

SUPPLEMENTARY MATERIAL

Table S1. Summary of mutation accumulation experiments.

Table S2. Size of the genome used (G^*) for mutations calling.

Table S3. Complete list of base-substitution mutations in *O. tauri*.

Table S4. Complete list of base-substitution mutations in *O. mediterraneus*.

Table S5. Complete list of base-substitution mutations in *B. prasinos*.

Table S6. Complete list of base-substitution mutations in *M. pusilla*.

Table S7. Complete list of deletions in the four experiments.

Table S8. Complete list of insertions in the four experiments.

Table S9. RNAseq coverage of exons and non-exonic regions, in mutated and non-mutated sites in *B. prasinos* and *O. tauri*.

Table S10: GC bias between coding and intergenic sequences.

Table S11. Spontaneous base-substitution mutation rates estimated by mutation accumulation experiments or pedigree sequencing in Bacteria and Eukaryotes.

Table S12. Effect of GC departure from equilibrium GC content on the base substitution mutation rate.

Fig. S1. Mutation rate as a function of genome size.

Table S1. N_e is the average effective population size during the experiment (estimated as the harmonic mean of cell numbers). Total lines are the number of sequenced lines at the end of the experiment. Total generations are the number of independent generations obtained with all sequenced MA lines. T_0 to T_f is the duration of the experiment since the inoculation of the MA lines.

Species	Total line	Total generations	Mean generations per MA line	N_e	T_0 to T_f (days)
<i>O. tauri</i> RCC4221	40	17 250	431	8.5	378
<i>O. mediterraneus</i> RCC2590	37	8 379	235	7	294
<i>M. pusilla</i> RCC299	37	4 145	112	6	299
<i>B. prasinos</i> RCC1105	36	4994	139	8.5	224

Table S2. Size of the genome used (G^*) for mutation calling. $G^*\text{min}$ is the minimum and $G^*\text{max}$ the maximum genome size for each species.

Species	G (Mb)	$G^*\text{average}$ (%)	$G^*\text{min}$ (%)	$G^*\text{max}$ (%)
<i>O. tauri</i> RCC4221	13.03	12.60 (97.5)	11.82 (91.54)	12.84 (99.45)
<i>O. mediterraneus</i> RCC2590	13.48	13.10 (97.2)	12.28 (91.10)	13.35 (99.08)
<i>B. prasinos</i> RCC1105	15.07	15.02 (99.6)	14.66 (97.23)	15.03 (99.73)
<i>M. pusilla</i> RCC299	21.11	21.01 (99.5)	20.55 (97.36)	21.07 (99.79)

Table S3. Complete list of base-substitution mutations in *O. tauri*. Ns: non-synonymous mutation, Syn: synonymous mutations, - : intergenic mutations.

Chromosome	Position	Reference	Mutation	Effect
1	281349	T	C	Syn
1	462543	T	A	-
1	462544	A	T	-
1	536352	G	A	-
1	688195	A	G	-
1	762227	A	G	Ns
1	764912	G	A	-
1	934994	C	T	Syn
2	202377	C	A	Ns
2	744253	G	A	Syn
2	759388	C	T	Ns
2	817108	C	T	Ns
2	820022	C	A	Ns
2	868393	G	A	Syn
2	1070559	A	G	Ns
3	41080	C	T	Syn
3	127716	G	A	Ns
3	210680	A	G	-
3	353325	C	T	Ns
3	379647	C	T	Ns
3	779548	T	C	Syn
3	971494	C	T	Ns
4	117	C	A	-
4	366731	C	T	Syn
4	524836	G	A	Ns
4	735752	C	T	Ns
5	5699	C	T	Ns
5	405535	G	A	Syn
5	498490	T	G	Ns
5	600053	T	A	-
5	600056	G	A	-
5	658543	G	A	Ns
5	668815	C	T	Ns
6	12972	C	T	Ns
6	18960	C	T	-
6	46323	T	G	Ns
6	162319	T	C	Ns
6	334481	G	C	-
6	661927	C	T	-
6	716579	C	T	Ns
7	475871	A	G	Ns
7	612454	C	T	Syn
7	702913	A	C	Ns
7	744212	G	A	Ns
8	66027	G	A	-
8	185592	G	C	-
9	31962	G	A	Ns
9	126121	C	T	Syn
9	393218	C	T	Ns

9	398617	C	T	Syn
9	514297	C	T	Ns
9	643568	A	G	Ns
10	108529	G	A	Ns
10	510428	C	T	Ns
10	562914	G	A	Syn
10	587147	G	A	-
11	222249	A	G	-
11	446073	T	C	Syn
11	452825	C	A	Ns
11	472590	G	A	Stop gained
11	474928	A	G	-
12	94	A	G	-
12	419488	C	A	Ns
12	460233	T	G	-
13	145936	G	A	Syn
13	365169	C	T	Ns
13	496008	C	T	Ns
14	75583	G	A	Intron
14	210547	G	A	Ns
14	231150	G	T	-
14	257724	C	A	Ns
14	374547	G	C	Syn
14	495621	G	A	Syn
14	500741	C	T	Ns
15	165137	C	T	Ns
15	177872	G	T	Ns
15	352817	A	G	Syn
16	460708	C	T	Syn
16	325515	G	T	Syn
16	521001	C	T	Ns
17	35411	T	G	Ns
17	410925	C	T	-
17	410926	G	A	-
17	410927	A	G	-
17	410928	C	G	-
18	122316	A	G	-
18	252024	T	G	-
18	345570	T	G	-
18	345641	G	A	-
20	49667	C	T	Ns
20	165267	T	G	-

Table S4. Complete list of base-substitution mutations in *O. mediterraneus*. Ns: non-synonymous mutation, Syn: synonymous mutations, - : intergenic mutations.

Chromosome	Position	Reference	Mutation	Effect
1	240410	T	C	Syn
1	281668	A	T	Ns
1	384332	T	C	-
1	500961	T	G	-
1	504598	G	A	-
1	960787	G	T	Ns
2	783209	C	A	Syn
3	168617	T	G	Ns
3	194253	A	T	Ns
4	4863	A	G	Ns
4	250707	T	C	-
4	271238	A	G	-
4	271242	G	T	-
4	271243	A	G	-
4	271249	A	C	-
4	271258	C	T	-
4	271264	G	T	-
5	204024	G	A	Ns
5	237455	G	A	Ns
5	369175	C	T	Ns
5	561353	C	A	Ns
6	167738	C	T	Ns
6	365873	C	T	Syn
6	638629	C	A	Ns
7	136735	A	C	Ns
7	298488	C	T	Ns
8	59263	G	A	Ns
8	106639	G	A	Ns
8	165454	G	A	Syn
8	246199	T	C	Ns
8	285631	G	A	Syn
8	390757	T	A	Ns
8	425236	T	G	Ns
8	437922	G	C	Ns
8	699362	G	A	Syn
9	552630	G	T	Ns
9	701807	A	C	Ns
9	717500	C	A	-
9	737678	G	T	Ns
10	165842	G	A	Ns
13	107450	T	C	Ns
13	162729	C	T	Syn

13	586916	T	C	Ns
14	60175	T	A	Ns
14	60217	G	A	Ns
14	98081	A	C	Ns
15	112247	C	A	Start gained
16	222414	C	T	Syn
16	297544	C	T	Syn
17	368001	G	T	Ns
17	422490	C	T	-
18	272777	A	G	Ns
18	299852	A	G	Ns
19	81732	C	G	Ns

Table S5. Complete list of base-substitution mutations in *B. prasinos*. Ns: non-synonymous mutation, Syn: synonymous mutations, - : intergenic mutations.

Chromosome	Position	Reference	Mutation	Effect
1	42082	C	A	Ns
1	79489	C	T	Syn
1	418914	T	G	Ns
1	1305044	G	C	Syn
11	525583	C	T	Ns
12	50	T	A	-
12	122406	T	C	-
13	208600	T	A	Ns
17	72	G	C	-
18	999	C	A	Ns
18	235306	T	G	-
19	69819	A	T	-
19	145968	A	T	-
2	556426	G	C	Ns
4	135852	C	T	Syn
4	531741	C	G	Ns
5	87328	C	T	Ns
5	145129	A	T	Intron
5	602981	C	G	Syn
5	797176	A	T	Ns
8	574299	C	A	Ns
9	93038	C	T	Syn

Table S6. Complete list of base-substitution mutations in *M. pusilla*. Ns: non-synonymous mutation, Syn: synonymous mutations, - : intergenic mutations.

Chromosome	Position	Reference	Mutation	Effect
1	101651	G	T	-
1	319728	T	A	-
1	319729	C	A	-
1	356850	G	C	Ns
1	868786	A	G	-
1	907594	A	G	-
1	989553	G	A	Ns
1	1166944	A	G	-
1	1311961	A	G	Ns
1	1328941	C	T	-
1	1480609	A	C	-
1	1531771	T	C	-
1	1771291	T	C	-
2	296	C	T	-
2	197240	G	C	Ns
2	318653	C	G	Ns
2	318654	A	C	Ns
2	318655	G	C	Ns
2	371001	G	T	Ns
2	629587	T	G	Ns
2	1231590	G	A	Syn
2	1439488	C	T	Ns
2	1898300	G	T	Syn
2	1898302	A	C	Ns
2	1898303	C	A	Ns
2	1898304	T	A	Ns
2	1898305	T	C	Syn
3	1370371	C	G	Ns
3	1375097	G	A	Ns
3	1708043	C	T	Ns
4	3470	C	A	Syn
4	334275	G	A	Syn
4	571514	G	A	Ns
4	1224092	G	A	Syn
5	486431	C	A	Syn
5	508099	G	C	Syn
5	556716	C	G	Ns
5	896616	C	T	-
5	1108981	G	A	Ns
6	1235633	G	A	Ns
7	570664	G	T	Ns

7	570665	A	C	Syn
7	570666	G	T	Ns
7	672236	G	A	-
7	746112	G	C	Syn
8	93285	G	T	Ns
8	273349	C	T	Ns
8	284318	G	T	Ns
8	303063	G	A	Syn
8	310233	T	A	Ns
8	310234	G	C	Ns
8	1077655	G	T	Ns
9	1126769	A	T	Syn
11	173723	G	A	-
11	430804	C	T	Ns
11	899300	A	G	Ns
11	899309	G	A	Ns
12	323466	T	C	Ns
12	797741	C	T	Syn
12	823088	T	G	Ns
13	688257	C	G	Syn
14	346000	G	T	Syn
14	346390	G	C	Syn
14	761603	G	A	Ns
15	146889	A	T	Ns
15	146890	T	G	Ns
15	320895	A	G	Syn
15	408470	A	C	-
15	410306	C	T	Ns
15	504682	G	T	Ns
16	329517	C	T	Ns

Table S7. Complete list of deletions in the four species.

Chromosome	Position	Reference	Mutation	Effect
<i>Bathycoccus prasinus</i>				
1	164711	AGGCAGCAGCTG	A	-
1	164726	AATTCAATTCAATA	A	-
5	622569	GA	G	Frame shift
5	963834	AAATATCTATTG	A	-
14	329038	TG	T	-
<i>Micromonas pusilla</i>				
1	1787228	TTATTCCTCGAAGCTTACGTACG	T	-
1	1924354	CA	C	Frame shift
1	319750	GTACCTTCGAAGGTATAA	G	-
3	214	CT	C	-
5	140463	CGCGAGACCTCG	C	Frame shift
6	1235612	AGGAGGAGGAGGGGGAGGGAGGG	A	Codons deletion
8	310219	CGATCTGCGTCCGGTG	C	Codons deletion
10	1045182	GCGC	G	-
10	1160336	TCA	T	-
11	899291	ACAGACGGCACAGCTGGCG	A	Codons deletion
14	474	AACCCTTCGT	A	-
16	287228	AACTCGAGTTGACAAGACC	A	-
<i>Ostreococcus tauri</i>				
1	601169	GA	G	-
6	46321	CT	C	Frame shift
8	273361	AAC	A	Frame shift
8	72783	GACACCCCGCGTGTACGGGACCGCGACCC	G	-
11	222762	GA	G	Frame shift
12	403601	CGCGAGACCGGCCGACATGCCCGTGTGCCACCGTCGGAAACT	C	Frame shift
13	135	CA	C	-
17	87	CT	C	-
<i>Ostreococcus mediterraneus</i>				
6	284415	CG	C	Frame shift
7	156017	GT	G	-
7	1775	TGTTGCC	T	-
8	350046	CGTT	C	Codons deletion
9	185017	CACGGCGACGACGAACGATGGCG	C	Frame shift
12	567140	TCG	T	Frame shift
12	328305	GT	G	-
13	112767	ATCTATCGTCGCGACGGCGGTCGTCTCTATG	A	Codons deletion

Table S8. Complete list of insertions in the four species.

Chromosome	Position	Reference	Mutation	Effect
<i>Bathycoccus prasinus</i>				
2	102	A	AC	-
9	521140	A	AAG	-
13	570953	T	TGCC	Codons insertion
14	272	A	ACC	-
18	523	A	AC	-
<i>Micromonas pusilla</i>				
1	319727	A	AC	-
2	378439	C	CCG	Frame shift
<i>Ostreococcus tauri</i>				
4	76071	C	CG	-
10	455685	T	TCGTCGG	Codons insertion
15	109437	C	CG	-
17	226384	C	CG	-
20	153964	G	GA	-
<i>Ostreococcus mediterraneus</i>				
1	384327	G	GATT	-
8	729406	C	CA	Frame shift
12	432782	C	CTACTG	-

Table S9: RNAseq coverage of coding (cds) and intergenic (non-cds) sequences, in mutated and non-mutated sites in *B. prasinus* and *O. tauri*. The removal of the 10 mutations located in the subtelomeric regions does not change the significance of the transcription bias (Wilcoxon test, P-value=0.016).

Species	Sequence types	n mutations	Mutated site coverage	Non-mutated site coverage	P. value
					Wilcoxon test
<i>Bathycoccus prasinus</i>	non-cds	13	247	646	0.0004
	cds	19	505	667	0.123
<i>Ostreococcus tauri</i>	non-cds	38	26	149	3.45×10^{-7}
	cds	64	127	126	0.382

Table S10: GC bias between coding and intergenic sequences. Data from *B. prasinus* are not presented because of low number of mutations.

Species	Coding sequences			Intergenic sequences		
	n (GC to AT)	n (AT to GC)	$\mu_{(GC \text{ to } AT)} / \mu_{(AT \text{ to } GC)}$	n (GC to AT)	n (AT to GC)	$\mu_{(GC \text{ to } AT)} / \mu_{(AT \text{ to } GC)}$
<i>O. tauri</i>	48	13	2.57	12	11	0.76
<i>O. mediterraneus</i>	24	12	1.57	6	6	0.79
<i>M. pusilla</i>	30	11	1.43	7	7	0.79

Table S11. Spontaneous base-substitution mutation rates estimated by mutation accumulation experiments and pedigree. G is the genome size in Mb, μ is the mutation rate per nucleotide and U is the number of mutations per genome per generation. The data from Ness et al. (2015) is the average of 6 strains. Effective population size come from Lynch supplementary material (Lynch 2010), except for *Mus musculus* (Phifer-Rixey et al. 2012), *Heliconius melpomene* (Keightley, Pinharanda, et al. 2014), *Ficedula albicollis* (Backström et al. 2013), *Arabidopsis thaliana* (Cao et al. 2011), *Caenorhabditis elegans* (Cutter 2006), *Caenorhabditis briggsae* (Cutter et al. 2006), *Drosophila melanogaster* (Shapiro et al. 2007) and *O. tauri* (Romain Blanc-Mathieu, personal communication).

Species	G	μ	U	Ne	References
<i>Homo sapiens</i>	3309.0	1.29E-08	38.5500	2.00E+04	(Besenbacher et al. 2016)
<i>Mus musculus</i>	2671.0	5.40E-09	14.5800	2.00E+05	(Uchimura et al. 2015)
<i>Ficedula albicollis</i>	1118.0	4.60E-09	5.0600	4.50E+05	(Smeds et al. 2016)
<i>Arabidopsis thaliana</i>	134.4	7.00E-09	1.0990	2.50E+05	(Ossowski et al. 2010)
<i>Caenorhabditis elegans</i>	100.3	1.48E-09	0.1479	8.00E+04	(Denver et al. 2012)
<i>Caenorhabditis briggsae</i>	108.4	1.34E-09	0.1447	6.00E+04	(Denver et al. 2012)
<i>Pristionchus pacificus</i>	133.1	2.00E-09	0.2663	-	(Weller et al. 2014)
<i>Drosophila melanogaster</i>	148.0	5.49E-09	0.6698	1.15E+06	(Schridler et al. 2013)
<i>Heliconius melpomene</i>	273.8	2.90E-09	0.7940	2.00E+06	(Keightley, Pinharanda, et al. 2014)
<i>Daphnia pulex</i>	197.2	2.30E-09	0.4536	-	(Flynn et al. 2016)
<i>Ostreococcus tauri</i>	13.0	4.19E-10	0.0054	2.00E+07	This study
<i>Ostreococcus mediterraneus</i>	13.5	4.92E-10	0.0065	-	This study
<i>Bathycoccus prasinos</i>	15.1	3.07E-10	0.0046	-	This study
<i>Micromonas pusilla</i>	21.1	8.15E-10	0.0172	-	This study
<i>Chlamydomonas reinhardtii</i>	112.0	9.63E-10	0.1079	3.10E+07	(Ness et al. 2015)
<i>Saccharomyces cerevisiae</i>	12.3	1.67E-10	0.0021	6.20E+06	(Zhu et al. 2014)
<i>Schizoaccharomyces pombe</i>	12.6	2.00E-10	0.0025	2.60E+06	(Farlow et al. 2015)
<i>Paramecium tetraurelia</i>	72.1	1.94E-11	0.0014	1.24E+08	(Sung, Tucker, et al. 2012)
<i>Tetrahymena thermophila</i>	104.0	7.61E-12	0.0008	1.12E+08	(Long et al. 2016)
<i>Dictyostelium discoideum</i>	34.2	2.90E-11	0.0010	-	(Saxer et al. 2012)
<i>Bacillus subtilis</i>	4.2	3.28E-10	0.0014	6.30E+07	(Sung et al. 2015)
<i>Escherichia coli</i>	4.6	2.20E-10	0.0010	1.80E+08	(Lee et al. 2012)
<i>Mesoplasma florum</i>	0.8	9.78E-09	0.0078	1.10E+06	(Sung, Ackerman, et al. 2012)
<i>Burkholderia cenocepacia</i>	7.7	1.33E-10	0.0010	-	(Dillon et al. 2015)
<i>Pseudomonas aeruginosa</i>	6.6	7.92E-11	0.0005	2.00E+07	(Dettman et al. 2016)
<i>Salmonella typhimurium</i>	4.8	7.00E-10	0.0034	-	(Lind and Andersson 2008)
<i>Mycobacterium tuberculosis</i>	4.4	2.58E-10	0.0011	-	(Ford et al. 2011)
<i>Deinococcus radiodurans</i>	3.2	4.99E-10	0.0016	-	(Long et al. 2015)
<i>Vibrio cholerae</i>	4.02	1.07E-10	0.00043	-	(Dillon et al. 2016)
<i>Vibrio fischeri</i>	4.23	2.07E-10	0.00089	-	(Dillon et al. 2016)

Table S12. Effect of GC deviation from equilibrium GC content on the base substitution mutation rate.

R_1 , R_2 , R_3 and R_4 (see methods) were used to calculate GC_{eq} and mutation rate at equilibrium μ_{eq} . The ratio μ/μ_{eq} is an estimation of the elevation of the mutation rate due to the distance from equilibrium GC content, i.e. in *O. tauri* the mutation rate increases by ~12%. We only included μ_{eq} estimated from more than 8 mutation events per category to avoid spurious variance in GC_{eq} estimates.

Species	μ_{bs}	GC	GC_{eq}	GC_r	GC>AT relative to AT>GC	μ/μ_{eq}	References
<i>Homo sapiens</i>	1.29E-08	0.415	0.326	1.272	2.064	1.106	(Besenbacher et al. 2016)
<i>Mus musculus</i>	5.40E-09	0.424	0.207	2.051	3.842	1.295	(Uchimura et al. 2015)
<i>Ficedula albicollis</i>	4.60E-09	0.443	0.311	1.426	2.219	1.116	(Smeds et al. 2016)
<i>Arabidopsis thaliana</i>	7.00E-09	0.367	0.138	2.663	6.255	1.640	(Ossowski et al. 2010)
<i>Daphnia pulex</i>	2.30E-09	0.422	0.141	2.997	6.101	1.629	(Flynn et al. 2016)
<i>Caenorhabditis elegans</i>	1.48E-09	0.354	0.193	1.837	4.189	1.225	(Denver et al. 2012)
<i>Caenorhabditis briggsae</i>	1.34E-09	0.377	0.211	1.784	3.732	1.227	(Denver et al. 2012)
<i>Pristionchus pacificus</i>	2.00E-09	0.427	0.157	2.711	5.350	1.381	(Weller et al. 2014)
<i>Drosophila melanogaster</i>	5.49E-09	0.419	0.188	2.228	4.318	1.324	(Schrider et al. 2013)
<i>Ostreococcus tauri</i>	4.19E-10	0.590	0.365	1.615	1.737	1.118	This study
<i>Ostreococcus mediterraneus</i>	3.73E-10	0.560	0.433	1.293	1.310	1.017	This study
<i>Micromonas pusilla</i>	8.15E-10	0.638	0.462	1.382	1.166	1.030	This study
<i>Chlamydomonas reinhardtii</i>	9.63E-10	0.619	0.259	2.392	2.864	1.428	(Ness et al. 2015)
<i>Saccharomyces cerevisiae</i>	1.67E-10	0.384	0.311	1.235	2.216	1.067	(Zhu et al. 2014)
<i>Schizoaccharomyces pombe</i>	2.00E-10	0.360	0.264	1.364	2.790	1.122	(Farlow et al. 2015)
<i>Bacillus subtilis</i>	3.28E-10	0.437	0.443	0.986	1.256	0.999	(Sung et al. 2015)
<i>Escherichia coli</i>	2.20E-10	0.506	0.450	1.124	1.222	1.010	(Lee et al. 2012)
<i>Mesoplasma florum</i>	9.78E-09	0.270	0.059	4.582	15.970	2.628	(Sung et al. 2012a)
<i>Burkholderia cenocepacia</i>	1.33E-10	0.669	0.551	1.213	0.814	0.978	(Dillon et al. 2015)
<i>Pseudomonas aeruginosa</i>	7.92E-11	0.662	0.396	1.671	1.524	1.083	(Dettman et al. 2016)
<i>Deinococcus radiodurans</i>	4.99E-10	0.668	0.643	1.039	0.555	0.983	(Long et al. 2015)
<i>Vibrio cholerae</i>	1.07E-10	0.476	0.281	1.694	-	-	(Dillon et al. 2016)
<i>Vibrio fischeri</i>	2.07E-10	0.384	0.202	1.901	-	-	(Dillon et al. 2016)

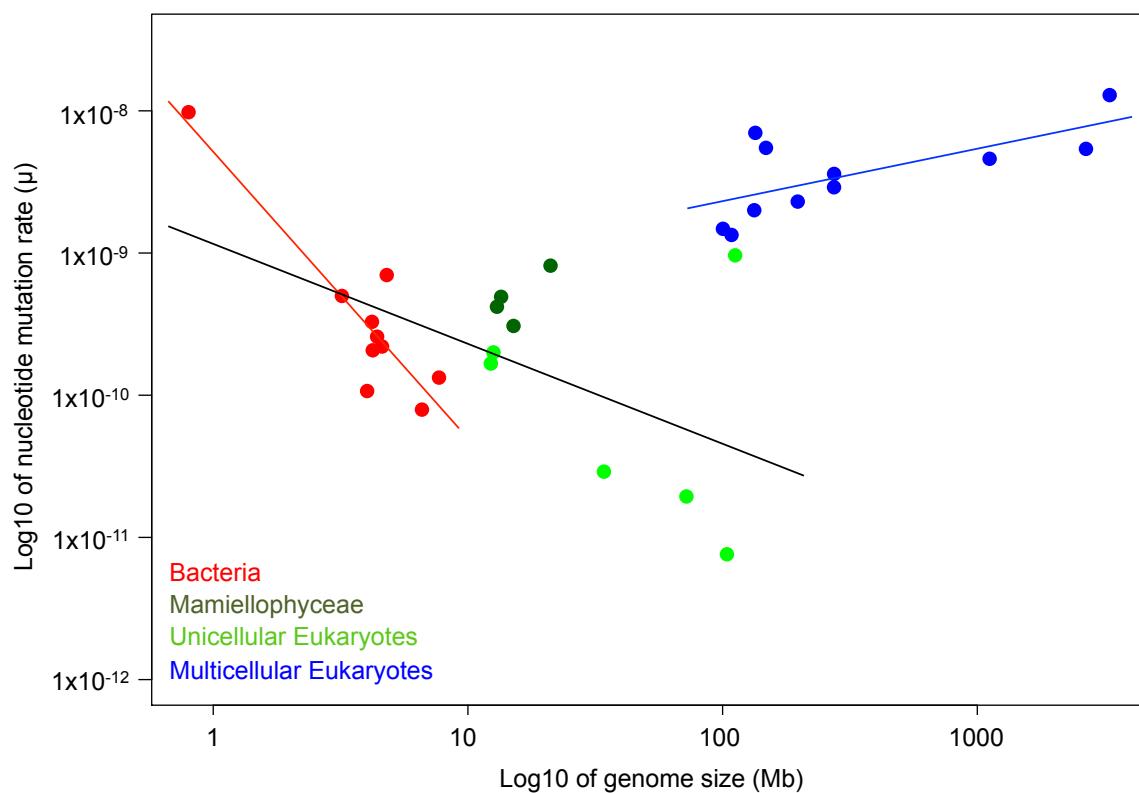


Fig. S1. Mutation rate as a function of genome size G (data available Table S11). *Blue*: positive correlation between μ and G in multicellular eukaryotes ($n=10$, Pearson correlation, $\rho=0.67$, P-value<0.03). *Black*: negative correlation between μ and G in microorganisms ($n=19$, Pearson correlation, $\rho=-0.57$, P-value=0.01). *Red*: negative correlation between μ and G in bacteria ($n=10$, Pearson correlation, $\rho=-0.95$, P-value=0.001).

References

- Backström N, Sætre G-P, Ellegren H. 2013. Inferring the demographic history of European Ficedula flycatcher populations. *BMC Evol. Biol.* 13:2.
- Besenbacher S, Sulem P, Helgason A, Helgason H, Kristjansson H, Jonasdottir A, Jonasdottir A, Magnusson OT, Thorsteinsdottir U, Masson G, et al. 2016. Multi-nucleotide de novo Mutations in Humans. *PLoS Genet.* 12:e1006315.
- Cao J, Schneeberger K, Ossowski S, Günther T, Bender S, Fitz J, Koenig D, Lanz C, Stegle O, Lippert C, et al. 2011. Whole-genome sequencing of multiple *Arabidopsis thaliana* populations. *Nat. Genet.* 43:956–963.
- Cutter AD. 2006. Nucleotide polymorphism and linkage disequilibrium in wild populations of the partial selfer *Caenorhabditis elegans*. *Genetics* 172:171–184.

- Cutter AD, Félix M-A, Barrière A, Charlesworth D. 2006. Patterns of Nucleotide Polymorphism Distinguish Temperate and Tropical Wild Isolates of *Caenorhabditis briggsae*. *Genetics* 173:2021–2031.
- Denver DR, Wilhelm LJ, Howe DK, Gafner K, Dolan PC, Baer CF. 2012. Variation in base-substitution mutation in experimental and natural lineages of *Caenorhabditis* nematodes. *Genome Biol. Evol.* 4:513–522.
- Dettman JR, Sztepanacz JL, Kassen R. 2016. The properties of spontaneous mutations in the opportunistic pathogen *Pseudomonas aeruginosa*. *BMC Genomics* [Internet] 17. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4702332/>
- Dillon MM, Sung W, Lynch M, Cooper VS. 2015. The Rate and Molecular Spectrum of Spontaneous Mutations in the GC-Rich Multi-Chromosome Genome of *Burkholderia cenocepacia*. *Genetics*.
- Dillon MM, Sung W, Sebra R, Lynch M, Cooper VS. 2016. Genome-wide biases in the rate and molecular spectrum of spontaneous mutations in *Vibrio cholerae* and *Vibrio fischeri*. *Mol. Biol. Evol.*:msw224.
- Farlow A, Long H, Arnoux S, Sung W, Doak TG, Nordborg M, Lynch M. 2015. The Spontaneous Mutation Rate in the Fission Yeast *Schizosaccharomyces pombe*. *Genetics:genetics*.115.177329.
- Flynn JM, Chain FJJ, Schoen DJ, Cristescu ME. 2016. Spontaneous mutation accumulation in *Daphnia pulex* in selection-free versus competitive environments. *Mol. Biol. Evol.*:msw234.
- Ford CB, Lin PL, Chase MR, Shah RR, Iartchouk O, Galagan J, Mohaideen N, loerger TR, Sacchettini JC, Lipsitch M, et al. 2011. Use of whole genome sequencing to estimate the mutation rate of *Mycobacterium tuberculosis* during latent infection. *Nat. Genet.* 43:482–486.
- Keightley PD, Pinharanda A, Ness RW, Simpson F, Dasmahapatra KK, Mallet J, Davey JW, Jiggins CD. 2014. Estimation of the spontaneous mutation rate in *Heliconius melpomene*. *Mol. Biol. Evol.*
- Lee H, Popodi E, Tang H, Foster PL. 2012. Rate and molecular spectrum of spontaneous mutations in the bacterium *Escherichia coli* as determined by whole-genome sequencing. *Proc. Natl. Acad. Sci. U. S. A.* 109:E2774-2783.
- Lind PA, Andersson DI. 2008. Whole-genome mutational biases in bacteria. *Proc. Natl. Acad. Sci. U. S. A.* 105:17878–17883.

Long H, Kucukyildirim S, Sung W, Williams E, Lee H, Ackerman M, Doak TG, Tang H, Lynch M. 2015. Background Mutational Features of the Radiation-Resistant Bacterium *Deinococcus radiodurans*. *Mol. Biol. Evol.* 32:2383–2392.

Long H, Winter DJ, Chang AY-C, Sung W, Wu SH, Balboa M, Azevedo RBR, Cartwright RA, Lynch M, Zufall RA. 2016. Low base-substitution mutation rate in the germline genome of the ciliate *Tetrahymena thermophila*. *Genome Biol. Evol.*:evw223.

Lynch M. 2010. Evolution of the mutation rate. *Trends Genet. TIG* 26:345–352.

Ness RW, Morgan AD, VasanthaKrishnan RB, Colegrave N, Keightley PD. 2015. Extensive de novo mutation rate variation between individuals and across the genome of *Chlamydomonas reinhardtii*. *Genome Res.* 25:1739–1749.

Ossowski S, Schneeberger K, Lucas-Lledó JI, Warthmann N, Clark RM, Shaw RG, Weigel D, Lynch M. 2010. The rate and molecular spectrum of spontaneous mutations in *Arabidopsis thaliana*. *Science* 327:92–94.

Phifer-Rixey M, Bonhomme F, Boursot P, Churchill GA, Piálek J, Tucker PK, Nachman MW. 2012. Adaptive Evolution and Effective Population Size in Wild House Mice. *Mol. Biol. Evol.* 29:2949–2955.

Saxer G, Havlak P, Fox SA, Quance MA, Gupta S, Fofanov Y, Strassmann JE, Queller DC. 2012. Whole genome sequencing of mutation accumulation lines reveals a low mutation rate in the social amoeba *Dictyostelium discoideum*. *PLoS One* 7:e46759.

Schrider DR, Houle D, Lynch M, Hahn MW. 2013. Rates and genomic consequences of spontaneous mutational events in *Drosophila melanogaster*. *Genetics* 194:937–954.

Shapiro JA, Huang W, Zhang C, Hubisz MJ, Lu J, Turissini DA, Fang S, Wang H-Y, Hudson RR, Nielsen R, et al. 2007. Adaptive genic evolution in the *Drosophila* genomes. *Proc. Natl. Acad. Sci. U. S. A.* 104:2271–2276.

Smeds L, Qvarnstrom A, Ellegren H. 2016. Direct estimate of the rate of germline mutation in a bird. *Genome Res.*

Sung W, Ackerman MS, Gout J-F, Miller SF, Williams E, Foster PL, Lynch M. 2015. Asymmetric Context-Dependent Mutation Patterns Revealed through Mutation–Accumulation Experiments. *Mol. Biol. Evol.* 32:1672–1683.

Sung W, Ackerman MS, Miller SF, Doak TG, Lynch M. 2012. Drift-barrier hypothesis and mutation-rate evolution. *Proc. Natl. Acad. Sci. U. S. A.* 109:18488–18492.

Sung W, Tucker AE, Doak TG, Choi E, Thomas WK, Lynch M. 2012. Extraordinary genome stability in the ciliate *Paramecium tetraurelia*. Proc. Natl. Acad. Sci. U. S. A. 109:19339–19344.

Uchimura A, Higuchi M, Minakuchi Y, Ohno M, Toyoda A, Fujiyama A, Miura I, Wakana S, Nishino J, Yagi T. 2015. Germline mutation rates and the long-term phenotypic effects of mutation accumulation in wild-type laboratory mice and mutator mice. Genome Res. 18:6148–6114.

Weller AM, Rödelsperger C, Eberhardt G, Molnar RI, Sommer RJ. 2014. Opposing Forces of A/T-Biased Mutations and G/C-Biased Gene Conversions Shape the Genome of the Nematode *Pristionchus pacificus*. Genetics 196:1145–1152.

Zhu YO, Siegal ML, Hall DW, Petrov DA. 2014. Precise estimates of mutation rate and spectrum in yeast. Proc. Natl. Acad. Sci. U. S. A. 111:E2310–E2318.