

Table S1 Gene accession numbers for phylogenetic analysis

Organism	Accession number	Reference
<i>Streptococcus pyogenes</i> PmtA	AAZ51785.1	(1)
<i>Streptococcus agalactiae</i>	KLL21255.1	
<i>Streptococcus canis</i>	WP_003044256.1	
<i>Streptococcus dysgalactiae</i>	WP_046159323.1	
<i>Streptococcus iniae</i>	WP_003101088.1	
<i>Streptococcus mutans</i>	WP_002304112.1	
<i>Streptococcus pneumoniae</i>	CVM74562.1	
<i>Streptococcus porcinus</i>	WP_003083554.1	
<i>Streptococcus suis</i>	WP_029187041.1	
<i>Streptococcus thermophilus</i>	WP_049555285.1	
<i>Enterococcus faecalis</i>	WP_034863010.1	
<i>Enterococcus faecium</i>	WP_058138288.1	
<i>Listeria monocytogenes</i> FrvA	WP_014929421.1	(2-4)
<i>Bacillus subtilis</i> PfeT	CUB56232.1	(4, 5)
<i>Mycobacterium smegmatis</i> CtpD		(6)
<i>Sulfitobacter</i> sp. <i>NAS 14.1</i> sCoaT	ZP_00964573.1	(7)
<i>Cupriavidus metallidurans</i> CzcP	WP_011514822.1	(8)
<i>Mycobacterium tuberculosis</i> CtpD	WP_057125145.1	(9)
<i>Mycobacterium tuberculosis</i> CtpJ	WP_031727628.1	(9)
<i>Chlamydia trachomatis</i>	CRH88065.1	

Table S2 List of strains and plasmids used in this study

Bacterial strains	Description	Reference
<i>Escherichia coli</i>		
MC1061	<i>E. coli</i> laboratory cloning strain	(10)
Top10	<i>E. coli</i> laboratory cloning strain	
<i>Streptococcus pyogenes</i>		
5448	<i>S. pyogenes</i> invasive MIT1 strain	(11)
5448 Δ <i>pmtA</i>	<i>S. pyogenes</i> 5448 Δ <i>pmtA</i> :: <i>km</i> ^R deletion mutant	This study
5448 Δ <i>pmtA</i> :: <i>pmtA</i>	<i>S. pyogenes</i> 5448 <i>pmtA</i> complemented strain	This study
5448 Δ <i>perR</i>	<i>S. pyogenes</i> 5448 Δ <i>perR</i> :: <i>spec</i> ^R deletion mutant	(12)
5448 Δ <i>perR</i> :: <i>perR</i>	<i>S. pyogenes</i> 5448 Δ <i>perR</i> complemented strain	(12)
Plasmids	Description	Reference
pHY304	Temperature sensitive shuttle plasmid:: <i>Em</i> ^R	(13)
pJRS233	Temperature sensitive shuttle plasmid:: <i>Em</i> ^R	(14)
pUC4 Ω Km2	Template for <i>Km</i> ^R	(15)
pHY304- <i>pmtA</i> -KO	pHY304 + <i>pmtA</i> knockout construct inserted at the <i>Xho</i> I and <i>Bam</i> HI site	This study
pJRS233- <i>pmtA</i>	pJRS233 + <i>pmtA</i> complement construct inserted at the <i>Xho</i> I and <i>Bam</i> HI site	This study

Table S3 List of primers used in this study

Primer	Sequence (5'-3')	
<i>Deletion mutation and reverse complementation constructs</i>		
pmtA-KO-1	CCCGGCTCGAGTTGATAGCGACAAGGTTCTTTTT	XhoI
pmtA-KO-2	CAAAGTTGGCGTATAACATAACCGATAGCTGCCAAAATCAT	<i>Km^R</i> 5'
pmtA-KO-3	TTACTGGATGAATTGTTTTAGAGAACGAACCGCCAATTATG	<i>Km^R</i> 3'
pmtA-KO-4	GGGCCC GATCC CTTTCATAAGTCTTACTATTATTATAAACAC	BamHI
km-F	TATGTTATACGCCAACTTTG	
km-R	CTAAAACAATTCATCCAGTAA	
<i>Gene expression studies</i>		
pmtA-RT-F	GAAAAGCAAAACCGCCACCT	
pmtA-RT-R	GGGCACATGGTGAAGCTACT	
gyrA-RT-F	GAAGTGATCCCTGGACCTGA	
gyrA-RT-R	CCCGACCTGTTTGAGTTGTT	

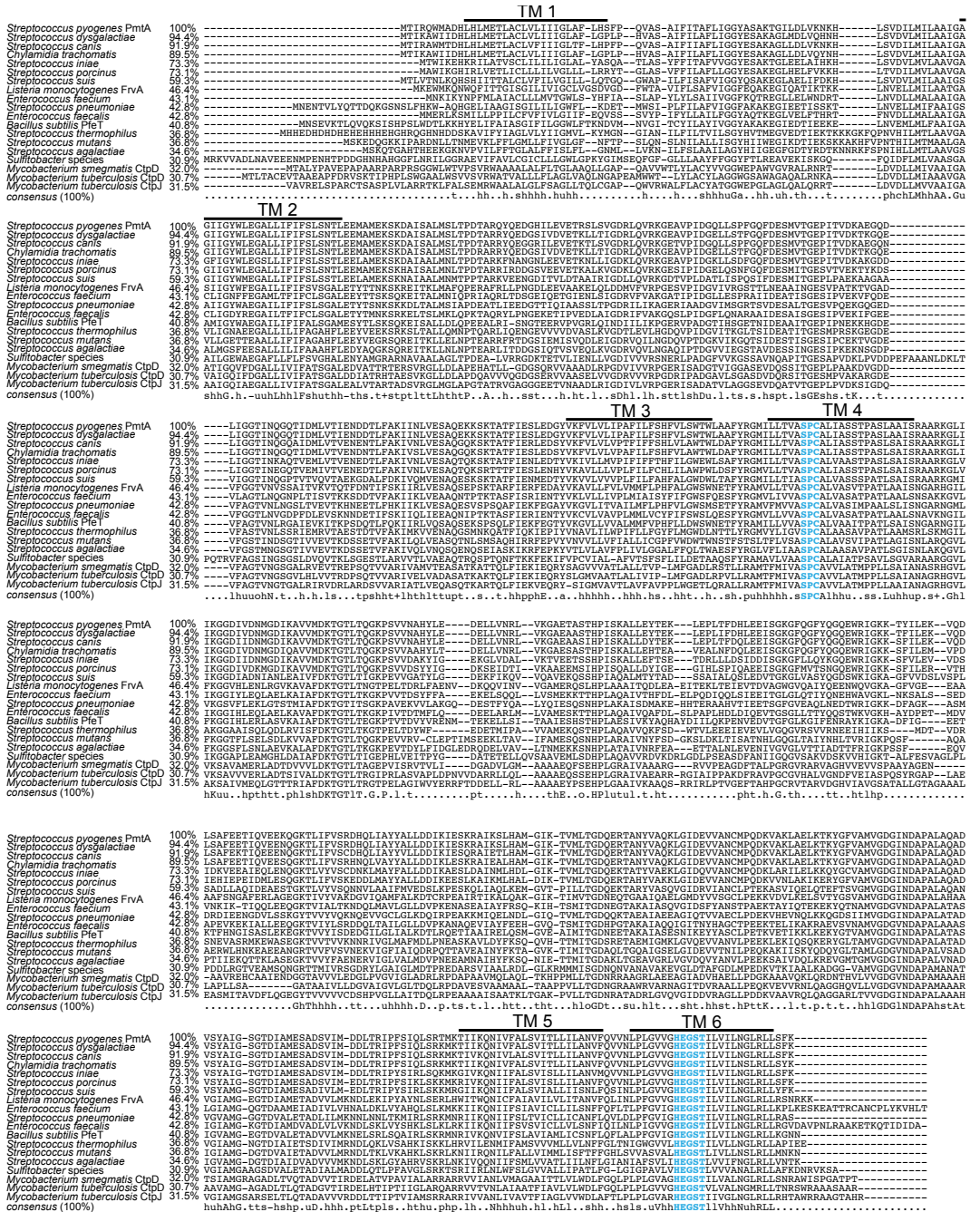


Figure S1: Multiple sequence alignment of *S. pyogenes* PmtA and homologous P_{1-B4} type ATPase family sequences. Relative location of six core purported transmembrane helices based on the coordinates from *Bacillus subtilis* PfeT transmembrane profiles (reference). Key conserved motifs within TM helices 4 and 6 are highlighted in blue and the essential serine within TM 6 is highlighted in red. Consensus patterns based on discriminating equivalence class at a 100% threshold are

indicated under the alignment. Percent amino acid identity relative to *S. pyogenes*
PmtA (TM 1-6) are indicated next to the species name.

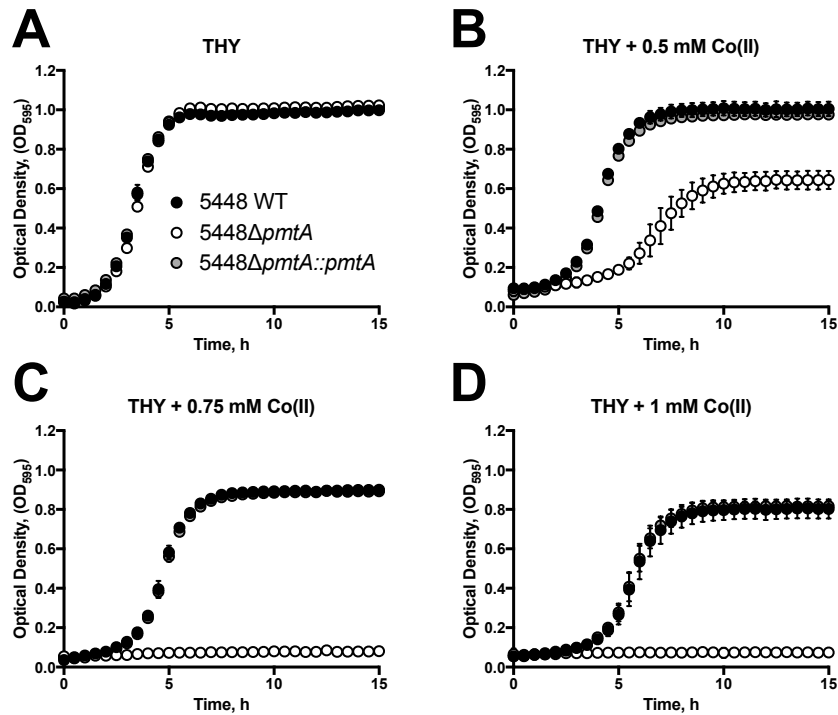


Figure S2: Growth analysis in the presence of Co(II). Overnight cultures of 5448 WT (black circles), 5448 Δ pmtA (open circles) and 5448 Δ pmtA::pmtA (grey circles) were diluted to OD₆₀₀ = 0.05 into THY broth alone (A) or THY broth with 0.5 mM (B), 0.75 mM (C) or 1 mM (D) Co(II). Growth was monitored at 37°C by optical density recording at 595 nm (OD₅₉₅). Graphs represent mean \pm standard deviation of 3 independent biological replicates.

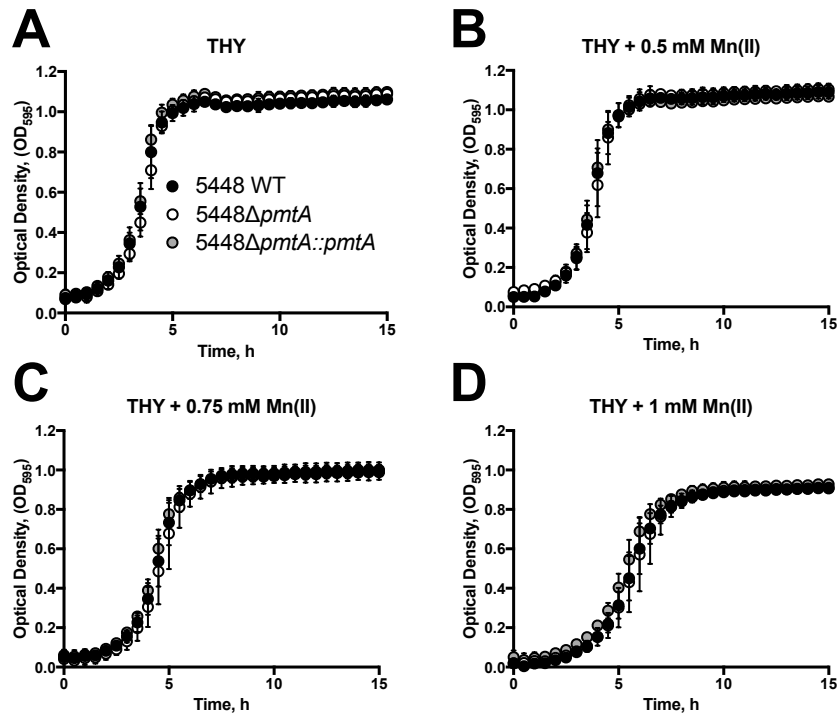


Figure S3: Growth analysis in the presence of Mn(II). Overnight cultures of 5448 WT (black circles), 5448 Δ pmtA (open circles) and 5448 Δ pmtA::pmtA (grey circles) were diluted to OD₆₀₀ = 0.05 into THY broth alone (A) or THY broth with 0.5 mM (B), 0.75 mM (C) or 1 mM (D) Mn(II). Growth was monitored at 37°C by optical density recording at 595 nm (OD₅₉₅). Graphs represent mean \pm standard deviation of 3 independent biological replicates.

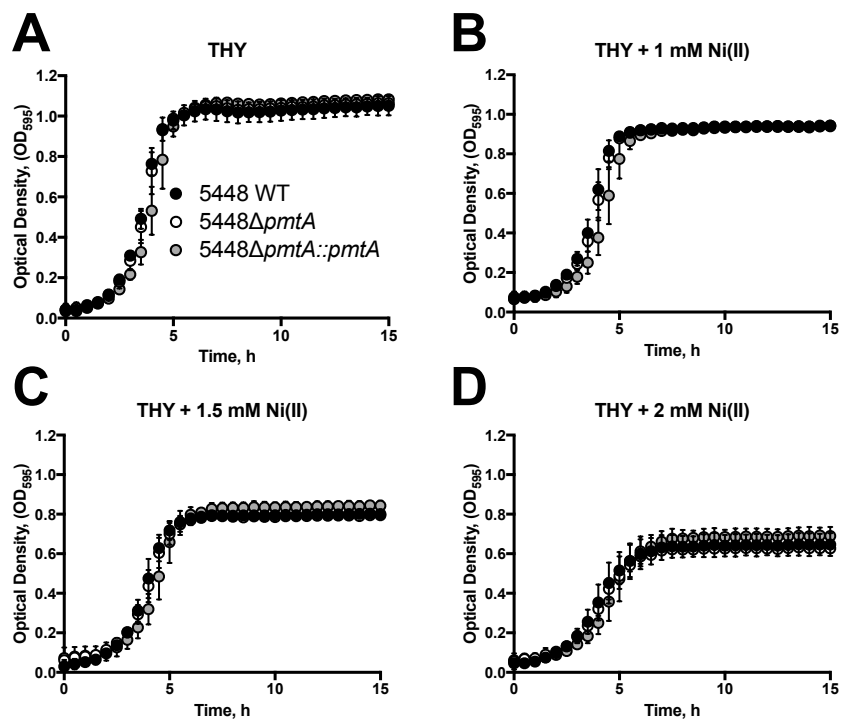


Figure S4: Growth analysis in the presence of Ni(II). Overnight cultures of 5448 WT (black circles), 5448Δ*pmtA* (open circles) and 5448Δ*pmtA*::*pmtA* (grey circles) were diluted to OD₆₀₀ = 0.05 into THY broth alone (A) or THY broth with 1 mM (B), 1.5 mM (C) or 2 mM (D) Ni(II). Growth was monitored at 37°C by optical density recording at 595 nm (OD₅₉₅). Graphs represent mean ± standard deviation of 3 independent biological replicates.

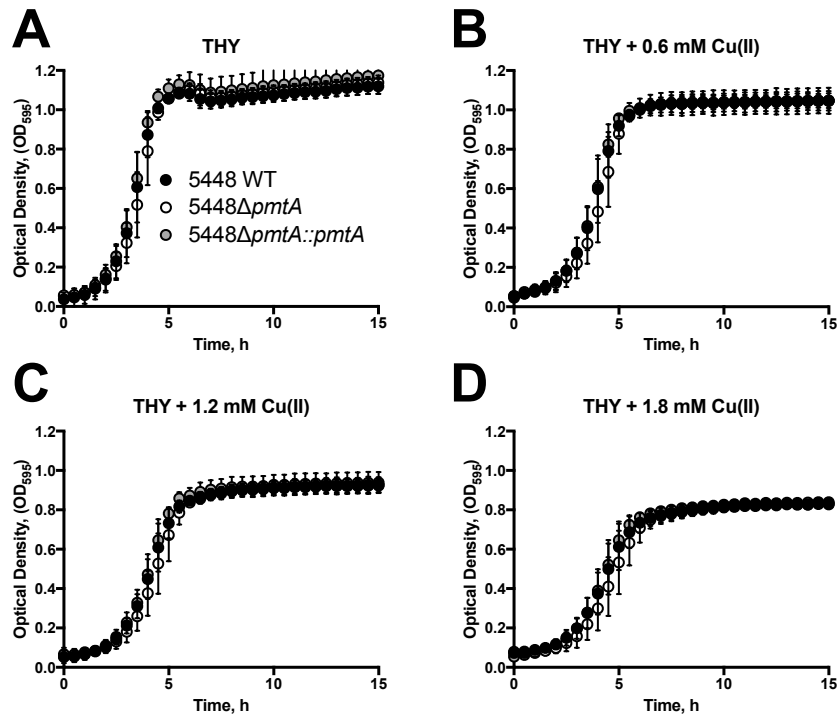


Figure S5: Growth analysis in the presence of Cu(II). Overnight cultures of 5448 WT (black circles), 5448Δ*pmtA* (open circles) and 5448Δ*pmtA*::*pmtA* (grey circles) were diluted to OD₆₀₀ = 0.05 into THY broth alone (A) or THY broth with 0.6 mM (B), 1.2 mM (C) or 1.8 mM (D) Cu(II). Growth was monitored at 37°C by optical density recording at 595 nm (OD₅₉₅). Graphs represent mean ± standard deviation of 3 independent biological replicates.

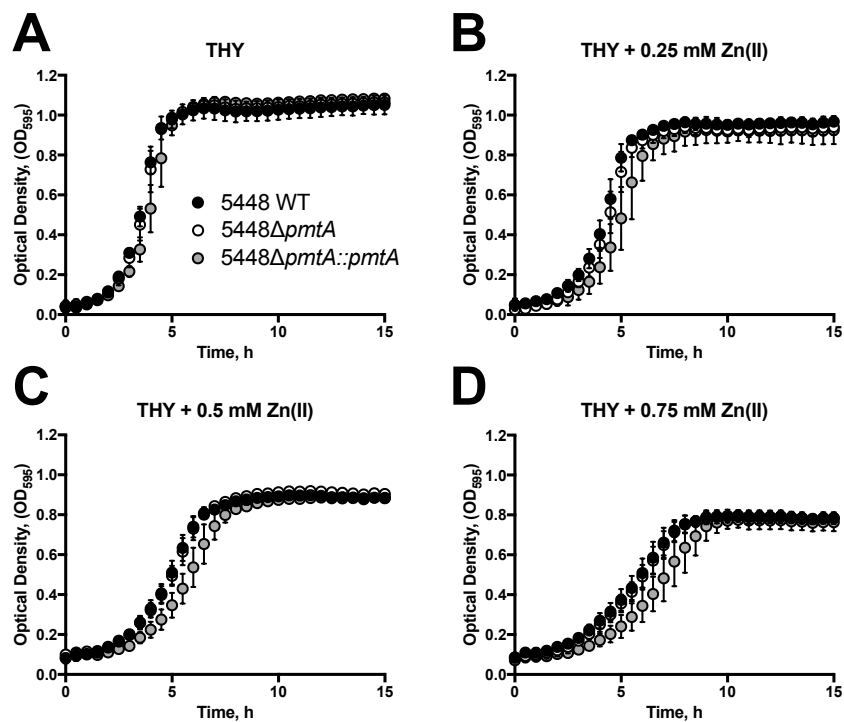


Figure S6: Growth analysis in the presence of Zn(II). Overnight cultures of 5448 WT (black circles), 5448Δ*pmtA* (open circles) and 5448Δ*pmtA*::*pmtA* (grey circles) were diluted to OD₆₀₀ = 0.05 into THY broth alone (A) or THY broth with 0.25 mM (B), 0.5 mM (C) or 0.75 mM (D) Zn(II). Growth was monitored at 37°C by optical density recording at 595 nm (OD₅₉₅). Graphs represent mean ± standard deviation of 3 independent biological replicates.

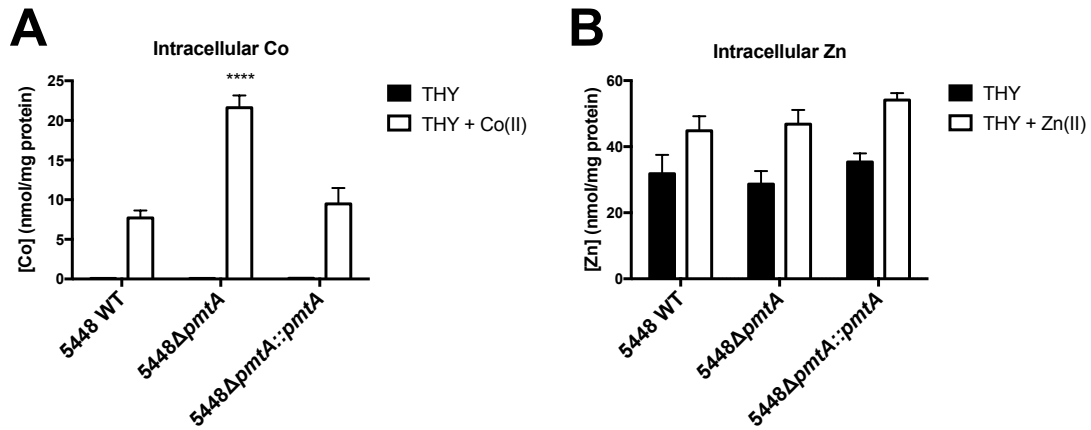


Figure S7: Intracellular metal accumulation of Co and Zn. Mid-exponential cultures (OD_{600} 0.6-0.8) of 5448 WT, 5448Δ*pmtA* and 5448Δ*pmtA*::*pmtA* were challenged with either sterile water (black bars) or 0.50 mM Co(II) (A) or 1 mM Zn(II) (B) (white bars) for 1 h at 37°C. Cells were analyzed by inductively-coupled plasma mass spectrometry (ICP-MS). Total metal content was normalized to protein content of the sample. Graph represents mean + standard deviation of 3 independent biological experiments (2-way ANOVA used comparing all to 5448 WT of that condition, **** $P < 0.0001$)

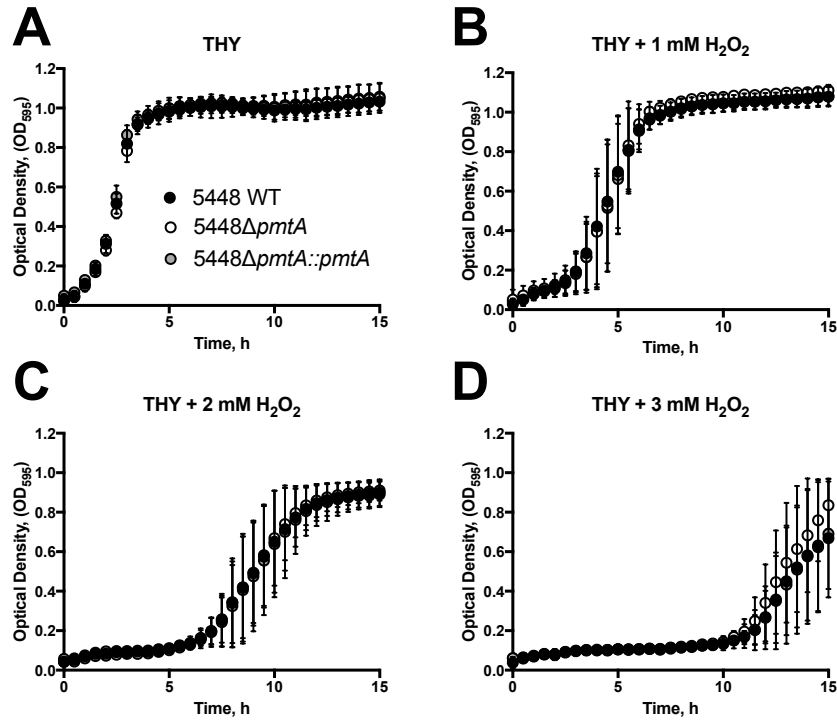


Figure S8: Growth curve analysis of strains 5448 WT (black circles), 5448Δ*pmtA* (open circles) and 5448Δ*pmtA*::*pmtA* (grey circles) in hydrogen peroxide. Strains were grown in THY to mid-exponential phase (OD 0.6-0.8) and diluted to OD₆₀₀ = 0.05 in THY broth (A) or THY broth containing 1 mM (B), 2 mM (C) or 3 mM (D) H₂O₂. Growth was monitored at 37°C by optical density recording at 595 nm (OD₅₉₅). Graphs represent mean ± standard deviation of 3 independent biological replicates.

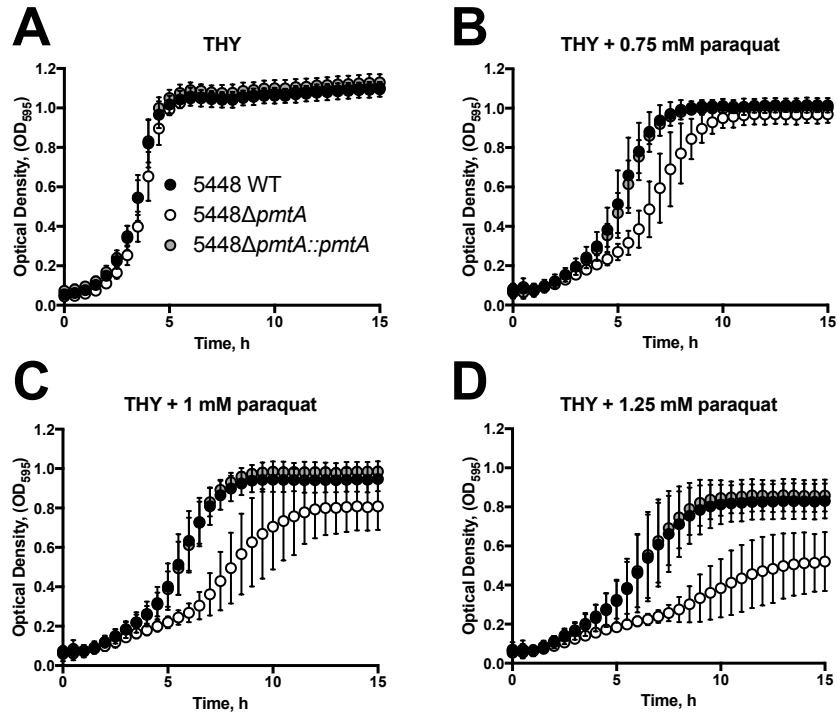


Figure S9: Growth analysis in the presence of the superoxide generator, paraquat.

Overnight cultures of 5448 WT (black circles), 5448 Δ *pmtA* (open circles) and 5448 Δ *pmtA::pmtA* (grey circles) were diluted to OD₆₀₀ = 0.05 THY broth alone (A) or THY broth with 0.75 mM (B), 1 mM (C) or 1.25 mM (D) paraquat. Growth was monitored at 37°C by optical density recording at 595 nm (OD₅₉₅). Graphs represent mean \pm standard deviation of 3 independent biological replicates.

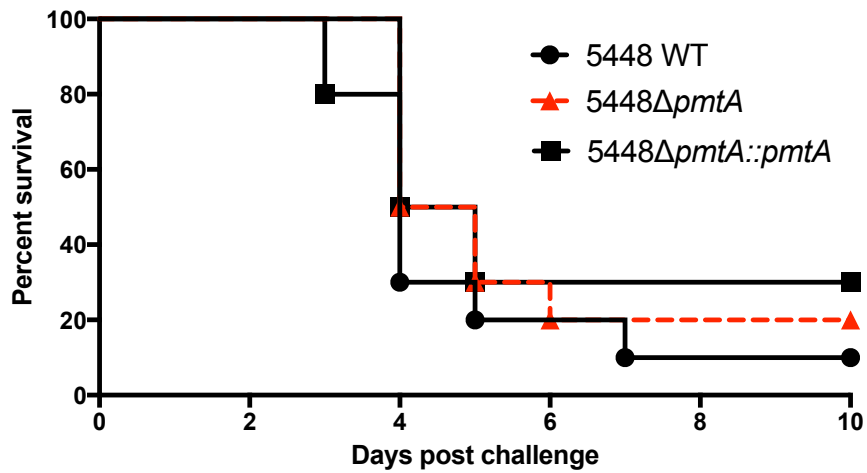


Figure S10: Virulence of 5448ΔpmtA in murine model of infection. Survival of mice after subcutaneous challenge of transgenic humanized plasminogen *AlbPLG1* C57BL/J6 mice with 5448 WT (black circles), 5448ΔpmtA (red triangles) and 5448ΔpmtA::pmtA (black squares). Infecting dose 2×10^8 colony-forming units (CFU). Mantel-cox log rank test was performed comparing 5448ΔpmtA to 5448 WT ($P = 0.5162$) and 5448ΔpmtA::pmtA ($P = 0.9723$) (Graphpad Prism 7)

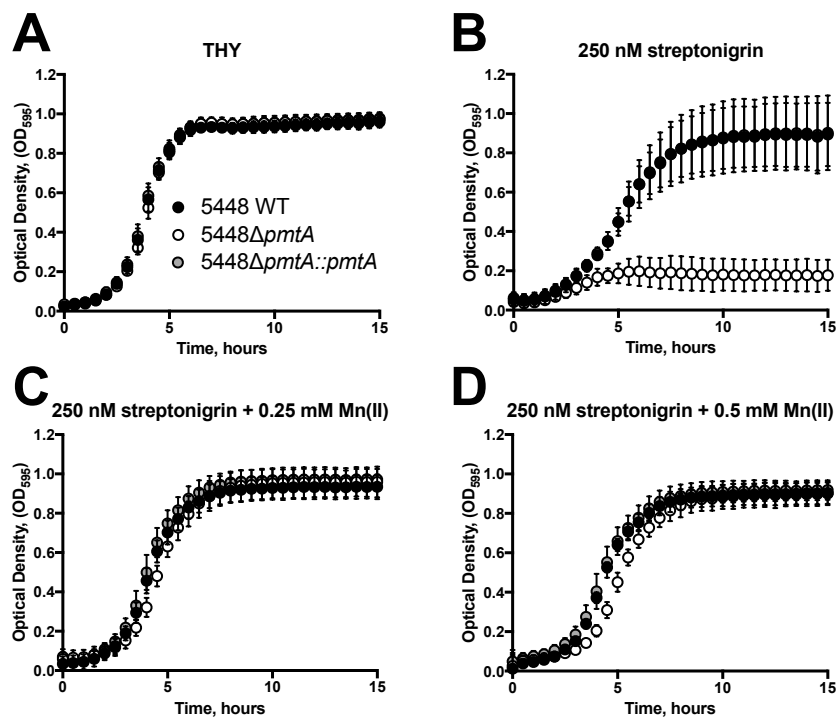


Figure S11: Growth curve analysis of streptonigrin rescue by Mn(II). Overnight cultures of strains 5448 WT (black circles), 5448 Δ *pmtA* (open circles) and 5448 Δ *pmtA::pmtA* (grey circles) were diluted to OD₆₀₀ = 0.05 in THY broth (A) or THY broth with 250 nM streptonigrin (B), 250 nM streptonigrin + 0.25 mM Mn(II) (C) or 250 nM streptonigrin + 0.5 mM Mn(II) (D). Growth was monitored at 37°C by optical density recording at 595 nm (OD₅₉₅). Graphs represent mean \pm standard deviation of 3 independent biological replicates.

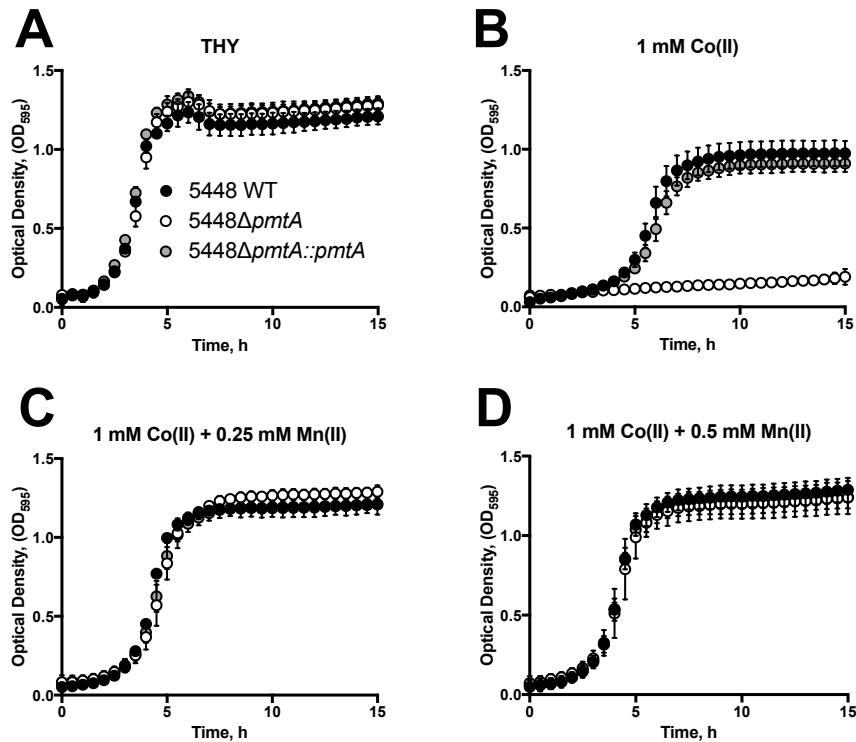


Figure S12: Growth curve analysis of Co(II) rescue by Mn(II). Overnight cultures of strains 5448 WT (black circles), 5448Δ*pmtA* (open circles) and 5448Δ*pmtA*::*pmtA* (grey circles) were diluted to OD₆₀₀ = 0.05 in THY broth (A) or THY broth with 1 mM Co(II) (B), 1 mM Co(II) + 0.25 mM Mn(II) (C) or 1 mM Co(II) + 0.5 mM Mn(II) (D). Growth was monitored at 37°C by optical density recording at 595 nm (OD₅₉₅). Graphs represent mean ± standard deviation of 3 independent biological replicates.

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