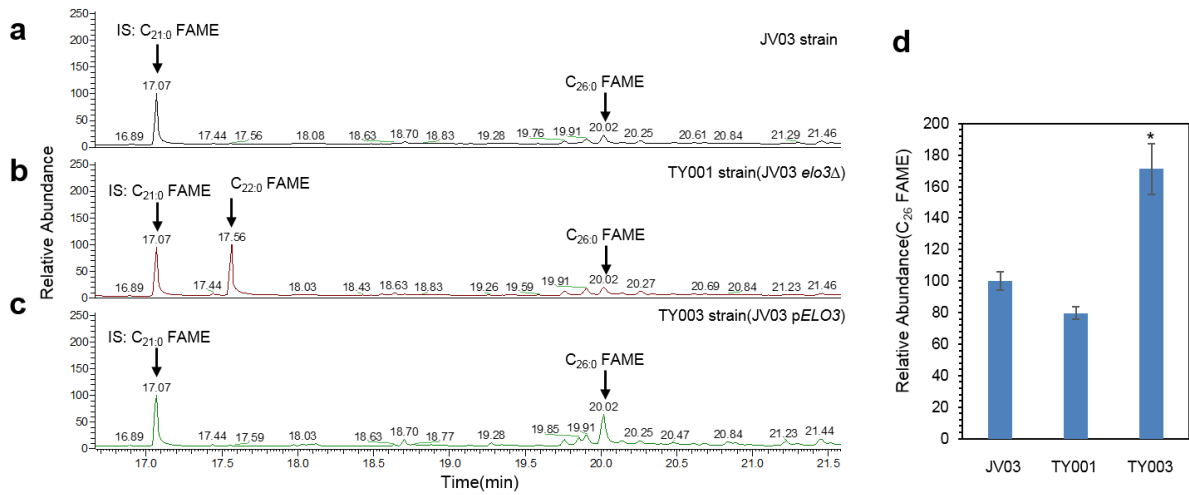


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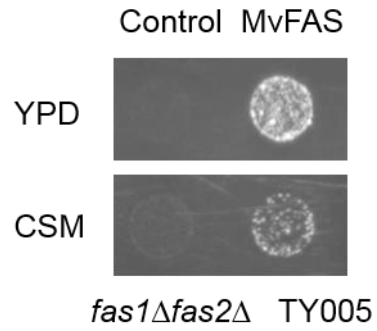


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3 Supplementary Fig. 1. Fatty acid chain length profiles of VLCFA-producing JV03 and derived strains.
4 Typical GC-MS total ion chromatograms (TIC) of total fatty acids extracted from JV03 cells (a), TY001
5 (JV03 *elo3*Δ) (b) and strain TY003 (JV03 *pELO3*) (c). (d) Relative quantification of total FA (C₂₆ FAME)
6 generated by yeast elongation system in strains JV03, TY001 and TY003. Statistical analysis was
7 performed using a Student's t-Test (one-tailed; *, $p < 0.05$; **, $p < 0.01$ and ***, $p < 0.001$; Two-sample
8 unequal variance). At least two independent measurements were performed for each experiment, and
9 the the mean \pm s.d of 3 biological replicates of a representative measurement is shown. Total fatty
10 acids were identified by retention time and comparison with the mass spectral library. All cells were
11 grown as described in experimental procedures.

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15 Supplementary Fig. 2. Complementation with the MvFAS restores the growth of a *fas1Δ/fas2Δ* strain.

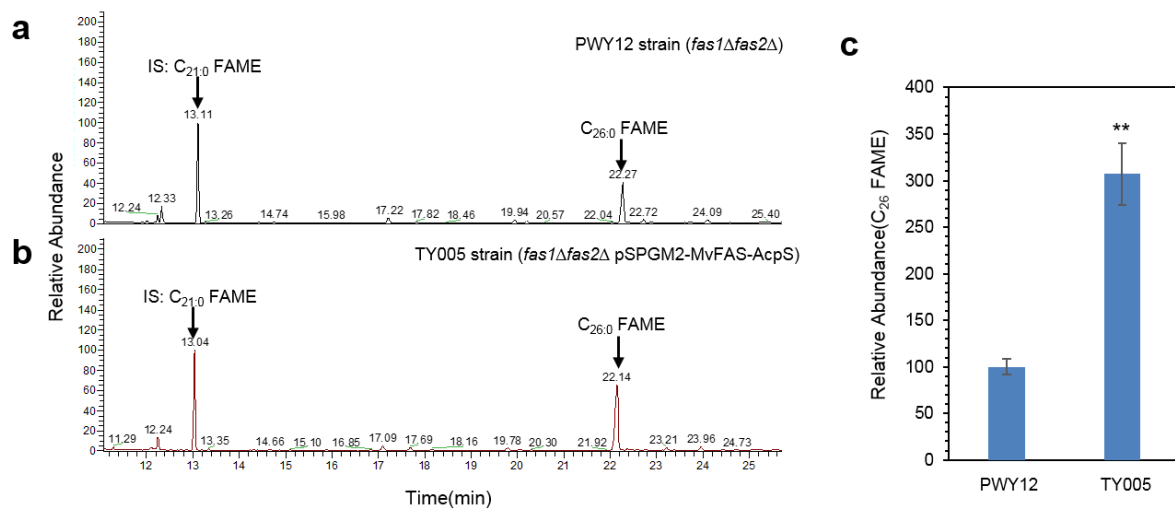
16 Spot assays of the *fas1Δ/fas2Δ* strain without or with expression of *mvfas* (strain TY005) on YPD or

17 CSM plates. The plates were incubated at 30°C and recorded photographically 3 days after inoculation.

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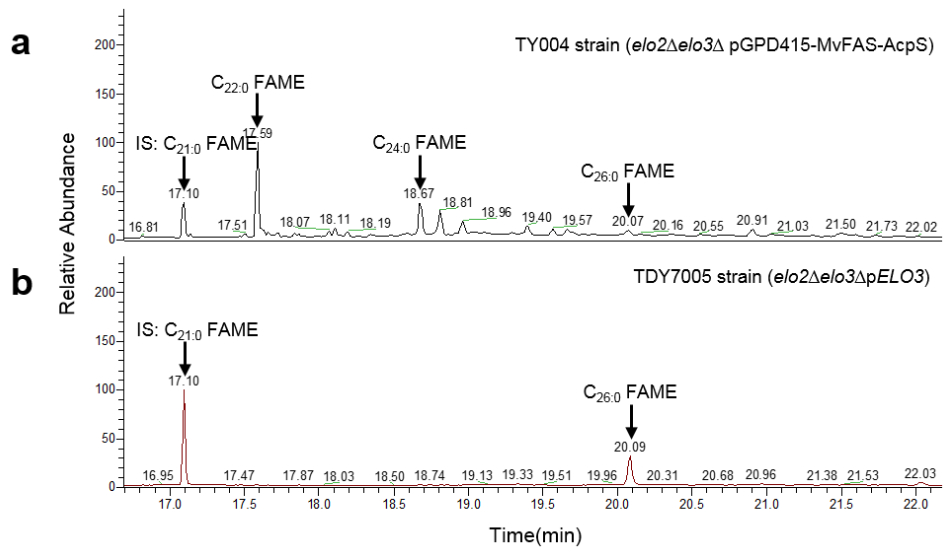


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22 Supplementary Fig. 3. Fatty acid chain length profiles of VLCFA-producing *fas1Δ/fas2Δ* strains. Typical
23 GC-MS total ion chromatograms (TIC) of total fatty acids extracted from PWY12 (*fas1Δ/fas2Δ*) (**a**), or
24 with plasmid pSPGM2-MvFAS-AcpS (strain TY005) (**b**). Total fatty acids were identified by retention
25 time and comparison with the mass spectral library. (**c**) Relative quantification of total FA (C₂₆ FAME)
26 generated by strains PWY12 and TY005. Statistical analysis was performed using a Student's t-Test
27 (one-tailed; *, $p < 0.05$; **, $p < 0.01$ and ***, $p < 0.001$; Two-sample unequal variance). At least two
28 independent measurements were performed for each experiment, and the the mean \pm s.d of 3
29 biological replicates of a representative measurement is shown. All cells were grown as described in
30 experimental procedures.

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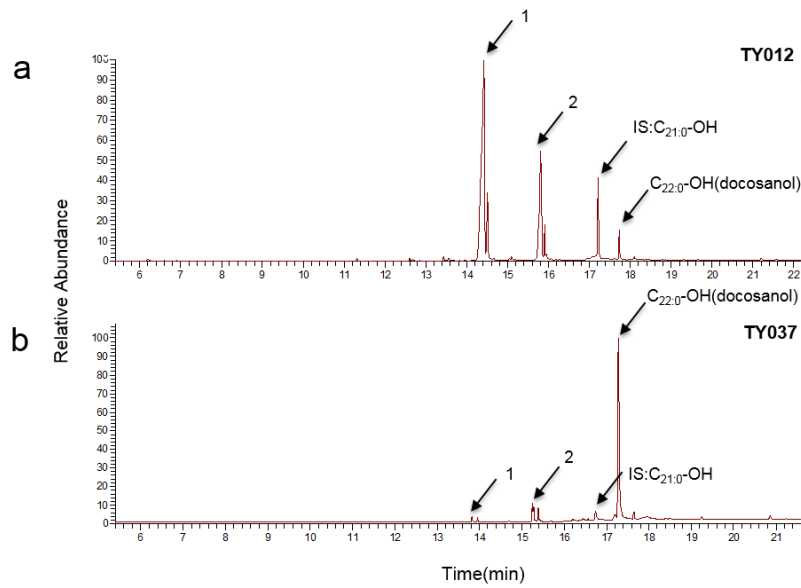


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34 Supplementary Fig. 4. Fatty acid chain length profiles of VLCFA-producing *elo2Δ/elo3Δ* strains. Typical
 35 GC-MS total ion chromatograms (TIC) of total fatty acids extracted from TDY7005 (*elo2Δ/elo3Δ*)
 36 carrying p415GPD-MvFAS-AcpS (strain TY004) **(a)**, or the control plasmid pELO3 (strain TDY7005) **(b)**.
 37 Total fatty acids were identified by retention time and comparison with the mass spectral library. All
 38 cells were grown as described in experimental procedures.

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42 Supplementary Fig. 5. Fatty alcohol profiles of docosanol producing strains. **(a)** Typical GC-MS total ion
 43 chromatogram (TIC) of fatty alcohol extracted from starting strain TY012 (JV03 *elo3Δ* pELO2 pAtFAR).
 44 **(b)** Typical GC-FID total ion chromatograms (TIC) of fatty alcohol extracted from the best docosanol
 45 producer strain TY037 (MATa *ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 gal1Δ::ACC1***
 46 *elo3Δ::AtFAR ELO2 ELO1/pSPGM2-MvFAS-AcpS*) (Fig 5b). Both samples were spiked with the same
 47 amounts of internal standard. 1, C_{16:0} and C_{16:1} fatty acid. 2, C_{18:0} and C_{18:1} fatty acid. Fatty alcohols were
 48 identified by retention time and comparison with the mass spectral library. All cells were grown and
 49 induced as described in experimental procedures.

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51

S. cerevisiae FAS

β -chain (1888aa)



α -chain (2052aa)



M. vaccae FAS (3089aa)



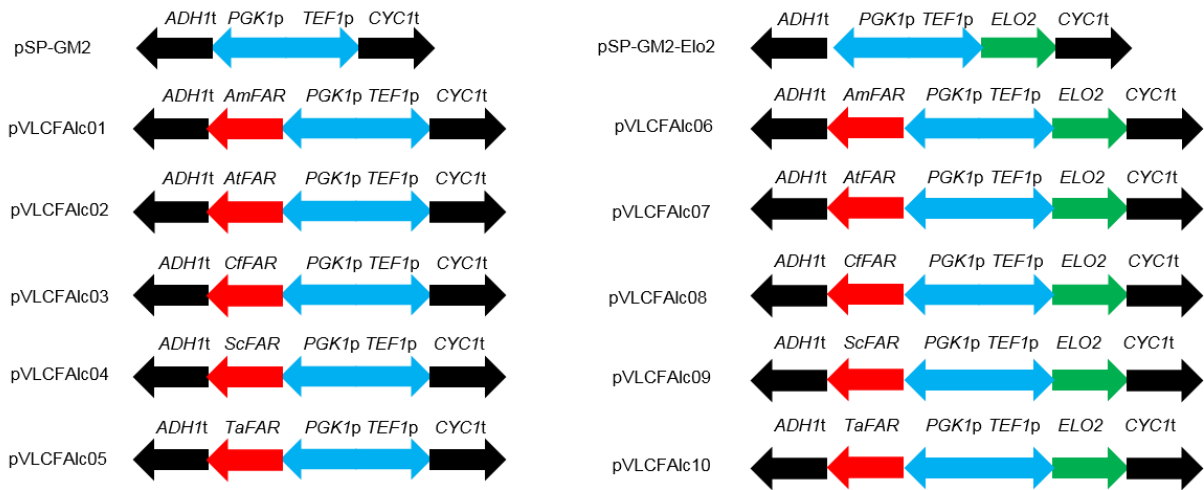
AcpS (134aa)

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53 Supplementary Fig. 6. Schematic alignment of *MvFAS* and *ScFAS* drawn to scale (modified from¹). The
54 major domains of the two FASs are labelled. The PPT domain of *MvFAS* is encoded by gene *acps*. *S.*
55 *cerevisiae* (*Saccharomyces cerevisiae*); *M. vaccae* (*Mycobacterium vaccae*); ACP (Acyl carrier protein);
56 KR (Ketoacyl reductase); KS (Ketoacyl synthase); PPT (Phosphopantetheinyl transferase); AT (Acetyl
57 transferase); ER (Enoyl reductase); DH (Dehydratase); MPT (Malonyl/palmitoyl transferase).

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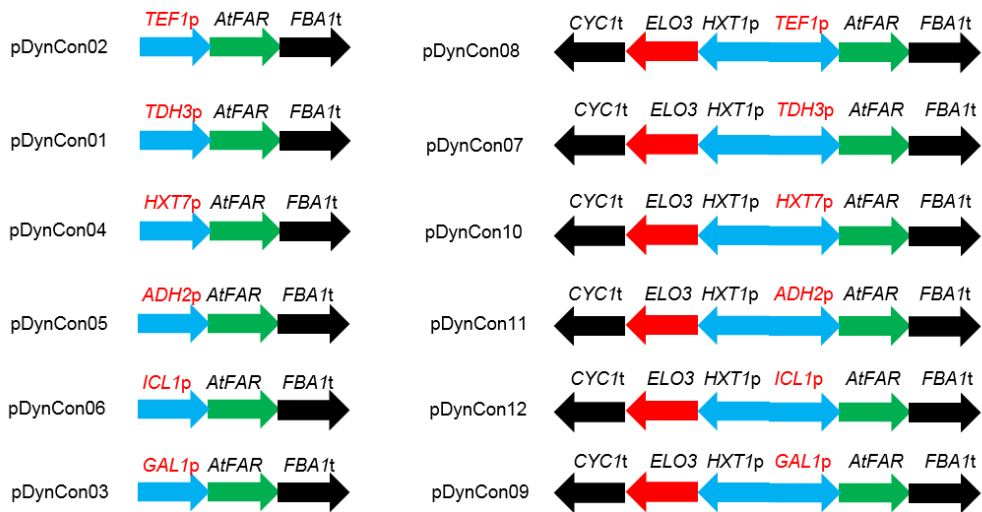


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61 Supplementary Fig. 7. Genetic constructs used in FAR screening. Graphical representation of the
 62 *amfar*, *atfar*, *cffar*, *scfar* and *tafar* gene constructs used for screening C₂₂ specifically FAR enzyme.
 63 The genetic sequences for *amfar*, *atfar*, *cffar*, *scfar* and *tafar* were codon optimized for expression in
 64 *S. cerevisiae* and synthesized by GenScript, Inc.

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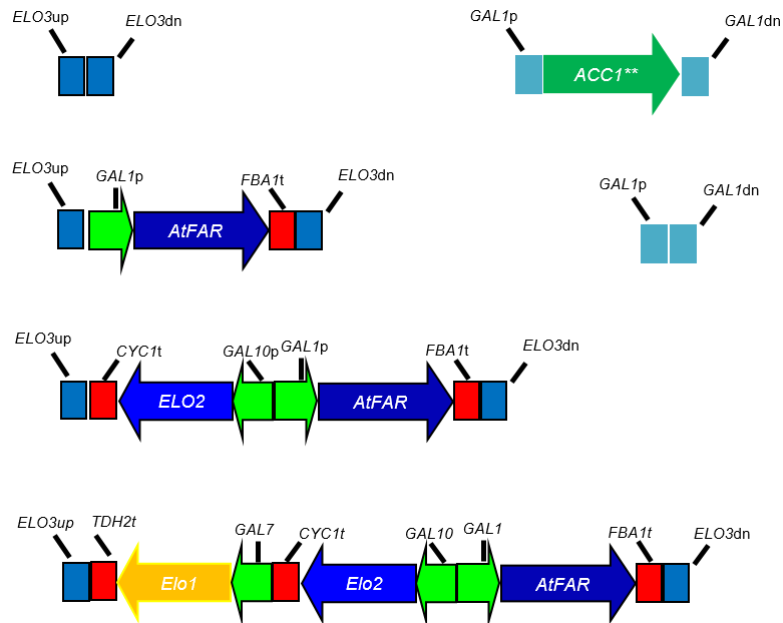
68 Supplementary Fig. 8. Genetic constructs used in promoter screening (dynamic control). Graphical
 69 representation of the *TEF1p*, *TDH3p*, *HXT7p*, *ADH2p*, *ICL1p* and *GAL1p* constructs used to screen for
 70 the best promoter combinations for the dynamic control system. All the promoters were amplified
 71 from *S. cerevisiae* CEN.PK113-5D genome DNA.

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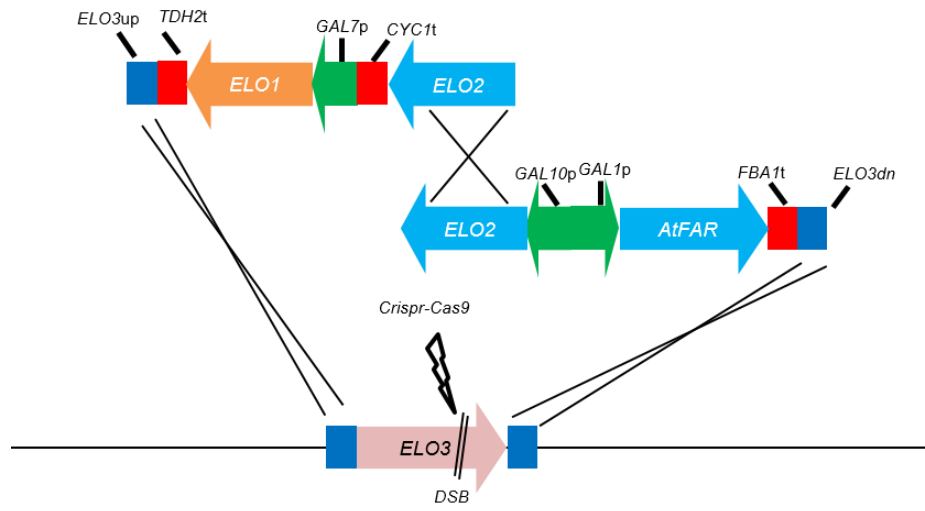
78 Supplementary Fig. 9. Schematic illustration summarizing the genetic arrangement of genomic
79 integration constructs of target pathway genes. All the promoters and terminators were amplified
80 from *S. cerevisiae* CEN.PK113-5D genome DNA.

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87 Supplementary Fig. 10. Schematic illustration of genome engineering strategy for gene deletion
88 coupled with gene integration via CRISPR/Cas9.

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91 Supplementary Table 1. Product specificity of FAS I system from different *Mycobacterium* species^{2, 3}

92	species	Pyrolytic cleavage products of a-mycolate		species	Pyrolytic cleavage products of a-mycolate	
93	aurum	C22:0		thermoresistibile	C22:0	C24:0
	austroafricanum	C22:0		tokaiense	C22:0	C24:0
94	brumane	C22:0		ulcerans	C22:0	C24:0
	diernhoferi	C22:0		vaccae	C22:0	C24:0
95	fallax	C22:0		absessus		C24:0
	gilvum	C22:0		agri		C24:0
96	neoaurum	C22:0		agri		C24:0
	parafortuitum	C22:0		asiaticum		C24:0
97	alvei	C22:0	C24:0	avium subsp. Avium		C24:0
				avium subsp.		
98	chitae	C22:0	C24:0	Paratuberculosis		C24:0
	chubuense	C22:0	C24:0	chelonae		C24:0
99	confluentis	C22:0	C24:0	farcinogenes		C24:0
	cookii	C22:0	C24:0	fortuitum		C24:0
100	duvalii	C22:0	C24:0	kansasii		C24:0
	flavescens	C22:0	C24:0	komossense		C24:0
101	gadium	C22:0	C24:0	lepramurim		C24:0
	gastri	C22:0	C24:0	marinum		C24:0
102	gilvum	C22:0	C24:0	phlei		C24:0
	gordonae	C22:0	C24:0	shimoidei		C24:0
103	haemophilum	C22:0	C24:0	simiae		C24:0
	hibernae	C22:0	C24:0	szulgai		C24:0
104	intermedium	C22:0	C24:0	terrae		C24:0
	intracellulare	C22:0	C24:0	thermoresistibile lab		C24:0
105	leprae	C22:0	C24:0	triviale		C24:0
	mucogenicum	C22:0	C24:0	celatum		C24:0
106	nonchromogenicum	C22:0	C24:0	interjectum		C24:0
	obuense	C22:0	C24:0	malmoense		C24:0
107	pulveris	C22:0	C24:0	microti		C24:0
	rhodesiae	C22:0	C24:0	tuberculosis		C24:0
108	rhodesiae	C22:0	C24:0	africanum		C26:0
	scrofulaceum	C22:0	C24:0	bovis		C26:0
109	smegmatis	C22:0	C24:0	bovis BCG		C26:0
	sphagni	C22:0	C24:0	xenopi		C26:0

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112 Supplementary Table 2. Fatty acyl-CoA reductase screening in different background strains

FARs	JV03 <i>elo3</i> Δ strain					control	JV03 <i>elo3</i> Δ pELO2 strain				
	<i>AmFAR</i>	<i>AtFAR</i>	<i>CfFAR</i>	<i>ScFAR</i>	<i>TaFAR</i>		<i>AmFAR</i>	<i>AtFAR</i>	<i>CfFAR</i>	<i>ScFAR</i>	<i>TaFAR</i>
OD ₆₀₀	3.20	NS*	4.34	5.29	0.53	5.41	3.15	0.29	3.13	3.47	3.46
Docosanol(mg/L)	ND [†]	ND	ND	ND	ND	ND	ND	1.13	ND	ND	ND
Strain name	TY006	TY007	TY008	TY009	TY010	TY001	TY011	TY012	TY013	TY014	TY015

113 ND*: Not detected; NS[†]: No survival colony

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117 Supplementary Table 3. Evaluation of different designs for dynamic control strategies

Promoters	JV03 <i>elo3Δ</i>						control	JV03 <i>elo3Δ</i> <i>HXT1p-ELO3</i>					
	<i>TDH3p</i>	<i>TEF1p</i>	<i>GAL1p</i>	<i>HXT7p</i>	<i>ADH2p</i>	<i>ICL1p</i>		<i>TDH3p</i>	<i>TEF1p</i>	<i>GAL1p</i>	<i>HXT7p</i>	<i>ADH2p</i>	<i>ICL1p</i>
OD ₆₀₀	0	0	6.59±0.19	0	0	0	8.71±0.38	7.86±0.20	7.7±0.28	8.45±0.25	8.11±0.47	7.21±1.4	8.43±0.12
Docosanol(mg/L)	0	0	4.19±0.20	0	0	0	0	0	0	0	0.80±0.07	0.15±0.02	0
Docosanol(mg/OD ₆₀₀)	0	0	12.85±0.55	0	0	0	0	0	0	0	2.09±0.19	0.54±0.04	0
Strains	TY016	TY017	TY018	TY019	TY020	TY021	TY001	TY022	TY023	TY024	TY025	TY026	TY027

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119 Supplementary Table 4. Primers used in this study

Primer	Description	Sequence(5'-3')
pTY001	Del1-F	GACAACCCTTAATTACCGTTCGTATAATG
pTY002	Del1-R	CAACCTATTAATTTCCCCTCGTCA
pTY003	Del2-F	GGCAAAACAGCATTCCAGGT GGATCTGATATACCGTTCGTATAGCATAACATTATACGAAGTTATTCATA TAGGGCGAATTGGGTAC
pTY004	Del2-R	AGCAGAATTATGAATAATAAACATTGATGACAATAG
pTY005	Elo3up-F	ACATTATACGAACGTAATTAAGGGTTGTCGACTGTTTATGTTTCGTAA
pTY006	Elo3up-R	ACAAACGG TGTATGCTATACGAACGGTATATCAGATCC ATA GGA AGC GAG AAT
pTY007	Elo3dw-F Primer	TTT TGA CAG CG
pTY008	Elo3dw-R Primer	TAGATAATGATGCTGGGCCGATAATTAG
pTY009	Chromosome XII before Elo3JD-F	TGCGTCGTTAGATTTCTGGGGTTAAGATAGCAAC
pTY010	Chromosome XII behind Elo3JD-R	TATTGGAGAGCTAGAACAACCAATGAACTTCATTGAAGAG
pTY011	p-vectorBamH1-R	TGTCATTGTTTTGTGAGTCGTATTACGGATCC
pTY012	p-vectorSma1fusion-F	TTGCAGTATTGGAAGAAAGATAAGCGTCGACATGGAACAGAAG
pTY013	P-MvFAS2-Gibson-F	GGATCCGTAATACGACTCACAAAACAATGACAATCAACGATCAACATAG
pTY014	P-MvFAS2-Gibson-R	TGTATGATCAGCGACCTTTTCTGGGTCCTTCAACGGAGA
pTY015	P-MvFAS3-Gibson-F	TCTCCGTTGAATGGGACCCAGAAAAGGTCGCTGATCATACA
pTY016	P-MvFAS3-Gibson-R	GCAATACTACCTTTGGAGGCACCTGTGACTACTGCAACTTCATCA
pTY017	P-MvFAS4-Gibson-F	TGATGAAGTTGCAGTAGTCACAGGTGCCTCCAAAGGTAGTATTGC
pTY018	P-MvFAS4-AcpS-R	CTGAACCGCCACCACCTGATCTTCTGTAAACACCATTTTTAC TCAGGTGGTGGCGGTTTCAGGCGGAGGTGGCTCTGGCGGTGGCGGATC GATGTTGCCAGTCGGTATAGTT
pTY019	AcpSfusion-F	CTTCTGTTCCATGTCGACGCTTATCTTTCTTCCAATACTGCAA
pTY020	AcpSfusion-R	ATCATTATCAATACTGCCATTTT
pTY021	GPD-F	CTATGTTGATCGTTGATTGTCATTAGTTCTAGATCCGTCGAAAC
pTY022	GPD-R	GTTTCGACGGATCTAGAATAATGACAATCAACGATCAACATAG
pTY023	MvFAS-fragment2-F	CATAGCAATCTAATCTAAGTTTTAATTACAAGGATCAAACAATGAATTC ACTCGTTACTCAATA
pTY024	ELO2-F	TCGACGCCCCGGCCCTATAGTGAGTCGTATTACGGATCTTACCTTTTTCT
pTY025	ELO2-R	TCTGTGTTGAG ATCTTATCGTCGTCATCCTTGTAATCCATCGATACTAGTTTACATGTATCT
pTY026	CfFAR-F	TCTTGAACCTG TCTACTTTTTACAACAAATATAAAACAAGCGGCCGCAAACAATGGAAG
pTY027	CfFAR-R	TTAACAACAACCTCAC ATCTTATCGTCGTCATCCTTGTAATCCATCGATACTAGTTTATTGTTTCAA
pTY028	AT5g22500-F	AACATGTGTGAT TCTACTTTTTACAACAAATATAAAACAAGCGGCCGCAAACAATGGAAT
pTY029	AT5g22500-R	CTAATTGTGTTCAATTC ATCTTATCGTCGTCATCCTTGTAATCCATCGATACTAGTGTTCATACAT
pTY030	ScFAR-F	GTTCAACGACA

pTY031	ScFAR-R	TCTACTTTTTACAACAAATATAAAACAAGCGGCCGCAAACAATGGAAG AAATGGGTTCAATCT
pTY032	TaFAR-F	ATCTTATCGTCGTCATCCTTGTAATCCATCGATACTAGTTTTCAACATGTA TTTTATGACACC
pTY033	TaFAR-R	TCTACTTTTTACAACAAATATAAAACAAGCGGCCGCAAACAATGGTTG ATACATTGTCTGAAG
pTY034	AmFAR-F	ATCTTATCGTCGTCATCCTTGTAATCCATCGATACTAGTAATTTGATAGT AAGACGTGTAAG
pTY035	AmFAR-R	TCTACTTTTTACAACAAATATAAAACAAGCGGCCGCAAACAATGTCAA CTATTTCCGATAATCAATG
pTY036	Kpn1ADH2p-F	CAGTGAGCGCGCGTAATACGACTCACTATAGGGCGAATTGGCAAACG TAGGGGCAAACA
pTY037	HXT1ADH2p-F	TAGTCGGTGCATATGAATTGAAGTTTCATTGTGGCGCAAACGTAGGG GCAAACA
pTY038	ADH2p-R	GTCTTATTACCCAAGAATTGAACACAATTAGATTCCATTGTTTTGTGTA TTACGATATAGTTAATAG
pTY039	Kpn1ICL1p-F	CAGTGAGCGCGCGTAATACGACTCACTATAGGGCGAATTGTGGAAATG TAAAGGATAATGAGTG
pTY040	HXT1ICL1p-F	TAGTCGGTGCATATGAATTGAAGTTTCATTGTGGCTGGAAATGTAAAGG ATAATGAGTG
pTY041	ICL1p-R	GTCTTATTACCCAAGAATTGAACACAATTAGATTCCATTGTTTTCGTTGA CTTTTTGTTATGTTATG
pTY042	Kpn1GAL1p-F	CAGTGAGCGCGCGTAATACGACTCACTATAGGGCGAATTGCGGATTAG AAGCCGCCGAG
pTY043	HXT1GAL1p-F	TAGTCGGTGCATATGAATTGAAGTTTCATTGTGGCCGATTAGAAGCCG CCGAG
pTY044	GAL1p-R	GTCTTATTACCCAAGAATTGAACACAATTAGATTCCATTGTTTTCTCCTTG ACGTTAAAGTATAGAG
pTY045	Kpn1TEF1p-F	CAGTGAGCGCGCGTAATACGACTCACTATAGGGCGAATTGATAGCTTCA AAATGTTTCTACTC
pTY046	HXT1TEF1p-F	TAGTCGGTGCATATGAATTGAAGTTTCATTGTGGCATAGCTTCAAATG TTTCTACTC
pTY047	Kpn1HXT7-F	CAGTGAGCGCGCGTAATACGACTCACTATAGGGCGAATTGAATAGTAC TCTCATCGCTAAGATC
pTY048	HXT1HXT7-F	TAGTCGGTGCATATGAATTGAAGTTTCATTGTGGCATAGCTTCAAATG TTTCTACTC
pTY049	Kpn1TDH3p-F	CAGTGAGCGCGCGTAATACGACTCACTATAGGGCGAATTGTCGAGTTTA TCATTATCAATACTG
pTY050	HXT1TDH3p-F	TAGTCGGTGCATATGAATTGAAGTTTCATTGTGGCATAGCTTCAAATG TTTCTACTC
pTY051	AtFAR-F	ATGGAATCTAATTGTGTTCAATTC
pTY052	AtFARFBA1-R	AAACTATATCAATTAATTTGAATTAACCTATTGTTTCAAACATGTGTGA TC
pTY053	5FBA1-F	GATCACACATGTTTTGAAACAATAAGTTAATTCAAATTAATTGATATAGT TT

pTY054	5FBA1-R	CAAGCGCGCAATTAACCCCTCACTAAAGGGAACAAAAGCTGAGTAAGCT ACTATGAAAGACTT
pTY055	Hind111Elo3-F	GACATAACTAATTACATGACTCGAGGTCGACGGTATCGATTTAAGCTTT CCTGGAAGAGAC
pTY056	Hind111Elo3-R	CAACTAGTTGATATACGTAAAATCAAACAATGAACACTACCACATCTA CTG
pTY057	3HXT1p-F	CAGTAGATGTGGTAGTGTTTCATTGTTTTGATTTTACGTATATCAACTAGT TG
pTY058	3HXT1p-R	GCCACAATGAAACTTCAATTCA
pTY059	HXT1TEF1p-F	TAGTCGGTCGATATGAATTGAAGTTTCATTGTGGCATAGCTTCAAATG TTTCTACTC
pTY060	TEF1p-R	GTCCTATTACCCAAGAATTGAACACAATTAGATTCCATTTTGTAATTA ACTTAGATTAG
pTY061	HXT1TDH3p-F	TAGTCGGTCGATATGAATTGAAGTTTCATTGTGGC
pTY062	TDH3-R	TCGAGTTTATCATTATCAATACTG
pTY063	HXT1HXT7-F	GTCCTATTACCCAAGAATTGAACACAATTAGATTCCATTTTGTTGTTTAT GTGTGTTTATTC
pTY064	HXT7-R	TAGTCGGTCGATATGAATTGAAGTTTCATTGTGGCAATAGTACTCTCATC GCTAAGATC
pTY065	Elo3repair-F	GTCCTATTACCCAAGAATTGAACACAATTAGATTCCATTTTTTGATTA ATTAATAAAAACT
pTY066	Elo3repair-R	AGCTTACTTCTAGTTTATTTATTCGGCTTTTTTCCGTTTGTTACGAAACA TAAACAGTCATAGGAAGCGAGAATTTTGACAGCGAATGAAAAAGAAA AAAATTAATGTTAACATTTT
pTY067	GAL1-gRNA1	AAAATGTTAACATTTAATTTTTTCTTTTTTATTTCGCTGTCAAAAATTCTC GCTTCCTATGACTGTTTATGTTTCGTAACAACCGAAAAAAGCCGAAT AAATAAACTAGAAGTAAGCT
pTY068	GAL1-gRNA2	TGCGCATGTTTCGGCGTTCGAAACTTCTCCGCAGTGAAAGATAAATGAT CAACCAACTATAATTTAAGAGGTTTTAGAGCTAGAAATAGCAAGTTAAA ATAAGGCTAGTCCGTTATCAAC
pTY069	ELO3-gRNA1	TGCGCATGTTTCGGCGTTCGAAACTTCTCCGCAGTGAAAGATAAATGAT CTACATCCAATCTTACAAGAAGTTTTAGAGCTAGAAATAGCAAGTTAAA ATAAGGCTAGTCCGTTATCAAC
pTY070	ELO3-gRNA2	TGCGCATGTTTCGGCGTTCGAAACTTCTCCGCAGTGAAAGATAAATGAT CTACTCGTTATTATTGCGTATTTTGTGATGCTAAAGTTATGAGTAGAAAA AAATGAGAAGTTGTTCTGAACAAAGTAAAAAAGAAGTATACTTATTT CAAAGTCTTCAACAATTTTTC
pTY071	ACC1repair-F	TTTCGGTTTGTATTACTTCTTATTCAAATGTCATAAAAAGTATCAACAAAA AATTGTTAATATACTCTATACTTTAACGTCAAGGAGAAAAAAGTATAAT GAGCGAAGAAAGCTTATTCG
pTY072	ACC1repair-R	GAGCGAAGAAAGCTTATTCG
pTY073	ACC1GAL1-upJDF	ACAAAGGGTTCTCGTAGAGTC

pTY074	ACC1GAL1-upJDR	AGGAACTGGAATGGACCGAG
pTY075	ACC1-DWjdf	CTAGTTTATCTACTGTATTGAGG
pTY076	Elo3 JDF	GTACTIONTACGAGGATTATCG
pTY077	Elo3 JDR	GCAATTTTATTAGAGATAGTGG
pTY078	ELO3up-F	CTGGGGTTAAGATAGCAACG CGCTCGAAGGCTTTAATTTGCGGCCGGTACCCGACTGTTTATGTTTCGTA AACAAAC
pTY079	ELO3up-R	AACAAAC
pTY080	CYC1-F	GATACCGTCGACCTCGAGTC
pTY081	CYC1-R	GGGTACCGGCCGCAAATTAAG CTAATTACATGACTCGAGGTCGACGGTATC
pTY082	Elo2-F	TTACCTTTTTCTTCTGTGTTG CAAAAAAAAAAGTAAGAATTTTTGAAAATTCAATATAAAAAACAATGAAT TCACTCGTTACTC
pTY083	Elo2-R	TCACTCGTTACTC
pTY084	Gal1andGal10-F	TTATATTGAATTTTCAAAAATT
pTY085	Gal1andGal10-R	TATAGTTTTTCTCCTTGACGT CTATACTTTAACGTCAAGGAGAAAAACTATAAAAAACAATGGAATCTAA TTGTGTTCAATTC
pTY086	AtFAR-F	CTCATTAAAAAACTATATCAATTAATTTGAATTAACCTATTGTTTCAAAAC ATGTGTGATC
pTY087	AtFAR-R	ATGTGTGATC
pTY088	FBA1-F	GTTAATTCAAATTAATTGATATAG
pTY089	FBA1-R	AGTAAGCTACTATGAAAGACTTTAC CGGCTTTTTCCGTTTGTTTACGAAACATAAACAGTC
pTY090	Elo3updown-F	ATAGGAAGCGAGAATTTTTGAC CTTCGAGTTCTTTGTAAAGCTTTTCATAGTAGCTTACTATAGGAAGCGAG AATTTTTGAC
pTY091	Elo3down-F	AATTTTTGAC
pTY092	Elo3down-R	AGTTACTTGATATCCATGTTCC CTTAGTATCACACTAATTGGCTTTTCGCGACTGTTTATGTTTCGTAAACA AAC
pTY093	Elo3up-TDH2-R	AAC
pTY094	Elo3up-Fxin	CTGGGGTTAAGATAGCAACG
pTY095	Elo3up-Rxin	GACTGTTTATGTTTCGTAAACAAAC GCTTTTTTCCGTTTGTTTACGAAACATAAACAGTCCGGATTAGAAGCCGC CGAG
pTY096	Elo3upGAL1p-F	CGAG
pTY097	GAL1p-R	TATAGTTTTTCTCCTTGACGTT GTGCATATTTTCAAGAAGGATAGTAAGCTGGCAAACGGATTAGAAGCC GCCGAG
pTY098	GAL7GAL1p-F	GCCGAG CGCTCGAAGGCTTTAATTTGCGGCCGGTACCCGACTGTTTATGTTTCGTA AACAAAC
pTY099	ELO3upcyc-R	AACAAAC
pTY100	GAL7-F	TTTTGAGGGAATATTCAACTGTT CGCTCGAAGGCTTTAATTTGCGGCCGGTACCCTTGCCAGCTTACTATCC TTC
pTY101	GAL7 CYC1-R	TTC
pTY102	primer 6005	GATCATTTATCTTTCCTGCGGAGAAG
pTY103	TDH2t-F	ATTTAACTCCTTAAGTTACTTTAATGATTTAG
pTY104	TDH2t-R	GCGAAAAGCCAATTAGTGTGATAC CTAAATCATTAAAGTAACTTAAGGAGTTAAATTTAATTGTTTTGTTGAT CTTCTTC
pTY105	Elo1-F	CTTCTTC

pTY106 Elo1-R GATAAAAAAAAAACAGTTGAATATTCCTCAAAAAAAAAACAATGGTAAGTG
ATTGGAAAAATTTTG

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122 Supplementary Table 5 *S. cerevisiae* strains used in this study.

Strain	Genotype and plasmid	Source
JV03	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ</i>	Ref ⁴
TY001	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ</i>	This work
TY002	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pELO2</i>	This work
TY003	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ/pELO3</i>	This work
TDY7005	Mata <i>lys2 ura3-52 trp1Δ leu2Δ elo2Δ::KAN elo3Δ::TRP1/pELO3</i>	Ref ⁵
TY004	Mata <i>lys2 ura3-52 trp1Δ leu2Δ elo2Δ::KAN elo3Δ::TRP1/pGPD415-MvFAS-AcpS</i>	This work
PWY12	MATα <i>ura3Δ leu2Δ his3Δ trp1Δ can1Δ fas1Δ::HIS3 fas2Δ::LEU2</i>	Ref ⁶
TY005	MATα <i>ura3Δ leu2Δ his3Δ trp1Δ can1Δ fas1Δ::HIS3 fas2Δ::LEU2/pSPGM2-MvFAS-AcpS</i>	This work
TY006	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pVLCFAlc01</i>	This work
TY007	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pVLCFAlc02</i>	This work
TY008	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pVLCFAlc03</i>	This work
TY009	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pVLCFAlc04</i>	This work
TY010	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pVLCFAlc05</i>	This work
TY011	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pVLCFAlc06</i>	This work
TY012	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pVLCFAlc07</i>	This work
TY013	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pVLCFAlc08</i>	This work
TY014	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pVLCFAlc09</i>	This work
TY015	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pVLCFAlc10</i>	This work
TY016	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pDynCon01</i>	This work
TY017	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pDynCon02</i>	This work
TY018	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pDynCon03</i>	This work
TY019	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pDynCon04</i>	This work
TY020	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pDynCon05</i>	This work
TY021	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ/pDynCon06</i>	This work

TY022	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ</i> /pDynCon07	This work
TY023	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ</i> /pDynCon08	This work
TY024	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ</i> /pDynCon09	This work
TY025	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ</i> /pDynCon10	This work
TY026	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ</i> /pDynCon11	This work
TY027	MATa MAL2-8c <i>SUC2 ura3-52 HIS3 are1Δ dga1Δ are2Δ lro1Δ pox1Δ elo3Δ</i> /pDynCon12	This work
IMX581	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3</i>	Ref ⁷
TY028	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 elo3Δ::AtFAR</i> /pROS10-gRNA:elo3	This work
TY029	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 elo3Δ::AtFAR ELO2</i> /pROS10-gRNA:elo3	This work
TY030	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 elo3Δ::AtFAR ELO2 ELO1</i> /pROS10-gRNA:elo3	This work
TY031	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 gal1Δ::ACC1**</i> /pROS13-gRNA:gal1	This work
TY032	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 gal1Δ::ACC1** elo3Δ</i> /pROS10-gRNA:elo3	This work
TY033	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 gal1Δ::ACC1** elo3Δ::AtFAR</i> /pROS10-gRNA:elo3	This work
TY034	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 gal1Δ::ACC1** elo3Δ::AtFAR ELO2</i> /pROS10-gRNA:elo3	This work
TY035	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 gal1Δ::ACC1** elo3Δ::AtFAR ELO2 ELO1</i> /pROS10-gRNA:elo3	This work
TY036	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 gal1Δ::ACC1** elo3Δ::AtFAR</i> /pSPGM2-MvFAS-AcpS	This work
TY037	MATa <i>ura3-52 can1Δ::cas9-natNT2 TRP1 LEU2 HIS3 gal1Δ::ACC1** elo3Δ::AtFAR ELO2 ELO1</i> /pSPGM2-MvFAS-AcpS	This work

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124 Note: For PWY12, in detail, for the $\Delta fas1::HIS3$ disruption, about 1.2 kb of the *FAS1* upstream region
125 together with 1.4 kb of the N-terminal part of the *FAS1* reading frame were replaced by the *HIS3*
126 insertion marker. Similarly, 0.9kb of the *FAS2* upstream region together with 1.3 kb of the N-terminal
127 part of the *FAS2* reading frame were replaced by the *LEU2* insertion giving the $\Delta fas2::LEU2$ construct.

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Synthesized genes	Sequence (5'-3')
<i>MvFAS</i>	<p>ATGACAATCAACGATCAACATAGAGCCGCCGCTGGTAGTGACTCAAGAAGTGGTCAAGACTT CGAACCTTTATCTGGTACACACGCCTTGGTAGATAGATTACATGCTGGTGAACCATACGCTGT CGCATTGGTGGTCAAGGTGGTCCTTGGTTAGAAAATTTGGAAGAATTAGTAACTCAGCTGG TATCGAATCTGAAATTCACAATTGGTAGCCGAAGCTGAATTAATGTTGGAACCATTGGCTAG AGAATTAGTTGTAGTCAGACCAATTGTTTCGAACCTATGAAGTGGATAAGAGCATTAGCTGC TGATGAACCATTGCCTGCCGCTAAAGATTTGACTACAGCAGCCATATCTGGTCCAGGTATCTT GTTAACACAAATGGCTGCACAAAGAGCTTTGAAGAGACAAGGTTTAGATTTGGACGCACACC CACCTGTTGCAATTGCCGGTCATTCACAAGGTGTAAGTCTGTCGAATCCTAAAAGCCGGTG GTGCTAGAGATGTTGAATTGTTAGCAATAGGTCAATTGATCGGTGCCGCTGGTTCCTTAGTTA GTAGAAGATGTGGTATGGTTGGTAGAGGTGACAGATCCCCTATGGTTAGTGTATTGAATGTT GACCCAGCAAGAATGGCCGAATTGTTAGACGAATTTGCTCAAGATGTTAGAAGTGTATTGCCA CCTGCATTATCAATCAGAAACGGTAGAAGATCCGTTGTAATTACTGGTACACCAGAACAATTA GCTAGATTCGAATTGTATTGCGAAAAGATTACCGAAAAGGAAGAAGCTGAAAGAAAAGAATAA GACTAGAGGTGGTGAATTTTACCAATATTCAATCAATTGAACGTGCAAGTTGGTTTTCA TACACCTAGATTGGCTGGTGGTATTGATTTGGTTAACGAATGGGCAGCCAGAAGTGGTTGG ATAGAGAATTAAGTAGAGAATTGACCGAAACTATCTTCGTAAGCCAGTTGATTGGGTATCAG AAGTCGAAGGTTTGGCTGCTGCTGGTGTAAATGGATTGTTGACTTAGTCCATCCGATACAG TCACCAGATTGACTGCACCTGTTATAAGAGGTTTAGGTATTGGTATAGTTGCTGCTGCTACAA GAGCTGGTCAAAGATCATTGTTTACTGTAGGTGCAGAACCAGCTGTCGCACCTGCCCTGGTCTT CTTATGCTCCAAAGCCTGTAGCCTTGCCAGATGGTTCTGTCAAAGCATCTACTAAGTTCACTAG ATTAACAGGTAGATCACCTATCTTGTTAGCTGGTATGACACCAACCACTGTCGATGCTAAAATT GTTGCTGCTGCTGCTAATGCAGGTCACTGGGCTGAATTGGCTGGTGGTCAAGTTACCGA AGAAATTTTTGATGCCAGAATCGCTGAATTAAGTCAATTGTTAGAACCAGGTAGAGCTGTTCA ATCAATAGTTTGTGTTTATGATCCATACTTGTGGAAGTTACAATTGGGTGGTAAAAGATTAGTT CAAAGGCTAGACAATCTGGTGCACCAATAGATGGTGTGCTGTTGTAAGTCTGGTATCCCTGAC TTAGAAGAAGCAGTTGATTGATCGAAGAATTGCATACAGTAGGTATATCAAACGTCGTTTTT AAACCAGGTACTGTTGATCAAATTAATCCGTAATCAAGATTGCAGCCGAAGTTCCTGGTAGA GATGTAATAGTCCACGTTGAAGGTGGTAGAGCTGGTGGTCATCACTCTGGGAAGATTTGGA TGACTTGTATTGTCTACTTATGGTGACTTGAGAAAGCATTCCAACATAACTATCTGTGTTGGT GGTGGTATAGGTACACCAGAAAGAGCTGCTGAATATTTGTCTGGTACTGGGCCAAAACCTTA CGGTTTTCCAGCTATGCCTGTAGATGGTATCTTGGTCGGTACTGCTGCTATGGCTACTTTAGAA GCCACAACCTCTCCAGCTGTTAAGCAATTATTGGTAGACACTACAGGTACTGATACATGGGTT GGTGTGTTAAAGCCATTTCTGGTATGGCTTCTGGTAGATCACAATTAGGTGCAGACATCCAT GAAATTGATAATACTGCTTCCAGATGCGGTAGATTATTGGATGAAGTTGCAGGTGACGCTGA CGCAGTAGCCGAAAGAAGAGACGAAATTATAGCAGCTATGGCTGATACTGCAAAACCACTACT TTGGTGACGTCGCTGATATGACATATTTGCAATGGTTACAAAGATACGTTGAATTGGCTATTG GTGACGGTGACAGTACCGCAGATACTGCTGCACCAGGTTCTCCTTGGTTAGCTGATACATGGA GAGAAAGATTTGAAGAAATGTTGACCAGAGCTGAAGCAAGATTACATGAAAAGGATAGTGG TCCTATCGAATCTTTGTTCCGCCGCTGGTCCAGAATCACAAGCATTGTTGGATAATCCTGATACA GCAATTGCAGCCTTATTGGCCAGATATCCAGATGCTGAAACAGTTAAATTGCACCCAGCTGAC GTCCATTTTTCGTTACCTTATGTAAAAAGCCAGGTAAACCTGTAAACTTTGTACCAGTCATTG ATAAGGACGTCAGAAGATGGTGGAGATCAGATTCATTATGGCAAGCCCATGACGCTAGATAC ACTGCTGATCAAGTCTGCATCATTCCAGGTACACAAGCCGTTGCTGGTATTACCAGAGTAGAC GAACCTGTCGGTGAATTATTGGATAGATTCAACAAGAAATCGTTGACAGAATTTGGCAACA</p>

GGTGCCGAACCAGTACCTGTAGTCAGTAGAAGACAAGCAAGAGCCGATGTAGCTGGTCCTT
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GAAGAAGAAGAAGAGATGTTGTAGTCAACGCACCTGTTGACATGTCAGCCTTCGCTGTTGTAT
CCGGTGACCACAATCCAATACATACTGATAGAGCCGCTGCATTATTGGCTGGTTGAAAATGC
CTATCGTTCACGGTATGTGGTTATCTGCCGCTGCACAACATGCAGTTACCGCCACTGATGGTA
GAGCTACCCACCTGCAAGATTGGTTGGTTGGACTTCAAGATTTTTGGGTATGGTATTGCCAG
GTGACGAAATTGAGTTTAGAGTAGACAGAGTTGGTATCGATAGAGGTGCTGAAATTGTTGAA
GTAGCCGCTAAAGTAGGTGGTGAATTGGTCATGTCTGCAACTGCCCAATTAGCAGCCCCTAAA
ACAGTTTATGCTTTCCAGGTCAAGGTATTCAATCAAAGGGTATGGGTATGGATGTCAGAGCA
AGATCCAAGGCTGCAAGAGCTGTTTGGGACACAGCAGATAAGTTTACTAGAGAAACCTTAGG
TTTCTCTGTTTTGCACGTCGTTAGAGATAACCCAAGTTTAAAGCTTCTGGTGTCCATTATC
ATCACCTGAAGGTGTTTTATACTTGACTCAATCACTCAAGTTGCTATGGCAACAGTAGCCGC
TGCACAAGTCGCCGAAATGAGAGAACAAGGTCTTTCGTAGAAGGTGCCATTGCTTGTGGTC
ATAGTGTGCGGTGAATATACTGCATTGGCCTGCGTTTCTGGTGTATACGAATTACCAGCTTTATT
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GAAGATGCAACCTCTGCCCCAGCTGATGACGCTGAAGATGACGAAGCTGCTGATCATTGATT
GCCGCTTTGCCATCACCACTAGACCTGCTGTTACAGCACCAGCCCTGAATGGGCAGATTTG
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AGTAGAACTAGATACGAAATGGAAGTCGATAATGAATTGTCTGCTGCTGGTGTGTTTGAATTG
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CTAACAGATCATTGGACTGCGTTGATGACGAATTAGCCACTTCCGCTCATTTTGTGGCCAAAG
AGAAACATTAAGATTGGGTGACAAATATCCTTTGAAGGCTGGTTTAGTTACAAGTTTGGGTTT
TGGTCACGTTTCTGGTTTGATTGCCTTAGTACATCCACAAGCATTTTTGGCCGCTTTAGCACCT
GAACAAAGAGCAGCCTACACTGCACAAGCCCAAGAAAGAGCTTTGGCAGGTCAAAGAAGATT
AGCCTCTGCTATTGCAGGTGGTAGACCAATGTTTAAAAGACCTGCTGATAGAAGATTTCGACG
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GAAAATGGTGTTCACAGAAGATGA

AcpS ATGTTGCCAGTCGGTATAGTTGGTATTGGTATTGATTTGGTTTCTATCCCAGAATTTGCAGAAC
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CTGATAAATCTTCATCCGCCGCTAGACATTTGGCAGCCAGATGGGCTGCAAAAAGAAGCAGTA
ATTAAGGCCTGGAGTGGTTCAAGATTCGCCAAGAGACCAGTCTTACCTGAAGCTATCCATAGA
GATATCGAAGTTATTACAGACATGTGGGGTAGACCAAGAGTTAGATTGTCAGGTGCCGTAGC
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AtFAR ATGGAATCTAATTGTGTTCAATTCTTGGGTAATAAGACTATCTTGATCACAGGTGCACCAGGTT
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TCGAAGGTTTGAAGACATTGGATTGATTGTTGCTTATGGTAAAGGTAGATTGAAGTGTT
TCTTGGCTGATTCTAATTCAGTTTTCGATTTGATCCCAGCAGATATGGTTGTTAATGCTATGGT
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ATGAGATTGGTTGAATTGTACCAACCATACTTATTGTTTAAAGGTATTTTTGATGATTTGAATA
CTGAAAGATTGAGAATGAAGAGAAAAGGAAAACATCAAAGAATTGGATGGTTCTTTTGAATTT
GATCCAAAGTCAATCGATTGGGATAACTACATCACTAACACACATATCCCAGGTTTATCACA
CATGTTTTGAAACAATAA

AmFAR ATGTCAACTATTTCCGATAATCAATGCACTTCCGTCAGAGACTTTTACAAGGACAGATCCATAT
TCATCACTGGTGGTACAGGTTTCATGGGTAAAGTCTTGGTAGAAAAGTTGTTAAGATCCTGTC
CAGGTATTAATAATATCTATATCTTGATGAGACCTAAAAAGAGTCAAGATATACAACAAAGAT
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TGAAGATCATCCCTATTGCTGGTGACGTTACCGAACATGAATTAGGTATCTCTGAAGCTGATC
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CfFAR

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ScFAR

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TaFAR

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AAACTATAGATTGGGACGAATACTTCTACAGAGTCCACATACCAAGGTGTCATAAAAATACATGT
TGAAATAA

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138 Supplementary Table 7. Plasmids used in this study.

Plasmids	Genotype or characteristics	Base plasmid	Source
pSPGM2	pSPGM2	pSPGM2	R&D systems
pELO2	pSPGM2-ELO2	pSPGM2	This work
pELO3	pRS316-ELO3	pRS316	Ref ⁸
p415GPD	p415GPD	p415GPD	R&D systems
pGPD415-MvFAS-AcpS	pGPD415-MvFAS-AcpS	p415GPD	This work
pSPGM2-MvFAS-AcpS	pSPGM2-MvFAS-AcpS	pSPGM2	This work
pVLCFAIc01	pSPGM2-AmFAR	pSPGM2	This work
pVLCFAIc02	pSPGM2-AtFAR	pSPGM2	This work
pVLCFAIc03	pSPGM2-CfFAR	pSPGM2	This work
pVLCFAIc04	pSPGM2-ScFAR	pSPGM2	This work
pVLCFAIc05	pSPGM2-TaFAR	pSPGM2	This work
pVLCFAIc06	pSPGM2-AmFAR-Elo2	pSPGM2-ELO2	This work
pVLCFAIc07	pSPGM2-AtFAR-Elo2	pSPGM2-ELO2	This work
pVLCFAIc08	pSPGM2-CfFAR-Elo2	pSPGM2-ELO2	This work
pVLCFAIc09	pSPGM2-ScFAR-Elo2	pSPGM2-ELO2	This work
pVLCFAIc10	pSPGM2-TaFAR-Elo2	pSPGM2-ELO2	This work
p416GPD	p416GPD	p416GPD	R&D systems
pDynCon01	p416TDH3p-AtFAR	p416GPD	This work
pDynCon02	p416TEF1p-AtFAR	p416GPD	This work
pDynCon03	p416GAL1p-AtFAR	p416GPD	This work
pDynCon04	p416HXT7p-AtFAR	p416GPD	This work
pDynCon05	p416ADH2p-AtFAR	p416GPD	This work
pDynCon06	p416ICL1p-AtFAR	p416GPD	This work
pDynCon07	p416-ELO3-HXT1p-TDH3p-AtFAR	p416GPD	This work
pDynCon08	p416-ELO3-HXT1p-TEF1p-AtFAR	p416GPD	This work
pDynCon09	p416-ELO3-HXT1p-GAL1p-AtFAR	p416GPD	This work
pDynCon10	p416-ELO3-HXT1p-HXT7p-AtFAR	p416GPD	This work
pDynCon11	p416-ELO3-HXT1p-ADH2p-AtFAR	p416GPD	This work
pDynCon12	p416-ELO3-HXT1p-ICL1p-AtFAR	p416GPD	This work
pROS10	pROS10	pROS10	Ref ⁷
pROS10-gRNA:elo3	pROS10-gRNA:elo3	pROS10	This work
pROS13	pROS13	pROS13	Ref ⁷
pROS13-gRNA:gal1	pROS13-gRNA:gal1	pROS13	This work

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144 Supplementary Table 8 Overview Fatty Acyl Reductase (FARs).

Gene	Species	NCBI accession	Primary substrate specificity	Reference
<i>AmFAR</i>	<i>Apis mellifera</i>	CAD67815	C22:0 C24:0 C26:0	Ref ⁹
<i>AtFAR</i>	<i>Arabidopsis thaliana</i>	NP_197642.1	C22:0	Ref ⁹
<i>CfFAR</i>	<i>Calanus finmarchicus</i>	AEO89345.1	C20:0 C22:0 C24:0 C26:0	Ref ¹⁰
<i>ScFAR</i>	<i>Simmondsia chinensis</i>	AAD38039	C20:0 C22:1	Ref ⁹
<i>TaFAR</i>	<i>Triticum aestivum</i>	CAD30692.1	C22:0	Ref ¹¹

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