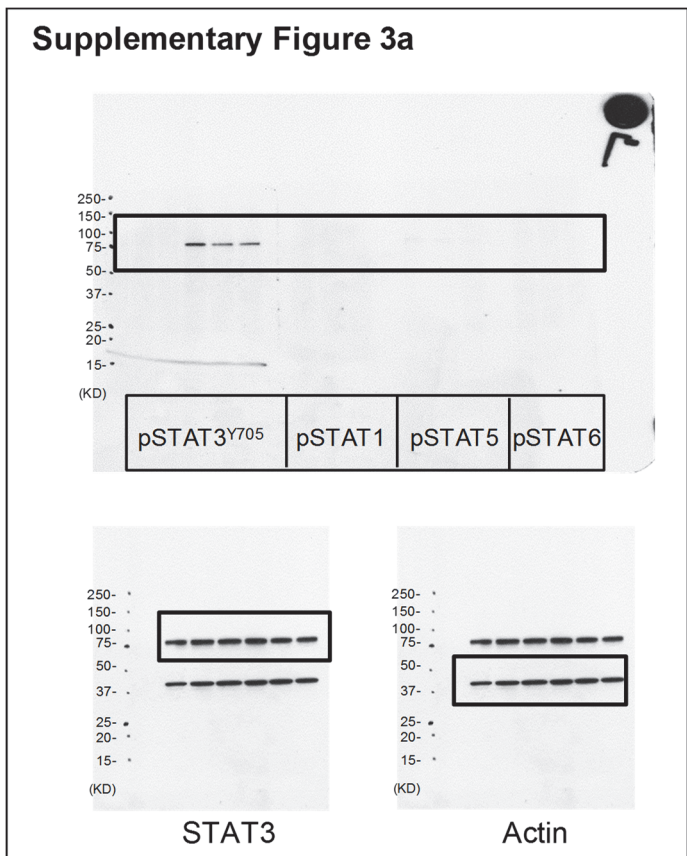
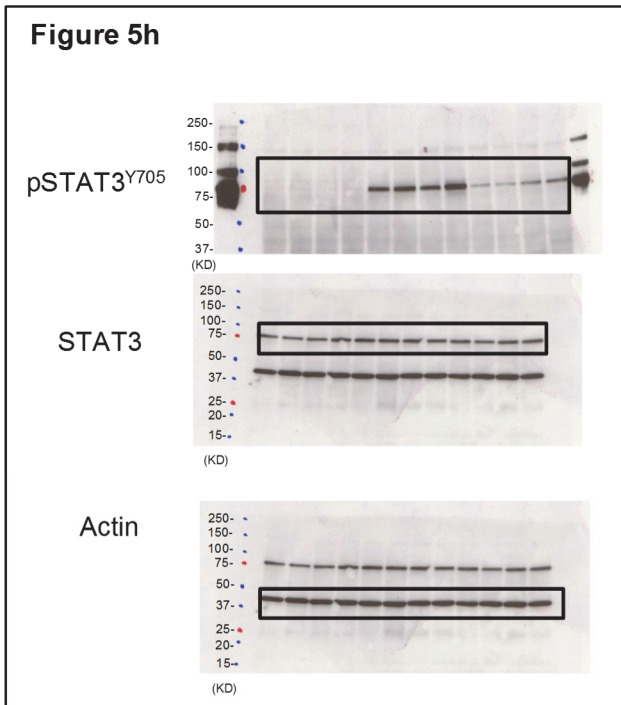
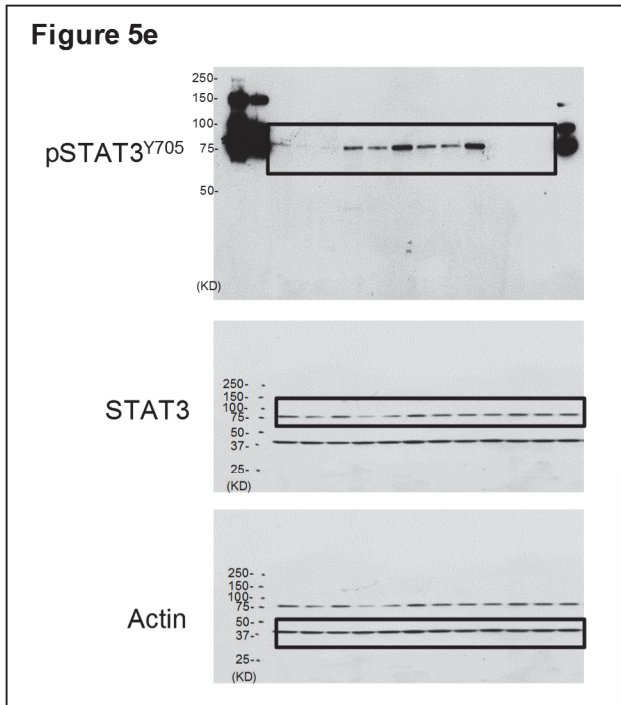


**Figure 3d**

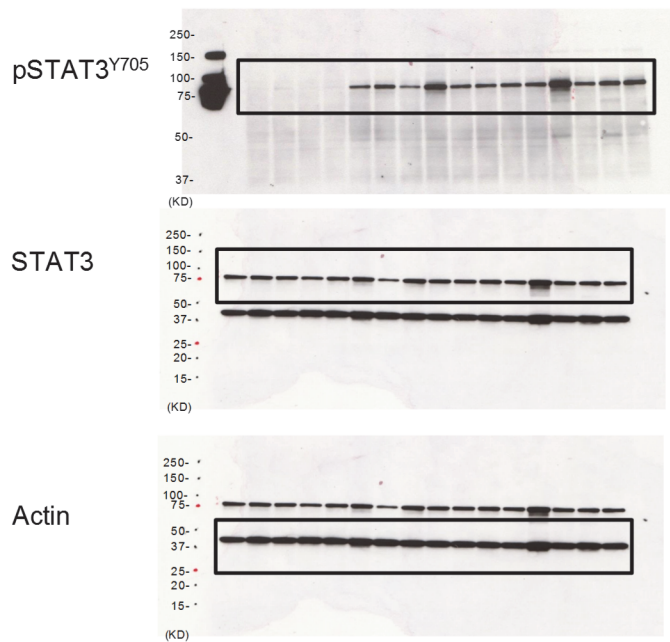


**Figure 3e**

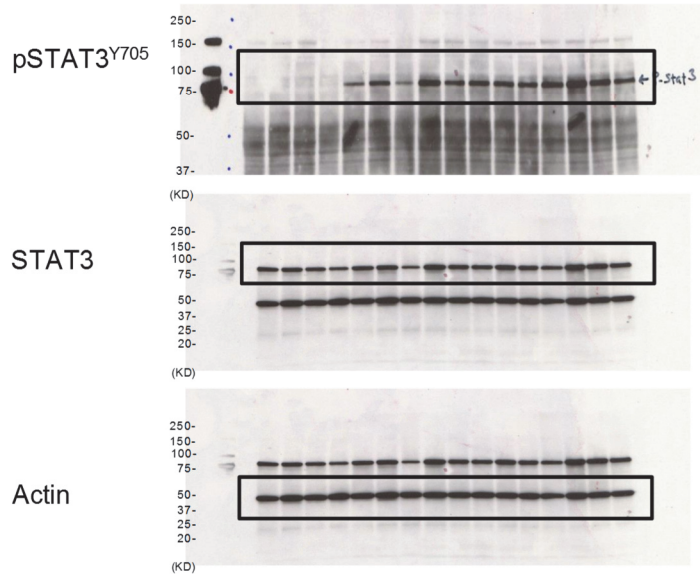




**Supplementary Figure 5d**



**Supplementary Figure 5e**



**Supplementary Figure 9. Uncropped immunoblots with markers of molecular weights.**

**Supplementary Table 1. TCGA datasets used in the current study**

<b>Tumor Type Abbreviations</b>	<b>Tumor Type Full Name</b>	<b>Number of Patient Samples</b>
ACC	Adrenocortical carcinoma	184
BLCA	Bladder Urothelial Carcinoma	846
BRCA	Breast invasive carcinoma	2298
CESC	Cervical squamous cell carcinoma and endocervical adenocarcinoma	620
CHOL	Cholangiocarcinoma	115
COAD	Colon adenocarcinoma	989
DLBC	Lymphoid Neoplasm Diffuse Large B-cell Lymphoma	116
ESCA	Esophageal carcinoma	377
GBM	Glioblastoma multiforme	1174
HNSC	Head and Neck squamous cell carcinoma	1123
KICH	Kidney Chromophobe	226
KIRC	Kidney renal clear cell carcinoma	1101
KIRP	Kidney renal papillary cell carcinoma	614
LAML	Acute Myeloid Leukemia	699
LGG	Brain Lower Grade Glioma	1051
LIHC	Liver hepatocellular carcinoma	796
LUAD	Lung adenocarcinoma	1288
LUSC	Lung squamous cell carcinoma	1083
MESO	Mesothelioma	174
OV	Ovarian serous cystadenocarcinoma	1221
PAAD	Pancreatic adenocarcinoma	377
PCPG	Pheochromocytoma and Paraganglioma	366
PRAD	Prostate adenocarcinoma	1063
READ	Rectum adenocarcinoma	349
SARC	Sarcoma	531
SKCM	Skin Cutaneous Melanoma	951
STAD	Stomach adenocarcinoma	940
TGCT	Testicular Germ Cell Tumors	306
THCA	Thyroid carcinoma	1053
THYM	Thymoma	250
UCEC	Uterine Corpus Endometrial Carcinoma	1131
UCS	Uterine Carcinosarcoma	114
UVM	Uveal Melanoma	160



**Supplementary Table 2. Patient information for duplex (FGF19 + BIRC5) RNAScope *in situ* hybridization on formalin-fixed, paraffin-embedded normal liver tissues**

Subject ID	Gender	Age	Organ	Pathology	Grade	Stage	TNM	Type
1	F	14	Liver	Normal hepatic tissue	-	-	-	Normal
2	F	50	Liver	Normal hepatic tissue	-	-	-	Normal
3	F	35	Liver	Normal hepatic tissue	-	-	-	Normal
4	M	40	Liver	Normal hepatic tissue	-	-	-	Normal
5	M	40	Liver	Normal hepatic tissue	-	-	-	Normal
6	M	38	Liver	Normal hepatic tissue	-	-	-	Normal
7	M	45	Liver	Normal hepatic tissue	-	-	-	Normal
8	F	18	Liver	Normal hepatic tissue	-	-	-	Normal
9	M	43	Liver	Normal hepatic tissue	-	-	-	Normal
10	M	16	Liver	Normal hepatic tissue	-	-	-	Normal

**Supplementary Table 3. Patient information for quantitative RT-PCR analysis of STAT3 target genes on RNA extracted from frozen normal liver tissues, FGF19-expressing, and FGF19-non-expressing HCCs**

Subject ID	Gender	Age	Organ	Pathology	Source	Catalog Number	Type	FGF19
N1	M	53	Liver	Normal hepatic tissue	Bioreclamation	MHU-L-112210	Normal	
N2	M	49	Liver	Normal hepatic tissue	Bioreclamation	MHU-L-040211	Normal	
N3	M	38	Liver	Normal hepatic tissue	Bioreclamation	MHU-L-071907	Normal	
N4	M	37	Liver	Normal hepatic tissue	Bioreclamation	MHU-L-122108	Normal	
N5	M	62	Liver	Normal hepatic tissue	Bioreclamation	MHU-L-100510	Normal	
T1	F	51	Liver	Hepatocellular carcinoma	CrownBio	LI0574	Malignant	FGF19-expressing
T2	M	59	Liver	Hepatocellular carcinoma	CrownBio	LI1054	Malignant	FGF19-expressing
T3	M	53	Liver	Hepatocellular carcinoma	CrownBio	LI1081	Malignant	FGF19-expressing
T4	M	65	Liver	Hepatocellular carcinoma	CrownBio	LI6619	Malignant	FGF19-expressing
T5	M	45	Liver	Hepatocellular carcinoma	CrownBio	LI6677	Malignant	FGF19-expressing
T6	M	NA	Liver	Hepatocellular carcinoma	CrownBio	LI0801	Malignant	FGF19-non-expressing
T7	M	53	Liver	Hepatocellular carcinoma	CrownBio	LI1025	Malignant	FGF19-non-expressing
T8	F	56	Liver	Hepatocellular carcinoma	CrownBio	LI1037	Malignant	FGF19-non-expressing
T9	M	33	Liver	Hepatocellular carcinoma	CrownBio	LI1097	Malignant	FGF19-non-expressing
T10	M	48	Liver	Hepatocellular carcinoma	CrownBio	LI6686	Malignant	FGF19-non-expressing

**Supplementary Table 4. List of antibodies used for immunohistochemistry and immunoblotting**

<b>Antigen</b>	<b>Vendor</b>	<b>Catalog Number</b>	<b>Host Species</b>	<b>Dilution</b>
<i>For immunohistochemistry:</i>				
BrdU	Dako	M0744	Mouse IgG	1:500
Glutamine synthetase	Abcam	Ab49873	Rabbit IgG	1:2000
Ki-67	Dako	M7249	Mouse IgG	1:500
PCNA	Dako	M0879	Mouse IgG	1:200
STAT3 (total)	Cell Signaling Technology	4904	Rabbit IgG	1:1000
Biotinylated anti-mouse IgG	Vector Laboratories	BA-2000	Horse IgG	1:500
Biotinylated anti-rabbit IgG	Vector Laboratories	BA-1000	Goat IgG	1:500
<i>For immunoblotting:</i>				
$\beta$ -Actin	Sigma	A5441	Mouse IgG	1:10000
ERK1/2 (total)	Santa Cruz	sc-94	Rabbit IgG	1:200
pERK1/2 <sup>T202Y204</sup>	Cell Signaling Technology	4370	Rabbit IgG	1:2000
pSTAT1 <sup>Y701</sup>	Cell Signaling Technology	7649	Rabbit IgG	1:1000
pSTAT3 <sup>Y705</sup>	Cell Signaling Technology	9145	Rabbit IgG	1:1000
pSTAT5 <sup>Y694</sup>	Cell Signaling Technology	4322	Rabbit IgG	1:1000
pSTAT6 <sup>Y641</sup>	Cell Signaling Technology	9361	Rabbit IgG	1:1000
STAT3 (total)	Cell Signaling Technology	4904	Rabbit IgG	1:2000

**Supplementary Table 5. List of antibodies used for flow cytometry**

<b>Antigen</b>	<b>Clone</b>	<b>Vendor</b>	<b>Catalog Number</b>	<b>Dilution</b>
CD3-BV421	17A2	BioLegend	100228	1:40
Rat IgG2b-BV421		BioLegend	400655	1:40
CD11b-AF700	M1/70	eBioscience	56-0112	1:40
Rat IgG2b-AF700		BioLegend	400628	1:40
CD19-BV605	6D5	BioLegend	115540	1:40
Rat IgG2a-BV605		BioLegend	400540	1:40
CD45-FITC	30-F11	BioLegend	103108	1:100
Rat IgG2b-FITC		BioLegend	400606	1:100
F4/80-BV650	BM8	BioLegend	123149	1:40
Rat IgG2a-BV650		BioLegend	400542	1:40
IL-6-APC	MP5-20F3	BioLegend	504507	1:40
Rat IgG1-APC		BioLegend	400411	1:40
Ly-6G-APC/Cy7	1A8	BioLegend	127624	1:40
Rat IgG2a-APC/Cy7		BioLegend	400524	1:40
NK1.1-PE/Cy7	PK136	BioLegend	108714	1:40