SUPPLEMENTARY METHODS

In Silico Simulations

The CPVT phenotype was modelled as described in our recent study using computationally reduced CASQ2 by modifying the Soltis-Saucerman model¹ and the Morotti-Grandi mouse cardiac cell model.²

We reduced the CASQ2 total protein ($B_{max_{CASQ2}}$) concentration by 25%, 50% and 75% in the following equation.

Calsequestrin buffer:

$$d\frac{[CASQ2]}{dt} = k_{on_CASQ2}[Ca]_{SR} (B_{max_CASQ2} - [CASQ2]) - k_{off_CASQ2}[CASQ2]$$

Buffer	B _{max_CASQ2} [mM]	$K_{off_CASQ2} (ms^{-1})$	$K_{on_{CASQ2}} (mM^{-1}ms^{-1})$
Calsequestrin	2.7	65	100

Cellular and tissue simulations

Mouse ventricular myocyte model: The I_{Na} channel was replaced in the Morotti-Grandi mouse cardiac cell model ² with our published Markov model.³ We then adjusted three transition rates for the mouse model to simulate the mouse I_{Na} kinetics:

Transition rates	
0→C1	β13 * 0.45
O→IF	α2 * 0.45
O→IS	αx * 0.45

Virtual myocytes were paced using a -9.5 pA/pF current stimulus for 5 ms in single cells for 2 mins at 2Hz pacing frequency in the presence of 1.0µM ISO.

Rabbit ventricular myocyte model: The I_{Na} channel was replaced in the Soltis-Saucerman rabbit cardiac cell model.¹ Virtual whole cells and tissues were allowed

to "rest" without external stimuli for 10 minutes to establish initial conditions. Cells were then virtually paced using a -80 pA/pF current stimulus for 0.5 ms in single cells and -500 pA/pF stimulus for 2.0 ms in tissues. Cells were paced for 2 minutes in the presence of 1 μ M ISO. Parameters including upstroke velocity, action potential duration (APD), and the number of early and delayed after-depolarizations (DADs) were tracked over the course of each simulation.

One-dimensional tissue was simulated as a fibre of 165 cells $(1.65 \text{cm})^4$ with reflective boundary conditions. Transmural heterogeneity was incorporated into the tissue by a linear decrease to 25% maximal I_{to} conductance,⁵ corresponding to a linear transition from epicardial to endocardial tissue ⁶ and an APD gradient of 205-224ms. The diffusion coefficient D_x was set to 0.002 cm²/ms to establish a conduction velocity of 61-73 cm/s (epicardium-endocardium in wild type conditions).⁷

REFERENCES

- **1.** Soltis AR, Saucerman JJ. Synergy between CaMKII substrates and beta-adrenergic signaling in regulation of cardiac myocyte Ca(2+) handling. Biophys J 2010;99:2038-2047.
- 2. Morotti S, Edwards AG, McCulloch AD, Bers DM, Grandi E. A novel computational model of mouse myocyte electrophysiology to assess the synergy between Na+ loading and CaMKII. J Physiol 2014;592:1181-1197.
- **3.** Moreno JD, Yang PC, Bankston JR, Grandi E, Bers DM, Kass RS, Clancy CE. Ranolazine for congenital and acquired late INa-linked arrhythmias: in silico pharmacological screening. Circ Res 2013;113:e50-61.
- **4.** Glukhov AV, Fedorov VV, Lou Q, Ravikumar VK, Kalish PW, Schuessler RB, Moazami N, Efimov IR. Transmural dispersion of repolarization in failing and nonfailing human ventricle. Circ Res 2010;106:981-991.
- **5.** Myles RC, Bernus O, Burton FL, Cobbe SM, Smith GL. Effect of activation sequence on transmural patterns of repolarization and action potential duration in rabbit ventricular myocardium. Am J Physiol Heart Circ Physiol 2010;299:H1812-1822.
- **6.** Fedida D, Giles WR. Regional variations in action potentials and transient outward current in myocytes isolated from rabbit left ventricle. J Physiol 1991;442:191-209.
- **7.** Brugada J, Boersma L, Kirchhof C, Allessie M. [Anisotropy and reentrant ventricular tachycardia: experimental model in the isolated rabbit heart]. Rev Esp Cardiol 1990;43:558-568.