

Fig. S2. In vitro growth profiles of the $\Delta ptsP$, $\Delta ptsO$ and $\Delta ptsP-ptsO$ mutants. Growth rates of wild-type, $\Delta ptsP$, $\Delta ptsO$ and $\Delta ptsP-ptsO$ mutant strains of *L. pneumophila* in AYE broth cultures. The optical density (OD) at 600 nm was measured every hour from triplicate samples.

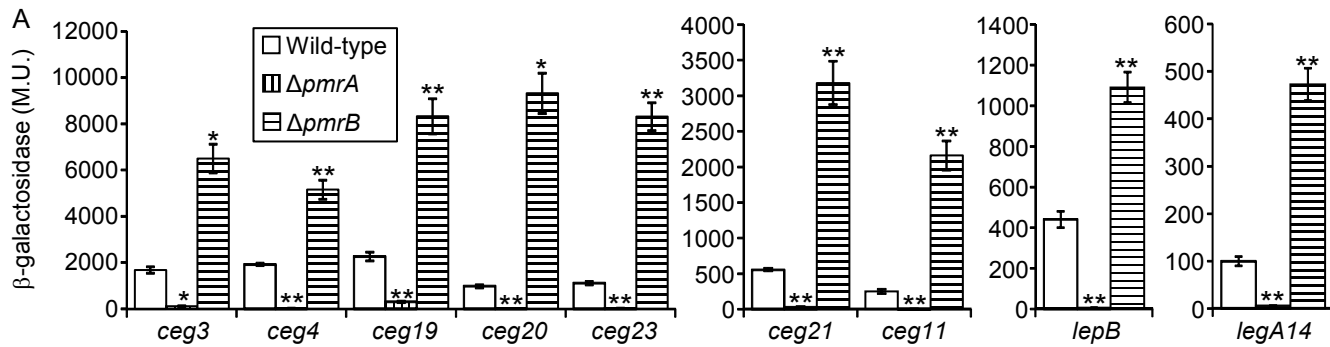


Fig. S3. The effect of the *L. pneumophila* PmrAB TCS on its target effector-encoding genes. The expression of translational *lacZ* fusions of effector-encoding genes activated by PmrA was examined in the wild-type strain (JR32) (white bars), in the *pmrA* deletion mutant (HK-PQ1) (vertical line bars), and in the *pmrB* deletion mutant (EA-*pmrB*) (horizontal line bars) at the stationary phase. The levels of expression of the *lacZ* fusions were found to be significantly different (*, $P < 10^{-4}$; **, $P < 10^{-5}$, Student's *t* test), when comparing the expression in the wild-type strain and the *pmrA* or *pmrB* deletion mutants. The genes were grouped according to their levels of expression.

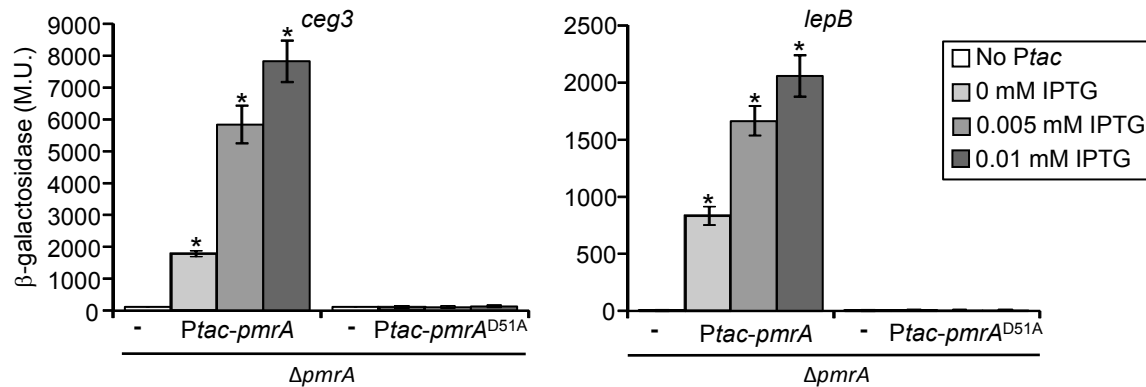


Fig. S4. PmrA must be phosphorylated in order to activate gene expression.

The levels of expression of two PmrA regulated effector-encoding genes (*ceg3* and *lepB*) were examined in the *L. pneumophila pmrA* deletion mutant (HK-PQ1). The bacteria contained a plasmid with the *pmrA* gene or the mutated *pmrA*^{D51A} gene cloned under the control of the IPTG inducible *Ptac* promoter. The plasmids containing the corresponding *lacZ* fusions of the examined genes without the *pmrA* gene were used as a control (white bars). The levels of expression of the *lacZ* fusions were found to be significantly different (*, P < 10⁻⁵, Student's *t* test), when comparing the expression of the *lacZ* fusions containing the wild-type *pmrA* gene expressed from the *Ptac* promoter and the expression of the fusions without the *pmrA* gene.