

## Supporting Information:

### **Air Pollution and Climate Change Effects on Allergies in the Anthropocene: Abundance, Interaction and Modification of Allergens and Adjuvants**

Kathrin Reinmuth-Selzle<sup>1,#,\*</sup>, Christopher J. Kampf<sup>1,2,#,\*</sup>, Kurt Lucas<sup>1</sup>, Naama Lang-Yona<sup>1</sup>, Janine Fröhlich-Nowoisky<sup>1</sup>, Manabu Shiraiwa<sup>1,3</sup>, Pascale S. J. Lakey<sup>1</sup>, Senchao Lai<sup>1,4</sup>, Fobang Liu<sup>1</sup>, Anna T. Kunert<sup>1</sup>, Kira Ziegler<sup>1</sup>, Fangxia Shen<sup>1</sup>, Rossella Sgarbanti<sup>1</sup>, Bettina Weber<sup>1</sup>, Iris Bellinghausen<sup>5</sup>, Joachim Saloga<sup>5</sup>, Michael G. Weller<sup>6</sup>, Albert Duschl<sup>7</sup>, Detlef Schuppan<sup>8,9</sup>, and Ulrich Pöschl<sup>1,\*</sup>

<sup>1</sup>Multiphase Chemistry Department, Max Planck Institute for Chemistry, Mainz, 55128, Germany

<sup>2</sup>Institute of Inorganic and Analytical Chemistry, Johannes Gutenberg University, Mainz, 55128, Germany

<sup>3</sup>Department of Chemistry, University of California, Irvine, California, 92697-2025, United States

<sup>4</sup>South China University of Technology, School of Environment and Energy, Guangzhou, 510006, China

<sup>5</sup>Department of Dermatology, University Medical Center, Johannes Gutenberg University, Mainz, 55131, Germany

<sup>6</sup>Division 1.5 Protein Analysis, Federal Institute for Materials Research and Testing (BAM), Berlin, 12489, Germany

<sup>7</sup>Department of Molecular Biology, University of Salzburg, Salzburg, 5020, Austria

<sup>8</sup>Institute of Translational Immunology and Research Center for Immunotherapy, Institute of Translational Immunology, University Medical Center, Johannes Gutenberg University, Mainz, 55131 Germany

<sup>9</sup>Division of Gastroenterology, Beth Israel Deaconess Medical Center and Harvard Medical School, Boston, Massachusetts, 02215, United States

<sup>#</sup>These authors contributed equally

\*Address correspondence to U. Pöschl, K. Reinmuth-Selzle, and C.J. Kampf, Multiphase Chemistry Department, Max Planck Institute for Chemistry, Hahn-Meitner Weg 1, D-55128 Mainz, Germany. Tel: +49 6131 305 7001. Emails: u.poschl@mpic.de, k.selzle@mpic.de, c.kampf@mpic.de

The supplementary material includes: Supplementary Section S1, Supplementary Tables S1-S2, Supplementary Figures S1-S5

## **Section S1. Allergic Sensitization and Response**

Essential steps and influencing factors in the development of IgE-mediated allergies (type I hypersensitivities) are outlined in Figure S1. Here, the term atopy refers to the genetic predisposition to mount IgE-responses to common allergens.<sup>1</sup> Genetic and epigenetic effects can also influence the further development of allergies that can be triggered by lifestyle and environmental factors. Among the lifestyle and environmental risks are allergen exposure, reduced childhood exposure to pathogens and parasites (hygiene hypothesis), diet/nutrition, psychological/social stress, and environmental pollution, including outdoor and indoor air pollutants (ozone, nitrogen oxides, diesel exhaust particles, tobacco smoke, etc.), which may effectively lead to an over- or under-stimulation of the immune system.<sup>2-22</sup> Figure S2 shows a simplified overview of cellular and molecular interactions that are central to the processes of allergic sensitization and response, involving elements of both adaptive and innate immunity.<sup>2</sup>

### ***Allergic sensitization and Th2 cells***

The term allergic sensitization describes the first induction of an allergic immune response upon exposure to an allergen, which is a multifactorial process that depends on the status of the immune system, the epithelium, the type and concentration of allergens and adjuvants, as well as the timing of exposure.<sup>2, 23-28</sup> During the sensitization process, the allergens cross epithelial barriers and are internalized by antigen presenting cells (APC), primarily dendritic cells (DC) as sentinels of the immune system. In the endolysosomal compartments of the DC, allergenic proteins are enzymatically cleaved, the allergen-derived immunodominant peptides are bound by the major histocompatibility complex (MHC) class II, and the MHC class II-peptide complex is presented on the surface of the DC. Depending on cytokines and other factors derived from epithelial cells and the surrounding micromilieu (e.g., IL-25, IL-33, TSLP), the DC are activated and migrate to secondary lymphoid organs such as the local draining lymph nodes or mucosal-associated lymphoid tissues.<sup>29-32</sup> There the DC interact with naïve T helper cells, and at least one of the allergen-derived immunodominant peptides presented by the DC is recognized as a T cell epitope binding to a T cell receptor (TCR). Recognition by the TCR induces T helper cell activation and differentiation into Th cells with a Th2 cytokine profile (mainly IL-4 and IL-13) depending on additional costimulatory signals from the DC and the

surrounding micromilieu, which are influenced by adjuvants as detailed below. These secondary signals involve various types of molecules (e.g., CD80, CD86, PD-1, ICOS, CLTA-4, CD40, OX-40) that can promote the development of an inflammatory allergic reaction or favor immune regulation leading to tolerance of the allergen.<sup>33</sup>

Through interaction with complement molecules of the innate immune system (opsonization) and with follicular dendritic cells, allergens are also collected in the follicles/germinal centers of secondary lymphoid organs.<sup>34-44</sup> There, the allergenic proteins interact with B cells via the B cell receptors (BCR), which recognize epitopes on the protein surface (B cell epitopes). The BCR contains the same antigen-binding fragment (Fab) as the antibodies produced by the B cell. Th2 cytokines and ligation of CD40 on the B cell surface with CD40 ligand on the Th2 cell surface induce an immunoglobulin class switch to the production of allergen-specific IgE antibodies. The binding between the allergen and the Fab region is optimized in the course of affinity maturation by somatic hypermutation of the Fab region and clonal selection of B cells, leading to a differentiation of B cells into antibody secreting plasma cells and memory cells.<sup>25</sup> The IgE production by B cells is controlled by T helper cells and regulatory T cells (Treg), whereby Th1 and Treg cells secreting IFN-gamma, IL-10 and TGF-beta dominate the regulation in healthy individuals and during successful allergen-specific immunotherapy.<sup>45-46</sup> Allergic diseases are mainly characterized by Th2 cell-mediated inflammation, whereas bacterial or viral infections normally lead to Th1 cell-mediated inflammation.<sup>47</sup>

### ***Allergic response and IgE antibodies***

Upon re-exposure of sensitized individuals to the allergen, the allergenic proteins (B cell epitopes) are recognized by specific IgE antibodies bound to the high affinity IgE-receptors (FcεRI) on effector cells like mast cells and basophils. The recognition and binding of allergens induces IgE cross-linking and activation of the FcεRI receptors, which leads to effector cell degranulation with release of highly vasoactive mediators like histamine that are responsible for the allergic symptoms, such as vascular leakage, edema, and inflammatory cell attraction and activation. Multivalent allergens like protein oligomers exhibiting multiple IgE binding sites (epitopes) on the protein surface are particularly effective in IgE cross-linking and FcεRI

activation, thus initiating especially strong cellular responses over a wide concentration range.<sup>48-50</sup>

Once the allergy is established, enhanced mucosal and tissue levels of IgE favor an immediate or early-phase reaction to the allergen (within minutes), explaining the immediate onset of symptoms upon allergen exposure. The rapid release of mast-cell derived mediators (histamine, leukotrienes, prostaglandins, tumor necrosis factor  $\alpha$ ) in the early-phase contributes to the rapid influx of other cells characteristic of promoting allergy (e.g. eosinophils, basophils, monocytes). The influx of these cells characterizes the late-phase allergic reaction (which can start 4-12 hours after allergen exposure and persist up 48 hours) with the release of other pro-inflammatory mediators such as lipid mediators and reactive oxygen and nitrogen species (ROS/RNS). The term chronic phase is used to describe persistent inflammation induced by prolonged or repetitive allergen exposure and the presence of large numbers of innate and adaptive immune cells leading to damage of the affected tissues and epithelia (e.g., the pulmonary mucosa in allergic asthma).<sup>51-52</sup> As IgE antibodies and related immune reactions are normally involved in the defense against parasitic infections, allergic reactions can be regarded as “false alarms” of the immune system.<sup>53-56</sup>

### ***Adjuvants and pattern recognition receptors***

Adjuvants and their interaction with the immune system play a critical role in the development of allergies. Here we use the term adjuvant generically for substances that are promoting pro-allergic innate and adaptive immune responses. Adjuvants can trigger the immune system by inducing tissue damage and subsequent enhanced uptake of allergens, by inducing oxidative stress and activation of immune cells, by co-exposure with the allergen favoring Th2 responses, or by modification of allergens enhancing their allergic potential. An overview of biogenic and anthropogenic adjuvants and their effects on the immune system is given in Table 2.<sup>26, 57-107</sup> Major substance groups are pollen-associated and microbial compounds (pollen associated lipid mediators (PALMs), lipopolysaccharides (LPS),  $\beta$ -glucans, etc.) and anthropogenic air pollutants (particulate matter, ozone and nitrogen oxides).

Many allergenic proteins have intrinsic adjuvant functions which are mediated, e.g., by protease activity, lipid-binding functions, and carbohydrate residues interacting with the innate

immune system.<sup>26, 62, 108-111</sup> More than half of the known major allergens are lipid-binding proteins, and many allergens can bind ligands exhibiting pathogen-associated molecular patterns (PAMPs).<sup>109</sup> PAMPs are structural characteristics of microbial substances like lipopolysaccharides (LPS), flagellin, glucanes, and other biomolecules that are recognized by pattern recognition receptors (PRR) such as transmembrane Toll like receptors (TLR) and C-type lectin receptors.<sup>109, 112-113</sup> For example, the major house dust mite allergen Der p 2 can mimic the function of MD-2, which is a key protein involved in TLR4 activation by LPS.<sup>114</sup>

Besides pathogen-associated molecular patterns, also damage-associated molecular patterns (DAMPs) formed upon oxidative, radiative or heat-induced stress can be recognized by PRR and activate the innate immune system.<sup>93, 112-113, 115</sup> PRR play a critical role in regulating the function of APC and the type of adaptive immune response<sup>110, 116</sup>, and the involvement of TLR and other PRR may enhance endocytosis, drive adaptive immune responses, and determine a cytokine bias, linking adaptive and innate immunity.<sup>112, 117</sup> Moreover, TLR and other PRR can mediate self-sustaining cycles of inflammation and may be involved in the development of allergies and related diseases.<sup>93, 115</sup> These cyclic processes comprise the release and formation of cytokines, ROS/RNS, and DAMPs, which in turn can reactivate PRR (see Figure S5, supporting information). They can be triggered by external influencing factors including air pollutants and related oxidative stress.<sup>93, 115</sup>

By recognizing characteristic structures of allergens through PRR, epithelial cells can influence the activation of DC as outlined above as well as the activation of other innate immune cells such as eosinophils, basophils and innate lymphoid cells (ILC), also called non-T, non-B effector cells or natural helper cells. Depending on their capacity to produce Th1, Th2, and other cytokine patterns, ILC are grouped into different subsets. The subset ILC2 can generate high amounts of IL-13, IL-5 and IL-9 in response to IL-25, IL-33 and TSLP leading to eosinophilia and mucus production. They are increased in lung parenchyma, nasal polyps and peripheral blood from asthmatic patients as well as in the skin from patients with atopic dermatitis.<sup>118-123</sup>

Because of the wide variety and complex nature of the cellular and molecular interactions involved in the development of allergies, air pollutants and climate parameters may influence the underlying physiological mechanisms and chemical reactions in many ways.

**Table S1.** Air pollution effects on the allergenic potential of plant pollen and fungal spores.

<b>Allergenic Species</b>	<b>Exposure</b>	<b>Effects</b>
<i>Ambrosia artemisiifolia</i> (ragweed)	high-traffic roads vs. vegetated areas (Lombardy region, Italy)	increased IgE-binding capacity of pollen protein extracts from polluted environments <sup>124</sup>
	NO <sub>2</sub> exposure	increased IgE reactivity, modified cell wall morphology, increased allergen content, and changed proteomic pattern including S-nitrosylation products and upregulation of other proteins (e.g., 14-3-3 protein) <sup>125</sup>
	O <sub>3</sub> exposure	stimulates NADPH oxidase activity (ROS production) leading to pollen damaging and decreased viability <sup>126</sup>
	O <sub>3</sub> exposure	enhances stress-induced RNA transcripts, cell wall components and wax of the pollen, Amb a 1 content not affected <sup>127</sup>
<i>Canna indica</i> (indian shot)	city vs. rural (Tehran, Iran)	increased number of eosinophils and IgE levels induced by pollen extracts from polluted environments <sup>128</sup>
<i>Cupressus arizonica</i> (arizona cypress)	high-traffic roads vs. vegetated areas (Spain)	increased Cup a 3 content of pollen collected in polluted environments <sup>129-130</sup>
<i>Phleum pratense</i> L. (timothy grass)	high-traffic roads vs. rural site (Bavaria, Germany)	decreased Phl p 5 content of pollen collected in polluted environments <sup>131</sup>
		Increased release of eicosanoid-like pro-inflammatory substances in pollen collected near roads with high traffic <sup>132</sup>
<i>Zinnia elegans</i> (common zinnia)	city vs. rural (Tehran, Iran)	increased skin prick test response observed in guinea pigs, decreased protein content of pollen collected in polluted environments <sup>133</sup>
<i>Betula pendula</i> (birch)	city vs. rural (Munich, Germany)	increased ozone levels increase allergenicity (skin prick test) and Bet v 1 content of pollen <sup>134</sup>
		changes in pollen proteome (e.g., upregulation of 14-3-3 protein), no difference in Bet v 1 content, and enhanced chemotactic activity of pollen extracts from polluted environments <sup>135</sup>
		differences in pollen microbiome, higher bacterial diversity was correlated with the allergen and PALMs content <sup>136</sup>
<i>Acer negundo</i> (maple), <i>Platanus x acerifolia</i> (London plane)	NO <sub>2</sub> exposure	decrease in pollen viability, decrease in soluble protein content, increased IgE reactivity <sup>137</sup>
	O <sub>3</sub> exposure	increased IgE reactivity <sup>138</sup>

<i>Aspergillus fumigatus</i> (mold)	exposure to outdoor air (urban area, Israel), and O <sub>3</sub> /NO <sub>2</sub> /O <sub>3</sub> +NO <sub>2</sub>	exposure time dependent changes of IgE binding capacity, suggested links to chemical modifications of Asp f 1 (e.g., nitration and deamidation) <sup>139</sup>
	combined exposure of mice to DEP and allergen	changes in methylation of genes favoring Th2 polarization and increased IgE production <i>in vivo</i> (epigenetics) <sup>92</sup>

---

**Table S2** Research activities proposed to address the key questions raised in the outlook section (Sect. 5, Q1-Q4).

---

*ENVIRONMENTAL STUDIES (Q1)*

- Quantify the atmospheric abundance and mixing state of allergenic bioparticles, major allergenic proteins and adjuvant substances (DEP, LPS, PALMs, flavonoids, glucans ...) in relation to air quality, meteorological, climate and ecosystem parameters (T, RH, PM, O<sub>3</sub>, NO<sub>x</sub>, CO<sub>2</sub>, vegetation cover, nutrient status ...) using advanced physical, chemical and biological measurement techniques (mass spectrometry, fluorescence spectroscopy and microscopy; genomic and proteomic analyses; immunological assays, etc.)
- Identify and quantify the products, kinetics and molecular mechanisms of air pollutant interactions with natural allergens and adjuvants (nitrated, oxidized, and oligomerized proteins, lipids, flavonoids, etc.) in field studies and laboratory experiments under conditions relevant for outdoor and indoor environments
- Establish and maintain long-term observations to document ongoing and future changes in the release, distribution, and allergenic potential of aeroallergens and adjuvants

*IMMUNOLOGICAL STUDIES (Q2)*

- Characterize and quantify air pollutant effects on epithelial transport and signaling (permeability, cytokine and mediator release)
- Determine air pollutant effects on the uptake, processing, and presentation of allergens by APCs, changes in allergen epitopes and adjuvant functions, and generation of multivalent allergens
- Investigate the role of air pollutant interactions with airway mast cells and basophils as a potential cause of initial IL-4 release polarizing immunity to Th2 responses
- Explore which components of the immune system could be modulated to prevent adverse effects of air pollution, e.g. whether therapeutic monoclonal antibodies against relevant cytokines (e.g., IL-4, IL-5, IL-13) or IgE antibodies could make a difference.
- Investigate immunological effects of co-exposure to realistic mixtures of allergens and adjuvants (e.g., O<sub>3</sub>/NO<sub>x</sub>; SOA, DEP, PALMs, LPS, microbes, ... etc.) and identify mixtures/synergies leading to particularly strong effects/(immune) responses, including complement interactions and epigenetic effects (e.g., activation of C3 proteins; regulation of genes associated with Th2-polarization and epithelial barrier functions)
- Quantify the effects of chemical modification on the potency of allergens using atmospherically and physiologically relevant rather than artificial reactants and conditions (e.g., allergenic protein and bioparticle exposure to O<sub>3</sub>/NO<sub>x</sub> and ONOO<sup>-</sup> rather than TNM) and resolve potential interferences between different reaction products and pathways, (e.g., nitration vs. dimerization or oligomerization or degradation of protein macromolecules exposed to oxidizing and nitrating agents)
- Explore how chemical modifications of food components due to environmental pollution and related physiological processes may influence the development of chronic inflammation and hypersensitivities in the gastrointestinal tract (e.g., through nitration of amylase/trypsin inhibitors, ATIs)



*EPIDEMIOLOGICAL STUDIES (Q3 & Q4)*

- Establish and optimize (micro)monitoring networks and air quality models for the assessment and prediction of regional and local sources, concentration levels, and effective allergenic potential of allergens and adjuvants as well as the resulting personal exposure as input for refined epidemiological studies
  - Assess the relative importance and potential interrelations of air pollution, other forms of environmental pollution and other influencing factors/risk factors for allergies (e.g., potential effects of air pollution and fertilization and pesticides on the allergenic potential of foods)
  - Establish characteristic levels of allergen and air pollutant co-exposure (exposure-response relations) that constitute a substantial risk for the development of allergic diseases
-

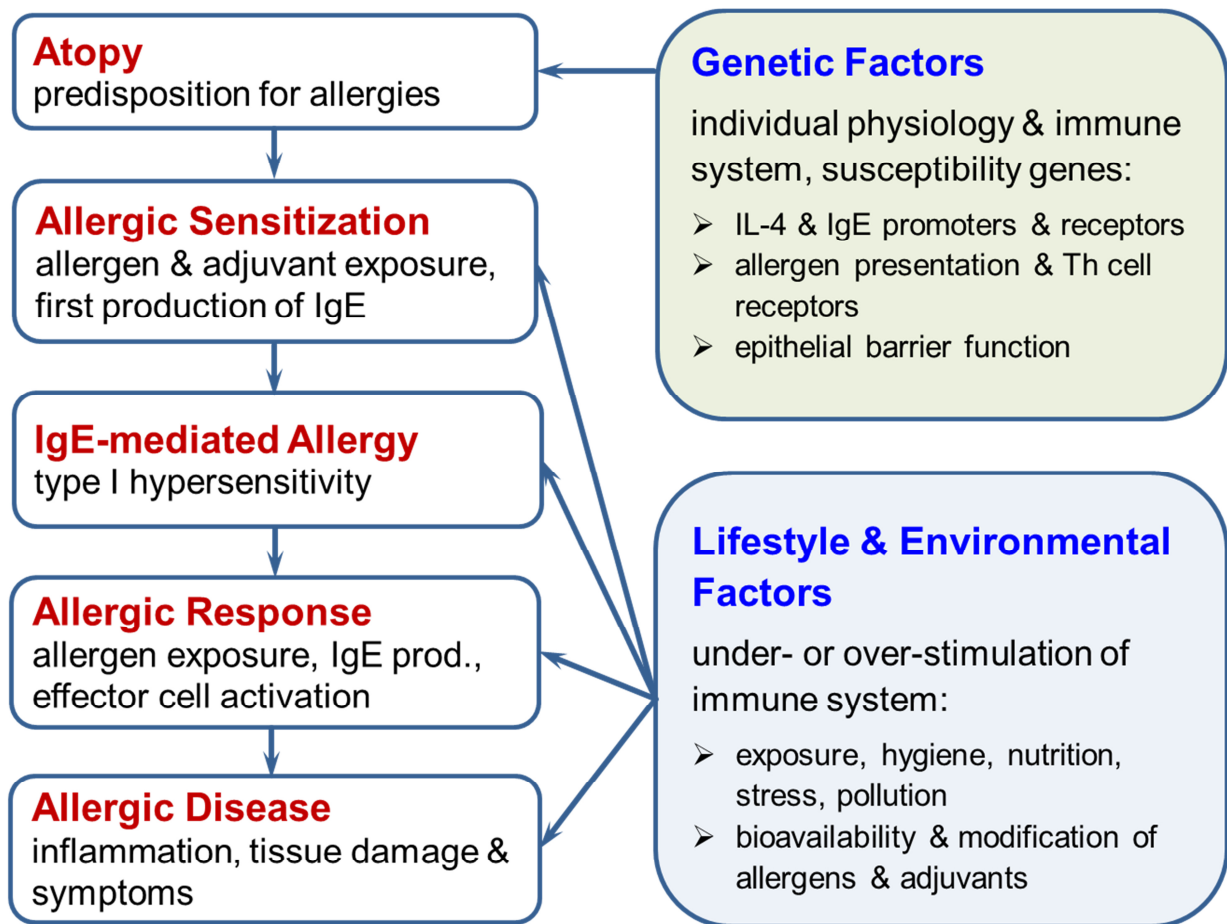


Figure S1. Essential steps and influencing factors in the development of IgE-mediated allergies (type I hypersensitivities) and related diseases.

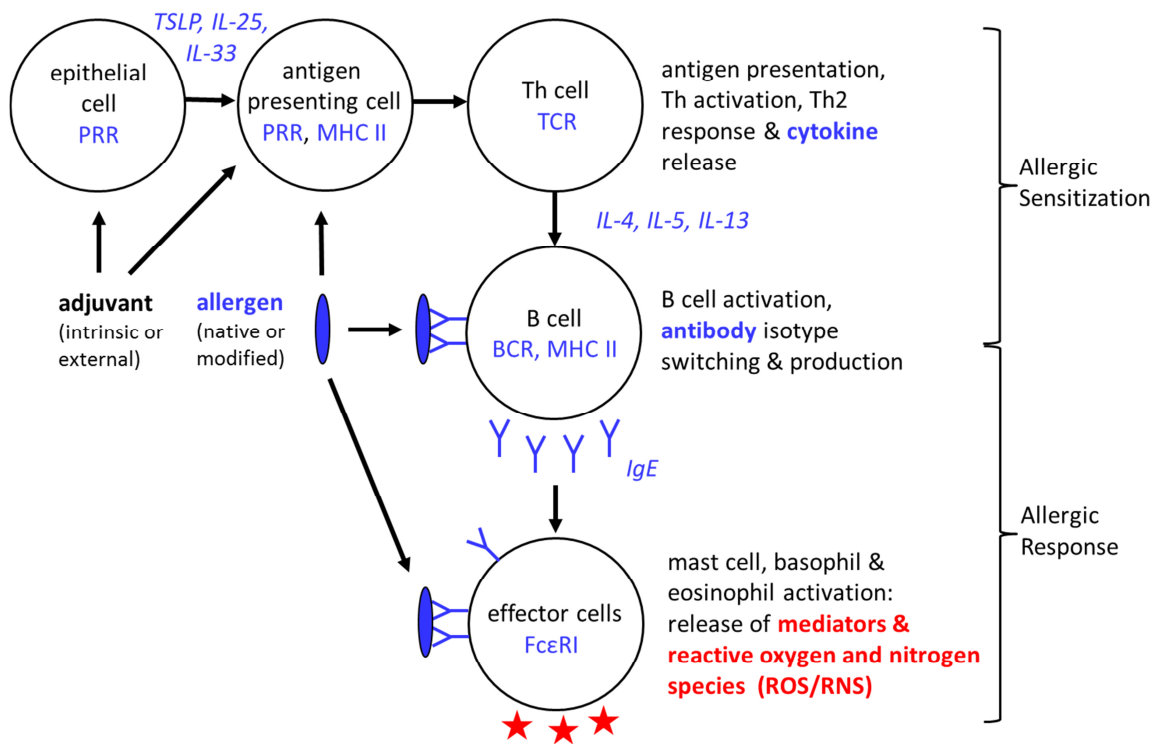


Figure S2.

Simplified scheme of major processes involved in allergic sensitization and response: molecular and cellular interactions of the innate and adaptive immune system with allergens and adjuvants (blue: proteins; red: pro-inflammatory mediators & ROS/RNS).

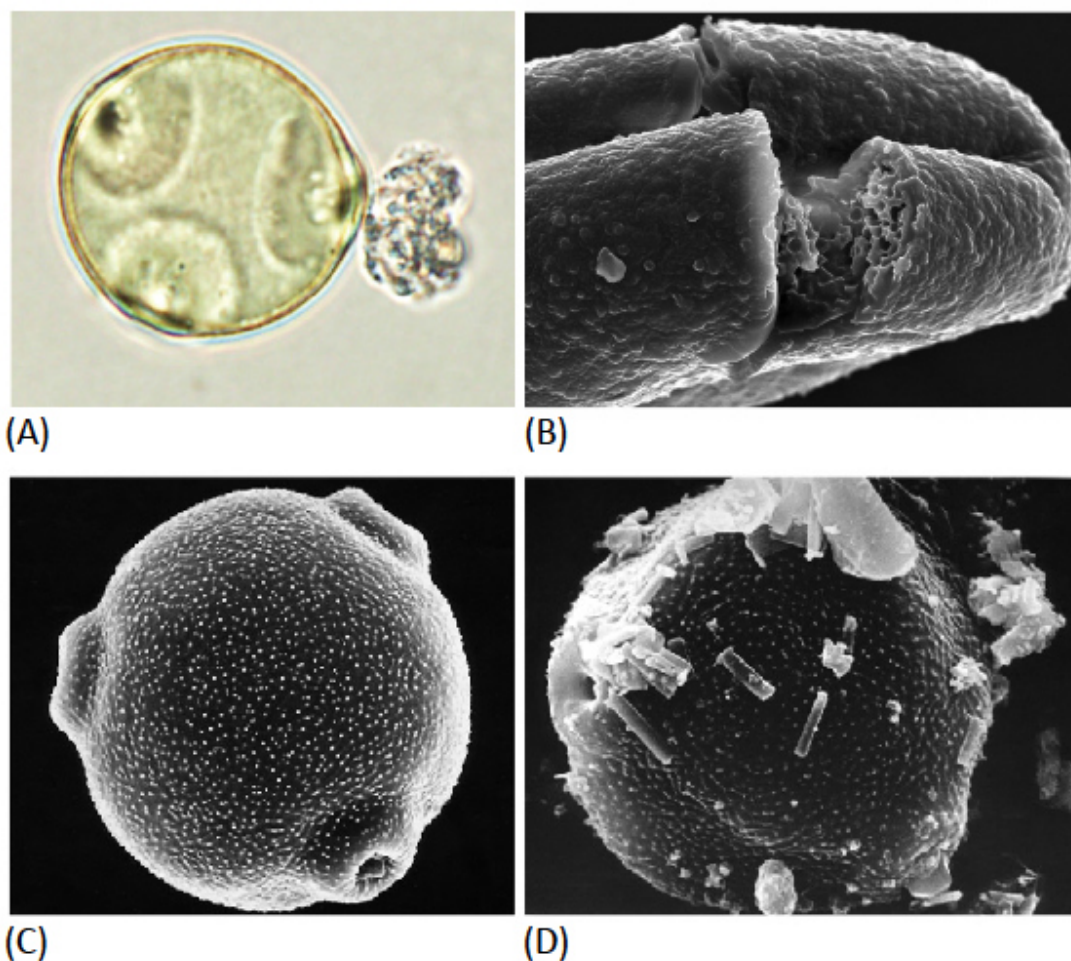


Figure S3.

Scanning electron micrographs of oak and birch pollen. (A) Release of pollen cytoplasmic granules (PCG) from oak pollen grain from pores after the contact with acid solution<sup>140</sup>; (B) oak pollen grains after exposure to 1ppm NO<sub>2</sub> (×4500 magnification)<sup>140</sup>; (C) birch pollen from a rural (C)<sup>141</sup> and (D) an urban collection site<sup>142</sup>. (A) and (B) are reproduced with permission from John Wiley and Sons<sup>140</sup>, (C) is reproduced with permission from Elsevier<sup>141</sup>, and (D) is reproduced with the permission from Hogrefe and Huber Publishers (now Hogrefe Publishing).<sup>142</sup>

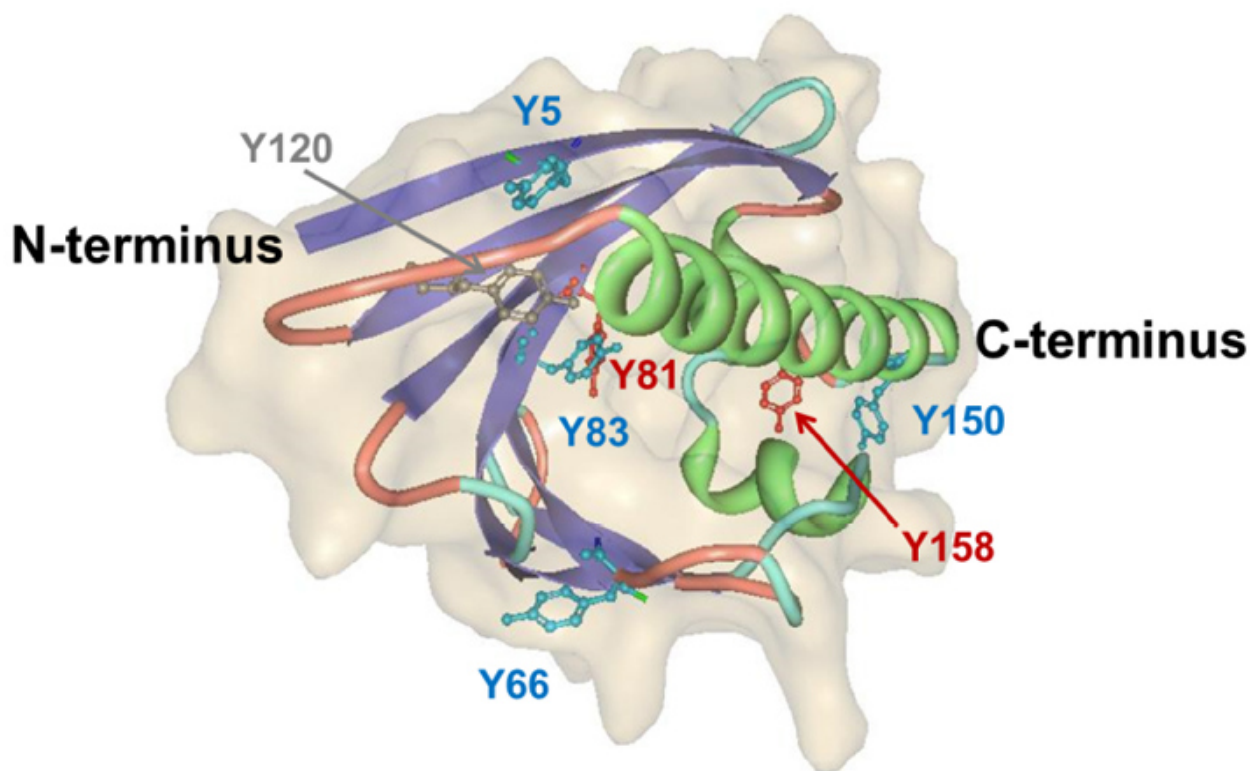


Figure S4.

3D-structure of the major birch pollen allergen Bet v 1.0101 highlighting the position of tyrosine residues in the amino acid sequence of the protein. Tyrosine residues mainly nitrated during exposure to environmentally relevant concentrations of  $O_3/NO_2$  are given in red. Reprinted from <sup>104</sup>. Copyright 2014 (American Chemical Society).

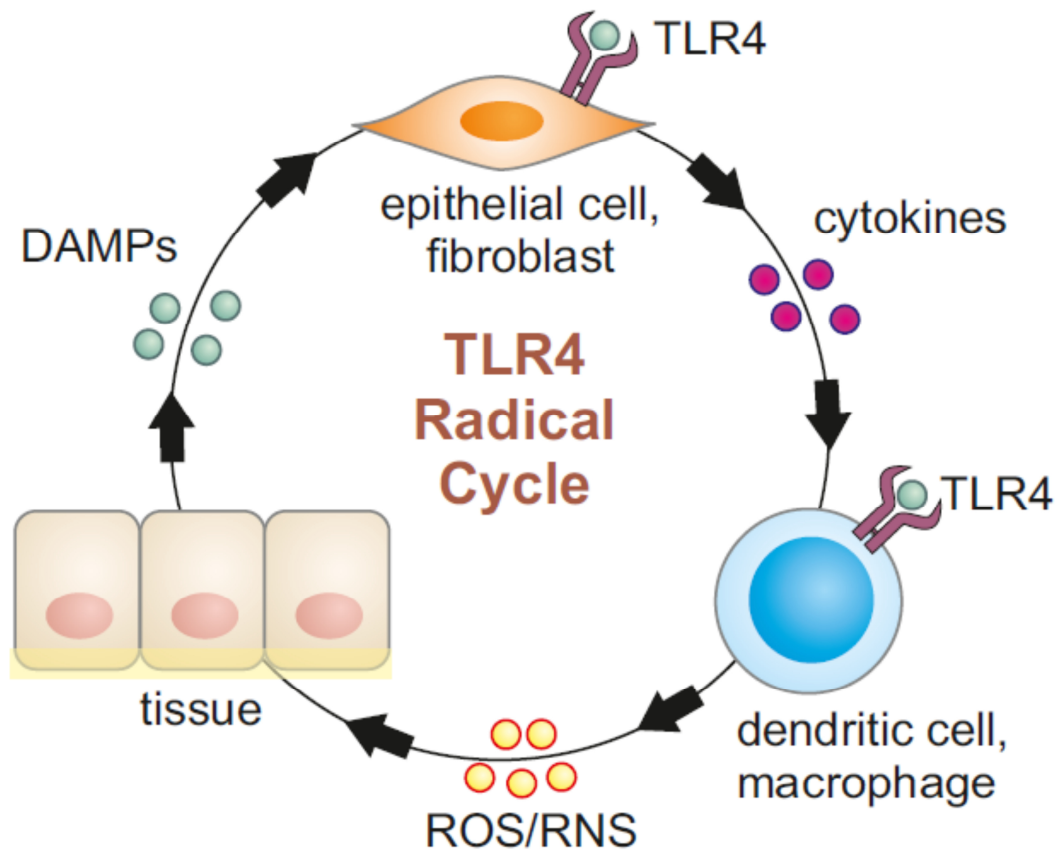


Figure S5.

TLR4-radical cycle of inflammation as an example of cyclic immunochemical processes that may be involved in the development of allergies and triggered by external influencing factors including air pollutants and related oxidative stress. Atmospheric and physiological reactive oxygen and nitrogen species (ROS/RNS) can cause the formation of damage-associated molecular patterns (DAMPs) that can activate the Toll-like receptor 4 (TLR4) and other pattern recognition receptors, leading to the release of more ROS/RNS and formation of more DAMPs.

## References:

1. Cookson, W., The immunogenetics of asthma and eczema: A new focus on the epithelium. **2004**, *4* (12), 978-988. DOI: 10.1038/nri1500
2. Adkinson Jr, N. F.; Bochner, B. S.; Burks, A. W.; Busse, W. W.; Holgate, S. T.; Lemanske Jr, R. F.; O'Hehir, R. E., *Middleton's Allergy Principles and Practice*. 8th ed.; Elsevier: 2014; Vol. 1 and 2, pp 1896.
3. Portelli, M. A.; Hodge, E.; Sayers, I., Genetic risk factors for the development of allergic disease identified by genome-wide association. *Clin. Exp. Allergy* **2015**, *45* (1), 21-31. DOI: 10.1111/cea.12327
4. Evans, H.; Mitre, E., Worms as therapeutic agents for allergy and asthma: Understanding why benefits in animal studies have not translated into clinical success. *J. Allergy Clin. Immunol.* **2015**, *135* (2), 343-353. DOI: 10.1016/j.jaci.2014.07.007
5. Ring, J.; Kramer, U.; Schafer, T.; Behrendt, H., Why are allergies increasing? *Curr. Opin. Immunol.* **2001**, *13* (6), 701-708. DOI: 10.1016/s0952-7915(01)00282-5
6. Heinrich, J.; Popescu, M. A.; Wjst, M.; Goldstein, I. F.; Wichmann, H. E., Atopy in children and parental social class. *Am. J. Public Health* **1998**, *88* (9), 1319-1324. DOI: 10.2105/ajph.88.9.1319
7. Larrick, J. W.; Buckley, C. E.; Machamer, C. E.; Schlagel, G. D.; Yost, J. A.; Blessingmoore, J.; Levy, D., Does hyperimmunoglobulinemia-E protect tropical populations from allergic disease? *J. Allergy Clin. Immunol.* **1983**, *71* (2), 184-188. DOI: 10.1016/0091-6749(83)90097-0
8. Olesen, A. B.; Juul, S.; Birkebaek, N.; Thestrup-Pedersen, K., Association between atopic dermatitis and insulin-dependent diabetes mellitus: a case-control study. *Lancet* **2001**, *357* (9270), 1749-1752. DOI: 10.1016/s0140-6736(00)04896-0
9. Coca, A. F.; Cooke, R. A., On the classification of the phenomena of hypersensitiveness. *J. Immunol.* **1923**, *8* (3), 163-182.
10. Holt, P. G.; Thomas, W. R., Sensitization to airborne environmental allergens: unresolved issues. *Nat. Immunol.* **2005**, *6* (10), 957-960. DOI: 10.1038/ni1005-957
11. D'Amato, G.; Holgate, S. T.; Pawankar, R., et al., Meteorological conditions, climate change, new emerging factors, and asthma and related allergic disorders. A statement of the World Allergy Organization. *World Allergy Organ. J.* **2015**, *8* (25). DOI: 10.1186/s40413-015-0073-0
12. Bégin, P.; Nadeau, K. C., Epigenetic regulation of asthma and allergic disease. *Allergy, Asthma, Clin. Immunol.* **2014**, *10* (1), 1-12. DOI: 10.1186/1710-1492-10-27
13. Ring, J.; Akdis, C.; Lauener, R., et al., Global Allergy Forum and Second Davos Declaration 2013 Allergy: Barriers to cure - challenges and actions to be taken. *Allergy* **2014**, *69* (8), 978-982. DOI: 10.1111/all.12406

14. Kramer, U.; Koch, T.; Ranft, U.; Ring, J.; Behrendt, H., Traffic-related air pollution is associated with atopy in children living in urban areas. *Epidemiology (Cambridge, Mass.)* **2000**, *11* (1), 64-70.
15. Martino, D. J.; Prescott, S. L., Progress in understanding the epigenetic basis for immune development, immune function, and the rising incidence of allergic disease. *Curr. Allergy Asthma Rep.* **2013**, *13* (1), 85-92. DOI: 10.1007/s11882-012-0312-1
16. Peden, D. B., Does air pollution really cause allergy? *Clin. Exp. Allergy* **2015**, *45* (1), 3-5. DOI: 10.1111/cea.12414
17. Miller, R. L.; Peden, D. B., Environmental Impacts on Immune Responses in Atopy and Asthma. *J. Allergy Clin. Immunol.* **2014**, *134* (5), 1001-1008. DOI: 10.1016/j.jaci.2014.07.064
18. Gaffin, J. M.; Kanchongkittiphon, W.; Phipatanakul, W., Perinatal and early childhood environmental factors influencing allergic asthma immunopathogenesis. *Int. Immunopharmacol.* **2014**, *22* (1), 21-30. DOI: 10.1016/j.intimp.2014.06.005
19. Wahn, U., What drives the allergic march? *Allergy* **2000**, *55* (7), 591-599.
20. Krämer, U.; Behrendt, H.; Dolgner, R.; Ranft, U.; Ring, J.; Willer, H.; Schlipkoter, H. W., Airway diseases and allergies in East and West German children during the first 5 years after reunification: time trends and the impact of sulphur dioxide and total suspended particles. *Int. J. Epidemiol.* **1999**, *28* (5), 865-873.
21. Castro-Rodriguez, J. A.; Forno, E.; Rodriguez-Martinez, C. E.; Celedon, J. C., Risk and Protective Factors for Childhood Asthma: What Is the Evidence? *J. Allergy Clin. Immunol.-Pract.* **2016**, *4* (6), 1111-1122. DOI: 10.1016/j.jaip.2016.05.003
22. Bernstein, J. A.; Alexis, N.; Barnes, C., et al., Health effects of air pollution. *J. Allergy Clin. Immunol.* **2004**, *114* (5), 1116-1123. DOI: 10.1016/j.jaci.2004.08.030
23. Valenta, R.; Hochwallner, H.; Linhart, B.; Pahr, S., Food Allergies: The Basics. *Gastroenterology* **2015**, *148* (6), 1120-1131. DOI: 10.1053/j.gastro.2015.02.006
24. Papazian, D.; Hansen, S.; Wurtzen, P. A., Airway responses towards allergens - from the airway epithelium to T cells. *Clin. Exp. Allergy* **2015**, *45* (8), 1268-1287. DOI: 10.1111/cea.12451
25. Galli, S. J.; Tsai, M.; Piliponsky, A. M., The development of allergic inflammation. *Nature* **2008**, *454* (7203), 445-454.
26. Lambrecht, B. N.; Hammad, H., Allergens and the airway epithelium response: Gateway to allergic sensitization. *J. Allergy Clin. Immunol.* **2014**, *134* (3), 499-507. DOI: 10.1016/j.jaci.2014.06.036
27. Lambrecht, B. N.; Hammad, H., The airway epithelium in asthma. *Nat. Med.* **2012**, *18* (5), 684-692. DOI: 10.1038/nm.2737
28. Mahlaköiv, T.; Artis, D., Allergen Exposure: When Timing Is Everything. **2016**, *45* (6), 1188-1190. DOI: 10.1016/j.immuni.2016.12.007



29. Berin, M. C.; Sampson, H. A., Mucosal immunology of food allergy. *Curr. Biol.* **2013**, *23* (9), 389-400. DOI: 10.1016/j.cub.2013.02.043
30. Holmgren, J.; Czerkinsky, C., Mucosal immunity and vaccines. *Nature medicine* **2005**, *11* (4 Suppl), S45-53. DOI: 10.1038/nm1213
31. Neurath, M. F.; Finotto, S.; Glimcher, L. H., The role of Th1/Th2 polarization in mucosal immunity. *Nat. Med.* **2002**, *8* (6), 567-573. DOI: 10.1038/nm0602-567
32. Divekar, R.; Kita, H., Recent advances in epithelium-derived cytokines (IL-33, IL-25 and TSLP) and allergic inflammation. **2015**, *15* (1), 98-103. DOI: 10.1097/ACI.0000000000000133
33. Lombardi, V.; Singh, A. K.; Akbari, O., The Role of Costimulatory Molecules in Allergic Disease and Asthma. *Int. Arch. Allergy Immunol.* **2010**, *151* (3), 179-189. DOI: 10.1159/000242355
34. Tangye, S. G.; Ma, C. S.; Brink, R.; Deenick, E. K., The good, the bad and the ugly - T-FH cells in human health and disease. *Nat. Rev. Immunol.* **2013**, *13* (6), 412-426. DOI: 10.1038/nri3447
35. Cyster, J. G., B cell follicles and antigen encounters of the third kind. **2010**, *11* (11), 989-996.
36. Heesters, B. A.; Myers, R. C.; Carroll, M. C., Follicular dendritic cells: dynamic antigen libraries. **2014**, *14* (7), 495-504.
37. Recaladin, T.; Fear, D., Transcription factors regulating B cell fate in the germinal centre. **2016**, *183* (1), 65-75.
38. Wills-Karp, M.; Koehl, J., New insights into the role of the complement pathway in allergy and asthma. *Curr. Allergy Asthma Rep.* **2005**, *5* (5), 362-369.
39. Köhl, J.; Wills-Karp, M., A dual role for complement in allergic asthma. **2007**, *7* (3), 283-289.
40. Zhang, X.; Köhl, J., A complex role for complement in allergic asthma. *Expert Rev. Clin. Immunol.* **2010**, *6* (2), 269-277.
41. Carroll, M. C.; Isenman, D. E., Regulation of humoral immunity by complement. **2012**, *37* (2), 199-207.
42. Schmutte, I.; Laumonnier, Y.; Köhl, J. In *Anaphylatoxins coordinate innate and adaptive immune responses in allergic asthma*, Seminars in immunology, Elsevier: 2013; pp 2-11.
43. Pandya, P. H.; Wilkes, D. S., Complement system in lung disease. *Am. J. Respir. Cell Mol. Biol.* **2014**, *51* (4), 467-473.
44. Khan, M. A.; Assiri, A. M.; Broering, D. C., Complement mediators: key regulators of airway tissue remodeling in asthma. *J. Transl. Med.* **2015**, *13* (1), 1-9.
45. Akdis, C. A., Therapies for allergic inflammation: refining strategies to induce tolerance. *Nat. Med.* **2012**, *18* (5), 736-749. DOI: 10.1038/nm.2754
46. Pellerin, L.; Jenks, J. A.; Begin, P.; Bacchetta, R.; Nadeau, K. C., Regulatory T cells and their roles in immune dysregulation and allergy. *Immunol. Res.* **2014**, *58* (2-3), 358-368. DOI: 10.1007/s12026-014-8512-5

47. Sompayrac, L., *How the Immune System Works*. 3rd ed.; Blackwell Publishing: 2008.
48. Posner, R. G.; Savage, P. B.; Peters, A. S.; Macias, A.; DelGado, J.; Zwartz, G.; Sklar, L. A.; Hlavacek, W. S., A quantitative approach for studying IgE–FcεRI aggregation. *Mol. Immunol.* **2002**, *38* (16–18), 1221-1228. DOI: 10.1016/S0161-5890(02)00067-6
49. Garman, S. C.; Wurzburg, B. A.; Tarchevskaya, S. S.; Kinet, J. P.; Jardetzky, T. S., Structure of the Fc fragment of human IgE bound to its high-affinity receptor Fc epsilon RI alpha. *Nature* **2000**, *406* (6793), 259-266.
50. Huby, R. D. J.; Dearman, R. J.; Kimber, I., Why are some proteins allergens? *Toxicol. Sci.* **2000**, *55* (2), 235-246. DOI: 10.1093/toxsci/55.2.235
51. Greiner, A. N.; Hellings, P. W.; Rotiroti, G.; Scadding, G. K., Allergic rhinitis. *Lancet* **2011**, *378* (9809), 2112-2122. DOI: 10.1016/S0140-6736(11)60130-X
52. Skoner, D. R., Allergic rhinitis: Definition, epidemiology, detection, and pathophysiology, diagnosis. *J. Allergy Clin. Immunol.* **2001**, *108* (1), S2-S8. DOI: 10.1067/mai.2001.115569
53. Tyagi, N.; Farnell, E. J.; Fitzsimmons, C. M.; Ryan, S.; Tukahebwa, E.; Maizels, R. M.; Dunne, D. W.; Thornton, J. M.; Furnham, N., Comparisons of Allergenic and Metazoan Parasite Proteins: Allergy the Price of Immunity. **2015**, *11* (10), e1004546. DOI: 10.1371/journal.pcbi.1004546
54. Palm, N. W.; Rosenstein, R. K.; Medzhitov, R., Allergic host defences. *Nature* **2012**, *484* (7395), 465-472. DOI: 10.1038/nature11047
55. Chinen, J.; Fleisher, T. A.; Shearer, W. T., 2 - Adaptive Immunity A2 - Adkinson, N. Franklin. In *Middleton's Allergy (Eighth Edition)*, Bochner, B. S.; Burks, A. W.; Busse, W. W.; Holgate, S. T.; Lemanske, R. F.; O'Hehir, R. E., Eds. Content Repository Only!: London, 2014; 20-29.
56. Smith, P. K.; Masilamani, M.; Li, X.-M.; Sampson, H. A., The false alarm hypothesis: Food allergy is associated with high dietary advanced glycation end-products and proglycating dietary sugars that mimic alarmins. **2017**, *139* (2), 429-437. DOI: 10.1016/j.jaci.2016.05.040
57. Knutsen, A. P.; Bush, R. K.; Demain, J. G., et al., Fungi and allergic lower respiratory tract diseases. *J. Allergy Clin. Immunol.* **2012**, *129* (2), 280-291. DOI: 10.1016/j.jaci.2011.12.970
58. Runswick, S.; Mitchell, T.; Davies, P.; Robinson, C.; Garrod, D. R., Pollen proteolytic enzymes degrade tight junctions. *Respirology* **2007**, *12* (6), 834-842. DOI: 10.1111/j.1440-1843.2007.01175.x
59. Reed, C. E.; Kita, H., The role of protease activation of inflammation in allergic respiratory diseases. *J. Allergy Clin. Immunol.* **2004**, *114* (5), 997-1008. DOI: 10.1016/j.jaci.2004.07.060
60. Millien, V. O.; Lu, W.; Shaw, J., et al., Cleavage of fibrinogen by proteinases elicits allergic responses through Toll-like receptor 4. *Science* **2013**, *341* (6147), 792-796. DOI: 10.1126/science.1240342

61. Gilles, S.; Mariani, V.; Bryce, M.; Mueller, M. J.; Ring, J.; Jakob, T.; Pastore, S.; Behrendt, H.; Traidl-Hoffmann, C., Pollen-derived E1-phytoprostanes signal via PPAR-gamma and NF-kappaB-dependent mechanisms. *J. Immunol.* **2009**, *182* (11), 6653-8. DOI: 10.4049/jimmunol.0802613
62. Deifl, S.; Bohle, B., Factors influencing the allergenicity and adjuvanticity of allergens. *Immunotherapy* **2011**, *3* (7), 881-893. DOI: 10.2217/imt.11.69
63. Boldogh, I.; Bacsi, A.; Choudhury, B. K.; Dharajiya, N.; Alam, R.; Hazra, T. K.; Mitra, S.; Goldblum, R. M.; Sur, S., ROS generated by pollen NADPH oxidase provide a signal that augments antigen-induced allergic airway inflammation. *J. Clin. Invest.* **2005**, *115* (8), 2169-2179. DOI: 10.1172/JCI24422
64. Wimmer, M.; Alessandrini, F.; Gilles, S., et al., Pollen-derived adenosine is a necessary cofactor for ragweed allergy. *Allergy* **2015**, *70* (8), 944-954. DOI: 10.1111/all.12642
65. Seutter von Loetzen, C.; Hoffmann, T.; Hartl, M. J.; Schweimer, K.; Schwab, W.; Rosch, P.; Hartl-Spiegelhauer, O., Secret of the major birch pollen allergen Bet v 1: identification of the physiological ligand. *Biochem. J.* **2014**, *457* (3), 379-390. DOI: 10.1042/bj20130413
66. Berrens, L.; Lopez, B. D., Complement activating agents in allergenic extracts. *Inflamm. Res.* **1997**, *46* (11), 455-460. DOI: 10.1007/s000110050224
67. Eisenbarth, S. C.; Piggott, D. A.; Huleatt, J. W.; Visintin, I.; Herrick, C. A.; Bottomly, K., Lipopolysaccharide-enhanced, toll-like receptor 4-dependent T helper cell type 2 responses to inhaled antigen. *J. Exp. Med.* **2002**, *196* (12), 1645-1651. DOI: 10.1084/jem.20021340
68. Heydenreich, B.; Bellinghausen, I.; Koenig, B.; Becker, W. M.; Grabbe, S.; Petersen, A.; Saloga, J., Gram-positive bacteria on grass pollen exhibit adjuvant activity inducing inflammatory T cell responses. *Clin. Exp. Allergy* **2012**, *42* (1), 76-84. DOI: 10.1111/j.1365-2222.2011.03888.x
69. Inamdar, A. A.; Bennett, J. W., A common fungal volatile organic compound induces a nitric oxide mediated inflammatory response in *Drosophila melanogaster*. *Sci. Rep.* **2014**, *4*, 3833. DOI: 10.1038/srep03833
70. Provoost, S.; Maes, T.; Joos, G. F.; Tournoy, K. G., Monocyte-derived dendritic cell recruitment and allergic T(H)2 responses after exposure to diesel particles are CCR2 dependent. *J. Allergy Clin. Immunol.* **2012**, *129* (2), 483-91. DOI: 10.1016/j.jaci.2011.07.051
71. Maes, T.; Provoost, S.; Lanckacker, E. A.; Cataldo, D. D.; Vanoirbeek, J. A.; Nemery, B.; Tournoy, K. G.; Joos, G. F., Mouse models to unravel the role of inhaled pollutants on allergic sensitization and airway inflammation. *Respir. Res.* **2010**, *11*, 7. DOI: 10.1186/1465-9921-11-7
72. Saxon, A.; Diaz-Sanchez, D., Air pollution and allergy: you are what you breathe. *Nat. Immunol.* **2005**, *6* (3), 223-226. DOI: 10.1038/ni0305-223
73. Riedl, M. A.; Landaw, E. M.; Saxon, A.; Diaz-Sanchez, D., Initial high-dose nasal allergen exposure prevents allergic sensitization to a neoantigen. *J. Immunol.* **2005**, *174* (11), 7440-7445.

74. Diaz-Sanchez, D.; Garcia, M. P.; Wang, M.; Jyrala, M.; Saxon, A., Nasal challenge with diesel exhaust particles can induce sensitization to a neoallergen in the human mucosa. *J. Allergy Clin. Immunol.* **1999**, *104* (6), 1183-1188.
75. Devouassoux, G.; Saxon, A.; Metcalfe, D. D.; Prussin, C.; Colomb, M. G.; Brambilla, C.; Diaz-Sanchez, D., Chemical constituents of diesel exhaust particles induce IL-4 production and histamine release by human basophils. *J. Allergy Clin. Immunol.* *109* (5), 847-853. DOI: 10.1067/mai.2002.122843
76. Pandya, R. J.; Solomon, G.; Kinner, A.; Balmes, J. R., Diesel exhaust and asthma: Hypotheses and molecular mechanisms of action. *Environ. Health Perspect.* **2002**, *110*, 103-112.
77. Dick, C. A.; Brown, D. M.; Donaldson, K.; Stone, V., The role of free radicals in the toxic and inflammatory effects of four different ultrafine particle types. *Inhal. Toxicol.* **2003**, *15* (1), 39-52. DOI: 10.1080/089583703044454
78. Hiura, T. S.; Li, N.; Kaplan, R.; Horwitz, M.; Seagrave, J. C.; Nel, A. E., The role of a mitochondrial pathway in the induction of apoptosis by chemicals extracted from diesel exhaust particles. *J. Immunol.* **2000**, *165* (5), 2703-2711.
79. Hiura, T. S.; Kaszubowski, M. P.; Li, N.; Nel, A. E., Chemicals in diesel exhaust particles generate reactive oxygen radicals and induce apoptosis in macrophages. *J. Immunol.* **1999**, *163* (10), 5582-5591.
80. Siegel, P. D.; Saxena, R. K.; Saxena, Q. B.; Ma, J. K. H.; Ma, J. Y. C.; Yin, X. J.; Castranova, V.; Al-Humadi, N.; Lewis, D. M., Effect of diesel exhaust particulate (DEP) on immune responses: Contributions of particulate versus organic soluble components. *J. Toxicol. Environ. Health, Part A* **2004**, *67* (3), 221-231. DOI: 10.1080/15287390490266891
81. Yang, H. M.; Antonini, J. M.; Barger, M. W.; Butterworth, L.; Roberts, J. R.; Ma, J. K. H.; Castranova, V.; Ma, J. Y. C., Diesel exhaust particles suppress macrophage function and slow the pulmonary clearance of *Listeria monocytogenes* in rats. *Environ. Health Perspect.* **2001**, *109* (5), 515-521. DOI: 10.2307/3454711
82. Bleck, B.; Tse, D. B.; Gordon, T.; Ahsan, M. R.; Reibman, J., Diesel Exhaust Particle-Treated Human Bronchial Epithelial Cells Upregulate Jagged-1 and OX40 Ligand in Myeloid Dendritic Cells via Thymic Stromal Lymphopoietin. *J. Immunol.* **2010**, *185* (11), 6636-6645. DOI: 10.4049/jimmunol.1000719
83. Fukuoka, A.; Matsushita, K.; Morikawa, T.; Takano, H.; Yoshimoto, T., Diesel exhaust particles exacerbate allergic rhinitis in mice by disrupting the nasal epithelial barrier. *Clin. Exp. Allergy* **2016**, *46* (1), 142-152. DOI: 10.1111/cea.12597
84. Bayram, H.; Devalia, J. L.; Sapsford, R. J.; Ohtoshi, T.; Miyabara, Y.; Sagai, M.; Davies, R. J., The effect of diesel exhaust particles on cell function and release of inflammatory mediators from human bronchial epithelial cells in vitro. *Am. J. Respir. Cell Mol. Biol.* **1998**, *18* (3), 441-448.

85. Li, N.; Buglak, N., Convergence of air pollutant-induced redox-sensitive signals in the dendritic cells contributes to asthma pathogenesis. *Toxicol. Lett.* **2015**, *237* (1), 55-60. DOI: 10.1016/j.toxlet.2015.05.017
86. Kang, X. D.; Li, N.; Wang, M. Y.; Boontheung, P.; Sioutas, C.; Harkema, J. R.; Bramble, L. A.; Nel, A. E.; Loo, J. A., Adjuvant effects of ambient particulate matter monitored by proteomics of bronchoalveolar lavage fluid. *Proteomics* **2010**, *10* (3), 520-531. DOI: 10.1002/pmic.200900573
87. Xiao, G. G.; Nel, A. E.; Loo, J. A., Nitrotyrosine-modified proteins and oxidative stress induced by diesel exhaust particles. *Electrophoresis* **2005**, *26* (1), 280-292. DOI: 10.1002/elps.200406145
88. Walters, D. M.; Breyse, P. N.; Schofield, B.; Wills-Karp, M., Complement factor 3 mediates particulate matter-induced airway hyperresponsiveness. *Am. J. Respir. Cell Mol. Biol.* **2002**, *27* (4), 413-418.
89. Kanemitsu, H.; Nagasawa, S.; Sagai, M.; MORI, Y., Complement activation by diesel exhaust particles (DEP). *Biol. Pharm. Bull.* **1998**, *21* (2), 129-132.
90. Tezza, G.; Mazzei, F.; Boner, A., Epigenetics of allergy. *Early Hum. Dev.* **2013**, *89*, Supplement 1, S20-S21. DOI: 10.1016/S0378-3782(13)70007-0
91. Sofer, T.; Baccarelli, A.; Cantone, L.; Coull, B.; Maity, A.; Lin, X.; Schwartz, J., Exposure to airborne particulate matter is associated with methylation pattern in the asthma pathway. *Epigenomics* **2013**, *5* (2), 147-154. DOI: 10.2217/epi.13.16
92. Liu, J.; Ballaney, M.; Al-alem, U.; Quan, C.; Jin, X.; Perera, F.; Chen, L. C.; Miller, R. L., Combined inhaled diesel exhaust particles and allergen exposure alter methylation of T helper genes and IgE production in vivo. *Toxicol. Sci.* **2008**, *102* (1), 76-81. DOI: 10.1093/toxsci/kfm290
93. Zuo, L.; Lucas, K.; Fortuna, C. A.; Chuang, C. C.; Best, T. M., Molecular Regulation of Toll-like Receptors in Asthma and COPD. *Front. Physiol.* **2015**, *9* (6), 312. DOI: 10.3389/fphys.2015.00312
94. Pöschl, U.; Shiraiwa, M., Multiphase Chemistry at the Atmosphere-Biosphere Interface Influencing Climate and Public Health in the Anthropocene. *Chem. Rev.* **2015**, *115* (10), 4440-4475. DOI: 10.1021/cr500487s
95. Ni, L.; Chuang, C.-C.; Zuo, L., Fine particulate matter in acute exacerbation of COPD. *Front. Physiol.* **2015**, *6* (294). DOI: 10.3389/fphys.2015.00294
96. Jiang, L.; Diaz, P. T.; Best, T. M.; Stimpfl, J. N.; He, F.; Zuo, L., Molecular characterization of redox mechanisms in allergic asthma. *Ann. Allergy Asthma Immunol.* **2014**, *113* (2), 137-142. DOI: 10.1016/j.anai.2014.05.030
97. Charrier, J. G.; McFall, A. S.; Richards-Henderson, N. K.; Anastasio, C., Hydrogen Peroxide Formation in a Surrogate Lung Fluid by Transition Metals and Quinones Present in Particulate Matter. *Environ. Sci. Technol.* **2014**, *48* (12), 7010-7017. DOI: 10.1021/es501011w

98. Zuo, L.; Otenbaker, N. P.; Rose, B. A.; Salisbury, K. S., Molecular mechanisms of reactive oxygen species-related pulmonary inflammation and asthma. *Mol. Immunol.* **2013**, *56* (1-2), 57-63. DOI: 10.1016/j.molimm.2013.04.002
99. Kampf, C. J.; Liu, F.; Reinmuth-Selzle, K.; Berkemeier, T.; Meusel, H.; Shiraiwa, M.; Pöschl, U., Protein Cross-Linking and Oligomerization through Dityrosine Formation upon Exposure to Ozone. *Environ. Sci. Technol.* **2015**, *49* (18), 10859-10866. DOI: 10.1021/acs.est.5b02902
100. Shiraiwa, M.; Selzle, K.; Yang, H.; Sosedova, Y.; Ammann, M.; Pöschl, U., Multiphase Chemical Kinetics of the Nitration of Aerosolized Protein by Ozone and Nitrogen Dioxide. *Environ. Sci. Technol.* **2012**, *46* (12), 6672-6680. DOI: 10.1021/es300871b
101. Shiraiwa, M.; Sosedova, Y.; Rouviere, A.; Yang, H.; Zhang, Y. Y.; Abbatt, J. P. D.; Ammann, M.; Pöschl, U., The role of long-lived reactive oxygen intermediates in the reaction of ozone with aerosol particles. *Nat. Chem.* **2011**, *3* (4), 291-295. DOI: 10.1038/nchem.988
102. Bayram, H.; Rusznak, C.; Khair, O. A.; Sapsford, R. J.; Abdelaziz, M. M., Effect of ozone and nitrogen dioxide on the permeability of bronchial epithelial cell cultures of non-asthmatic and asthmatic subjects. *Clin. Exp. Allergy* **2002**, *32* (9), 1285-1292.
103. Park, J.-W.; Taube, C.; Joetham, A., et al., Complement activation is critical to airway hyperresponsiveness after acute ozone exposure. *Am. J. Respir. Crit. Care Med.* **2004**, *169* (6), 726-732.
104. Reinmuth-Selzle, K.; Ackaert, C.; Kampf, C. J., et al., Nitration of the Birch Pollen Allergen Bet v 1.0101: Efficiency and Site-Selectivity of Liquid and Gaseous Nitrating Agents. *J. Proteome Res.* **2014**, *13* (3), 1570-1577. DOI: 10.1021/pr401078h
105. Ezratty, V.; Guillosoy, G.; Neukirch, C., et al., Repeated nitrogen dioxide exposures and eosinophilic airway inflammation in asthmatics: a randomized crossover study. *Environ. Health Perspect.* **2014**, *122* (8), 850-855. DOI: 10.1289/ehp.1307240
106. Bevelander, M.; Mayette, J.; Whittaker, L. A., et al., Nitrogen dioxide promotes allergic sensitization to inhaled antigen. *J. Immunol.* **2007**, *179* (6), 3680-3688. DOI:
107. Franze, T.; Weller, M. G.; Niessner, R.; Pöschl, U., Protein nitration by polluted air. *Environ. Sci. Technol.* **2005**, *39* (6), 1673-1678. DOI: 10.1021/es0488737
108. Scheurer, S.; Toda, M.; Vieths, S., What makes an allergen? *Clin. Exp. Allergy* **2015**, *45*, 1150-1161. DOI: 10.1111/cea.12571
109. Thomas, W. R., Innate affairs of allergens. *Clin. Exp. Allergy* **2013**, *43* (2), 152-163. DOI: 10.1111/j.1365-2222.2012.04059.x
110. Iwasaki, A.; Medzhitov, R., Regulation of Adaptive Immunity by the Innate Immune System. *Science* **2010**, *327* (5963), 291-295. DOI: 10.1126/science.1183021
111. Gómez-Casado, C.; Díaz-Perales, A., Allergen-Associated Immunomodulators: Modifying Allergy Outcome. *Arch. Immunol. Ther. Exp.* **2016**, *64* (5), 1-9. DOI: 10.1007/s00005-016-0401-2

112. Thomas, W. R., Allergen Ligands in the Initiation of Allergic Sensitization. *Curr. Allergy Asthma Rep.* **2014**, *14* (5), 10. DOI: 10.1007/s11882-014-0432-x
113. Bianchi, M. E., DAMPs, PAMPs and alarmins: all we need to know about danger. *J. Leukocyte Biol.* **2007**, *81* (1), 1-5. DOI: 10.1189/jlb.0306164
114. Trompette, A.; Divanovic, S.; Visintin, A., et al., Allergenicity resulting from functional mimicry of a Toll-like receptor complex protein. *Nature* **2009**, *457* (7229), 585-588. DOI: 10.1038/nature07548
115. Lucas, K.; Maes, M., Role of the Toll Like receptor (TLR) radical cycle in chronic inflammation: possible treatments targeting the TLR4 pathway. *Mol. Neurobiol.* **2013**, *48* (1), 190-204. DOI: 10.1007/s12035-013-8425-7
116. Karp, C. L., Guilt by intimate association: What makes an allergen an allergen? *J. Allergy Clin. Immunol.* **2010**, *125* (5), 955-960. DOI: 10.1016/j.jaci.2010.03.002
117. Holgate, S. T., Innate and adaptive immune responses in asthma. *Nat. Med.* **2012**, *18* (5), 673-683. DOI: 10.1038/nm.2731
118. Lambrecht, B. N.; Hammad, H., The immunology of asthma. *Nat. Immunol.* **2015**, *16* (1), 45-56. DOI: 10.1038/ni.3049
119. Bernink, J. H.; Germar, K.; Spits, H., The role of ILC2 in pathology of type 2 inflammatory diseases. *Curr. Opin. Immunol.* **2014**, *31*, 115-120. DOI: 10.1016/j.coi.2014.10.007
120. Ho, J.; Bailey, M.; Zaunders, J.; Mrad, N.; Sacks, R.; Sewell, W.; Harvey, R. J., Group 2 innate lymphoid cells (ILC2s) are increased in chronic rhinosinusitis with nasal polyps or eosinophilia. *Clin. Exp. Allergy* **2015**, *45* (2), 394-403. DOI: 10.1111/cea.12462
121. Scanlon, S. T.; McKenzie, A. N., The messenger between worlds: the regulation of innate and adaptive type-2 immunity by innate lymphoid cells. *Clin. Exp. Allergy* **2015**, *45* (1), 9-20. DOI: 10.1111/cea.12464
122. Salimi, M.; Barlow, J. L.; Saunders, S. P., et al., A role for IL-25 and IL-33-driven type-2 innate lymphoid cells in atopic dermatitis. *J. Exp. Med.* **2013**, *210* (13), 2939-2950. DOI: 10.1084/jem.20130351
123. Bartemes, K. R.; Kephart, G. M.; Fox, S. J.; Kita, H., Enhanced innate type 2 immune response in peripheral blood from patients with asthma. *J. Allergy Clin. Immunol.* **2014**, *134* (3), 671-678.e4. DOI: 10.1016/j.jaci.2014.06.024
124. Ghiani, A.; Aina, R.; Asero, R.; Bellotto, E.; Citterio, S., Ragweed pollen collected along high-traffic roads shows a higher allergenicity than pollen sampled in vegetated areas. *Allergy* **2012**, *67* (7), 887-894. DOI: 10.1111/j.1398-9995.2012.02846.x
125. Zhao, F.; Elkelish, A.; Durner, J., et al., Common ragweed (*Ambrosia artemisiifolia* L.): allergenicity and molecular characterization of pollen after plant exposure to elevated NO<sub>2</sub>. *Plant, Cell Environ.* **2016**, *39* (1), 147-164. DOI: 10.1111/pce.12601

126. Pasqualini, S.; Tedeschini, E.; Frenguelli, G.; Wopfner, N.; Ferreira, F.; D'Amato, G.; Ederli, L., Ozone affects pollen viability and NAD(P)H oxidase release from *Ambrosia artemisiifolia* pollen. *Environ. Pollut.* **2011**, *159* (10), 2823-2830. DOI: 10.1016/j.envpol.2011.05.003
127. Kanter, U.; Heller, W.; Durner, J., et al., Molecular and Immunological Characterization of Ragweed (*Ambrosia artemisiifolia* L.) Pollen after Exposure of the Plants to Elevated Ozone over a Whole Growing Season. *PLoS One* **2013**, *8* (4), 12. DOI: 10.1371/journal.pone.0061518
128. Majd, A.; Chehregani, A.; Moin, M.; Gholami, M.; Kohno, S.; Nabe, T.; Shariatzade, M. A., The effects of air pollution on structures, proteins and allergenicity of pollen grains. *Aerobiologia* **2004**, *20* (2), 111-118. DOI: 10.1023/b:aero.0000032950.12169.38
129. Cortegano, I.; Civantos, E.; Aceituno, E.; del Moral, A.; Lopez, E.; Lombardero, M.; del Pozo, V.; Lahoz, C., Cloning and expression of a major allergen from *Cupressus arizonica* pollen, Cup a 3, a PR-5 protein expressed under polluted environment. *Allergy* **2004**, *59* (5), 485-490. DOI: 10.1046/j.1398-9995.2003.00363.x
130. Suarez-Cervera, M.; Castells, T.; Vega-Maray, A., et al., Effects of air pollution on Cup a 3 allergen in *Cupressus arizonica* pollen grains. *Ann. Allergy Asthma Immunol.* **2008**, *101* (1), 57-66. DOI: 10.1016/j.annall.2007.09.011
131. Behrendt, H.; Tomczok, J.; Sliwa-Tomczok, W.; Kasche, A.; von Eschenbach, C. E.; Becker, W. M.; Ring, J., Timothy grass (*Phleum pratense* L.) pollen as allergen carriers and initiators of an allergic response. *Int. Arch. Allergy Immunol.* **1999**, *118* (2-4), 414-418. DOI: 10.1159/000024151
132. Behrendt, H.; Kasche, A.; Ebner von Eschenbach, C.; Risse, U.; Huss-Marp, J.; Ring, J., Secretion of proinflammatory eicosanoid-like substances precedes allergen release from pollen grains in the initiation of allergic sensitization. *International archives of allergy and immunology* **2001**, *124* (1-3), 121-5.
133. Chehregani, A.; Majde, A.; Moin, M.; Gholami, M.; Shariatzadeh, M. A.; Nassiri, H., Increasing allergy potency of *Zinnia* pollen grains in polluted areas. *Ecotox. Environ. Safe.* **2004**, *58* (2), 267-272. DOI: 10.1016/j.ecoenv.2003.12.004
134. Beck, I.; Jochner, S.; Gilles, S., et al., High Environmental Ozone Levels Lead to Enhanced Allergenicity of Birch Pollen. *PLoS One* **2013**, *8* (11), 7. DOI: 10.1371/journal.pone.0080147
135. Bryce, M.; Drews, O.; Schenk, M. F., et al., Impact of Urbanization on the Proteome of Birch Pollen and Its Chemotactic Activity on Human Granulocytes. *Int. Arch. Allergy Immunol.* **2010**, *151* (1), 46-55. DOI: 10.1159/000232570
136. Obersteiner, A.; Gilles, S.; Frank, U., et al., Pollen-Associated Microbiome Correlates with Pollution Parameters and the Allergenicity of Pollen. *PLoS One* **2016**, *11* (2). DOI: 10.1371/journal.pone.0149545



137. Cuinica, L. G.; Abreu, I.; da Silva, J. E., Effect of air pollutant NO<sub>2</sub> on *Betula pendula*, *Ostrya carpinifolia* and *Carpinus betulus* pollen fertility and human allergenicity. *Environ. Pollut.* **2014**, *186*, 50-55. DOI: 10.1016/j.envpol.2013.12.001
138. Ribeiro, H.; Duque, L.; Sousa, R.; Cruz, A.; Gomes, C.; da Silva, J. E.; Abreu, I., Changes in the IgE-reacting protein profiles of *Acer negundo*, *Platanus x acerifolia* and *Quercus robur* pollen in response to ozone treatment. *Int. J. Environ. Health Res.* **2014**, *24* (6), 515-527. DOI: 10.1080/09603123.2013.865716
139. Lang-Yona, N.; Shuster-Meiseles, T.; Mazar, Y.; Yarden, O.; Rudich, Y., Impact of urban air pollution on the allergenicity of *Aspergillus fumigatus* conidia: Outdoor exposure study supported by laboratory experiments. *Sci. Total Environ.* **2015**, *541*, 365-371. DOI: 10.1016/j.scitotenv.2015.09.058
140. Ouyang, Y.; Xu, Z.; Fan, E.; Li, Y.; Zhang, L., Effect of nitrogen dioxide and sulfur dioxide on viability and morphology of oak pollen. *Int. Forum Allergy Rhinol.* **2016**, *6* (1), 95-100. DOI: 10.1002/alr.21632
141. Behrendt, H.; Beckert, W. M., Localization, release and bioavailability of pollen allergens: the influence of environmental factors. *Curr. Opin. Immunol.* **2001**, *13* (6), 709-715. DOI: 10.1016/s0952-7915(01)00283-7
142. Behrendt, H.; Friedrichs, K. H.; Krämer, U.; Hitzfeld, B.; Becker, W. M.; Ring, J., The role of indoor and outdoor air pollution on allergic diseases. In *Progress in Allergy and Clinical Immunology*, Johanson, S. G., Ed. Hogrefe & Huber Publisher (now Hogrefe Publishing): Stockholm, 1995; Vol. 3, 83-89.