Cancer Type	# Samples with Mutation & Copy Number	# Samples with Mutation & Expression	
Acute Myeloid Leukemia	-	181	
Bladder	99	38	
Breast	746	698	
Colorectal	144	137	
Glioblastoma	122	109	
Head & Neck	291	248	
Lower Grade Glioma	208	205	
Lung Adeno	450	121	
Lung Squam	157	157	
Renal	388	354	
Ovarian	308	261	
Uterine	157	220	



b

С



Supplementary Figure 1. Primary Tumor Samples and Cancer Types Used in MiSL. (a) Summary of Samples in 12 TCGA Cancers Analyzed. (b) HI-HI Boolean implication (if amplification B is present, then mutation X is present) represents a subset relationship between the two attributes. (c) HI-LO Boolean implication (if deletion B is present, then mutation X is NOT present) represents a mutual exclusion relationship between the two attributes. (d) Distribution of number of MiSL candidates for each MiSL-targetable mutation in each cancer type. (e) Sensitivity analysis by varying the p-value threshold for differential expression for copy number filter (blue) and expression filter (red) in MiSL pipeline (dashed line shows current threshold). (f) Distribution of shared synthetic lethal predictions across different cancers. The green barplot shown the number of MiSL-targetable mutations that have at least one common MiSL candidate in different number of cancers. The red barplot shows the number of synthetic lethal interactions (i.e., a {mutation, MiSL candidate} pair) that are common across different numbers of cancers.

-	Cancer	# of	Evaluable Mutation	h_			
d		Evaluable Mutations		D Mutation		p-value	NES
	Breast 2		MAP3K1, PIK3CA		APC	0.003	1.55
	Colorectal	5	APC, CSMD3, KRAS,		CSMD3	> 0.1	-
			PIK3CA, TP53	KRAS PIK3CA		0.04	1.48
	Glioblastoma	1	PTEN			0.03	1.35
	Kidney	0	-	1	TP53	>0.1	-
	Leukemia	1	TP53		APC&KRAS	0.07	1.35
	Lung Squam	0	-				
	Ovarian	0	-				
С	× v score	COPE	[™] 0 POLR2E [№] 0 [№] 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0	SF3B5	° GI	PC1
	mary shRN.	0 0 0 0 0 0 0 0 0		9			
		+ Mutation	- + APC Mutation	- APC	+ Mutation	- + APC Mutation	l
a	A score	<i>KLK15</i>	° MARVELD2	o 0000	NQO1	° A	МН
		8	0 0 0	0	0 7 -	° °	
	ary sh	8 8 0		00	0 7 -	8 ē 0	
		•	° 8 - ▲			0	
	の KRAS Mutation		KRAS Mutation	KRA	S Mutation	KRAS Mutatic	n
•				7001	omatation		/11
e	° SCOLE	CDK16	[°] ₈ EPS8L1	0	LRSAM1	° NUC	CB1
	mary shRNA	0 9 - 8 0 9 - 7 -		0 80 80 80 8	- • • • • • • • • • • • • • • • • • • •		
	unities of the second s	+ A Mutation	 + PIK3CA Mutation 	- PIK3	T CA Mutation	- + PIK3CA Mutat	ion



shRNA shRNA

Non-SL

Hits

Supplementary Figure 2. Validation of MiSL. (a) Summary of Evaluable Mutations in Project Achilles for different cancers. (b) Summary of gene set enrichment analyses (normalized enrichment score and p-value) comparing enrichment of MiSL candidates in Achilles data for mutations in colorectal cancer. (c) Achilles shRNA summary scores for representative shRNAs in colorectal cell lines with APC mutation versus wild-type cell lines for 4 different MiSL candidates for APC mutation in colorectal cancer. (d) Achilles shRNA summary scores for representative shRNAs in colorectal cell lines with KRAS mutation versus wild-type cell lines for 4 different MiSL candidates for KRAS mutation in colorectal cancer. (e) Achilles shRNA summary scores for representative shRNAs in colorectal cell lines with PIK3CA mutation versus wild-type cell lines for 4 different MiSL candidates for PIK3CA mutation in colorectal cancer . (f) Contingency table showing overlap between MiSL candidates for IDH1 mutation in acute myeloid leukemia and shRNA synthetic lethals as per DECIPHER screen with a more stringent criterion (drop-out ratio of 0.6 instead of 0.8 for 2 or more shRNAs) for calling synthetic lethals in shRNA data. The overlap is statistically significant with the more stringent criterion for calling synthetic lethals. (g) MiSL analysis steps illustrated for VHL Mutation and GLS - (i) subset relationship between GLS gene amplification and VHL mutation (HI-HI Boolean implication), (ii) amplification of GLS is concordant with higher expression of the gene (p < 0.05), (iii) expression of GLS is higher in VHL-mutated kidney cancer (p < 0.05). (h) Cell viability of RCC4 and RCC4/VHL after siRNA treatment to GLS. All error bars represent the SEM (n = 3) (p = 0.024).



Adherens Junction

0

1

2

3

-log10(p-value)

4

5

6

BRCA1: FANCC,XRCC6

PIK3CA: AKT1, FOXO3, PSEN1

GATA3: EIF2B1,MSH3,RAD17

MAP3K1: ANAPC4,BTRC,CBL, CUL4A,CUL5,FZR1,HERC2, HERC3,HERC4,KLHL9,PIAS4, SKP1,STUB1,TCEB2,UBE2D3, UBE2G1,UBE2I

CDH1: PTPRM, SMAD2, SMAD4, TCF7L2, TGFBR1, NUP214, TSC1, XPO7, AP3B1, AP3M1, ARFIP1, BIN3, KIF13B, MXI1, SYNRG, TOPORS,GBF1, RAB14, RHOT1, SEC22C, STX18

Cancer Type	# of MiSL candidates	# of Recurrent MiSL
		Candidates
AML	601	21
Bladder	1842	0
Breast	1758	56
Colorectal	1321	33
Glioblastoma	335	10
Head & Neck	1924	4
Kidney	299	16
Lower grade glioma	1794	420
Lung adeno	2772	0
Lung squam	2425	0
Ovarian	534	20
Uterine	4481	225

Supplementary Figure 3. Pathway Analysis of MiSL Candidates for Recurrent Mutations and Recurrent MiSL Candidates. (a) Same pathway enrichment analyses using GO and KEGG gene sets for MiSL-targetable mutations in breast cancer. The x-axis shows the negative log of p-value of Fisher's exact test. MiSL candidates are said to be enriched in the same pathway as the mutated gene if (i) the MiSL candidates of the mutation are enriched for a pathway where overlap is measured using Fisher's exact test (ii) the mutation belongs to the same pathway. Only statistically significant results (p < 0.05) are reported. (b) Recurrent MiSL Candidates in the Different TCGA Cancers. A MiSL candidate is called recurrent in a cancer if it appears as a MiSL candidate for more than 5% of the recurrent mutations in the cancer.



Supplementary Figure 4. Drug Screen for MiSL candidates of IDH1 mutation in AML. (a) List of druggable MiSL candidates for the IDH1 mutation in AML. (b) Fraction IC_{50} for each drug tested in inducible IDH1 wildtype cells comparing (+ dox) to (- dox) as a control. (c) Table showing clinical characteristics of IDH1 mutant AML patient samples. (d) ACACA expression in sorted IDH1 mutant blasts (n=8) and IDH1 wildtype blasts (n=5) from AML patients compared to normal CD34+ cord blood determined by Taqman expression assay. Bars=SD of 4 replicates. (e) ACACA mRNA expression for 3 different shRNAs designed to target ACACA determined by real-time quantitative PCR in triplicate after transduction of THP-1 cells with shRNA lentivirus expressed in pRSI9 virus. shRNA #3 showed minimal knockdown. Error bars represent standard deviations, * p < 0.05, unpaired t-test. ns = non-significant. (f) Viable cell growth of THP-1 pTRIPZ IDH1 R132H-T2A-GFP in 0.5% serum transduced with shRNA #3 showing minimal defect in presence of IDH1 mutation (+dox). (g) Cell growth of parental THP-1 cells treated with 1 µg/ml doxycycline in 0.5% and 10% serum. (h) In-del mutation frequency reported by TIDE from genomic DNA extracted from IDH1 wildtype and R132H inducible THP-1 cells transduced with pLENTICRISPR v2 encoding guide RNA targeting exon 4 of ACACA. This guide was selected out of four gRNA sequences that were designed and tested in K562 cells. (i) Sequencing of ACACA exon 4 showing disruption of ACACA gene in both IDH1 wildtype and R132H THP-1 cell pools. (j) Cell sorting strategy after lentiviral CRISPR transduction showing RFP+ THP-1 cells marked lentiCRISPR integration and doxyxcyline-inducible GFP-T2A-IDH1 wildtype and R132H. (k) Western blot showing reduced ACACA protein expression after CRISPR cutting in pooled cells compared to untransduced IDH1 WT or R132H inducible THP-1 cells respectively. (1) Sequencing of Exon 4 of ACACA from the single cell-derived THP-1 clones showing in/dels. (m) Flow cytometry showing engraftment of RFP+, CD33+ AML blasts in NSG mice, gated on engrafted CD45+ cells nontargeting shRNA vs ACACA shRNA #1.

AML No.	Age	Sex	Primary / Secondary	Cytogenetics	IDH1	Other mutations
SU430	46	М	Primary	Der(7)t(7;11)	R132C	-
SU291.F	37	F	Primary	+8	R132H	FLT3-TKD, NPM1c+
SU277C	70	F	Primary	NK	R132G	
SU372	53	F	Primary	NK	R132H	FLT3-ITD
SU654	47	М	Primary	NK	R132C	NPM1c+
SU676	62	М	Primary	NK	R132H	NPM1c+, N-RAS, DNMT3A
SU582	71	F	Primary	NK	R132H	FLT3-ITD, N-RAS, NPM1c+
SU694	80	М	Secondary	+8	R132H	N-RAS
SU839	62	М	Primary	NK	R132H	FLT3-TKD
SU366	75	М	Secondary, MDS	NK	R132C	
SU223	18	F	Primary, Relapsed	t(9;11)	WT	MLL rearrangment
SU353	65	М	Primary	NK	WТ	FLT3-ITD, NPM1c+
SU354	65	М	Primary, relapsed	+19	WT	
SU463	56	F	Primary	Complex	WT	K-RAS
SU674	51	F	Primary	NK	WT	DMNT3A R882H





Sorted *IDH1* wildtype AML blasts





THP-1 **IDH1 WT** ACACA pLENTICRISPR v2

spPAM : Protospacer Adjacent Motif 20 nt : Specificity Determining Sequence ----- : Deletion ACTG : Insertions ACTG: Point Mutation:

THP-1 IDH1 R132H ACACA pLENTICRISPR v2

Insertions

A12

A14 GT AGAAGCCACAGTGAAATCTCGTTGAGAATCTATTTTCTT TCTGTCTCGGCCCCGCTTTT ACTAGGTGCAAGCCACAGCCACAGCCTATAAATATACAGAAGCCACAAACACCACCTATATTTTCTGGAGACCTGCTTTTCACTACCATACCA

---TAGGTGCAAGCCAGACATGCTGGACCTATAAAAATACAGAAGCCATAAAAACACCACCTATATTTTCTGGAGACCTGCTTTTCACCACCATACCCA

- AAAAATACAGAAGCCATAAAAACACCACCTATATTTTCTGGAGACCTGCTTTTCACTACCATACCCA

Deletions

BIO GTAACAATTITGCTGGAGAGCCACAGTGAAATCTGGTGAGAATCTATTITCTTGTGCTGCGCCGCGCTTTACTAGGTGCAAGCCAGCATGCTGGAGCTATAAAATACAGAAGCCATAAAAAAACACCACGCTATATTITCTGGAGACCTGCTTTTACTGGGAGCCTGCTTTTACTGGAGACTGCTGGCGCCGCTTTAACAAGCAGCAGCAAGCCATAAAAATACAGAAGCCATAAAAAATACAGCAGCCATAAAAATTCTGGAGACTGCTGCTGCTGCTGCTGCGCCGCTTTACCAAGCCA ТАВСТВСИИТСЯ СТАТИТСЯ СТАТИТИ ПО СТАТИ СТАТИ СО СТАТИ СТАТИТИ СТАТИТИ СТАТИТИ СТАТИТИ СТАТИ СТАТИТИ СТАТИСИ СТ ССТОВСИТ ТА ПЛАНИМИ СТАТИТИ СТА ПО СТАТИТИ СТАТИ АВСТ ССЛИССКИ СТАТИТИ СТАТИ СТАТИТИ СТАТИТИ.



TOPO Sequencing Exon 4 ACACA THP-1 Clones

THP1_IDH1_R132H_Clone4 ACACA Locus TRACTAGGETGCAAGCCAGACATGCTGGAACCTATAAAAATACAGAAGCCATAAAAAACACCACCATATTTTCTGGAGACCTGCTTTCACTACCATACC --------GTGCAAGCCAGACATGCTGGACCTATAAAAATACAGAAGCCATAAAAAACACCACCTATATTTTCTGGAGACCTGCTTTTCACTACCATACC THP1_IDH1_R132H_Clone5 ACACA Locus THP1_IDH1_WT_Clone3 ACACA Locus шин инде На Абласалаатстостобабалабсаластолатстобтобабалстатттеттетотетобо<mark>ссоркентт нетабетоблас</mark>аласатостобалетатлалалагасалабесаталаласалесалестобласталалагасалестобалестатлалагасалестобалестаталана В5 лоталелалатетостобабалассалетобабалетсатттесттетотетобосстоститтастаботобалессалестобалетаталалагасалестобале THP1_IDH1_WT_Clone 4 ACACA Locus AND LANTTO COGLAGOCALATICAL TO CALL TO THE CONTROL AND THE THE TO THE CONTROL AND THE AND THE CONTROL AND THE B14 B16 B17

m

1° AML SU372 IDH1 R132H 66% 104. huCD33-Transduce shRNA **RFP+** lentivirus 103 -**↓**48h transplant 12 weeks 104 5 RFP analysis



N				
Panobinostat	Target	Drug	Evaluable	p-value
HDAC1, HDAC2, HDAC3, HDAC4, HDAC5, HDAC6,	HDAC	Panobinostat	Yes	0.004
HDAC7, HDAC8, HDAC9, HDAC10, HDAC11, SIRT1, SIRT2, SIRT3, SIRT4,	MEK	AZD6244, PD-0325901	Yes	0.01
SIRT5, SIRT6, SIRT7	RTK	Sorafenib	Yes	0.09
AZD6244, PD-0325901 MAP2K1, MAP2K2, MAP2K3, MAP2K4,	CDK4	PD-0332991	No, very few sensitive lines	-
MAP2K5, MAP2K6, MAP2K7	RAF	PLX4720, RAF265	Yes	NS (> 0.1)
	EGFR	Erlotinib, Lapatinib, ZD-6474	Yes	NS (> 0.1)



h

a

С



Supplementary Figure 5. Validation of Predictive Biomarker Analysis using MiSL. (a) Targets of Panobinostat (HDAC inhibitor) (upper panel) and MEK inhibitors AZD6244 and PD-0325901 (lower panel) as per DGidb data. (b) Summary of comparisons between MiSL-predicted sensitive cell lines and sensitive cell lines using pharmacologic data for a variety of targets. There is a statistically significant overlap between the predictions and sensitivity based on actual pharmacological data. (c) Plot showing IC_{50} values for all the CCLE cell lines (ranked in decreasing order of sensitivity) tested by Barrentina et. al.. for Panobinostat (HDAC inhibitor). The horizontal line shows the IC_{50} threshold used to determine sensitivity to the drug – lower than threshold implies the cell line is sensitive. The MiSL-predicted sensitive cell lines are marked in black. (d) MiSL analysis steps illustrated for AKTI - (i) mutual exclusion of PIK3CA mutation and gene deletion across cancers (HI-LO Boolean implication), (ii) deletion of gene concordant with lower expression of gene (p < 0.05), (iii) expression of gene is higher in *PIK3CA*-mutated breast cancer (p < 0.05) (e) Breast cancer cells were seeded in the presence of increasing concentrations of MK-2206 with concentrations as indicated. All cells were fixed with 4% formaldehyde and stained with 0.1% crystal violet when wells containing untreated cells became confluent. (f) Western blot of cell extracts of PIK3CA mutant (MCF-7, T47D, CAL-148) and PIK3CA wild-type (SKBR-7, HCC-1806, CAL-120) breast cancer cells treated with 2 mM MK-2206 for 0, 4, 24 and 48 hours blotted with antibodies against cleaved-PARP, phospho-AKT, phospho-PRAS40, phospho-S6 kinase, phospho-4EBP1, phospho-BAD and HSP90.



Supplementary Figure 6. DAISY Comparison with shRNA Data. (a) Schematic for comparing DAISY candidates for *IDH1* mutation with synthetic lethal partners as per DECIPHER library screen generated using a doxycycline-inducible IDH1 R132H THP-1 cells. (b) Contingency table showing overlap between DAISY candidates for *IDH1* mutation and shRNA synthetic lethals as per DECIPHER screen with drop-out ratio of 0.8 for 2 or more shRNAs for calling synthetic lethals in shRNA data. The overlap is not statistically significant. (c) Contingency table showing overlap between DAISY candidates for *IDH1* mutation and shRNA synthetic lethals as per DECIPHER screen with a more stringent criterion (drop-out ratio of 0.6 for 2 or more shRNAs) for calling synthetic lethals in shRNA data. The overlap is still not statistically significant with the more stringent criterion for calling synthetic lethals.



Supplementary Figure 7 Uncropped images of Western blot of cell extracts of *PIK3CA* mutant (MCF-7, T47D, CAL-148) and *PIK3CA* wild-type (SKBR-7, HCC-1806, CAL-120) breast cancer cells treated with 2 mM MK-2206 for 0, 4, 24 and 48 hours blotted with antibodies against cleaved-PARP, phospho-AKT, phospho-PRAS40, phospho-S6 kinase, phospho-4EBP1, phospho-BAD and HSP90.