

S1 Appendix

Further properties of Mix²

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1 Parameter estimation for the Mix² model

1.1 Derivation of the EM update formulas

The Expectation Maximization (EM) algorithm [1] increases the likelihood $L(R|\theta)$ of a data set R under a model $p(R|\theta)$ by maximizing, or more generally increasing, the auxiliary function

$$Q(\theta'|\theta) = E_{Z|R,\theta}(\log p(R, z|\theta')) \quad (1)$$

Here, θ is the current parameter set of the model $p(R|\theta)$ and θ' is the new parameter set that needs to be optimized. In addition, $Z = (z_r)_{r \in R}$ is a sequence of random hidden variables z_r and, hence, the expression on the right hand side of (1) is the expected value of $\log p(R, z|\theta')$, where z is one realization of Z , with respect to the random variable Z given R and θ . The hidden variables in the Mix² model are the transcript variable, $t = i$, and the mixture variable, $b = j$.

A necessary condition for the maximization of $Q(\theta'|\theta)$ is that the gradient of $Q(\theta'|\theta)$ equals zero, i.e.

$$\frac{\partial}{\partial \theta'} Q(\theta'|\theta) = 0 \quad (2)$$

For the Mix² model this means that

$$\frac{\partial}{\partial \alpha_i} Q(\theta'|\theta) = 0 \quad (3)$$

and

$$\frac{\partial}{\partial \beta_{kj}} Q(\theta'|\theta) = 0 \quad (4)$$

where i is the index of transcript $t = i$ and k is the index of group $g = k$. As usual, the update formula of the relative abundances α_i is given by

$$\alpha_i^{(n+1)} = \frac{1}{|R|} \sum_r p^{(n)}(t = i|r) \quad (5)$$

where $\alpha_i^{(n+1)}$ and $p^{(n)}(t = i|r)$ are the relative abundance and posterior probability after the $n + 1$ -th and n -th iteration of the EM algorithm. In addition to (4), the β_{kj} have to satisfy the constraint

$$\sum_{j=1}^M \beta_{kj} = 1 \quad (6)$$

where M is the number of mixture components. This constraint can be enforced with the Lagrange method. Taking the derivative with respect to β_{kj} leads to

$$\sum_{r \in R} p(g = k, b = j|r) + \beta_{kj} \lambda = 0 \quad (7)$$

which after some rearrangement results in

$$\beta_{kj}^{(n+1)} = \frac{\sum_r p^{(n)}(g = k, b = j|r)}{\sum_r p^{(n)}(g = k|r)} \quad (8)$$

where, as previously, $\beta_{kj}^{(n+1)}$ and $p^{(n)}(\cdot)$ are the mixture components and posterior probabilities after the $n + 1$ -th and after the n -th iteration, respectively. The posterior probabilities in (8) are given by

$$p^{(n)}(g = k, b = j|r) = \sum_{i \in k} p^{(n)}(t = i, b = j|r) \quad (9)$$

and

$$p^{(n)}(g = k|r) = \sum_{i \in k} p^{(n)}(t = i|r) \quad (10)$$

where the sums in (9) and (10) extend over all transcripts $t = i$ in group $g = k$ and the posteriors on the right-hand side of these equations can be derived according to Bayes formula as follows

$$p^{(n)}(t = i, b = j|r) = \frac{\alpha_i^{(n)} \beta_{ij}^{(n)} p(r|t = i, b = j)}{\sum_{ij} \alpha_i^{(n)} \beta_{ij}^{(n)} p(r|t = i, b = j)} \quad (11)$$

and

$$p^{(n)}(t = i|r) = \frac{\sum_j \alpha_i^{(n)} \beta_{ij}^{(n)} p(r|t = i, b = j)}{\sum_{ij} \alpha_i^{(n)} \beta_{ij}^{(n)} p(r|t = i, b = j)} \quad (12)$$

The posterior probability $p(r|t = i, b = j)$ in (11) and (12) is independent of the iteration. In the main paper the $p(r|t = i, b = j)$ were chosen to be Gaussians which are equidistantly distributed across the transcript $t = i$.

Without any tying, the group $g = k$ consists of a single transcript $t = i$ and (8) therefore becomes

$$\beta_{ij}^{(n+1)} = \frac{\sum_r p^{(n)}(t = i, b = j|r)}{\sum_r p^{(n)}(t = i|r)} \quad (13)$$

For global tying, on the other hand, the group consists of all the transcripts within the locus and therefore

$$p(g = k|r) = 1 \quad (14)$$

As a result, the update formula (8) becomes

$$\beta_j^{(n+1)} = \frac{1}{|R|} \sum_r p^{(n)}(b = j|r) \quad (15)$$

It is interesting to note, that (15) is similar to the update formula for the relative abundances α_i , equation (5). This is the case, because for global tying the following holds

$$p(r) = \sum_j \beta_j p(r|b = j) \quad (16)$$

which is similar to the superposition

$$p(r) = \sum_i \alpha_i p(r|t = i) \quad (17)$$

Multi-mapping reads and sequence specific bias

The previous discussion assumes that a fragment r maps uniquely to the genomic reference. If, on the other hand, fragment r has multiple hits $H(r)$ on the reference, then

$$p(h|r) = \frac{p(h)}{\sum_{h \in H(r)} p(h)} \quad (18)$$

needs to be taken into account when estimating the parameters of the Mix² model. Rather than calculating (18) during parameter estimation $p(h|r)$ is often set to $1/\#H(r)$ [2]. Equation (18) can be extended to cover the situation of a sequence specific bias. In this case, the probability that a sequence $seq(r)$ within or surrounding fragment r is generated can be smaller than 1 and the right-hand side of equation (18) needs to be multiplied by this sequence specific probability, $p(generate|seq(r))$. The probability $p(generate|seq(r))$ can, for instance, be estimated as in [4] by calculating the ratio of the probability of the sequence $seq(r)$ under the biased model to the uniform model. Most commonly, $seq(r)$ is a sequence directly preceding or following r and $p(generate|seq(r))$ therefore reflects the probability that a primer with start sequence $seq(r)$ anneals to the sample. Details on how equation (18) and its generalization to a sequence specific bias fits into the parameter estimation of the Mix² model are given in Section "Parameter estimation". It should be noted that in our current implementation of the Mix² model we do not take sequence specific bias into consideration, nor do we use (18) to calculate the posterior probability of a hit.

If fragment r has multiple hits $H(r)$ and a sequence specific bias then

$$p(t = i, b = j|r) = \sum_{h \in H(r)} p(t = i, b = j|h) p(h|r) \quad (19)$$

and the update formula for β_{kj} , equation (8), becomes

$$\beta_{kj}^{(n+1)} = \frac{\sum_{r \in R} \sum_{h \in H(r)} p^{(n)}(g = k, b = j|h) p(h|r)}{\sum_{r \in R} \sum_{h \in H(r)} p^{(n)}(g = k|h) p(h|r)}. \quad (20)$$

Here $p(h|r)$ is given by equation (18) or the right-hand side of equation (18) multiplied by $p(generate|seq(r))$ the probability of generating the sequence $seq(r)$, which is either part of or surrounding fragment r .

1.2 Identifiability and uniqueness of maximum likelihood solution

The Mix² model is identifiable on the set of fragments R iff the mapping $\theta \rightarrow p_\theta(R)$ is injective, where, as in the previous section, θ is the vector of pairs of parameters

$$\theta = ((a_i, b_{i,j}))_{i=1,\dots,N \wedge j=1,\dots,M} \quad (21)$$

The mapping $\theta \rightarrow p_\theta(R)$ is given by the product of two mappings

$$p_\theta(R) = A \cdot M \cdot \theta \quad (22)$$

where A is the linear map given by

$$A = (a_{r,(i,j)})_{r \in R \wedge (i,j) \in (1,\dots,N) \times (1,\dots,M)} \quad (23)$$

with

$$a_{r,(i,j)} = p(r|t = i, b = j) \quad (24)$$

which is the value of the j -th Gaussian of transcript i for fragment r . Hence r is an index for the rows and the pair (i, j) is an index for the columns of A . The second mapping in (22) is componentwise multiplication of θ given by

$$M(\theta) \rightarrow ((a_i b_{i,j}))_{i=1,\dots,N \wedge j=1,\dots,M} \quad (25)$$

The mapping M is invertible on the parameters θ since

$$\sum_j \alpha_i \beta_{ij} = \alpha_i \quad (26)$$

and thus equation (22) is injective iff A is injective on the set $M\theta$, which is the $NM - 1$ simplex Δ^{NM-1} . This condition can be checked by first checking the stronger condition of injectivity of A on the full linear space $\mathbb{R}^{N \times M}$. If A is injective on $\mathbb{R}^{N \times M}$ then, clearly, A is injective on Δ^{NM-1} . If, on the other hand, A is not injective on $\mathbb{R}^{N \times M}$ then it is necessary to check whether differences of elements in Δ^{NM-1} other than 0 lie in the kernel of A on $\mathbb{R}^{N \times M}$. The latter will be the case if the dimension of the kernel of A is greater than 1, since then

$$\dim(\ker(A)) + \dim(\Delta^{NM-1}) > \dim(\mathbb{R}^{N \times M}) \quad (27)$$

The dimension of the kernel of A is, for instance, greater than 1 if two transcripts $t = i$ and $t = i'$ share the same Gaussian $b = j$ and $b = j'$, which happens only if the transcripts have the same length and their exons are properly aligned. This situation can be avoided by shifting the Gaussians $p(r|t = i, b = j)$, $p(r|t = i', b = j')$ away from each other, which ensures that

$$p(r|t = i, b = j) \neq p(r|t = i', b = j') \quad (28)$$

and removes therefore identical columns in A . Shifting the Gaussians means that some of them are not equidistantly distributed along a transcript but has otherwise a minor effect on the properties of the Mix² model. Summarizing, we state the following

Proposition 1. *A sufficient condition for the identifiability of the Mix² model is the injectivity on $\mathbb{R}^{N \times M}$ of the matrix A in equations (23) and (24). If the Mix² model fails to be identifiable because two transcripts $t = i$ and $t = i'$ share one Gaussian for two of their mixture components $b = j$ and $b = j'$, then the Mix² model can be made identifiable by shifting the Gaussians $p(r|t = i, b = j)$, $p(r|t = i', b = j')$ away from each other.*

Equation (26) shows further that the Mix² model is equivalent to a mixture model of the distributions $p(r|t = i, b = j)$ with mixture weights c_{ij} if no Gaussian is shared between two transcripts. In this case, the maximum likelihood solution for the c_{ij} is unique, since the log likelihood surface of mixture models is concave [3], and the c_{ij} and the parameters of the Mix² model stand in a one-to-one relationship. This can be summarized as follows.

Proposition 2. *The Mix² model is equivalent to a mixture of the distributions $p(r|t = i, b = j)$ with respective mixture weights c_{ij} if no two transcripts share the same Gaussian. Since the log likelihood function for a mixture*

is concave there exists a unique maximum likelihood solution for the c_{ij} to which the EM algorithm converges. The α_i and β_{ij} of the Mix^2 model can be derived, in this case, from the c_{ij} as follows.

$$\alpha_i = \sum_{j=1}^M c_{ij} \quad (29)$$

$$\beta_{ij} = \frac{c_{ij}}{\alpha_i} \quad (30)$$

2 Fragment start distributions in Cufflinks

The Mix² model in the main paper factorizes the transcript specific fragment distribution $p(r|t = i)$ as follows

$$p(r|t = i) = p(s(r)|t = i)p(l(r)|s(r), t = i) \quad (31)$$

where $s(r)$ and $l(r)$ are the start and length of fragment r . Cufflinks [5], on the other hand, reverses the order of $s(r)$ and $l(r)$ in (31) and factorizes $p(r|t = i)$ according to

$$p(r|t = i) = p(l(r)|t = i)p(s(r)|l(r), t = i) \quad (32)$$

The fragment length distribution $p(l(r)|t = i)$ in (32) is derived from the cumulative distribution of fragment lengths $p(l(r))$ for the complete data set. For this purpose, $p(l(r))$ is truncated to the possible fragment lengths for transcript $t = i$ and subsequently renormalized such that

$$\sum_{l=1}^{l(t=i)} p(l|t = i) = 1 \quad (33)$$

where $l(t = i)$ is the length of transcript $t = i$. The fragment start distribution $p(s(r)|l(r), t = i)$, on the other hand, is assumed to be uniform over the possible fragment starts $s(r)$ for transcript $t = i$ and fragment length $l(r)$, i.e.

$$p(s(r)|l(r), t = i) = \frac{1}{l(t = i) - l(r) + 1} \quad (34)$$

The fragment start distribution $p(s(r)|t = i)$ for $t = i$ according to the Cufflinks model can be derived by summing $l(r)$ out of (32). In the absence of fragment length information, e.g. for single-end RNA-Seq data, Cufflinks assumes by default a Gaussian with mean 200 and standard deviation 80 for the cumulative fragment length distribution $p(l(r))$. For this default setting the fragment start distribution $p(s(r)|t = i)$ is given in Figure 2 (a) of the main article for transcripts with length between 400 bps and 3000 bps. It can be seen that for long transcripts the Gaussian distribution $p(l(r))$ produces a short and steep tail at the end of $p(s(r)|t = i)$, whereas this tail shifts increasingly to the 5' end of the transcript for shorter transcripts. The assumption of a Gaussian with mean 200 and standard deviation 80 corresponds to a size selection of the fragments prior to sequencing. Thus, Figure 2 (a) in the main text shows that even for a uniform fragment distribution, size selection generates a transcript length specific bias.

References

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