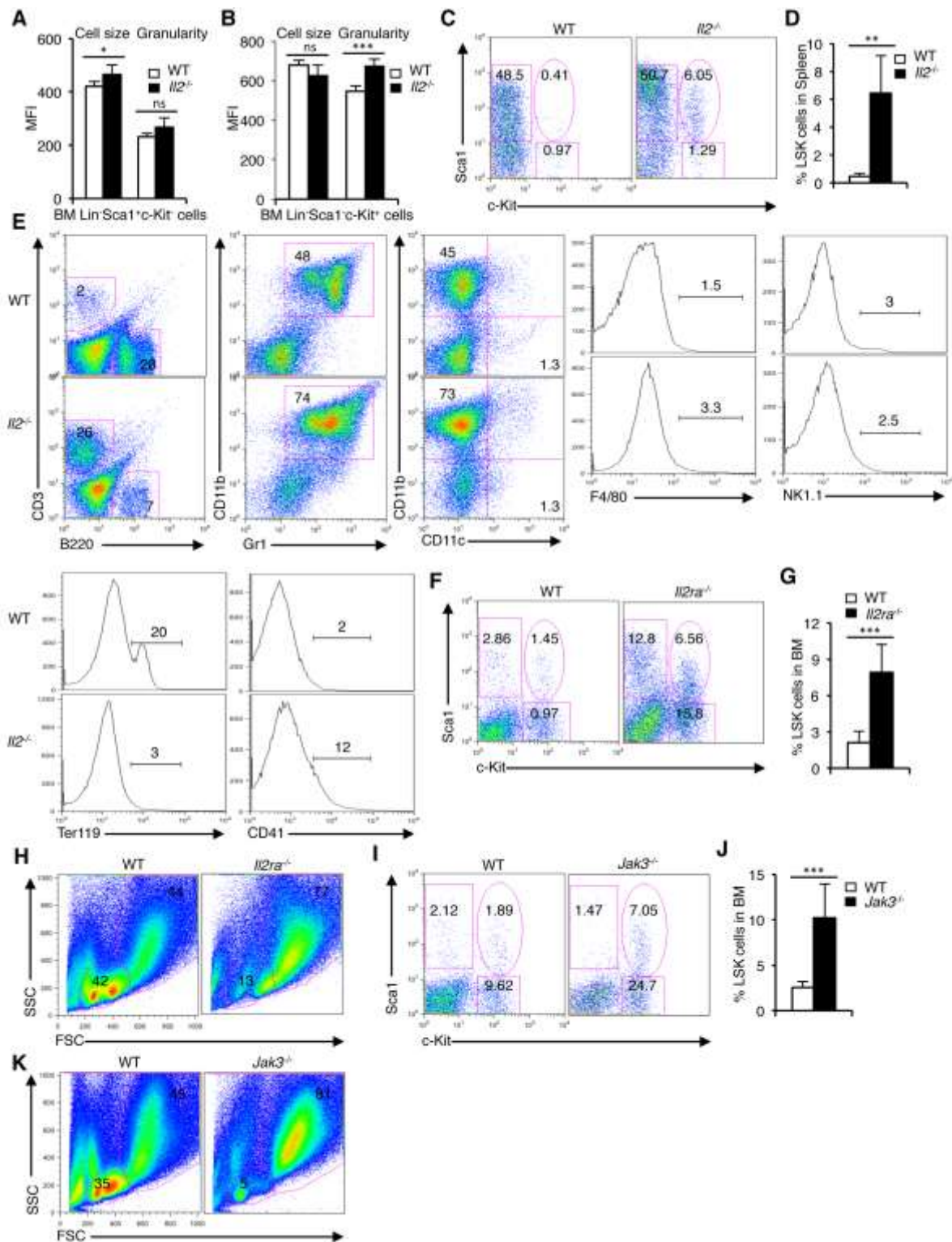


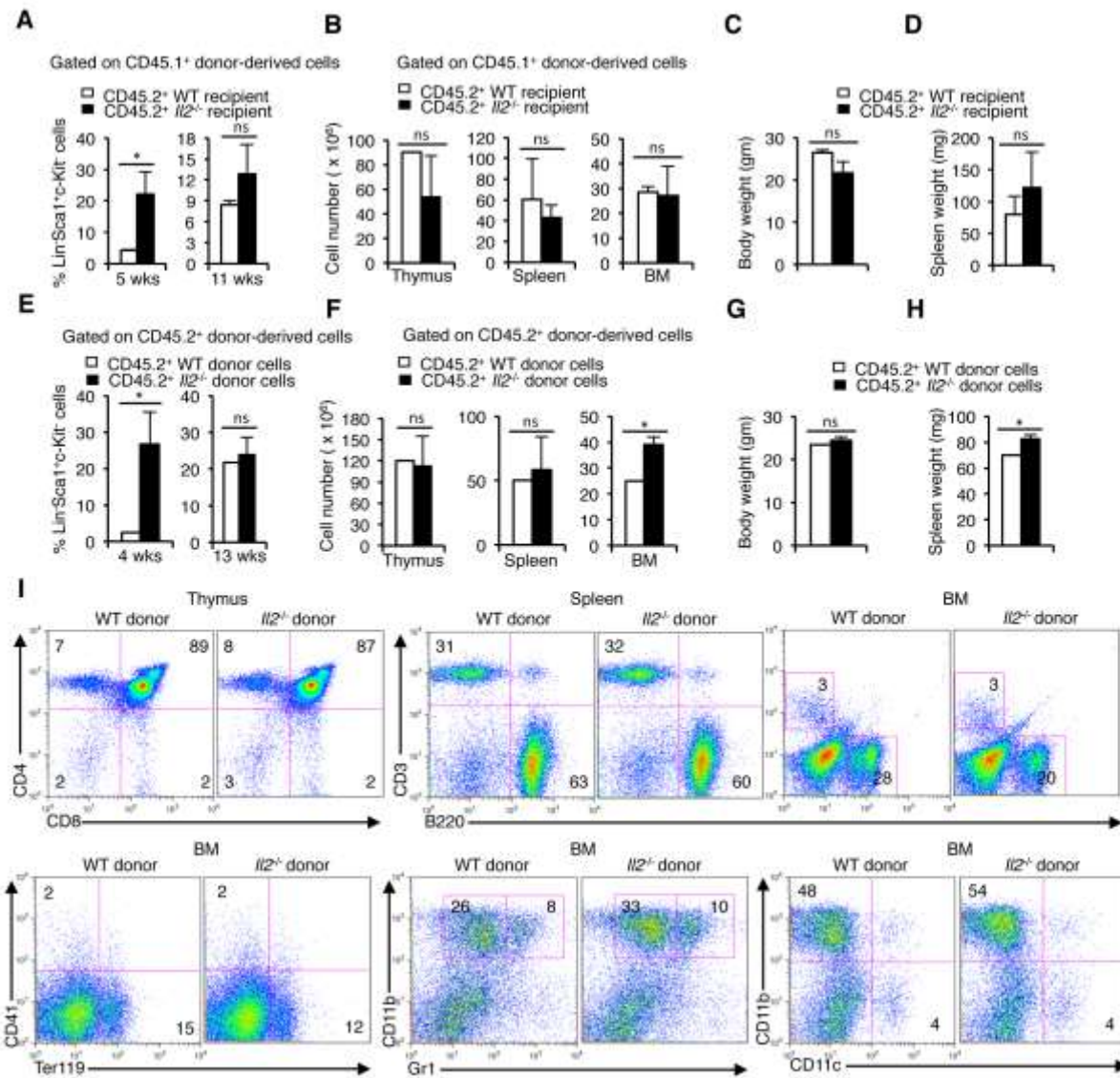
Interleukin-2-regulatory T cell axis critically regulates maintenance of hematopoietic stem cells

Supplementary Material



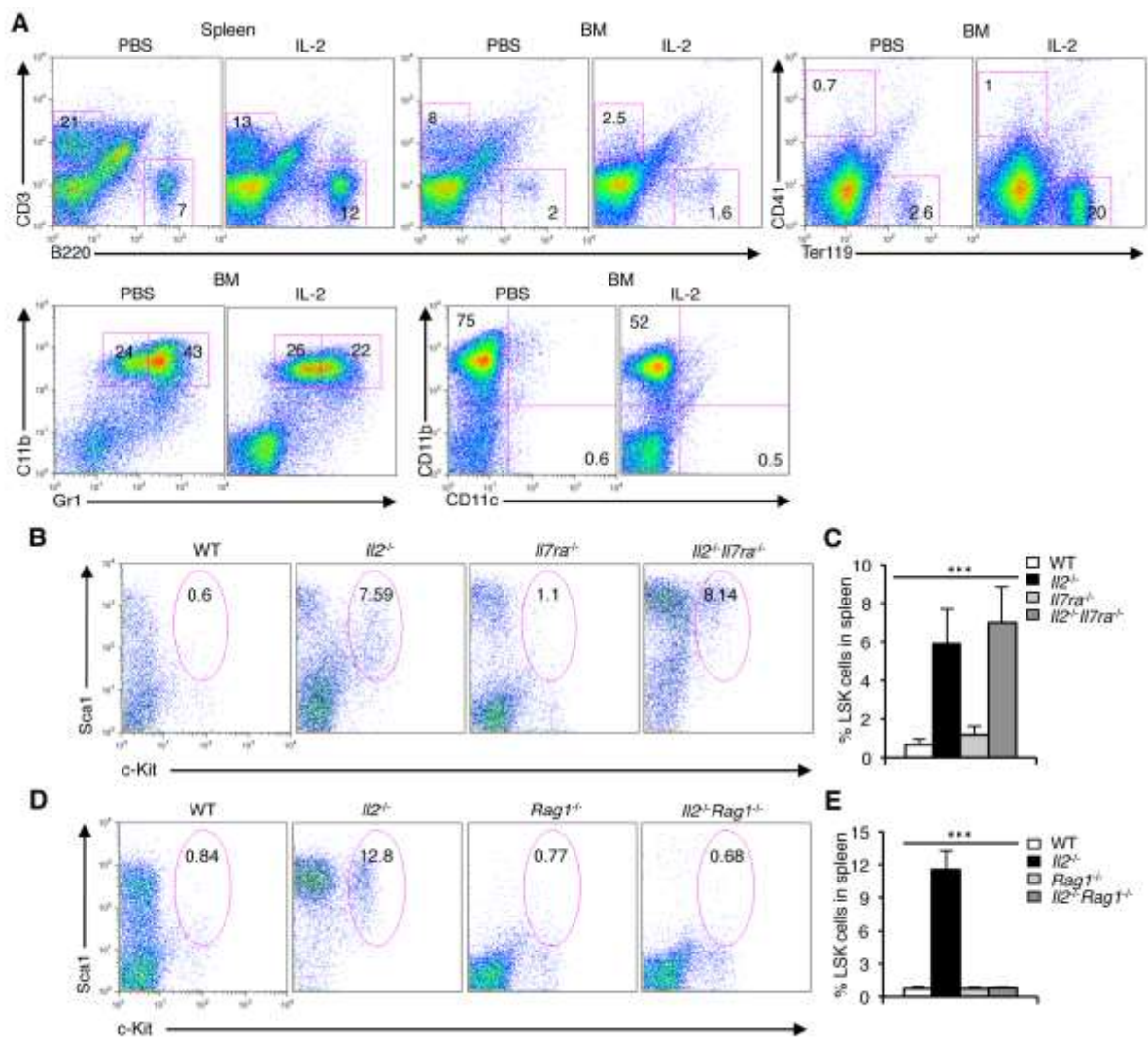
Supplementary Figure 1: Impaired HSC maintenance in IL-2 signaling-deficient mice.

A. Evaluation of cell size and granularity of BM Lin⁻Sca1⁺c-Kit⁻ cells from WT and *Il2*^{-/-} mice. **B.** Cell size and granularity of *Il2*^{-/-} BM Lin⁻Sca1⁻c-Kit⁺ cells compared to WT cells. **C.** Flow cytometry profiles of LSK cells distribution among splenocytes in *Il2*^{-/-} mice compared to WT mice. **D.** Quantification of percent LSK cells in the spleen of WT and *Il2*^{-/-} mice. **E.** Distribution of lymphoid (CD3⁺, B220⁺ and NK1.1⁺), myeloid (Gr1⁺, CD11b⁺, CD11c⁺ and F4/80⁺) and erythroid (Ter119⁺ and CD41⁺) cells in the BM of *Il2*^{-/-} mice compared to WT mice. **F.** Distribution of LSK cells in the BM of *Il2ra*^{-/-} mice compared to littermate WT mice. **G.** Quantification of percent LSK cells in the BM of WT and *Il2ra*^{-/-} mice. **H.** FSC/SSC distribution pattern of BM cells from *Il2ra*^{-/-} mice compared to WT control mice. **I.** LSK cells distribution in the BM of *Jak3*^{-/-} mice compared to littermate WT controls. **J.** Quantification of percent LSK cells in the BM of WT and *Jak3*^{-/-} mice. **K.** FSC/SSC distribution pattern of BM cells in WT and *Jak3*^{-/-} mice. Numbers inside each FACS plot represent percent respective population. Data are shown as mean ± s.d., in **(A)** **P* = 0.0439, **(B)** ****P* = 0.0002, **(D)** ***P* < 0.0012, **(G)** ****P* < 0.0001 and in **(J)** ****P* = 0.0006, ns = not significant, unpaired *t*-test and are representative of 5 independent experiments, (*n* = 3 per group).



Supplementary Figure 2: Impaired reconstitution potential of *Il2*^{-/-} HSCs. **A.** Evaluation of CD45.1⁺ donor-derived BM Lin⁻Sca1⁺c-Kit⁻ cells distribution in CD45.2⁺ WT or *Il2*^{-/-} recipient mice 5 or 11 weeks after cell transfer. **B.** Cellularity in the thymus, spleen and BM of irradiated CD45.2⁺ WT or *Il2*^{-/-} recipient mice 11 weeks after transfer of CD45.1⁺ WT BM cells. **C.** Evaluation of body weight of irradiated CD45.2⁺ WT or *Il2*^{-/-} recipient mice 11 weeks after transfer of CD45.1⁺ WT BM cells. **D.** Spleen weight in CD45.2⁺ WT or *Il2*^{-/-} recipient mice 11 weeks after transfer of CD45.1⁺ WT BM cells. **E.** Evaluation of CD45.2⁺ WT or *Il2*^{-/-} donor-derived BM Lin⁻Sca1⁺c-Kit⁻ cells distribution in CD45.1⁺ WT recipient mice 4 or 13 weeks after cell transfer. **F.** Cellularity in the thymus, spleen and BM of

irradiated CD45.1⁺ WT recipient mice 13 weeks after transfer of CD45.2⁺ WT or *Il2*^{-/-} BM cells. **G.** Evaluation of body weight of irradiated CD45.1⁺ WT recipient mice 13 weeks after transfer of CD45.2⁺ WT or *Il2*^{-/-} BM cells. **H.** Spleen weight in CD45.1⁺ WT recipient mice 13 weeks after transfer of CD45.2⁺ WT or *Il2*^{-/-} BM cells. **I.** Flow cytometry profiles of the distribution of various CD45.2⁺ WT or *Il2*^{-/-} donor-derived mature hematopoietic cell populations in the thymus, spleen and BM of CD45.1⁺ WT recipient mice 13 weeks after cell transfer. Numbers inside each FACS plot represent percent respective population. Data are representative of two independent experiments, (*n* = 4 per group). Data are shown as mean ± s.d., in (A) **P* = 0.0410, (E) **P* = 0.0345, (f) **P* = 0.0103, (H) **P* = 0.0319, ns = not significant, unpaired *t*-test.



Supplementary Figure 3: T cell activity in the BM influences LSK cell maintenance. A.

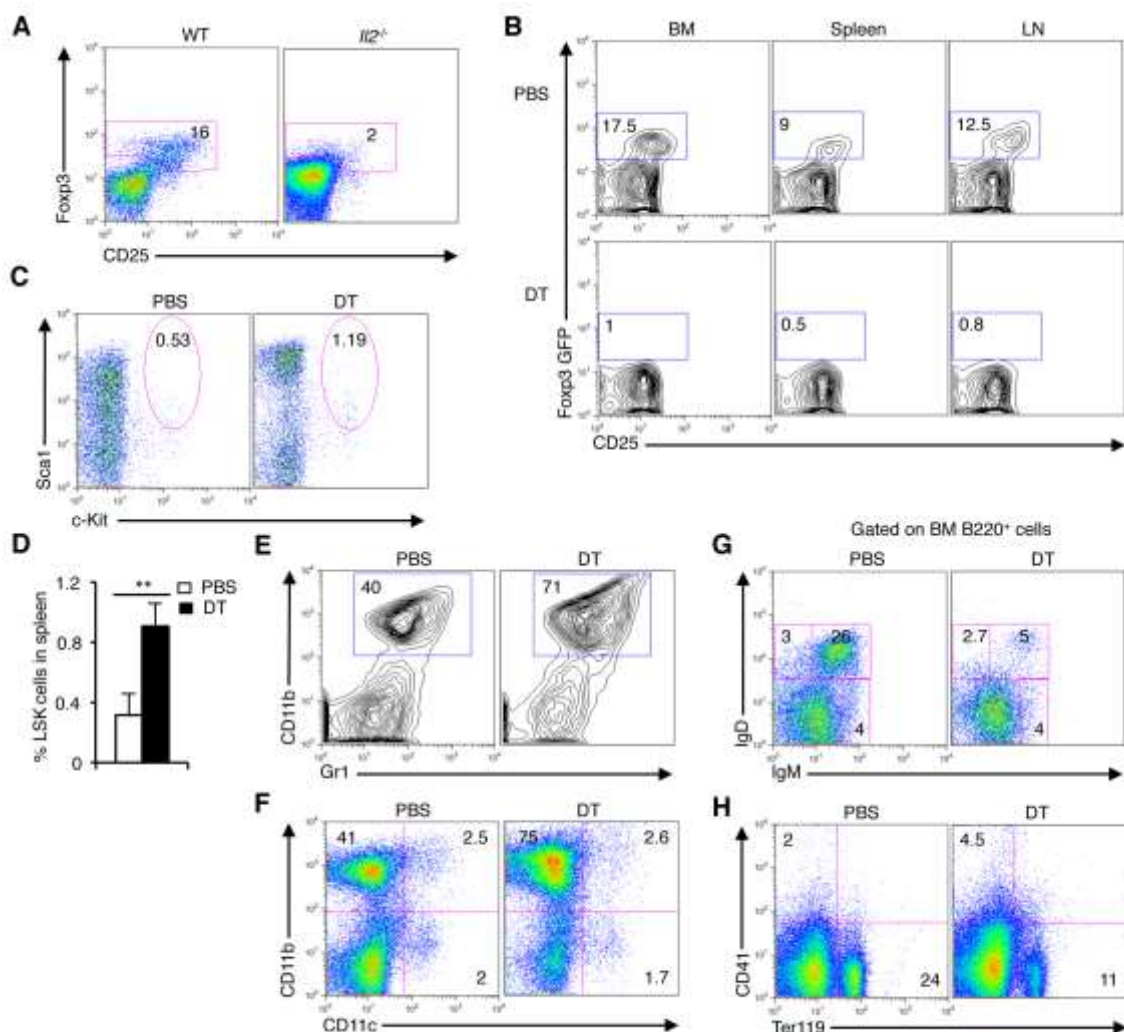
Distribution of lymphoid ($CD3^+$ and $B220^+$), erythroid ($Ter119^+$ and $CD41^+$) and myeloid ($Gr1^+$, $CD11b^+$ and $CD11c^+$) cells in the spleen and BM of PBS or IL-2 treated $Il2^{-/-}$ mice. **B.**

Flow cytometry profiles showing the distribution of LSK cells in the spleen of $Il2^{-/-}Il7ra^{-/-}$ mice compared to WT, $Il2^{-/-}$ and $Il7ra^{-/-}$ mice. **C.** Quantification of percent LSK population in the spleen of indicated mice. **D.** Flow cytometry profiles showing the distribution of LSK

cells in the spleen of $Il2^{-/-}Rag1^{-/-}$ mice compared to WT, $Il2^{-/-}$ and $Rag1^{-/-}$ mice. **E.** Quantification of percent LSK population in the spleen of indicated mice. Numbers inside

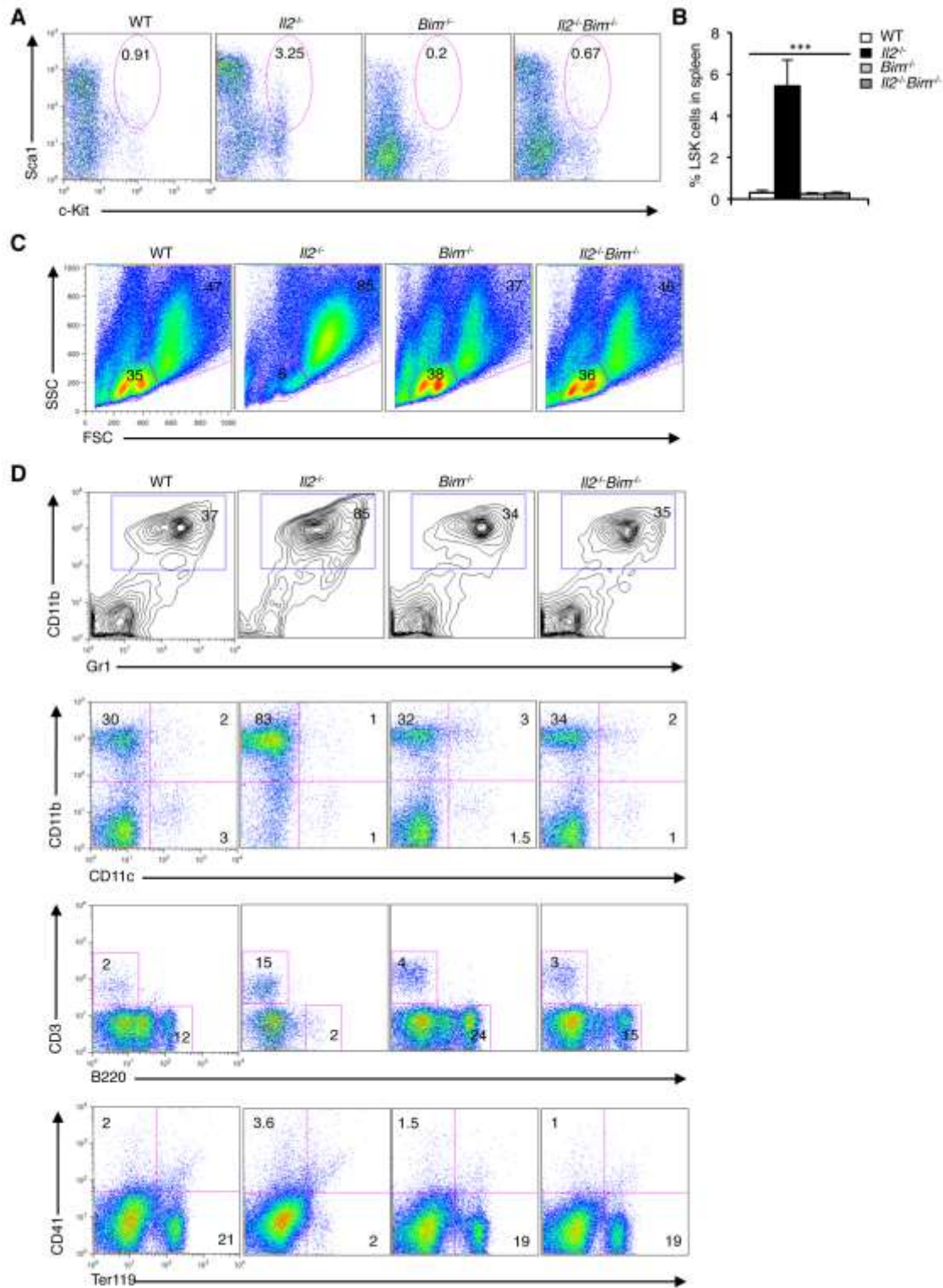
each FACS plot represent percent respective population. Data are shown as mean \pm s.d., $***P < 0.0001$, one-way ANOVA, $n = 4$ per group, data represent one out of three independent

experiments.



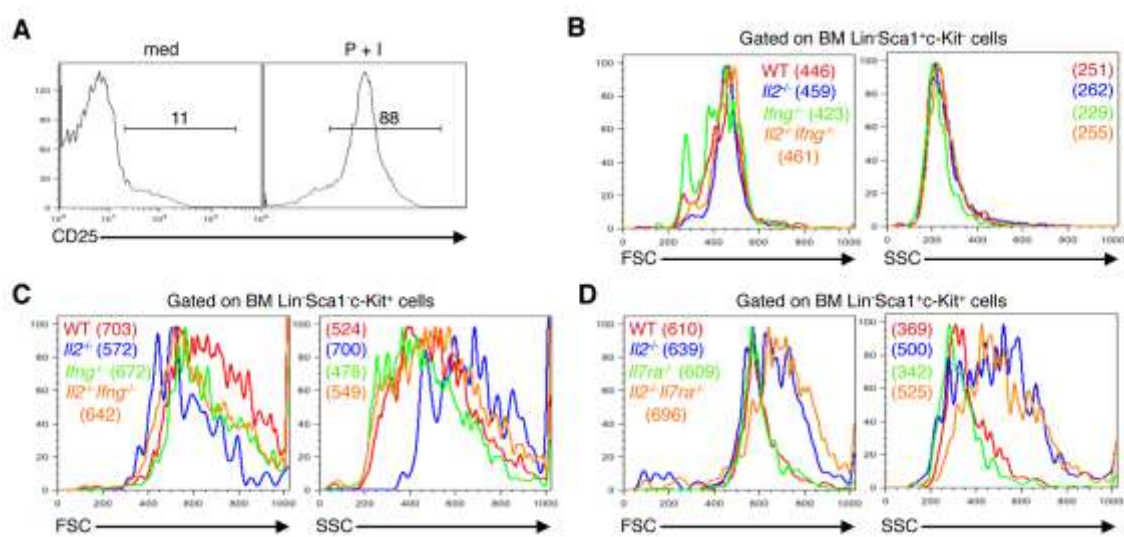
Supplementary Figure 4: T_{reg} cell activity is critical in maintaining BM hematopoiesis.

A. Flow cytometry analysis of CD4⁺CD25⁺Foxp3⁺ T_{reg} cell population in the spleen of WT or *Il2*^{-/-} mice. **B.** Analysis of the distribution of CD4⁺CD25⁺Foxp3⁺ T_{reg} cell population in the BM, spleen and LNs of PBS or DT treated DERE_G mice. **C.** Distribution of LSK cells in the spleen of PBS or DT treated DERE_G mice. **D.** Quantification of percent LSK population in the spleen of PBS or DT treated DERE_G mice. **E-H.** Distribution of CD11b⁺ and Gr1⁺ cells (**E**), CD11b⁺ and CD11c⁺ cells (**F**), IgM⁺ and IgD⁺ cells (**G**), and Ter119⁺ and CD41⁺ cells (**H**) in the BM of DERE_G mice treated with PBS or DT. Numbers inside each FACS plot represent percent respective population. Data are shown as mean ± s.d., ***P* = 0.0013, unpaired *t*-test, *n* = 4 per group, data represent one out of three independent experiments.

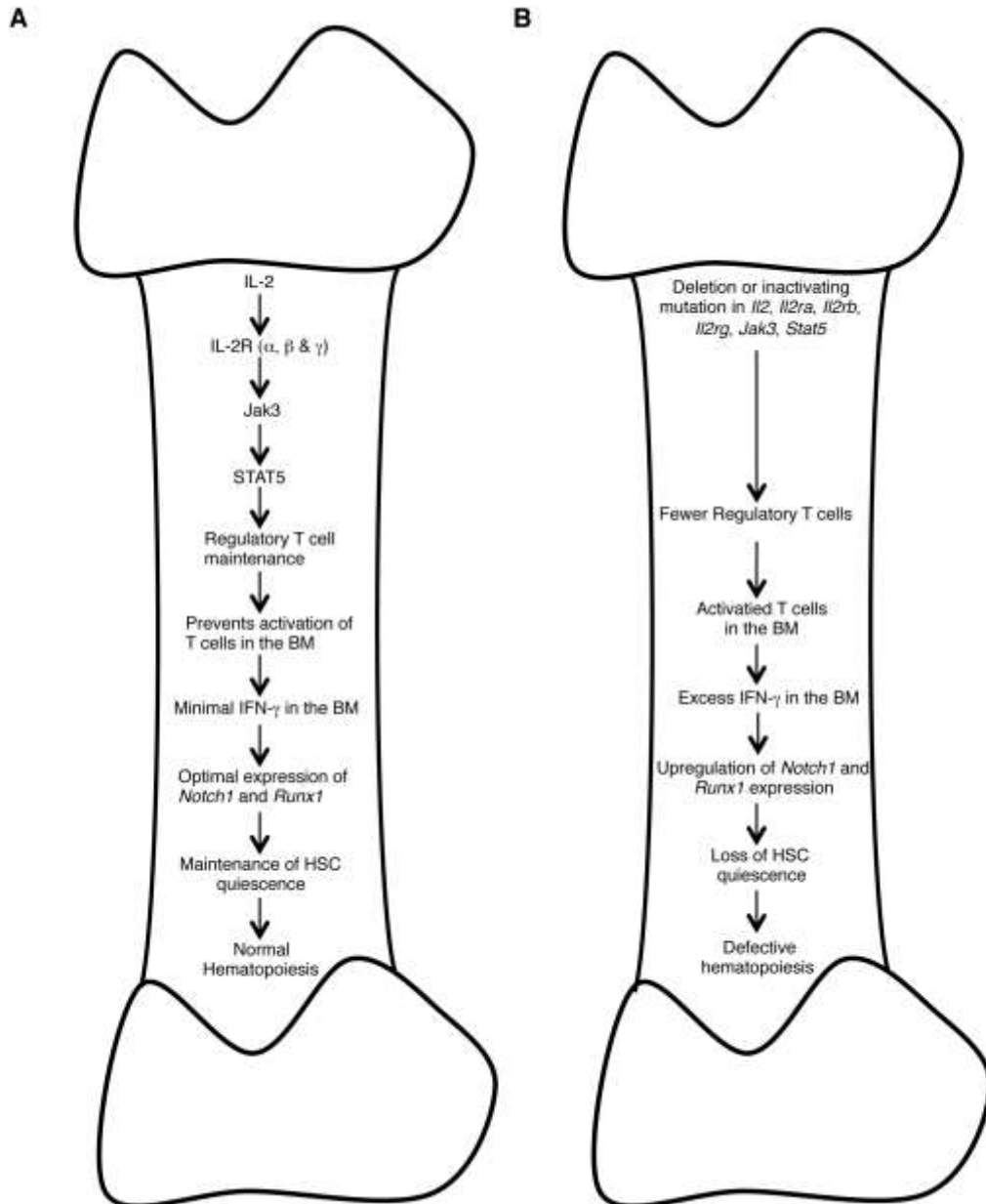


Supplementary Figure 5: Restoration of T_{reg} cells reverses hematopoietic defects in *Il2*^{-/-} mice. **A.** Distribution of LSK cells in the spleen of *Il2*^{-/-}*Bim*^{-/-} mice compared to WT, *Il2*^{-/-} and *Bim*^{-/-} mice as revealed by flow cytometry. **B.** Quantification of percent LSK population in the

spleen of indicated mice. **C.** Profiles of BM cells according to their FSC/SSC distribution pattern in *Il2^{-/-}Bim^{-/-}* mice compared to WT, *Il2^{-/-}* and *Bim^{-/-}* mice. **D.** Distribution of myeloid (Gr1^+ , CD11b^+ and CD11c^+), lymphoid (CD3^+ and B220^+), and erythroid (Ter119^+ and CD41^+) cells in the BM of *Il2^{-/-}Bim^{-/-}* mice compared to littermate control mice. Numbers inside each FACS plot represent percent respective population. Data are shown as mean \pm s.d., *** $P < 0.0001$, one-way ANOVA, $n = 4$ per group, data represent one out of three independent experiments.



Supplementary Figure 6: IFN- γ deficiency reverses HSC defects in *Il2^{-/-}* mice. **A.** Flow cytometry profiles of CD25 expression in WT CD4^+ T cells either left unstimulated or stimulated for 6 h with P+I. **B.** FSC, and SSC profiles of BM $\text{Lin}^- \text{Sca1}^+ \text{c-Kit}^-$ cells from *Il2^{-/-} Ifng^{-/-}* mice compared to WT, *Il2^{-/-}* and *Ifng^{-/-}* mice. **C.** FSC, and SSC profiles of BM $\text{Lin}^- \text{Sca1}^+ \text{c-Kit}^+$ cells from *Il2^{-/-} Ifng^{-/-}* mice compared to WT, *Il2^{-/-}* and *Ifng^{-/-}* mice. **D.** Profile of BM LSK cells from *Il2^{-/-} Il7ra^{-/-}* mice according to their FSC, and SSC distribution pattern compared to that from littermate WT, *Il2^{-/-}* and *Il7ra^{-/-}* mice. Numbers inside the histograms in **(A)** represent percent respective population, and in the histograms in **(B-D)** represent MFI. Data represent one out of three independent experiments, $n = 4$ per group.



Supplementary Figure 7: IL-2-dependent T_{reg} cell activity is essential for BM hematopoiesis. A. Scheme showing the essentiality of IL-2 signaling on the maintenance of HSCs in the bone marrow. **B.** Schematic representation of the effects of deficiency in IL-2 or any of the components of IL-2 signaling pathway on HSC maintenance and hematopoiesis in the bone marrow.

Supplementary Table 1
LIST OF RT-PCR PRIMERS

Gene	Primer Sequence	Product Size
<i>Actb</i>	For: 5'-CCAGGTCATCACTATTGGCAAGGA-3' Rev: 5'-GAGCAGTAATCTCCTTCTGCATCC-3'	223 bp
<i>Bcl2</i>	For: 5'-GGTGGTGGAGGAACTCTTCA-3' Rev: 5'-CTCACTTGTGGCCCAGGTAT-3'	326 bp
<i>Bcl2l1</i>	For: 5'-TTCGGGATGGAGTAAACTGG-3' Rev: 5'-TGTCTGGTCACTTCCGACTG-3'	318 bp
<i>Bmi1</i>	For: 5'-TGTCCTGTGTGGAGGGTACT-3' Rev: 5'-CAACTTCTCCTCGGTCTTCA-3'	299 bp
<i>Cdkn1b</i>	For: 5'-GCTCTGCTCCATTTGACTGT-3' Rev: 5'-GTCCCGGGTTAGTTCTTCAT-3'	293 bp
<i>Cebpa</i>	For: 5'-GGTGGACAAGAACAGCAACGAG-3' Rev: 5'-TAGAGATCCAGCGACCCGAAAC-3'	325 bp
<i>Csf1</i>	For: 5'-AGTCTGTCTTCCACCTGCTG-3' Rev: 5'-TGGTAGTGGTAGGCCACATT-3'	299 bp
<i>Csf1r</i>	For: 5'-ATGAGTCCCTCTTCACTCCG-3' Rev: 5'-ACCTTCAGCACTGCATCTTC-3'	306 bp
<i>Csf2ra</i>	For: 5'-CCTGCTCTTCTCCACGCTAC-3' Rev: 5'-ACTCCCCAGCAGTGACAAGT-3'	541 bp
<i>Csf3r</i>	For: 5'-AACTACACCCAGGCCTTCT-3' Rev: 5'-GAGCTCAAACCTGGTCCTTGC-3'	606 bp
<i>Cxcr4</i>	For: 5'-GTGGATGGTGGTGTTCAT-3' Rev: 5'-CTCGAAGTCACATCCTTGCT-3'	254 bp
<i>Egr2</i>	For: 5'-CTCCCATCTCTGCACCTAGA-3' Rev: 5'-ATAAGGAGGAGGAGGTGGTG-3'	284 bp
<i>Epor</i>	For: 5'-GGACACCTACTTGGTATTGG-3' Rev: 5'-GACGTTGTAGGCTGGAGTCC-3'	452 bp
<i>Flk2</i>	For: 5'-GACAAATCTCCCAATTGCAC-3' Rev: 5'-GTGGCAGATCAACACAATGA-3'	291 bp
<i>Gata1</i>	For: 5'-ATTCCTGGGGGCTCACCTTATG-3' Rev: 5'-TCCACAGTTCACACACTCTCTGGC-3'	366 bp
<i>Gata2</i>	For: 5'-AGCAAGGCTCGTTCCTGTTCAG-3' Rev: 5'-CCATAAGGTGGTGGTTGTCGTC-3'	231 bp
<i>Gfi1</i>	For: 5'-GAGCTTCAAGAGGTCATCCA-3' Rev: 5'-AGTCCATGCTGAGTCTCTCG-3'	306 bp
<i>Hax1</i>	For: 5'-GATGACGACGATGATGATGA-3' Rev: 5'-CTGGGTTGGTGA CTATCTGG-3'	330 bp
<i>Hoxa9</i>	For: 5'-TCACCAACCAAACACAACAG-3' Rev: 5'-CACAAATTAATCACGCCATCA-3'	299 bp
<i>Hoxb4</i>	For: 5'-TCACTCCCTTCCACCATAGA-3' Rev: 5'-ATAACAACGGGAGAGGGTTC-3'	342 bp

<i>Ifng</i>	For: 5'-CAATGAACGCTACACACTGC-3' Rev: 5'-CGCTTATGTTGTTGCTGATG-3'	279 bp
<i>Il2</i>	For: 5'-CCCTTGCTAATCACTCCTCA-3' Rev: 5'-TTCCAATTGAAAGCTTTTGC-3'	392 bp
<i>Il3ra</i>	For: 5'-GCCCTGCATGGACAACACT-3' Rev: 5'-GCACCGTAGCCACTGAAGTCA-3'	423 bp
<i>Il4</i>	For: 5'-GACGGCACAGAGCTATTGAT-3' Rev: 5'-AAAATATGCGAAGCACCTTG-3'	253 bp
<i>Il10</i>	For: 5'-GGACCAGCTGGACAACATAC-3' Rev: 5'-CACTCTTCACCTGCTCCACT-3'	263 bp
<i>Il12b</i>	For: 5'-TGTCTGCAGAGAAGGTCACA-3' Rev: 5'-CAAAGGCTTCATCTGCAAGT-3'	218 bp
<i>Il23a</i>	For: 5'-GATCTGAGAAGCAGGGAACA-3' Rev: 5'-CAGAACTGGCTGTTGTCCTT-3'	291 bp
<i>Klf1</i>	For: 5'-GATCGCCGGAGACGCAGGCT-3' Rev: 5'-TCCCCAGTCCTTGTGCAGGA-3'	363 bp
<i>Lef1</i>	For: 5'-CCAGACTGTCTCCACAGCTT-3' Rev: 5'-TGACAGGTCAGCACAGAAGA-3'	288 bp
<i>Mpl</i>	For: 5'-GAAATCTGCCTGCTGTGACT-3' Rev: 5'-GAACCAGGAAGGAAGGTGAT-3'	254 bp
<i>Meis1</i>	For: 5'-AATGCCTATCGATTTGGTGA-3' Rev: 5'-CCTTATCAGGGTCATCATCG-3'	266 bp
<i>Myc</i>	For: 5'-CATCCTGTCCATTCAAGC-3' Rev: 5'-TAATTCCAGCGCATCAGT-3'	200 bp
<i>Nfe2</i>	For: 5'-GAGCCCTGGCCATGAAGATTCC-3' Rev: 5'-CACCATCAGCAGCCTGTTGCAG-3'	391 bp
<i>Notch1</i>	For: 5'-TTGACGTCCTCTCCTGTGC-3' Rev: 5'-ACACAGGTGCCATTGTTGAA-3'	408 bp
<i>Notch2</i>	For: 5'-ACCCTTGTATGCACGGAGTC-3' Rev: 5'-CCAGGTTATTGCACGTTCCCT-3'	373 bp
<i>Pbx1</i>	For: 5'-AAACTGCCACAGAATGAAGC-3' Rev: 5'-CTTCTCCAGCTCTGTGTGGT-3'	298 bp
<i>Pu.1</i>	For: 5'-CGGATGACTTGGTTACTTACG-3' Rev: 5'-GTAGGAAACCTGGTGACTGAG-3'	292 bp
<i>Runx1</i>	For: 5'-GAGGCAAACCTCTGTCCTGAA-3' Rev: 5'-TTAGGCCTCAAAGACACCTG-3'	312 bp
<i>Scl</i>	For: 5'-TTCTACAGGACGTGCTTTCC-3' Rev: 5'-GAAACAGAAATGCCAGCCTA-3'	324 bp
<i>Slamf1</i>	For: 5'-TCCATGCCTCAGTTTCTCTC-3' Rev: 5'-GGCAACTTTACAGCAGCATT-3'	264 bp
<i>Tel/Etv6</i>	For: 5'-TAAACACAGTTTGGCCATT-3' Rev: 5'-AACGCACAGCAGCTCTAACT-3'	273 bp
<i>Tgfb</i>	For: 5'-GCTTCAGCTCCACAGAGAAG-3' Rev: 5'-CGTAGTAGACGATGGGCAGT-3'	255 bp
<i>Tnf</i>	For: 5'-TGGGTGTTTCATCCATTCTCT-3' Rev: 5'-TTTGAGTCCTTGATGGTGGT-3'	232 bp
<i>Trp53</i>	For: 5'-CAGGGCTGAGACACAATC-3' Rev: 5'-TGGGGTAGGGTGAGATTT-3'	201 bp