

Supplemental Information:

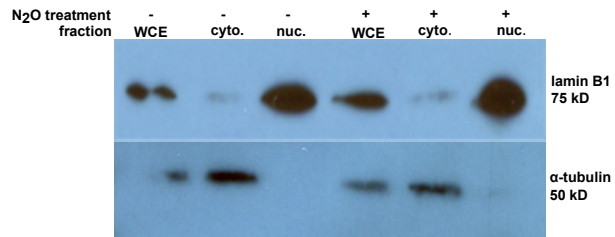


Figure S1. Western blot of whole cell, cytosolic, and nuclear fractions for nuclear (lamin B1) and cytosolic (α -tubulin) marker proteins from HeLa cells harvested in tandem with those cells used for quantifying nuclear one-carbon folate forms (refer to Fig. 2C). The absence of α -tubulin in the lanes corresponding to nuclear fractions confirms the lack of cytosolic contamination in nuclei. Lane designations: WCE, whole cell extract; cyto, cytosolic; nuc, nuclear. N₂O, nitrous oxide.

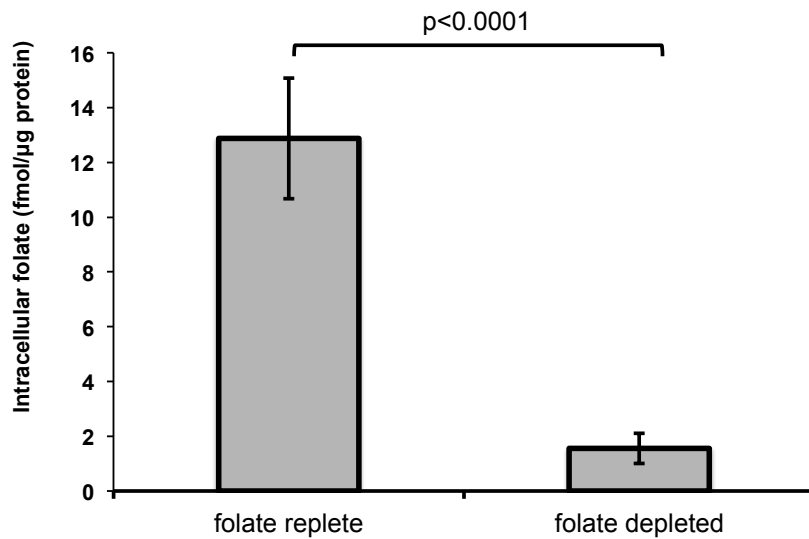


Figure S2. Intracellular folate concentrations in HeLa cells assayed for γ H2AX at the time of staining and quantification (refer to Fig. 3). The intracellular folate concentrations were 8-fold higher in HeLa cells maintained in folate-replete (25 nM) media compared to those cultured in folate-depleted (5 nM) conditions ($p < 0.0001$). Data are shown as the mean \pm S.D. of $n=4$ groups for each folate replete or folate depleted condition. Significance was determined by using a Student's two-tailed t-test.

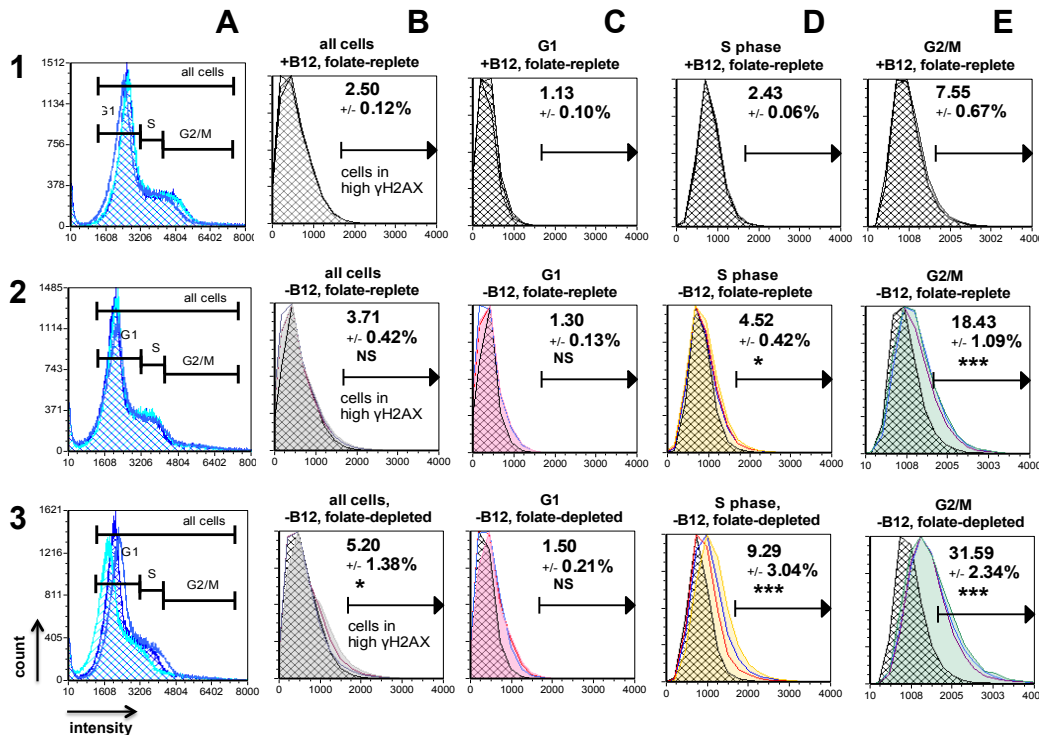


Figure S3. Vitamin B12 depletion induces changes in γ H2AX, a marker of DNA damage, in the S and G2/M phases of the cell cycle in HeLa cells. Cells were stained for DNA content (Vybrant Violet; 1-3A) and γ H2AX (FITC; 1-3 B-E); individual plots depict the cell count (Y-axis) versus fluorescence intensity (X-axis). The high γ H2AX parameter is a threshold defined by the mean top 2.5% of cells in the G1, S, and G2/M ('all cells') stained for γ H2AX in the vitamin B12- and folate-replete condition (1B), and this gate was uniformly applied to all conditions and cell cycle phases. Each plot shows the mean percent high γ H2AX \pm S.D. for triplicates for each experimental condition and cell cycle phase. Individual triplicates stained for γ H2AX in each condition are plotted relative to the corresponding mean γ H2AX values in the vitamin B12- and folate-replete condition (hatched histograms, 1B-E). Asterisks designate statistical significance in percent high γ H2AX values between treatment conditions and cell cycle phase compared to the corresponding phases in the vitamin B12- and folate-replete condition (1B-E). The greatest percentage of cells in high γ H2AX within conditions was observed in G2/M under vitamin B12- and folate-depleted conditions ($p < 0.001$; 3E). A combined vitamin B12 and folate depletion exacerbated the percent high γ H2AX observed in HeLa cells compared to cells maintained in vitamin B12-depleted and folate-replete culture conditions in all cells, S, and G2/M (compare 2A,D-E to 3A,D-E), and this difference in high γ H2AX between conditions was significant for S phase ($p = 0.01$) and G2/M ($p = 0.0003$). Statistical significance was determined using a one-way ANOVA. The dependent variable was log-transformed percent high γ H2AX, and the

independent variable was vitamin B12 and folate level. Folate-replete, 25 nM (6S) 5-formylTHF in culture media; Folate-depleted, 5 nM (6S) 5-formylTHF in culture media. The statistical significance is represented as follows: NS = Not significant ($p > 0.05$); * = $0.01 < p < 0.05$; ** = $0.01 < p < 0.001$; *** = $p < 0.001$

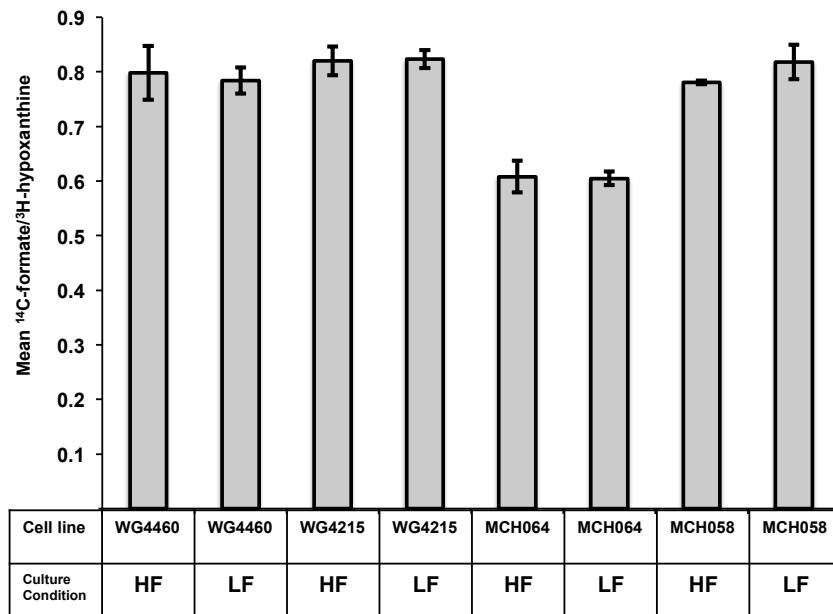


Figure S4. Mean ratio of ^{14}C -formate/ ^3H -hypoxanthine incorporation into nuclear DNA in cblG (WG4215 and WG4460) and control (MCH064 and MCH058) fibroblasts. Data are shown as mean \pm S.D. for fibroblast line and treatment. A two-way ANOVA revealed a non-significant effect of folate exposure ($p > 0.05$) and a significant effect of fibroblast genotype ($p = 0.02$) on the rank-transformed mean ratios of $^{14}\text{C}/^3\text{H}$ incorporated into nuclear DNA. HF, 25 nM (6S) 5-formylTHF; LF, 5 nM (6S) 5-formylTHF.

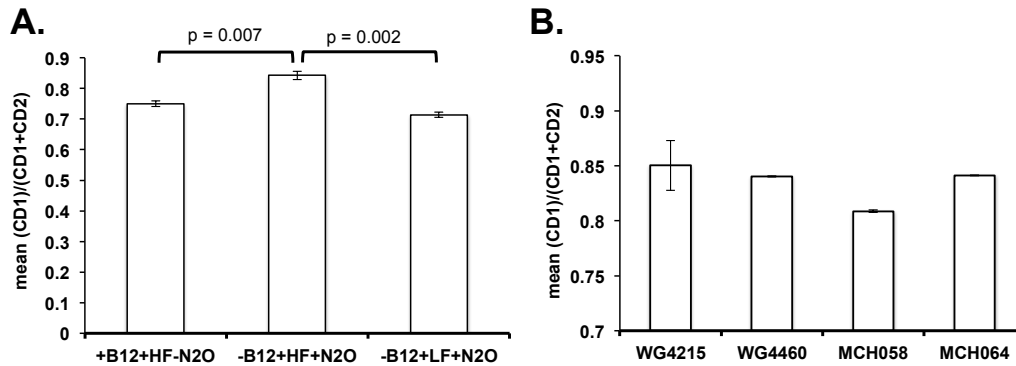


Figure S5. Mean ratio of isotopically labeled one-carbon units from MTHFD1 (CD1) to the total number of labeled one-carbons containing 1 or 2 deuterium atoms generated from MTHFD1 (CD1) or SHMT (CD2), respectively, into thymidine in nuclear DNA in (A) HeLa cells and (B) human fibroblasts. (A) Vitamin B12-depleted and folate-replete conditions increased the contribution of labeled one-carbon units from MTHFD1 relative to SHMT by 12% compared to vitamin B12 and folate-replete conditions in HeLa cells ($p=0.007$). The ratios between vitamin B12-depleted conditions were significant ($p=0.002$). (B) In human cblG (WG4215 and WG4460) and control fibroblasts (MCH058 and MCH064) grown under high folate conditions, there was no difference in the contributions of one-carbon units from MTHFD1 ($p>0.05$). For (A) and (B), cells were plated in duplicate, and the data are shown as mean \pm S.D. Statistical significance between conditions was assessed using a one-way ANOVA. The dependent variable was the log-transformed ratio of mean CD1/(CD1+CD2) and the independent variable was folate and vitamin B12 exposure. HF, 25 nM (6S) 5-formylTHF; LF, 5 nM (6S) 5-formylTHF.

TABLE S1. Fold differences in percent high γ H2AX¹ between control and experimental conditions² within cell cycle phase (to accompany Fig. 3).

| | All cells | G1 | S | G2/M |
|--|--|------------------------------------|------------------------------------|-------------------------|
| | Least Square (LS) Mean 95% CI (Lower, Upper) | LS Mean 95% CI (L,U) | LS Mean 95% CI (L,U) | LS Mean 95% CI (L,U) |
| Treatment | B | C | D | E |
| (1) B12- and folate-replete (control) | 2.50 | 0.76 | 2.46 | 8.99 |
| (2) B12-replete, folate-depleted | 2.10*** (1.65, 2.66) | 1.12 ^{NS} (0.89, 1.40) | 1.55 ^{NS} (1.11, 2.16) | 1.84** (1.39, 2.43) |
| (3) B12-depleted, folate-replete | 3.16*** (2.42, 4.13) | 2.25*** (1.74, 2.90) | 4.50*** (3.10, 6.52) | 2.80*** (2.05, 3.84) |
| (4) B12- and folate-depleted | 5.38*** (4.23, 6.83) | 4.83*** (3.85, 6.06) | 9.02*** (6.47, 12.59) | 4.85*** (3.66, 6.42) |
| (5) B12-depleted: folate-replete vs. depleted (row 3 vs. 4) | 1.70** (1.30, 2.23) | 2.15*** (1.67, 2.77) | 2.01* (1.38, 2.91) | 1.73* (1.26, 2.37) |

¹Percent (%) high γ H2AX refers to the percentage of HeLa cells stained for γ H2AX above the mean top 2.5% total γ H2AX intensity in 'all cells' in the vitamin B12- and folate-replete (control) condition (Figure 3, 1B). Columns show the back-transformed least-squares (LS) means and 95% confidence intervals (CI) for Log_e transformed mean % high γ H2AX for specific contrasts, indicating fold differences in % high γ H2AX. Values for LS means in row 1B-E for control conditions are those mean % high γ H2AX predicted by the model. LS means and 95% CI were back-transformed = exp¹(LS Mean) = geometric mean % high γ H2AX.

²Post-hoc comparisons (n=3) were made within cell cycle phase ('all cells', G1, S, G2/M) and compared the % high γ H2AX in the control to that in row: 2) B12-replete, folate-depleted, 3) B12-depleted, folate-replete, and 4) B12- and folate-depleted conditions. A final comparison considered the difference in % high γ H2AX between B12 depleted conditions (row 3 vs. 4). A Bonferroni correction was applied to all p-values to account for multiple comparisons (n=4).

Legend: NS = Not significant (p > 0.05); * = 0.01 < p < 0.05; ** = 0.01 < p < 0.001; *** = p < 0.001