Acetic Acid Promoted Redox-Annulations with Dual C–H Functionalization

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Supporting Information

General Information: Reagents and solvents were purchased from commercial sources and were purified by distillation or recrystallization prior to use. Purification of reaction products was carried out by flash column chromatography using EM Reagent silica gel 60 (230-400 mesh). Analytical thin layer chromatography was performed on EM Reagent 0.25 mm silica gel 60 F₂₅₄ plates. Visualization was accomplished with UV light, and potassium permanganate, Dragendorff-Munier and anisaldehyde stains, followed by heating. Melting points were recorded on a Thomas Hoover capillary melting point apparatus and are uncorrected. Infrared spectra were recorded on an ATI Mattson Genesis Series FT-Infrared spectrophotometer. Proton nuclear magnetic resonance spectra (¹H-NMR) were recorded on a Varian VNMRS-500 MHz instrument and are reported in ppm using solvent as an internal standard (CDCl₃ at 7.26 ppm, (CD₃)₂SO at 2.50 ppm). Data are reported as app = apparent, s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, br = broad, dd = doublet of doublets, dt = doublet of triplets, td = triplet of doublets, ddd = doublet of doublet of doublets, dddd = doublet of doublet of doublets, ddddd = doublet of doublet of doublet of doublets, m = multiplet, comp = complex; and coupling constant(s) in Hz. Proton-decoupled carbon nuclear magnetic resonance spectra (¹³C-NMR) were recorded on a Varian VNMRS-500 MHz instrument and are reported in ppm using solvent as an internal standard (CDCl₃ at 77.16 ppm, (CD₃)₂SO at 39.52 ppm). Mass spectra were recorded on a Finnigan LCQ-DUO mass spectrometer or on a Finnigan 2001 Fourier Transform Ion Cyclotron Resonance Mass Spectrometer. Products 1a,¹ 1m,² 3a,³ and 3b⁴ were previously reported and their published characterization data matched our own in all regards. Ratios of diastereomeric products were determined by ¹H-NMR analysis of the crude reaction mixture.

Synthesis of ethyl 2-alkylquinoline-3-carboxylates 11 and ethyl 2-alkylnicotinates 15

General procedure A

This method was adopted from a literature procedure.⁵ To a stirred solution of nitrobenzaldehyde **10** (20 mmol) and β -ketoester (1.2 equiv) in acetic acid (0.3 M) was added iron (5 equiv) at room temperature. The mixture was heated to 50 °C for 1 h. The resulting mixture was allowed to cool to room temperature and filtered through a short pad of celite and washed with EtOAc. The filtrate was basified with 1 M NaOH. The aqueous layer was extracted with EtOAc (2 x 75 mL). The combined organic layers were then washed with brine and dried over anhydrous Na₂SO₄. The solvent was removed under reduced pressure and the residue purified by silica gel chromatography. Products **11a**⁵ and **11**⁵ were reported previously and their published characterization data matched our own in all regards.



General procedure B

This method was adopted from a literature procedure.⁶ To a mixture of **13** (20 mmol) and β -ketoester (1.2 equiv) was added citric acid (0.5 equiv) at room temperature. The mixture was heated to 100 °C for 2 h. The reaction mixture was allowed to cool to room temperature and poured into distilled water (100 mL). In case of solid products, the mixture was stirred for another hour and then filtered. The solid was recrystallized from EtOH. For liquid products, the mixture was extracted with EtOAc (2 x 75 mL). The combined organic layers were then washed with brine and dried over anhydrous Na₂SO₄. The solvent was removed under reduced pressure and the residue purified by silica gel chromatography. Products **11g**⁶ and **11i**⁶ were reported previously and their published characterization data matched our own in all regards.



General Procedure C

This method was adopted from a literature procedure.⁷ To a stirred solution of **14** (1.5 equiv) and β -ketoester (20 mmol) in toluene (0.5 M) was added ammonium acetate (2 equiv) and 4 Å molecular sieves (10 g) at room temperature. The mixture was heated under reflux for 15 h. The reaction mixture was then allowed to cool to room temperature, filtered through a short pad of celite, and washed with EtOAc. The solvent was removed under reduced pressure and the residue purified by silica gel chromatography. Compound **15a**⁷ was reported previously and its published characterization data matched our own in all regards.



Ethyl 6-fluoro-2-methylquinoline-3-carboxylate (11b): Following the general procedure A, **11b** was obtained from 5-fluoro-2-nitrobenzaldehyde and ethyl acetoacetate as a white solid in 52% yield ($R_f = 0.39$ in hexane/EtOAc 80:20 v/v); mp = 85–87 °C; IR (KBr) 3055, 2989, 2938, 2364, 2346, 1720, 1603, 1561, 1493, 1480, 1403, 1391, 1378, 1271, 1246, 1210, 1148, 1122, 1066, 968, 843, 775, 758, 690, 618, 423; ¹H NMR (CDCl₃, 500 MHz) δ 8.63 (s, 1H), 8.01 (dd, J = 9.2, 5.2 Hz, 1H), 7.52 (ddd, J = 9.3, 8.3, 2.9 Hz, 1H), 7.44 (dd, J = 8.5, 2.9 Hz, 1H), 4.43 (q, J = 7.1 Hz, 2H), 2.95 (s, 3H), 1.44 (t, J =7.1 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 166.4, 160.4 (d, $J_{C-F} = 248.5$ Hz), 157.8 (d, $J_{C-F} =$ 2.7 Hz), 145.8, 139.1 (d, $J_{C-F} = 5.5$ Hz), 131.2 (d, $J_{C-F} = 9.0$ Hz), 126.4 (d, $J_{C-F} = 10.2$ Hz), 124.8, 121.8 (d, $J_{C-F} = 25.8$ Hz), 111.4 (d, $J_{C-F} = 21.7$ Hz), 61.7, 25.6, 14.4; m/z (ESI–MS) 233.9 [M + H]⁺.

Ethyl 6-methyl-[1,3]dioxolo[4,5-g]quinoline-7-carboxylate (11e): Following the general procedure A, **11e** was obtained from 6-Nitropiperonal and ethyl acetoacetate as a yellow solid in 54% yield ($R_f = 0.21$ in hexane/EtOAc 80:20 v/v); mp = 159–161 °C; IR (KBr) 3032, 2910, 1687, 1612, 1590, 1461, 1421, 1395, 1276, 1245, 1214, 1105, 1070, 1039, 1018, 943, 888, 863, 583; ¹H NMR (DMSO, 500 MHz) δ 8.64 (s, 1H), 7.47 (s, 1H), 7.37–7.23 (m, 1H), 6.23 (s, 2H), 4.35 (q, J = 7.1 Hz, 2H), 2.78 (s, 3H), 1.36 (t, J = 7.1 Hz, 3H); ¹³C NMR (125 MHz, DMSO) δ 166.1, 155.3, 152.4, 147.6, 146.9, 137.9, 122.2, 121.4, 104.3, 103.3, 102.3, 60.9, 24.8, 14.1; m/z (ESI–MS) 260.0 [M + H]⁺.

Ethyl 2-methyl-7-(trifluoromethyl)quinoline-3-carboxylate (11f): Following a modified



literature procedure,⁸ to a stirred solution of (2-amino-4-(trifluoromethyl)phenyl)methanol (20 mmol) and ethyl acetoacetate (1.2 equiv) in EtOH (0.3 M) was added MnO_2 (2 equiv) at room temperature. The mixture was heated under reflux for 15 h. The reaction mixture was

then allowed to cool to room temperature, filtered through a short pad of celite, and washed with EtOAc. The solvent was removed under reduced pressure and the residue purified by silica gel chromatography. **11f** was obtained as a yellow solid in 89% yield ($R_f = 0.52$ in hexane/EtOAc 80:20 v/v); mp = 42–43 °C; IR (KBr) 2987, 1730, 1728, 1603, 1563, 1501, 1368, 1326, 1285, 1233, 1147, 1110, 1058, 933, 900, 818, 775, 720, 683, 538, 478; ¹H NMR (CDCl₃, 500 MHz) δ 8.86–8.57 (m, 1H), 8.25 (dq, *J* = 1.7, 0.9 Hz, 1H), 7.87 (app dt, *J* = 8.6, 0.9 Hz, 1H), 7.61 (dd, *J* = 8.5, 1.8 Hz, 1H), 4.41 (q, *J* = 7.2 Hz, 2H), 2.94 (s, 3H), 1.42 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 166.0, 160.0, 147.6, 139.30 133.0 (q, *J*_{C-F} = 32.7 Hz), 129.6, 127.2, 126.5 (q, *J*_{C-F} = 4.5 Hz), 125.8, 123.8 (q, *J*_{C-F} = 272.7 Hz), 122.1 (q, *J*_{C-F} = 3.2 Hz), 61.8, 25.7, 14.3; *m*/z (ESI–MS) 284.3 [M + H]⁺.

Ethyl 4-ethyl-2-methylquinoline-3-carboxylate (11h): Following the general procedure B, **11h** was obtained from 1-(2-aminophenyl)propan-1-one and ethyl acetoacetate as a yellow liquid in 85% yield ($R_f = 0.30$ in hexane/EtOAc 80:20 v/v); IR (KBr) 3054, 2986, 2684, 2305, 1723, 1583, 1489, 1422, 1266, 1224, 1166, 1131, 1078, 1060, 896, 749, 705; ¹H NMR (CDCl₃, 500 MHz) δ 8.10–7.95 (comp, 2H), 7.69 (dddd, J = 8.3, 6.9, 1.3, 0.6 Hz, 1H), 7.60–7.45 (m, 1H), 4.48 (q, J = 7.1 Hz, 2H), 3.03 (q, J = 7.6 Hz, 2H), 2.70 (s, 3H), 1.43 (td, J = 7.2, 0.6 Hz, 3H). 1.35 (t, J = 7.6 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 169.3, 154.6, 147.8, 147.1, 130.0, 129.7, 127.4, 126.4, 124.8, 124.0, 61.7, 23.9, 23.5, 15.4, 14.4; m/z (ESI–MS) 244.0 [M + H]⁺.

Ethyl 2-benzylquinoline-3-carboxylate (11j): Following the general procedure A, **11j** was obtained from 2-nitrobenzaldehyde and ethyl 3-oxo-4-phenylbutanoate as a yellow solid in 62% yield ($R_f = 0.43$ in hexane/EtOAc 80:20 v/v); mp = 60-63 °C; IR (KBr) 3060, 2977, 2923, 1719, 1620, 1565, 1489, 1425, 1278, 1250, 1203, 1128, 1061, 1025, 868, 833, 785, 760, 748, 718, 695, 505; ¹H NMR (CDCl₃, 500 MHz) δ 8.67 (s, 1H), 8.13 (d, *J* = 8.4 Hz, 1H), 7.82 (d, *J* = 8.6 Hz, 1H), 7.77 (ddd, *J* = 8.4, 6.9, 1.5 Hz, 1H), 7.52 (ddd, *J* = 8.1, 7.0, 1.2 Hz, 1H), 7.40–7.21 (comp, 4H), 7.18–7.07 (m, 1H), 4.81 (s, 2H), 4.31 (q, *J* = 7.2 Hz, 2H), 1.31 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 166.4, 159.7, 148.6, 140.0, 139.6, 131.5, 129.02, 129.00, 128.4, 128.2, 126.8, 126.0, 125.9, 124.3, 61.4, 43.3, 14.2; *m/z* (ESI–MS) 292.2 [M + H]⁺.

Ethyl 2-ethylquinoline-3-carboxylate (11k): Following the general procedure A, **11k** was obtained from 2-nitrobenzaldehyde and ethyl 3-oxopentanoate as a yellow solid in 60% yield ($R_f = 0.47$ in hexane/EtOAc 80:20 v/v); mp = 61–63 °C; IR (KBr) 3043, 2973, 2936, 1711, 1619, 1560, 1478, 1421, 1275, 1254, 1204, 1132, 1071, 1052, 1015, 960, 878, 798, 755, 618, 590, 478; ¹H NMR (CDCl₃, 500 MHz) δ 8.68 (s, 1H), 8.05 (dd, J = 8.6, 0.9 Hz, 1H), 7.85 (ddd, J = 8.1, 1.4, 0.6 Hz, 1H), 7.80–7.66 (m, 1H), 7.52 (ddd, J = 8.1, 6.9, 1.1 Hz, 1H), 4.44 (q, J = 7.1 Hz, 2H), 3.34 (q, J = 7.5 Hz, 2H), 1.45 (t, J = 7.1 Hz, 3H), 1.38 (t, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 166.9, 163.1, 148.8,

140.0, 131.6, 128.9, 128.5, 126.6, 125.8, 124.0, 61.6, 31.2, 14.4, 14.2; m/z (ESI–MS) 230.1 [M + H]⁺.

Ethyl 2-benzylnicotinate (15b): Following the general procedure C, **15b** was obtained from ethyl 3-oxo-4-phenylbutanoate, acrolein and ammonium acetate as a yellow liquid in 23% yield ($R_f = 0.28$ in hexane/EtOAc 80:20 v/v); IR (KBr) 3049, 2984. 2684, 2359, 2304, 1721, 1421, 1261, 1156, 895, 750, 700; ¹H NMR (CDCl₃, 500 MHz) δ 8.73 (dd, J = 4.8, 1.8 Hz, 1H), 8.21 (dd, J = 7.9, 1.9 Hz, 1H), 7.34–7.24 (comp, 5H), 7.24–7.15 (m, 1H), 4.63 (s, 2H), 4.36 (q, J = 7.1 Hz, 2H), 1.36 (t, J = 7.1 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 166.6, 161.2, 151.9, 139.7, 138.8, 129.0, 128.3, 126.3, 126.2, 121.4, 61.5, 42.4, 14.3; m/z (ESI–MS) 242.1 [M + H]⁺.

Ethyl 2-ethylnicotinate (15c): Following the general procedure C, **15c** was obtained from ethyl 3-oxopentanoate, acrolein and ammonium acetate as a yellow liquid in 25% yield ($R_f = 0.30$ in hexane/EtOAc 80:20 v/v); IR (KBr) 3054, 2987, 2687, 2411, 2305, 1721, 1581, 1566, 1422, 1266, 1143, 1093, 1023, 896, 749, 706; ¹H NMR (CDCl₃, 500 MHz) δ 8.56 (dd, J = 4.8, 1.9 Hz, 1H), 8.06 (dd, J = 7.9, 1.9 Hz, 1H), 7.11 (dd, J = 7.9, 4.8 Hz, 1H), 4.30 (q, J = 7.1 Hz, 2H), 3.09 (q, J = 7.5 Hz, 2H), 1.32 (t, J = 7.1 Hz, 3H), 1.23 (t, J =7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 166.6, 164.3, 151.7, 138.3, 125.5, 120.7, 61.3, 30.3, 14.2, 14.0; m/z (ESI–MS) 180.1 [M + H]⁺.

Synthesis of (2-alkylquinolin-3-yl)methanol 12 and (2-alkylpyridin-3-yl)methanol 16

General Procedure D

To an ice-cooled suspension of LiAlH₄ (531 mg, 14 mmol, 2 equiv) in dry THF (15 mL) was slowly added a solution of compound **11** or **15** (7 mmol, 1 equiv) in dry THF (5 mL). The reaction was allowed to warm to room temperature and stirred for two hours. The mixture was then cooled in an ice bath, followed by quenching with water (2 mL) and 1M NaOH (1 mL). The lithium and aluminum hydroxide salts were then filtered through a short pad of celite and washed with EtOAc (6 x 50 mL). The solvent was removed under reduced pressure. For pyridine substrates, the residue was directly used in the next step without purification. For quinoline substrates, the residue was purified by silica gel chromatography.



General Procedure E

This method was adopted from a literature procedure.⁹ To an ice-cooled solution of NaBH₄ (2.5 equiv) in EtOH (10 mL) were added a solution of **11** (10 mmol) in EtOH (20 mL) and a solution of anhydrous CaCl₂ (1.2 equiv) in EtOH (20 mL). The reaction mixture was allowed to warm to room temperature and stirred for 18 h. After the consumption of **7** as indicated by TLC, the solvent was removed under reduced pressure. The residue was suspended in dichloromethane

(50 mL) and water (50 mL). After separation, the organic layer was washed with brine (30 mL x 3), then dried over NaSO₄. The solvent was removed under reduced pressure and the residue was directly used in the next step without purification.

Following the general procedure E, **11b**, **11d**, **11f**, **11j** were reduced to the corresponding alcohols and used in the next step without purification.



(6-chloro-2-methylquinolin-3-yl)methanol (12c): Following the general procedure D, 12c was obtained from 11c as a yellow solid in 60% yield ($R_f = 0.30$ in hexane/EtOAc 20:80 v/v); mp = 142–143 °C; IR (KBr) 3301, 2933, 2361, 2339, 1598, 1561, 1483, 1382, 1345, 1224, 1115, 1076, 1052, 987, 933, 926, 824, 720, 558, 487; ¹H NMR (CDCl₃, 500 MHz) δ 8.02 (s, 1H), 7.92 (d, J = 8.9 Hz, 1H), 7.71 (d, J = 2.3 Hz, 1H), 7.57 (dd, J = 8.9, 2.3 Hz, 1H), 4.85 (s, 2H), 2.76 (br s, 1H), 2.66 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 157.8, 145.3, 133.8, 132.7, 131.7, 130.2, 129.9, 127.8, 126.2, 62.3, 22.6; m/z (ESI–MS) (³⁵Cl) 208.1 [M + H]⁺, (³⁷Cl) 210.1 [M + H]⁺.

(6-methyl-[1,3]dioxolo[4,5-g]quinolin-7-yl)methanol (12e): Following the general procedure D, 12e was obtained from 11e as a yellow solid in 60% yield ($R_f = 0.24$ in hexane/EtOAc 20:80 v/v); mp = 198–200 °C; IR (KBr) 3097, 2912, 2823, 2364, 1624, 1499, 1468, 1370, 1241, 1204, 1099, 1043, 946, 918, 849, 799, 614; ¹H NMR (CDCl₃, 500 MHz) δ 8.02 (d, J = 1.1 Hz, 1H), 7.31 (s, 1H), 7.25 (s, 1H), 6.16 (s, 2H), 5.31 (t, J = 5.3 Hz, 1H) 4.60 (dd, J = 5.3, 1.0 Hz, 2H), 2.51 (s, 3H);

 $^{1.25}$ (s, 1H), 6.16 (s, 2H), 5.31 (t, J = 5.3 Hz, 1H) 4.60 (dd, J = 5.3, 1.0 Hz, 2H), 2.51 (s, 3H); 13 C NMR (125 MHz, DMSO) δ 154.4, 149.7, 146.7, 144.0, 132.1, 132.0, 123.2, 104.4, 102.7, 101.6, 60.4, 21.8; m/z (ESI–MS) 218.1 [M + H]⁺.

(2,4-dimethylquinolin-3-yl)methanol (12g): Following the general procedure D, 12g was obtained from 11g as a yellow solid in 78% yield ($R_f = 0.18$ in hexane/EtOAc 20:80 v/v); mp = 119–121 °C; IR (KBr) 3119, 2922, 2847, 2739, 2617, 1616, 1586, 1572, 1501, 1449, 1405, 1369, 1346, 1315, 1218, 1181, 1108, 1038, 1017, 873, 831, 765, 721, 675, 675, 648, 508; ¹H NMR (CDCl₃, 500 MHz) δ 8.00–7.85 (comp, 2H), 7.59 (ddd, J = 8.3, 6.8, 1.3 Hz, 1H), 7.44 (ddd, J = 8.3, 6.8, 1.2 Hz, 1H), 4.87

(s, 2H), 3.13 (br s, 1H), 2.70 (s, 3H), 2.65 (s, 3H); 13 C NMR (125 MHz, CDCl₃) δ 158.6, 146.6, 143.5, 129.9, 129.2, 128.9, 127.1, 125.8, 124.1, 59.0, 23.7, 14.2; m/z (ESI–MS) 188.2 [M + H]⁺.

(4-ethyl-2-methylquinolin-3-yl)methanol (12h): Following the general procedure D, 12h was obtained from 11h as a yellow solid in 83% yield ($R_f = 0.24$ in hexane/EtOAc 20:80 v/v); mp = 106–108 °C; IR (KBr) 3186, 2979, 2934, 2874, 1611, 1585, 1571, 1502, 1438, 1404, 1373, 1181, 1108, 1031, 1015, 865, 774, 650; ¹H NMR (CDCl₃, 500 MHz) δ 7.92 (app dt, J = 8.5, 1.7 Hz, 2H), 7.65–7.55 (m, 1H), 7.45 (app td, J = 7.8, 6.9, 1.9 Hz, 1H), 4.86 (s, 2H), 3.30 (br s, 1H), 3.13 (q, J = 7.5 Hz, 3H), 2.72 (s, 3H), 1.28 (t, J = 7.6 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 159.1, 149.1, 147.2, 129.2, 129.1, 126.0, 125.9, 124.0, 58.6, 23.6, 21.3, 15.8; m/z (ESI–MS) 202.1 [M + H]⁺.

(6-chloro-2-methyl-4-phenylquinolin-3-yl)methanol (12i): Following the general procedure Ph CI NMe NMe NMe NME NMR (125 MHz, CDCl₃) δ 159.9, 147.3, 145.6, 135.7, 131.8, 130.4, 130.3, 130.2, 129.5, 128.8, 120.2 methanol (12i): Following the general procedure D, 12i was obtained from 11i as a yellow solid in 55% yield ($R_f = 0.42$ in hexane/EtOAc 20:80 v/v); mp = 137–139 °C; IR (KBr) 3060, 2910, 2826, 2724, 2361, 1584, 1570, 1484, 1444, 1375, 1348, 1301, 1078, 1040, 950, 831, 707, 603, 516; ¹H NMR (CDCl₃, 500 MHz) δ 7.96 (dd, J = 8.9, 1.6 Hz, 1H), NMR (125 MHz, CDCl₃) δ 159.9, 147.3, 145.6, 135.7, 131.8, 130.4, 130.3, 130.2, 129.5, 128.8,

NMR (125 MHz, CDCl₃) δ 159.9, 147.3, 145.6, 135.7, 131.8, 130.4, 130.3, 130.2, 129.5, 128.8, 128.6, 127.5, 125.6, 60.1, 23.7; *m*/*z* (ESI–MS) (³⁵Cl) 284.0 [M + H]⁺, (³⁷Cl) 286.1 [M + H]⁺.

(2-ethylquinolin-3-yl)methanol (12k): Following the general procedure D, 12k was obtained from 11k as a yellow solid in 56% yield ($R_f = 0.56$ in hexane/EtOAc 20:80 v/v); mp = 87–90 °C; IR (KBr) 3134, 2970, 2832, 1619, 1603, 1566, 1495, 1458, 1420, 1353, 1217, 1187, 1136, 1100, 1048, 919, 845, 786, 743, 653, 621, 481; ¹H NMR (CDCl₃, 500 MHz) δ 8.13 (s, 1H), 8.05 (d, J = 8.5 Hz, 1H), 7.82–7.73 (m, 1H), 7.66 (ddd, J = 8.4, 6.9, 1.5 Hz, 1H), 7.48 (ddd, J = 8.2, 6.9, 1.2 Hz, 1H), 4.92 (s, 2H), 3.00 (q, J = 7.5 Hz, 2H), 1.39 (t, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 162.1, 134.3, 132.2, 129.3, 128.6, 127.6, 127.1, 126.1, 110.2, 62.4, 28.8, 13.5; m/z (ESI–MS) 188.1 [M + H]⁺.

Synthesis of 2-alkylquinoline-3-carbaldehydes and 2-alkylnicotinaldehydes 1

General Procedure F

To a stirred solution of compounds **12** or **16** (2 mmol, 1 equiv) in CH_2Cl_2 (15 mL) was added MnO_2 (869 mg, 10 mmol, 5 equiv) at room temperature. The resulting mixture was heated under reflux until **12** or **16** was consumed as indicated by TLC. The reaction mixture was then allowed to cool to room temperature, filtered through a short pad of celite and washed with CH_2Cl_2 . The solvent was removed under reduced pressure and the residue purified by silica gel chromatography.

6-fluoro-2-methylquinoline-3-carbaldehyde (1b): Following the general procedure F, 1b was obtained from 12b as a yellow solid in 75% yield (over 2 steps), ($R_f = 0.20$ in hexane/EtOAc 80:20 v/v); mp = 104–105 °C; IR (KBr) 1714, 1685, 1592, 1564, 1496, 1373, 1349, 1210, 1154, 1105, 965, 833, 750, 585, 488; ¹H NMR (CDCl₃, 500 MHz) δ 10.39 (s, 1H), 8.54 (s, 1H), 8.07 (dd, J = 9.2, 5.1 Hz, 1H), 7.60 (dddd, J = 9.1, 8.2, 2.8, 0.8 Hz, 1H), 7.55 (dd, J = 8.3, 2.8 Hz, 1H), 3.02 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.3, 160.5 (d, $J_{C-F} = 249.6$ Hz), 157.7 (d, $J_{C-F} = 2.8$ Hz), 146.4, 141.5 (d, $J_{C-F} = 5.4$ Hz), 131.4 (d, $J_{C-F} = 9.0$ Hz), 128.6, 126.7 (d, $J_{C-F} = 10.1$ Hz), 122.8 (d, $J_{C-F} = 25.7$ Hz), 111.9 (d, $J_{C-F} = 21.7$ Hz), 23.9; m/z (ESI–MS) 222.0 [M + MeOH +H]⁺.

6-chloro-2-methylquinoline-3-carbaldehyde (1c): Following the general procedure F, **1c** was obtained from **12c** as a yellow solid in 70% yield ($R_f = 0.24$ in hexane/EtOAc 80:20 v/v); mp = 130–132 °C; IR (KBr) 3033, 3002, 1709, 1691, 1616, 1592, 1560, 1482, 1397, 1338, 1226, 1155, 1121, 1075, 928, 830, 769, 728, 667, 615; ¹H NMR (CDCl₃, 500 MHz) δ 10.37 (s, 1H), 8.48 (s, 1H), 7.99 (d, *J* = 9.0 Hz, 1H), 7.90 (d, *J* = 2.4 Hz, 1H), 7.75 (dd, *J* = 8.9, 2.3 Hz, 1H), 3.01 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.2, 158.7, 147.7, 141.2, 133.5, 132.8, 130.6, 128.7, 127.6, 126.8, 24.1; *m/z* (ESI–MS) (³⁵Cl) 238.0 [M + MeOH +H]⁺, (³⁷Cl) 240.0 [M + MeOH +H]⁺.

6-bromo-2-methylquinoline-3-carbaldehyde (1d): Following the general procedure F, 1d was ^{Br} betained from 12d as a yellow solid in 60% yield (over 2 steps), (R_f = 0.23 in hexane/EtOAc 80:20 v/v); mp = 136–138 °C; IR (KBr) 2975, 1708, 1692, 1612, 1588, 1559, 1481, 1391, 1375, 1226, 1156, 1118, 1058, 923, 830, 765, 648; ¹H NMR (CDCl₃, 500 MHz) δ10.37 (s, 1H), 8.48 (s, 1H), 8.08 (d, *J* = 2.1 Hz, 1H), 7.92 (ddd, *J* = 9.2, 2.4, 1.0 Hz, 1H), 7.88 (dt, *J* = 9.0, 1.8 Hz, 1H), 3.01 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.12, 158.9, 147.9, 141.1, 136.1, 131.0, 130.6, 128.7, 127.3, 120.8, 24.1; *m/z* (ESI– MS) (⁷⁹Br) 282.0 [M + MeOH + H]⁺, (⁸¹Br) 283.9 [M + MeOH + H]⁺.

6-methyl-[1,3]dioxolo[4,5-g]quinoline-7-carbaldehyde (1e): Following the general procedure C, 1e was obtained from 12e as a yellow solid in 99% yield ($R_f = 0.11$ in hexane/EtOAc 80:20 v/v); mp = 171–173 °C; IR (KBr) 3043, 2927, 2718, 1692, 1590, 1501, 1465, 1389, 1245, 1205, 1170, 1136, 1035, 936, 801, 765, 595, 433; ¹H NMR (CDCl₃, 500 MHz) δ 10.30 (s, 1H), 8.33 (s, 1H), 7.32 (s, 1H), 7.10 (s, 1H), 6.13 (s, 2H), 2.93 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.4, 156.8, 153.5, 148.5, 148.2, 140.0, 126.5, 123.1, 105.6, 103.7, 102.3, 23.4; *m*/z (ESI–MS) 248.0 [M + MeOH + H]⁺.

2-methyl-7-(trifluoromethyl)quinoline-3-carbaldehyde (1f): Following the general procedure F₃C K_{Me} F, **1f** was obtained from **12f** as a yellow solid in 90% yield (over 2 steps), (R_f = 0.29 in hexane/EtOAc 80:20 v/v); mp = 64–66 °C; IR (KBr) 2896, 1697, 1685, 1599, 1558, 1351, 1323, 1284, 1162, 1125, 1055, 935, 895, 870, 825, 763, 690; ¹H NMR (CDCl₃, 500 MHz) δ 10.42 (s, 1H), 8.64 (s, 1H), 8.38 (dd, *J* = 1.7, 0.9 Hz, 1H), 8.07 (d, *J* = 8.4 Hz, 1H), 7.75 (dd, *J* = 8.5, 1.7 Hz, 1H), 3.06 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.1, 159.9, 148.4, 141.8, 134.1 (q, *J* = 33.0 Hz), 130.3, 129.5, 126.8 (q, *J* = 4.3 Hz), 123.74 (q, *J* = 272.8 Hz), 122.74 (q, *J* = 3.2 Hz), 24.2; *m/z* (ESI–MS) 271.9 [M + MeOH + H]⁺.

2,4-dimethylquinoline-3-carbaldehyde (1g): Following the general procedure F, **1g** was obtained from **12g** as a yellow solid in 80% yield, ($R_f = 0.29$ in hexane/EtOAc 70:30 v/v); mp = 111–113 °C; IR (KBr) 2924, 2356, 2344, 1689, 1608, 1570, 1496, 1441, 1398, 1374, 1188, 1138, 1110, 1030, 754, 645; ¹H NMR (CDCl₃, 500 MHz) δ 10.77 (s, 1H), 8.12 (dd, J = 8.3, 1.5 Hz, 1H), 8.06–7.95 (m, 1H), 7.77 (ddd, J = 8.4, 6.9, 1.4 Hz, 1H), 7.56 (ddd, J = 8.3, 6.8, 1.3 Hz, 1H), 2.93 (s, 3H), 2.92 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 193.3, 158.0, 148.8, 148.3, 131.8, 129.6, 126.9, 126.8, 126.7, 124.8, 24.9, 14.2.; m/z (ESI–MS) 218.1 [M + MeOH + H]⁺.

4-ethyl-2-methylquinoline-3-carbaldehyde (1h): Following the general procedure F, **1h** was obtained from **12h** as a yellow solid in 75% yield ($R_f = 0.35$ in hexane/EtOAc 70:30 v/v); mp = 78–80 °C; IR (KBr) 3057, 2967, 2925, 2864, 2762, 1689, 1611, 1561, 1498, 1406, 1238, 1181, 1133, 1103, 1063, 905, 853, 770, 743, 643, 508; ¹H NMR (CDCl₃, 500 MHz) δ 10.76 (s, 1H), 8.18–8.11 (m, 1H), 8.08–7.99 (m, 1H), 7.85–7.72 (m, 1H), 7.64–7.47 (m, 1H), 3.41 (q, *J* = 7.5 Hz, 2H), 2.92 (s, 3H), 1.39 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 193.0, 158.2, 154.8, 148.8, 131.7, 129.8, 126.8, 125.8, 125.7, 124.5, 25.1, 20.9, 16.0; *m/z* (ESI–MS) 232.0 [M + MeOH + H]⁺.

6-chloro-2-methyl-4-phenylquinoline-3-carbaldehyde (1i): Following the general procedure F, **1i** was obtained from **12i** as a yellow solid in 55% yield ($R_f = 0.54$ in hexane/EtOAc 70:30 v/v); mp = 220–222 °C; IR (KBr) 3052, 2999, 2869, 1685, 1559, 1478, 1406, 1379, 1338, 1296, 1148, 1123, 1073, 953, 833, 750, 707, 655, 618, 465; ¹H NMR (CDCl₃, 500 MHz) δ 9.96 (s, 1H), 8.03 (dd, J = 8.9, 0.5 Hz, 1H), 7.73 (dd, J = 8.9, 2.3 Hz, 1H), 7.65–7.55 (comp, 3H), 7.50 (dd, J = 2.3, 0.5 Hz, 1H), 7.41–7.34 (comp, 2H), 2.99 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 193.3, 158.3, 154.0, 147.0, 133.1, 133.0, 132.8, 130.8, 130.2, 129.5, 129.0, 126.8, 126.3, 125.7, 25.8; *m/z* (ESI–MS) (³⁵Cl) 314.0 [M + MeOH + H]⁺, (³⁷Cl) 316.0 [M + MeOH + H]⁺.

2-benzylquinoline-3-carbaldehyde (1j): Following the general procedure F, **1j** was obtained from **12j** as a yellow solid in 70% yield (over 2 steps), ($R_f = 0.52$ in hexane/EtOAc 70:30 v/v); mp = 114–116 °C; IR (KBr) 2918, 2363, 2345, 1698, 1619, 1561, 1419, 1430, 1171, 1108, 923, 786, 759, 723, 696, 463; ¹H NMR (CDCl₃, 500 MHz) δ 10.31 (s, 1H), 8.63 (d, J = 0.8 Hz, 1H), 8.16 (dd, J = 8.5, 0.9 Hz, 1H), 7.95 (dd, J = 8.1, 1.4 Hz, 1H), 7.87 (ddd, J = 8.5, 6.9, 1.5 Hz, 1H), 7.61 (ddd, J = 8.1, 6.9, 1.1 Hz, 1H), 7.32–7.22 (comp, 4H), 7.17 (ddt, J = 8.5, 6.1, 1.9 Hz, 1H), 4.77 (s, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 190.8, 160.0, 149.4, 142.4, 139.0, 132.7, 129.3, 129.2, 129.0, 128.6, 127.9, 127.3, 126.6, 126.4, 42.3; m/z (ESI–MS) 280.1 [M + MeOH + H]⁺.

2-ethylquinoline-3-carbaldehyde (1k): Following the general procedure F, **1k** was obtained from **12k** as a yellow solid in 70% yield ($R_f = 0.46$ in hexane/EtOAc 70:30 v/v); mp = 57-59 °C; IR (KBr) 2975, 2934, 2874, 2360, 2342, 1698, 1685, 1619, 1595, 1561, 1490, 1452, 1421, 1373, 1343, 1213, 1186, 1151, 1104, 1013, 955, 921, 803, 750, 598, 482; ¹H NMR (CDCl₃, 500 MHz) δ 10.39 (s, 1H), 8.61 (s, 1H), 8.09 (d, *J* = 8.5 Hz, 1H), 7.93 (d, *J* = 8.1 Hz, 1H), 7.87-7.72 (m, 1H), 7.57 (t, *J* = 7.5 Hz, 1H), 3.39 (q, *J* = 7.5 Hz, 2H), 1.40 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.3, 163.4, 149.6, 142.5, 132.7, 129.2, 129.1, 127.6, 127.0, 126.2, 29.8, 14.2; *m/z* (ESI-MS) 218.2 [M + MeOH + H]⁺.

2-benzylnicotinaldehyde (10): Following the general procedure F, **10** was obtained from **16b** as a yellow liquid in 80% yield (over 2 steps), ($R_f = 0.29$ in hexane/EtOAc 70:30 v/v); IR (KBr) 3410, 3054, 2922, 2852, 2356, 2341, 1696, 1633, 1581, 1493, 1453, 1266, 743; ¹H NMR (CDCl₃, 500 MHz) δ 10.34 (s, 1H), 8.77 (dd, J = 4.8, 1.9 Hz, 1H), 8.14 (dd, J = 7.8, 1.9 Hz, 1H), 7.37 (dd, J = 7.8, 4.8 Hz, 1H), 7.31–7.22 (comp, 4H), 7.21–7.15 (m, 1H), 4.61 (s, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 191.1, 162.4, 153.8, 139.1, 138.4, 129.6, 128.9, 128.8, 126.7, 122, 41.2; *m/z* (ESI–MS) 230.0 [M + MeOH + H]⁺.

2-ethylnicotinaldehyde (1p): Following the general procedure F, **1p** was obtained from **16c** as a yellow liquid in 80% yield (over 2 steps), ($R_f = 0.24$ in hexane/EtOAc 70:30 v/v); IR (KBr) 2969, 2934, 2874, 2371, 1701, 1696, 1681, 1581, 1453, 1261, 1218, 1098, 1063, 1033, 845, 758, 588; ¹H NMR (CDCl₃, 500 MHz) δ 10.34 (d, J = 0.5 Hz, 1H), 8.72 (dd, J = 4.8, 1.9 Hz, 1H), 8.11 (dd, J = 7.8, 1.9 Hz, 1H), 7.32 (ddd, J = 7.8, 4.8, 0.6 Hz, 1H), 3.23 (q, J = 7.6 Hz, 2H), 1.34 (t, J = 7.6 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 191.1, 165.6, 153.5, 138.2, 129.0, 121.9, 28.3, 14.6; m/z (ESI–MS) 168.2 [M + MeOH + H]⁺.

General Procedure G for the redox-annulation of 2-alkylquinoline-3-carbaldehydes with secondary amines

To a solution of the aldehyde (0.5 mmol, 1 equiv) in toluene (2.5 mL) and acetic acid (2.5 mL) was added the amine (0.75 mmol, 1.5 equiv). The resulting mixture was heated under reflux until the aldehyde was consumed as indicated by TLC. The reaction mixture was then allowed to cool to room temperature, diluted with EtOAc (15 mL), and washed with 1N NaOH (2 x 20 mL). The combined aqueous layers were extracted with EtOAc (2 x 15 mL), and the combined organic layers washed with brine (40 mL) and dried over anhydrous Na₂SO₄. Solvent was then removed under reduced pressure and the residue purified by silica gel chromatography.

General Procedure H for the redox-annulation of 2-methylpyridine-3-carbaldehyde with secondary amines

To a solution of the aldehyde (0.5 mmol, 1 equiv) in acetic acid (0.1 M, 5 mL) was added the amine (0.75 mmol, 1.5 equiv). The resulting mixture was heated under reflux until the aldehyde was consumed as indicated by TLC. The reaction mixture was then allowed to cool to room temperature, diluted with EtOAc (15 mL), and washed with 1N NaOH (2 x 20 mL). The combined aqueous layers were extracted with EtOAc (2 x 15 mL), and the combined organic layers washed with brine (40 mL) and dried over anhydrous Na₂SO₄. Solvent was then removed under reduced pressure and the residue purified by silica gel chromatography.

(±)-**2a**: To a solution of 2-methylquinoline-3-carbaldehyde (342 mg, 2 mmol, 1 equiv) in toluene (10 mL) and acetic acid (10 mL) was added 1,2,3,4-tetrahydroisoquinoline (0.377 mL, 3 mmol, 1.5 equiv). The resulting mixture was heated under reflux for 1.5 h. The reaction mixture was then allowed to cool to room temperature, diluted with EtOAc (40 mL), and

washed with 1N NaOH (2 x 50 mL). The combined aqueous layers were extracted with EtOAc (2 x 40 mL), and the combined organic layers washed with brine (50 mL) and dried over anhydrous Na_2SO_4 . Solvent was then removed under reduced pressure and the residue purified

by silica gel chromatography. Product (±)-**2a** was obtained as a yellow solid in 85% yield (486 mg). (R_f = 0.19 in hexane/EtOAc 70:30 v/v); mp = 173–175 °C; IR (KBr) 3016, 2923, 2816, 2748, 2360, 1671, 1584, 1493, 1466, 1427, 1315, 1270, 1250, 1163, 1144, 1099, 1045, 815, 738, 520, 478; ¹H NMR (CDCl₃, 500 MHz) δ 8.01 (app d, *J* = 8.4 Hz, 1H), 7.84 (s, 1H), 7.74 (app d, *J* = 8.1, 1H), 7.65 (ddd, *J* = 8.4, 6.8, 1.5 Hz, 1H), 7.47 (ddd, *J* = 8.2, 6.7, 1.2 Hz, 1H), 7.35 (app d, *J* = 7.7 Hz, 1H), 7.25 (app td, *J* = 7.6, 7.2, 1.7 Hz, 1H), 7.22–7.11 (comp, 2H), 4.24 (d, *J* = 14.9 Hz, 1H), 3.99–3.73 (comp, 3H), 3.38–3.14 (comp, 3H), 2.81 (dd, *J* = 14.7, 3.7 Hz, 1H), 2.75–2.61 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 156.7, 147.2, 137.4, 134.3, 132.6, 129.1, 129.0, 128.6, 128.3, 127.3, 127.1, 126.4, 126.0, 125.6, 60.2, 58.1, 51.4, 40.8, 29.6; *m/z* (ESI–MS) 287.1 [M + H]⁺.

X-ray quality crystals of (\pm) -2a were obtained from hexane/ethyl acetate through slow diffusion at room temperature.



The requisite CIF has been submitted to the journal and deposited with the CCDC (deposition # 1541944).

(±)-**2b**: Following general procedure G, product (±)-**2b** was obtained from **1b** and 1,2,3,4tetrahydroisoquinoline (reflux for 1 h) as a yellow solid in 85% yield (130 mg), ($R_f = 0.23$ in hexane/EtOAc 70:30 v/v); mp = 171–172 °C; IR (KBr) 2924, 2813, 2760, 1627, 1612, 1497, 1410, 1361, 1329, 1300, 1235, 1213, 1163, 1140, 1108, 1046, 984, 961, 901, 875, 829, 760, 736, 470; ¹H NMR (CDCl₃, 500 MHz) δ 8.00 (dd, J = 9.2, 5.3 Hz, 1H), 7.80 (s, 1H), 7.42 (ddd, J = 9.2, 8.3,

NMR (CDCl₃, 500 MHz) 6 8.00 (dd, J = 9.2, 5.5 Hz, 1H), 7.80 (s, 1H), 7.42 (ddd, J = 9.2, 8.5, 2.8 Hz, 1H), 7.38–7.32 (comp, 2H), 7.28–7.23 (m, 1H), 7.21 (app tdd, J = 7.1, 1.5, 0.7 Hz, 1H), 7.18–7.15 (m, 1H), 4.24 (d, J = 14.9, 1H), 3.94–3.84 (comp, 2H), 3.80 (dd, J = 17.1, 3.9 Hz, 1H), 3.33–3.15 (comp, 3H), 2.90–2.78 (m, 1H), 2.75–2.63 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 160.3 (d, $J_{C-F} = 246.9$ Hz), 156.1 (d, $J_{C-F} = 2.8$ Hz), 144.3, 137.3, 134.3, 132.0 (d, $J_{C-F} = 5.5$ Hz), 131.0 (d, $J_{C-F} = 9.2$ Hz), 129.2, 129.0, 127.6 (d, $J_{C-F} = 9.9$ Hz), 126.5 (d, $J_{C-F} = 3.4$ Hz), 125.7, 119.4 (d, $J_{C-F} = 25.8$ Hz), 110.2 (d, $J_{C-F} = 21.7$ Hz), 60.1, 58.0, 51.3, 40.7, 29.6; m/z (ESI–MS) 305.1 [M + H]⁺.

(±)-2c: Following general procedure G, product (±)-2c was obtained from 1c and 1,2,3,4tetrahydroisoquinoline (reflux for 1 h) as a yellow solid in 86% yield (137 mg), ($R_f = 0.31$ in hexane/EtOAc 70:30 v/v); mp = 179–181 °C; IR (KBr) 2926, 2810, 2757, 1601, 1484, 1407, 1372, 1325, 1299, 1160, 1142, 1104, 1075, 932, 919, 827, 757, 734, 670, 485; ¹H NMR (CDCl₃, 500 MHz) δ 7.93 (d, J = 8.9 Hz, 1H), 7.74 (s, 1H), 7.71 (d, J = 2.3 Hz, 1H), 7.57 (dd, J = 8.9, 2.3

Hz, 1H), 7.33 (d, J = 7.7 Hz, 1H), 7.28–7.23 (m, 1H), 7.23–7.18 (m, 1H), 7.16 (d, J = 7.2 Hz, 1H), 4.22 (d, J = 15.1 Hz, 1H), 3.93–3.83 (comp, 2H), 3.79 (dd, J = 17.2, 3.9 Hz, 1H), 3.33–3.14 (comp, 3H), 2.88–2.76 (m, 1H), 2.74–2.62 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 157.1, 145.6, 137.2, 134.2, 131.67, 131.66, 130.2, 130.0, 129.4, 129.0, 127.6, 126.52, 126.48, 125.9,

125.6, 60.0, 58.0, 51.3, 40.8, 29.6; m/z (ESI-MS) (³⁵Cl) 321.0 [M + H]⁺, (³⁷Cl) 323.0 [M + $H]^{+}$.

 (\pm) -2d: Following general procedure G, product (\pm) -2d was obtained from 1d and 1,2,3,4tetrahydroisoquinoline (reflux for 1.5 h) as a yellow solid in 85% yield (156 mg), ($R_f = 0.33$ in hexane/EtOAc 70:30 v/v); mp = 180–183 °C; IR (KBr) 2924, 2756, 2360, 1596, 1480, 1406, 1324, 1298, 1141, 1103, 913, 900, 827, 737; ¹H NMR (CDCl₃, 500 MHz) δ 7.90 (d, J = 2.2 Hz, 1H), 7.87 (d, J = 8.9 Hz, 1H), 7.75 (s, 1H), 7.71 (dd, J = 9.0, 2.2 Hz, 1H), 7.33 (d, J = 7.7 Hz, 1H), 7.28–7.23 (m, 1H), 7.23–7.19 (m, 1H), 7.17 (d, J = 7.2, 1H), 4.23 (d, J = 15.1 Hz, 1H), 3.93– 3.83 (comp, 2H), 3.79 (dd, J = 17.3, 3.9 Hz, 1H), 3.35–3.13 (comp, 3H), 2.90–2.78 (m, 1H), 2.75–2.63 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 157.3, 145.8, 137.2, 134.2, 132.6, 131.6, 130.4, 129.4, 129.3, 129.0, 128.2, 126.54, 126.50, 125.6, 119.8, 60.0, 58.0, 51.3, 40.8, 29.6; m/z (ESI-MS) (⁷⁹Br) 365.0 [M + H]⁺, (⁸¹Br) 366.9 [M + H]⁺.

(\pm)-2e: Following general procedure G, product (\pm)-2e was obtained from 1e and 1,2,3,4tetrahydroisoquinoline (reflux for 1.5 h) as a yellow solid in 83% yield (136 mg), ($R_f = 0.12$ in hexane/EtOAc 70:30 v/v); mp = 201–203 °C; IR (KBr) 2893, 2813, 1621, 1495, 1461, 1395, 1238, 1214, 1164, 1098, 1035, 947, 877, 800, 732, 585, 458; ¹H NMR (CDCl₃, 500 MHz) δ 7.67 (s, 1H), 7.34 (d, J = 7.7 Hz, 1H), 7.30 (s, 1H), 7.25–7.22 (m, 1H), 7.22–7.18 (m, 1H), 7.17–7.13

(m, 1H), 6.99 (s, 1H), 6.08 (d, J = 1.1 Hz, 1H), 6.07 (d, J = 1.2 Hz, 1H), 4.17 (d, J = 14.7 Hz, 1H), 3.90-3.80 (comp, 2H), 3.73 (dd, J = 17.0, 3.9 Hz, 1H), 3.34-3.20 (comp, 2H), 3.16 (dd, J = 17.0, 3.9 Hz, 1H), 3.34-3.20 (comp, 2H), 3.16 (dd, J = 17.0, 3.9 Hz, 1H), 3.34-3.20 (comp, 2H), 3.16 (dd, J = 17.0, 3.9 Hz, 1H), 3.34-3.20 (comp, 2H), 3.16 (dd, J = 17.0, 3.9 Hz, 1H), 3.34-3.20 (comp, 2H), 3.16 (dd, J = 17.0, 3.9 Hz, 1H), 3.34-3.20 (comp, 2H), 3.16 (dd, J = 17.0, 3.9 Hz, 1H), 3.34-3.20 (comp, 2H), 3.16 (dd, J = 17.0, 3.9 Hz, 100 Hz, 16.9, 11.5 Hz, 1H), 2.88–2.77 (m, 1H), 2.74–2.63 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 154.0, 150.5, 147.5, 145.4, 137.6, 134.3, 131.8, 129.0, 126.43, 126.40, 125.7, 123.9, 105.2, 102.3, 101.7, 60.2, 57.9, 51.4, 40.4, 29.6; m/z (ESI-MS) 331.1 [M + H]⁺.

 $(\pm)-2f:$

Following general procedure G, product (±)-2f was obtained from 1f and 1,2,3,4tetrahydroisoquinoline (reflux for 1 h) as a light yellow solid in 80% yield (142 mg), ($R_f = 0.26$ in hexane/EtOAc 70:30 v/v); mp = 159-160 °C; IR (KBr) 3027, 2930, 2815, 2748, 2361, 2344, 1673, 1616, 1561, 1493, 1436, 1334, 1319, 1297, 1224, 1197, 1171, 1141, 1123,

1104, 1059, 940, 905, 823, 738, 688, 483; ¹H NMR (CDCl₃, 500 MHz) δ 8.33 (dd, J = 2.0, 1.1Hz, 1H), 7.92 (s, 1H), 7.87 (d, J = 8.5 Hz, 1H), 7.65 (dd, J = 8.5, 1.8 Hz, 1H), 7.35 (d, J = 7.7 Hz, 1H), 7.30–7.25 (m, 1H), 7.25–7.20 (m, 1H), 7.18 (dd, J = 7.7, 1.5 Hz, 1H), 4.28 (d, J = 15.2 Hz, 1H), 3.97-3.88 (comp, 2H), 3.84 (dd, J = 17.3, 3.9 Hz, 1H), 3.36-3.21 (comp, 3H), 2.90-2.80 (m, 1H), 2.78–2.67 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 158.6, 146.1, 137.1, 134.2, 132.4, 130.8 (q, $J_{C-F} = 32.5$ Hz), 130.6, 129.0, 128.5, 126.59, 126.57, 126.54, 126.50, 125.6, 124.2 (q, $J_{C-F} = 272.4$ Hz), 121.7 (q, $J_{C-F} = 3.2$ Hz), 60.0, 58.0, 51.3, 40.8, 29.6; m/z (ESI–MS) $355.1 [M + H]^+$.

 (\pm) -2g: Following general procedure G, product (\pm) -2g was obtained from 1g and 1,2,3,4tetrahydroisoquinoline (reflux for 1 h) as a white solid in 87% yield (131 Me mg), ($R_f = 0.11$ in hexane/EtOAc 50:50 v/v); mp = 171–173 °C; IR (KBr) 3061, 2945, 2897, 2792, 1671, 1600, 1497, 1466, 1387, 1269, 1252, 1111, 1051, 813, 756, 744, 518; ¹H NMR (CDCl₃, 500 MHz) δ 8.01 (app d, J = 8.3 Hz, 1H), 7.92 (app d, J = 8.1 Hz, 1H), 7.64 (ddd, J = 8.4, 6.8, 1.4 Hz, 1H), 7.51 (ddd, J = 8.2, 6.8, 1.3 Hz, 1H), 7.40 (app d, J = 7.6 Hz, 1H), 7.31–7.27 (m, 1H), 7.26–7.22 (m, 1H), 7.21–7.17 (m, 1H), 4.16 (d, J = 15.6 Hz, 1H), 3.89–3.76 (comp, 2H), 3.72 (app dt, J = 15.6, 2.0 Hz, 1H), 3.37–3.21 (comp, 2H), 3.19–3.07 (m, 1H), 2.90–2.74 (comp, 2H), 2.65 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 156.2, 146.1, 139.2, 137.5, 134.7, 129.25, 129.17, 128.6, 126.8, 126.6, 126.4, 126.3, 125.9, 125.6, 122.5, 58.7, 56.4, 51.4, 33.6, 29.6, 22.8; m/z (ESI–MS) 300.9 [M + H]⁺.

(±)-**2h**: Following general procedure G, product (±)-**2h** was obtained from **1h** and 1,2,3,4tetrahydroisoquinoline (reflux for 1 h) as a yellow solid in 87% yield (137 mg), ($R_f = 0.24$ in hexane/EtOAc 70:30 v/v); mp = 116–119 °C; IR (KBr) 3059, 2970, 2939, 2742, 1588, 1496, 1458, 1408, 1371, 1291, 1156, 1101, 1036, 1013, 742, 477; ¹H NMR (CDCl₃, 500 MHz) δ 8.06–7.96 (comp, 2H), 7.64 (ddd, J = 8.4, 6.7, 1.4 Hz, 1H), 7.50 (ddd, J = 8.4, 6.6, 1.3 Hz, 1H), 7.35

(app d, J = 7.7 Hz, 1H), 7.29–7.13 (comp, 3H), 4.35 (d, J = 15.1 Hz, 1H), 3.94–3.67 (comp, 3H), 3.38–3.17 (comp, 3H) 3.15–2.95 (comp, 2H), 2.93–2.79 (m, 1H), 2.78–2.69 (m, 1H), 1.32 (t, J = 7.6 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 156.3, 147.1, 145.2, 137.4, 134.3, 129.4, 128.9, 128.6, 126.43, 126.40, 125.84, 125.79, 125.7, 125.3, 123.3, 59.5, 56.2, 51.6, 41.5, 29.6, 20.6, 13.9; m/z (ESI–MS) 315.2 [M + H]⁺.

(±)-2i: Following general procedure G, product (±)-2i was obtained from 1i and 1,2,3,4tetrahydroisoquinoline (reflux for 1 h) as a yellow solid in 93% yield (184 mg), ($R_f = 0.46$ in hexane/EtOAc 70:30 v/v); mp = 205–207 °C; IR (KBr) 3026, 2893, 2746, 1671, 1581, 1483, 1366, 1333, 1270, 1172, 1103, 953, 841, 758, 724, 704, 678, 613; ¹H NMR (CDCl₃, 500 MHz) δ 7.99 (d, J = 9.0 Hz, 1H), 7.64–7.48 (comp, 4H), 7.40–7.33 (comp, 2H),

7.32–7.28 (m, 1H), 7.28–7.23 (comp, 2H), 7.22–7.17 (m, 1H), 7.14 (app d, J = 7.5 Hz, 1H), 3.91–3.81 (comp, 3H), 3.55 (d, J = 15.8 Hz, 1H), 3.28 (dd, J = 17.7, 12.2 Hz, 1H), 3.17 (ddd, J = 16.7, 12.0, 5.2 Hz, 1H), 3.05 (ddd, J = 11.2, 5.3, 1.9 Hz, 1H), 2.79–2.70 (m, 1H), 2.57 (app td, J = 11.6, 3.4 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 156.9, 145.5, 144.2, 137.2, 135.4, 134.3, 131.8, 130.4, 129.9, 129.2, 129.1, 129.0, 128.9, 128.6, 127.3, 126.9, 126.51, 126.50, 125.6, 124.7, 59.8, 57.2, 51.4, 41.4, 29.6; m/z (ESI–MS) (³⁵Cl) 397.0 [M + H]⁺, (³⁷Cl) 399.0 [M + H]⁺.

(\pm)-2j and (\pm)-2j': Following the general procedure G, products (\pm)-2j and (\pm)-2j' were obtained from 1j and 1,2,3,4-tetrahydroisoquinoline (reflux for 18 h) in a 4:1 ratio (153 mg, 84% combined yield).

Characterization data for major diastereomer (±)-2j: Yellow liquid (68% yield, 123 mg), ($R_f =$

N Ph

0.20 in hexane/EtOAc 70:30 v/v); mp = 113–115 °C; IR (KBr) 3025, 2916, 1736, 1619, 1599, 1493, 1452, 1421, 1308, 1240, 1155, 1041, 783, 746, 699; ¹H NMR (CDCl₃, 500 MHz) δ 7.91 (s, 1H), 7.85 (d, *J* = 8.5 Hz, 1H), 7.74 (dd, *J* = 8.4, 1.5 Hz, 1H), 7.54 (ddd, *J* = 8.5, 6.8, 1.5 Hz, 1H), 7.44 (ddd, *J* =

8.2, 6.8, 1.2 Hz, 1H), 3.35–3.10 (comp, 3H), 7.27–7.20 (m, 1H), 7.20–7.09 (comp, 4H), 6.89 (ddd, J = 8.3, 5.8, 2.9 Hz, 1H), 6.35 (app d, J = 7.8 Hz, 1H), 4.66 (d, J = 15.9 Hz, 1H), 4.52 (comp, 2H), 4.19 (d, J = 15.9 Hz, 1H), 3.19 (ddd, J = 11.4, 8.0, 5.4 Hz, 1H), 3.13–2.98 (comp, 2H), 2.90 (ddd, J = 11.1, 6.3, 4.6 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 159.8, 147.9, 144.6,

137.2, 133.5, 133.2, 130.4, 129.3, 129.1, 128.8, 128.5, 127.7, 127.2, 127.01, 126.98, 126.8, 126.4, 126.2, 125.0, 66.1, 56.8, 52.9, 46.7, 29.1; *m*/*z* (ESI–MS) 363.0 [M + H]⁺.

X-ray quality crystals of (\pm) -2j were obtained from hexane/ethyl acetate through slow diffusion at room temperature. The relative stereochemistry was assigned by X-ray crystallography



The requisite CIF has been submitted to the journal and deposited with the CCDC (deposition # 1541946).

Characterization data for minor diastereomer (±)-**2j**': Yellow solid (17% yield, 30 mg), (R_f = 0.51 in hexane/EtOAc 70:30 v/v); mp = 162–165 °C; IR (KBr) 3059, 3024, 2912, 2804, 2759, 2361, 2345, 1654, 1600, 1560, 1493, 1459, 1426, 1377, 1314, 1226, 1147, 1095, 913, 746, 733, 700, 575, 474; ¹H NMR (CDCl₃, 500 MHz) δ 8.03–7.93 (comp, 2H), 7.77 (dd, *J* = 8.4, 1.3 Hz, 1H), 7.60 (ddd, *J* = 8.5, 6.8, 1.5 Hz, 1H), 7.53–7.41 (comp, 2H), 7.24–7.19 (m, 1H), 7.17–7.09 (comp, 2H), 7.04 (app tt, *J* = 7.5, 1.0 Hz, 1H), 7.00–6.89 (comp, 4H), 4.92 (d, *J* = 3.4 Hz, 1H), 4.43 (dd, *J* = 15.0, 1.0 Hz, 1H), 4.34 (d, *J* = 2.9 Hz, 1H), 4.02 (d, *J* = 15.1 Hz, 1H), 3.21 (ddd, *J* = 10.5, 4.4, 2.0 Hz, 1H), 2.77 (dddd, *J* = 14.8, 11.7, 4.5, 1.4 Hz, 1H), 2.68 (ddd, *J* = 12.2, 10.3, 2.2 Hz, 1H), 2.52 (app dt, *J* = 15.0, 2.2 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃). δ 159.8, 147.5, 141.4, 136.0, 135.4, 133.0, 131.0, 129.2, 128.9, 128.64, 128.55, 127.18, 127.16, 126.8, 126.7, 126.2, 125.85, 125.82, 64.6, 57.6, 54.4, 50.6, 29.7; *m/z* (ESI–MS) 363.1 [M + H]⁺.

(\pm)-2k and (\pm)-2k': Following the general procedure G, products (\pm)-2k and (\pm)-2k' were obtained from 1k and 1,2,3,4-tetrahydroisoquinoline (reflux for 2 h) in a 2:1 ratio (143 mg, 95% combined yield).

Characterization data for major diastereomer (±)-**2k**: Yellow oil (63% yield, 95 mg), (R_f = 0.14 in hexane/EtOAc 70:30 v/v); IR (KBr) 2967, 2908, 2878, 2849, 2363, 1601, 1561, 1492, 1420, 1380, 1306, 1218, 1159, 1140, 1123, 1102, 1013, 966, 941, 908, 758, 742, 592, 450; ¹H NMR (CDCl₃, 500 MHz) δ 8.02 (app d, *J* = 8.5 Hz, 1H), 7.81 (s, 1H), 7.73 (app d, *J* = 7.6 Hz, 1H), 7.63 (ddd, *J* = 8.5, 6.8, 1.5 Hz, 1H), 7.46 (ddd, *J* = 8.1, 6.8, 1.2 Hz, 1H), 7.36 (dd, *J* = 7.0, 2.2 Hz, 1H), 7.23–7.08 (comp, 3H), 4.26 (d, *J* = 15.4 Hz, 1H), 4.03 (d, *J* = 15.7 Hz, 1H), 3.92 (d, *J* = 7.8 Hz, 1H), 3.51 (app p, *J* = 7.2 Hz, 1H), 3.21–3.11 (m, 1H), 3.08–2.96 (comp, 2H), 2.93–2.83 (m, 1H), 1.73 (d, *J* = 7.1 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 161.1, 147.6, 138.1, 133.8, 132.9, 129.2, 128.9, 128.8, 127.4, 127.1, 127.0, 126.80, 126.78, 126.0, 125.6, 65.7, 56.1, 48.0, 40.1, 28.5, 22.0; *m*/z (ESI–MS) 301.1 [M + H]⁺. Characterization data for minor diastereomer (\pm)-2k': Yellow oil (32% yield, 48 mg), (R_f = 0.54



in hexane/EtOAc 70:30 v/v); IR (KBr) 3024, 2983, 2935, 2796, 2756, 1619, 1494, 1450, 1424, 1359, 1318, 1234, 1191, 1111, 1060, 1019, 894, 859, 762, 743, 615, 481; ¹H NMR (CDCl₃, 500 MHz) δ 8.07–8.03 (m, 1H), 7.86 (s, 1H), 7.79–7.73 (m, 1H), 7.66 (ddd, J = 8.4, 6.8, 1.5 Hz, 1H), 7.48 (ddd, J =

8.1, 6.8, 1.2 Hz, 1H), 7.31 (app d, J = 7.8 Hz, 1H), 7.28–7.22 (m, 1H), 7.22–7.13 (comp, 2H), 4.25 (d, J = 15.0 Hz, 1H), 4.03 (d, J = 2.6 Hz, 1H), 3.85 (d, J = 15.0 Hz, 1H), 3.74 (qd, J = 7.0, 3.3 Hz, 1H), 3.28–3.14 (m, 2H), 2.80–2.71 (m, 1H), 2.70–2.62 (m, 1H) 1.13 (d, J = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 162.3, 147.3, 136.1, 135.9, 132.8, 129.0, 128.9, 128.7, 128.0, 127.3, 127.1, 126.5, 126.03, 125.98, 125.8, 63.7, 58.1, 51.1, 42.8, 29.8, 17.2; m/z (ESI–MS) 301.1 [M + H]⁺.

(\pm)-**2l** and (\pm)-**2l'**: To a solution of 1-(2-methylquinolin-3-yl)ethan-1-one (93 mg, 0.5 mmol, 1 equiv) in toluene (0.1 M, 5 mL) were added 1,2,3,4-tetrahydroisoquinoline (94 µL, 0.75 mmol, 1.5 equiv) and acetic acid (0.572 mL, 10 mmol, 20 equiv). The mixture was heated under reflux for 24 h. The reaction was then allowed to cool to room temperature, diluted with EtOAc (15 mL), and washed with 1N NaOH (2 x 20 mL). The combined aqueous layers were extracted with EtOAc (2 x 15 mL), and the combined organic layers washed with brine (40 mL), and dried over anhydrous Na₂SO₄. Solvent was then removed under reduced pressure and the residue was purified by silica gel chromatography. Products (\pm)-**2l** and (\pm)-**2l'** were obtained in a 3:1 ratio (126 mg, 83% combined yield).

Characterization data for major diastereomer (\pm)-21: Yellow solid (63% yield, 95 mg), (R_f = 0.21



in hexane/EtOAc 70:30 v/v); mp = 154–156 °C; IR (KBr) 2969, 2941, 2904, 2821, 2364, 1622, 1604, 1496, 1450, 1381, 1381, 1297, 1171, 1143, 1104, 991, 908, 855, 761, 746, 479; ¹H NMR (CDCl₃, 500 MHz) δ 8.01 (d, J = 8.5Hz, 1H), 7.88 (s, 1H), 7.76 (dd, J = 8.1, 1.4 Hz, 1H), 7.65 (ddd, J = 8.4, 6.9, 1.5 Hz, 1H), 7.47 (ddd, J = 8.0, 6.9, 1.2 Hz, 1H), 7.31 (d, J = 7.6

Hz, 1H), 7.24 (app td, J = 7.3, 1.7 Hz, 1H), 7.19 (app td, J = 7.3, 1.4 Hz, 1H), 7.16 (dd, J = 7.6, 1.7 Hz, 1H), 4.53 (dd, J = 11.4, 4.3 Hz, 1H), 4.46 (q, J = 6.9 Hz, 1H), 3.64 (dd, J = 17.5, 4.3 Hz, 1H), 3.18 (dd, J = 17.5, 11.4 Hz, 1H), 3.14–3.03 (comp, 2H), 2.97 (ddd, J = 10.7, 9.7, 3.9 Hz, 1H), 2.91 (app dt, J = 16.2, 4.2 Hz, 1H), 1.48 (d, J = 6.8 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 156.2, 147.2, 138.4, 134.4, 133.9, 133.5, 129.10, 129.08, 128.5, 127.3, 127.2, 126.5, 126.2, 125.9, 59.1, 51.0, 47.3, 40.5, 30.2, 17.8; m/z (ESI–MS) 301.1 [M + H]⁺.

X-ray quality crystals of (\pm) -**2l** were obtained from hexane/ethyl acetate through slow diffusion at room temperature. The relative stereochemistry was assigned by X-ray crystallography



The requisite CIF has been submitted to the journal and deposited with the CCDC (deposition # 1541945).

Characterization data for minor diastereomer (±)-2l': Yellow oil (20% yield, 31 mg), ($R_f = 0.29$



in hexane/EtOAc 70:30 v/v); IR (KBr) 3024, 2983, 2935, 2796, 2756, 1619, 1494, 1450, 1424, 1359, 1318, 1234, 1191, 1111, 1060, 1019, 894, 859, 762, 743, 615, 481; ¹H NMR (CDCl₃, 500 MHz) δ 8.02 (app d, *J* = 8.5, 1H), 7.99 (s, 1H), 7.78 (dd, *J* = 8.1, 1.4 Hz, 1H), 7.67 (ddd, *J* = 8.4, 6.8, 1.4 Hz, 1H), 7.48 (ddd, *J* = 8.1, 6.8, 1.2 Hz, 1H), 7.34 (app d, *J* = 7.6 Hz, 1H), 7.23 (app

td, J = 7.3, 1.9 Hz, 1H), 7.21–7.15 (comp, 2H), 4.03 (dd, J = 11.5, 2.3 Hz, 1H), 3.98 (q, J = 6.4 Hz, 1H), 3.72 (dd, J = 16.8, 3.3 Hz, 1H), 3.44 (ddd, J = 11.3, 5.2, 3.1 Hz, 1H), 3.30 (dd, J = 16.8, 11.6 Hz, 1H), 3.17 (ddd, J = 15.8, 10.9, 4.9 Hz, 1H), 2.86 (app dt, J = 16.2, 3.4 Hz, 1H), 2.55 (app td, J = 11.1, 3.6 Hz, 1H), 1.69 (d, J = 6.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃). δ 156.8, 146.7, 138.0, 134.7, 133.5, 133.4, 129.3, 128.9, 128.4, 127.5, 127.4, 126.4, 126.3, 126.0, 59.5, 59.3, 46.6, 41.2, 30.2, 21.8; m/z (ESI–MS) 301.0 [M + H]⁺.

(±)-**2m**: Following general procedure H, product (±)-**2m** was obtained from **1m** and 1,2,3,4tetrahydroisoquinoline (reflux for 2 h) as a white solid in 89% yield (105 mg), (R_f = 0.15 in hexane/EtOAc 20:80 v/v); mp = 98–100 °C; IR (KBr) 2957, 2920, 2816, 2755, 1668, 1578, 1493, 1446, 1338, 1293, 1140, 1104, 976, 778, 740, 722, 490; ¹H NMR (CDCl₃, 500 MHz) δ 8.43 (app d, *J* = 4.8 Hz, 1H), 7.39 (app d, *J* = 7.7 Hz, 1H), 7.30 (app d, *J* = 7.8 Hz, 1H), 7.27–7.13 (comp, 3H), 7.09 (dd, *J* = 7.8, 4.8 Hz, 1H), 4.02 (d, *J* = 14.9 Hz, 1H), 3.79 (dd, *J* = 11.3, 3.1 Hz, 1H), 3.72 (d, *J* = 14.9 Hz, 1H), 3.59 (dd, *J* = 17.0, 4.0 Hz, 1H), 3.37–3.16 (comp, 2H), 3.06 (dd, *J* = 17.0, 11.4 Hz, 1H), 2.87–2.74 (m, 1H), 2.71–2.57 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 155.0, 147.9, 137.5, 134.3, 134.0, 129.9, 128.9, 126.39, 126.37, 125.6, 121.2, 59.9, 57.6, 51.2, 39.8, 29.6; *m/z* (ESI–MS) 237.2 [M + H]⁺.

(±)-**2n**: Following general procedure H, product (±)-**2n** was obtained from **1n** and 1,2,3,4tetrahydroisoquinoline (reflux for 2 h) as a white solid in 74% yield (93 mg), (R_f = 0.20 in hexane/EtOAc 20:80 v/v); mp = 152–155 °C; IR (KBr) 3009, 2938, 2892, 2795, 2750, 1601, 1576, 1496, 1468, 1360, 1287, 1248, 1141, 1108, 1015, 973, 873, 788, 751, 723, 560, 515, 478; ¹H NMR (CDCl₃, 500 MHz) δ 8.26 (s, 1H), 7.29 (app d, *J* = 7.7 Hz, 1H), 7.23 (app td, *J* = 7.4, 1.7 Hz, 1H), 7.20–7.16 (comp, 2H), 7.14 (app d, *J* = 7.3 Hz, 1H), 3.98 (d, *J* = 14.9 Hz, 1H), 3.76 (dd, *J* = 11.4, 4.0 Hz, 1H), 2.68 (d, *L* = 14.0 Hz, 1H), 2.54 (dd, *L* = 16.0, 2.0 Hz, 1H), 2.27, 2.12 (appr 2H), 2.00 (dd

1H), 3.68 (d, J = 14.9 Hz, 1H), 3.54 (dd, J = 16.9, 3.9 Hz, 1H), 3.27–3.12 (comp, 2H), 3.00 (dd, J = 16.8, 11.4 Hz, 1H), 2.83–2.73 (m, 1H), 2.65 (ddd, J = 12.0, 11.2, 3.5 Hz, 1H), 2.29 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 151.9, 148.3, 137.6, 134.4, 134.3, 130.5, 129.2, 128.9, 126.4, 126.3, 125.6, 60.0, 57.5, 51.2, 39.4, 29.6, 18.2; m/z (ESI–MS) 251.1 [M + H]⁺.

(\pm)-20 and (\pm)-20': Following the general procedure H, products (\pm)-20 and (\pm)-20' were obtained from 10 and 1,2,3,4-tetrahydroisoquinoline (reflux for 1.5 h) in a 3:1 ratio (125 mg, 81% combined yield).

Characterization data for major diastereomer (\pm)-20: Yellow oil (60% yield, 160 mg), (R_f = 0.34



in hexane/EtOAc 50:50 v/v; IR (KBr) 3027, 2922, 2891, 2781, 1576, 1490, 1437, 1388, 1356, 1287, 1206, 1143, 1105, 1078, 1036, 948, 789, 755, 732, 703, 604, 564, 465; ¹H NMR (CDCl₃, 500 MHz) δ 8.42–8.37 (m, 1H), 7.50–7.42 (m, 1H), 7.37–7.28 (comp, 2H), 7.28–7.19 (m, 1H), 7.17–7.09 (comp, 2H), 7.08–

7.00 (comp, 3H), 6.89–6.76 (m, 1H), 6.16 (app d, J = 7.8 Hz, 1H), 4.49 (d, J = 16.4 Hz, 1H), 4.37 (d, J = 9.6 Hz, 1H), 4.29 (d, J = 9.6 Hz, 1H), 3.97 (d, J = 16.0 Hz, 1H), 3.22 (ddd, J = 11.4, 7.7, 5.8 Hz, 1H), 3.13–2.99 (comp, 2H), 2.92–2.82 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 158.0, 148.7, 144.0, 136.8, 134.3, 133.6, 130.3, 129.0, 128.9, 128.6, 127.8, 126.6, 126.5, 124.7, 121.2, 65.7, 56.5, 52.1, 46.7, 29.3; m/z (ESI–MS) 313.1 [M + H]⁺.

Characterization data for minor diastereomer (±)-**20**': Yellow oil (21% yield, 32 mg), (R_f = 0.19 in hexane/EtOAc 50:50 v/v); mp = 142–144 °C; IR (KBr) 3026, 2890, 2837, 1575, 1490, 1437, 1388, 1356, 1211, 1143, 1105, 788, 755, 731, 703, 604, 564, 473; ¹H NMR (CDCl₃, 500 MHz) δ 8.48–8.39 (m, 1H), 7.49 (app d, *J* = 7.7 Hz, 1H), 7.37 (app d, *J* = 7.9 Hz, 1H), 7.24–7.14 (comp, 3H), 7.11 (app dd, *J* = 7.7, 4.7 Hz, 1H), 7.06–6.92 (comp, 4H), 6.89 (app d, *J* = 7.6 Hz, 1H), 4.68 (d, *J* = 3.6 Hz, 1H), 4.19 (d, *J* = 15.0 Hz, 1H), 3.83 (d, *J* = 15.0 Hz, 1H), 3.19 (ddd, *J* = 10.7, 4.8, 2.1 Hz, 1H), 2.82 (app tdd, *J* = 13.2, 5.1, 2.7 Hz, 1H), 2.65 (ddd, *J* = 11.9, 10.6, 2.7 Hz, 1H), 2.52 (app dt, *J* = 15.3, 2.3 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 158.2, 148.4, 141.2, 135.8, 135.4, 134.2, 130.9, 130.1, 128.6, 127.0, 126.7, 126.0, 125.74, 125.72, 121.4, 64.3, 57.2, 53.5, 50.6, 29.7; *m*/z (ESI–MS) 313.0 [M + H]⁺.

(\pm)-**2p and** (\pm)-**2p'**: Following the general procedure H, products (\pm)-**2p** and (\pm)-**2p'** were obtained from **1p** and 1,2,3,4-tetrahydroisoquinoline (reflux for 1.5 h) in a 2:1 ratio (120 mg, 96% combined yield).

Characterization data for major diastereomer (\pm)-**2p**: Yellow oil (63% yield, 79 mg), (R_f = 0.10



in hexane/EtOAc 50:50 v/v; IR (KBr) 3052, 2987, 2687, 2411, 2359, 2301, 1421, 1266, 893, 748, 689, ; ¹H NMR (CDCl₃, 500 MHz) δ 8.50–8.44 (m, 1H), 7.35 (ddd, J = 7.6, 1.7, 0.8 Hz, 1H), 7.29 (dd, J = 7.3, 1.5 Hz, 1H), 7.23–7.12 (comp, 3H), 7.07 (dd, J = 7.6, 4.8 Hz, 1H), 4.16 (d, J = 15.7 Hz, 1H), 3.85–3.78

(comp, 2H), 3.26 (app p, J = 7.2 Hz, 1H), 3.17 (ddd, J = 11.6, 7.1, 6.0 Hz, 1H), 3.09–2.99 (comp, 2H), 2.86 (app dt, J = 12.0, 6.1 Hz, 1H), 1.61 (d, J = 6.9 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 159.0, 148.0, 137.8, 134.3, 133.9, 129.3, 128.8, 127.1, 126.8, 125.4, 121.1, 65.1, 55.6, 47.5, 38.4, 28.4, 20.4; m/z (ESI–MS) 250.9 [M + H]⁺.

Characterization data for minor diastereomer (±)-**2p**': Yellow oil (33% yield, 41 mg), (R_f = 0.29 in hexane/EtOAc 50:50 v/v); IR (KBr) 3057, 2969, 2919, 1586, 1572, 1492, 1449, 1439, 1380, 1307, 1142, 1123, 1100, 1010, 941, 845, 795, 780, 762, 732, 455; ¹H NMR (CDCl₃, 500 MHz) δ 8.50–8.43 (m, 1H), 7.43–7.38 (m, 1H), 7.26–7.20 (comp, 2H), 7.19–7.12 (comp, 2H), 7.10 (dd, J = 7.7, 4.7 Hz, 1H), 4.01 (d, J = 15.0 Hz, 1H), 3.92 (d, J = 3.4, 1H), 3.68 (dd, J = 14.9, 1.0 Hz, 1H), 3.50 (qd, J = 7.0, 3.4 Hz, 1H), 3.28–3.10 (comp, 2H), 2.82–2.68 (m, 1H), 2.68–2.57 (m, 1H), 1.04 (d, J = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃). δ 160.5, 147.8, 136.1, 135.8, 134.0, 129.5, 128.8, 126.4, 125.90, 125.86, 121.1, 63.4, 57.6, 50.9, 41.7, 29.8, 16.6; m/z (ESI–MS) 251.0 [M + H]⁺. (±)-2q: Following general procedure G, product (±)-2q was obtained from 1a and 6,7dimethoxy-1,2,3,4-tetrahydroisoquinoline (reflux for 1.5 h) as a yellow solid in 93% yield (162 mg), ($R_f = 0.12$ in hexane/EtOAc 50:50 v/v); mp = 67-69 °C; IR (KBr) 2935, 2827, 1609, 1516 1498, 1464, 1430, 1375, 1363, 1260, 1230, 1209, 1134, 1110, 1021, 855, 786, 752; ¹H NMR (CDCl₃, 500 MHz) δ 7.95 (app d, J = 8.5 Hz, 1H), 7.74 (s, 1H),

7.66 (app d, J = 7.9 Hz, 1H), 7.58 (ddd, J = 8.4, 6.8, 1.5 Hz, 1H), 7.40 (ddd, J = 8.1, 6.9, 1.2 Hz, 1H), 6.76 (s, 1H), 6.57 (s, 1H), 4.13 (d, J = 14.9 Hz, 1H), 3.82 (s, 3H), 3.80 (s, 3H), 3.77–3.62 (comp, 3H), 3.24–3.04 (comp, 3H), 2.72–2.53 (comp, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 13C NMR (126 MHz, cdcl3) δ 156.4, 147.52, 147.48, 146.9, 132.5, 129.03, 128.95, 128.3, 128.1, 127.2, 126.9, 126.2, 125.8, 111.2, 108.2, 77.4, 59.6, 57.8, 55.82, 55.78, 51.4, 40.8, 28.9; m/z (ESI–MS) 347.0 [M + H]⁺.

(±)-2**r**: Following general procedure G, product (±)-2**r** was obtained from 1**a** and 2,3,4,9tetrahydro-1*H*-pyrido[3,4-*b*]indole (reflux for 1.5 h) as a yellow solid in 74% yield (120 mg), ($R_f = 0.29$ in hexane/EtOAc 50:50 v/v); mp (decomposition at 230 °C); IR (KBr) 3268, 2942, 2896, 2824, 1604, 1492, 1452, 1431, 1314, 1276, 1164, 1128, 1063, 983, 906, 858, 743, 617, 479; ¹H NMR (DMSO, 500 MHz) δ 10.99 (s, 1H), 8.07 (s, 1H),

7.94 (app d, J = 8.4 Hz, 1H), 7.88 (app d, J = 8.1 Hz, 1H), 7.71–7.65 (m, 1H), 7.52 (app t, J = 7.5 Hz, 1H), 7.43 (app d, J = 7.8 Hz, 1H), 7.37 (app d, J = 8.0 Hz, 1H), 7.08 (app t, J = 7.5 Hz, 1H), 6.99 (app t, J = 7.4 Hz, 1H), 4.27 (d, J = 15.3 Hz, 1H), 4.00–3.75 (comp, 3H), 3.28 (dd, J = 11.2, 5.2 Hz, 1H), 3.01 (dd, J = 16.5, 11.4 Hz, 1H), 2.89 (dddd, J = 14.3, 11.7, 5.5, 2.3 Hz, 1H), 2.80–2.72 (m, 1H), 2.65 (app td, J = 11.3, 4.0 Hz, 1H); ¹³C NMR (125 MHz, DMSO) δ 155.8, 146.4, 136.3, 134.9, 132.5, 128.9, 128.6, 128.1, 127.4, 126.6, 126.5, 125.9, 120.7, 118.4, 117.7, 111.1, 106.6, 56.4, 56.1, 51.6, 38.4, 21.2; m/z (ESI–MS) 326.0 [M + H]⁺.

(±)-2s: Following general procedure G, product (±)-2s was obtained from 1a and 2,3-dihydro-1*H*-benzo[*de*]isoquinoline (reflux for 1 h) as a white solid in 50% yield (80 mg), ($R_f = 0.24$ in hexane/EtOAc 50:50 v/v); mp (decomposition at 150 °C); IR (KBr) 3047, 2928, 2743, 2361, 1624, 1600, 1497, 1430, 1415, 1320, 1306, 1164, 1129, 1086, 1006, 976, 822, 776, 745, 479; ¹H NMR (CDCl₃, 500 MHz) δ 8.04 (app d, J = 8.5 Hz, 1H), 7.86 (s, 1H), 7.83–7.72 (comp, 3H), 7.67

(ddd, J = 8.4, 6.8, 1.5 Hz, 1H), 7.59–7.40 (comp, 4H), 7.28 (app d, J = 7.0 Hz, 1H), 4.41 (dd, J = 10.5, 4.7 Hz, 1H), 4.35 (app d, J = 14.9 Hz, 2H), 4.07 (app dd, J = 20.6, 14.9 Hz, 2H), 3.94 (dd, J = 17.5, 4.7 Hz, 1H), 3.45 (dd, J = 17.5, 10.4 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 155.8, 147.4, 136.1, 133.2, 132.8, 132.1, 129.2, 128.6, 127.5, 127.4, 127.34, 127.27, 126.7, 126.6, 126.11, 126.09, 125.7, 122.1, 121.8, 59.1, 56.7, 56.0, 38.6; m/z (ESI–MS) 323.2 [M + H]⁺.

(±)-2t: Following general procedure G, product (±)-2t was obtained from 1a and 1-phenyl-1,2,3,4-tetrahydroisoquinoline (reflux for 12 h) as a white solid in 63% yield (114 mg), ($R_f = 0.69$ in hexane/EtOAc 50:50 v/v); mp = 176–179 °C; IR (KBr) 3050, 2918, 2853, 2361, 2209, 1621, 1492, 1470, 1428, 1316, 1159, 1015, 916, 905, 771, 751, 727, 704, 479; ¹H NMR (CDCl₃, 500 MHz) δ

8.10–8.06 (m, 1H), 7.76–7.70 (comp, 2H), 7.68 (ddd, J = 8.4, 6.9, 1.5 Hz, 1H), 7.48 (ddd, J = 8.1, 6.9, 1.2 Hz, 1H), 7.25–7.12 (comp, 7H), 7.08 (ddd, J = 7.8, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1H), 6.87 (dd, J = 5.4, 6.9, 1.8 Hz, 1.8 Hz,

7.9, 1.2 Hz, 1H), 4.07 (d, J = 18.1 Hz, 1H), 3.94–3.77 (comp, 2H), 3.71 (d, J = 18.1 Hz, 1H), 3.35–3.22 (m, 1H), 3.22–3.11 (comp, 2H), 3.07 (app dt, J = 15.3, 5.0 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 156.6, 147.4, 143.9, 143.4, 133.1, 132.5, 129.0, 128.8, 128.6, 128.3, 127.9, 127.31, 127.29, 127.0, 126.3, 126.2, 126.0, 63.1, 52.8, 46.4, 40.9, 29.9; *m*/*z* (ESI–MS) 363.1 [M + H]⁺.

(±)-**2u**: Following general procedure G, product (±)-**2u** was obtained from **1a** and 1-phenyl-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole (reflux for 14 h) as a white solid in 80% yield (160 mg), ($R_f = 0.60$ in hexane/EtOAc 50:50 v/v); mp = 240–242 °C; IR (KBr) 3053, 2910, 2834, 1736, 1618, 1598, 1561, 1491, 1464, 1446, 1413, 1312, 1303, 1236, 1178, 1110, 1098, 1045, 988, 908, 744, 704, 435; ¹H NMR (CDCl₃, 500 MHz) δ 8.03 (app d, *J* = 8.6 Hz, 1H), 7.84 (br s, 1H), 7.80 (s, 1H), 7.77–7.72 (m, 1H), 7.67 (ddd, *J* = 8.4, 6.9, 1.4 Hz, 1H), 7.61–7.56 (m, 1H), 7.50 (ddd, *J* = 8.1, 6.8, 1.2 Hz, 1H), 7.29–7.23 (m, 1H), 7.21–7.11 (comp, 7H), 4.06 (d, *J* = 17.6 Hz, 1H), 3.98 (dd, *J* = 16.8 Hz, 1H), 3.85 (d, *J* = 7.1 Hz, 1H), 3.81 (d, *J* = 5.9 Hz, 1H), 3.26–3.17 (comp, 2H), 3.14–3.07 (comp, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 156.0, 147.3, 140.6, 138.7,

136.6, 132.8, 129.2, 128.8, 128.6, 128.5, 128.3, 127.9, 127.4, 127.3, 127.1, 126.2, 121.9, 119.6, 118.6, 111.3, 108.1, 60.4, 51.7, 47.1, 40.8, 21.8; *m/z* (ESI–MS) 402.0 [M + H]⁺.
(±)-4a: Following general procedure H, product (±)-4a was obtained from 3a and 1,2,3,4-



tetrahydroisoquinoline (reflux for 2 h) as a yellow solid in 57% yield (82 mg), ($R_f = 0.10$ in hexane/EtOAc 50:50 v/v); mp = 70–72 °C; IR (KBr) 3027, 2917, 2752, 2364, 2344, 1736, 1689, 1653, 1621, 1506, 1501, 1456, 1388, 1143, 1108, 753, 733; ¹H NMR (CDCl₃, 500 MHz) δ 8.67 (s, 1H), 8.09 (dd, J

= 8.5, 1.2 Hz, 1H), 7.95 (dd, J = 8.4, 1.3 Hz, 1H), 7.67 (ddd, J = 8.3, 6.8, 1.4 Hz, 1H), 7.56 (ddd, J = 8.2, 6.8, 1.3 Hz, 1H), 7.39 (app d, J = 7.5 Hz, 1H), 7.28 (app td, J = 7.5, 1.6 Hz, 1H), 7.25–7.21 (m, 1H), 7.19 (app d, J = 7.4 Hz, 1H), 4.19 (d, J = 15.4 Hz, 1H), 3.98–3.75 (comp, 3H), 3.32–3.19 (comp, 2H), 3.18–3.06 (m, 1H), 2.93–2.56 (comp, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 149.4, 146.8, 139.4, 137.5, 134.6, 130.2, 129.2, 128.6, 127.6, 127.2, 126.8, 126.6, 126.4, 125.6, 122.6, 59.2, 56.4, 51.2, 33.1, 29.6; m/z (ESI–MS) 287.0 [M + H]⁺.

(±)-**4b**: Following general procedure G, product (±)-**4b** was obtained from **3b** and 1,2,3,4tetrahydroisoquinoline (reflux for 1 h) as a yellow oil in 91% yield (107 mg), (R_f = 0.22 in hexane/EtOAc 20:80 v/v); IR (KBr) 3054, 2987, 2684, 2411, 2304, 1596, 1418, 1263, 1148, 1100, 888, 745, 698; ¹H NMR (CDCl₃, 500 MHz) δ 8.43–8.32 (comp, 2H), 7.26–7.17 (comp, 3H), 7.17–7.12 (m, 1H), 7.09 (app d, *J* = 5.0 Hz, 1H), 4.07 (d, *J* = 15.0 Hz, 1H), 3.78–3.66 (comp, 2H), 3.37 (dd, *J* = 17.0, 4.0 Hz, 1H), 3.27–3.14 (comp, 2H), 2.90 (app dddt, *J* = 17.0, 11.1, 1.9, 1.0 Hz, 1H), 2.84–2.75 (m,

1H), 2.73–2.60 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 147.8, 147.4, 143.8, 137.2, 134.6, 130.8, 129.1, 126.5, 126.3, 125.5, 123.6, 59.4, 56.0, 51.4, 36.2, 29.5; *m/z* (ESI–MS) 237.1 [M + H]⁺.

(±)-4c: Following general procedure G, product (±)-4c was obtained from 3b and 5,6,7,8tetrahydro-[1,3]dioxolo[4,5-g]isoquinoline (reflux for 1.5 h) as a yellow solid in 85% yield (119 mg), ($R_f = 0.24$ in MeOH/EtOAc 10:90 v/v); mp = 140– 142 °C; IR (KBr) 2945, 1915, 2794, 2734, 1597, 1501, 1486, 1420, 1393, 1361, 1339, 1303, 1246, 1225, 1127, 1036, 932, 861, 823, 760, 543, 453; ¹H NMR (CDCl₃, 500 MHz) δ 8.36 (app s, 2H), 7.06 (d, *J* = 4.8 Hz, 1H), 6.70 (app s, 1H), 6.59 (s, 1H), 5.93–5.91 (comp, 2H), 4.04 (d, *J* = 15.0 Hz, 1H), 3.68 (d, *J* = 15.0 Hz, 1H) 3.60 (dd *L* = 11, 1, 3.7 Hz, 1H) 3.26 (dd *L* = 16, 9, 3.9 Hz, 1H) 3.19–3.02 (comp, 2H), 2.85

1H), 3.60 (dd, J = 11.1, 3.7 Hz, 1H), 3.26 (dd, J = 16.9, 3.9 Hz, 1H), 3.19–3.02 (comp, 2H), 2.85 (dd, J = 16.9, 11.2 Hz, 1H), 2.70–2.56 (comp, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 147.8, 147.4, 146.4, 146.3, 143.6, 130.1, 127.8, 123.6, 108.6, 105.5, 101.0, 59.4, 55.9, 51.5, 36.4, 29.6; m/z (ESI–MS) 280.9 [M + H]⁺.

 (\pm) -5: A 10 mL microwave reaction tube was charged with a 10 x 8 mm SiC passive heating element, 2-methylquinoline-3-carbaldehyde (34.2 mg, 0.2 mmol, 1 equiv), acetic acid (2 mL), and pyrrolidine (81 µL, 1 mmol, 5 equiv). The reaction tube was sealed with a Teflon-lined snap cap, and heated in a microwave reactor at 200 °C (200 W, 70–100 psi) for 15 minutes. After cooling with compressed air flow, the reaction mixture was diluted with EtOAc (15 mL) and then washed with 1N NaOH (2 x 20 mL). The combined aqueous layers were extracted with EtOAc (2 x 15 mL), and the combined organic layers washed with brine (40 mL) and dried over anhydrous Na₂SO₄. Solvent was then removed under reduced pressure and the residue purified by silica gel chromatography. (\pm) -5 was obtained as a yellow solid in 59% yield (27 mg), ($R_f = 0.24$ in MeOH/EtOAc 10:90 v/v); mp = 115–118 °C; IR (KBr) 2964, 2827, 2799, 2364, 1621, 1558, 1491, 1456, 1413, 1303, 1161, 1126, 1023, 998, 955, 900, 780, 750, 615, 478; ¹H NMR (CDCl₃, 500 MHz) δ 8.01–7.95 (m, 1H), 7.81 (s, 1H), 7.75–7.69 (m, 1H), 7.63 (ddd, J = 8.4, 6.9, 1.5 Hz, 1H), 7.45 (ddd, J = 8.1, 1.5 (ddd, J = 8.1, 1. 6.8, 1.2 Hz, 1H), 4.32 (d, J = 14.7 Hz, 1H), 3.56 (d, J = 14.6 Hz, 1H), 3.39 (dd, J = 16.9, 3.8 Hz, 1H), 3.32 (app td, J = 8.6, 2.3 Hz, 1H), 3.00 (dd, J = 16.8, 11.1 Hz, 1H), 2.54 (dddd, J = 10.9, 9.7, 6.8, 3.8 Hz, 1H), 2.32 (app q, J = 8.9 Hz, 1H), 2.19 (dddd, J = 12.3, 9.8, 6.8, 4.2 Hz, 1H), 1.98 (app ddtd, J = 12.8, 11.2, 8.7, 4.2 Hz, 1H), 1.87 (ddddd, J = 12.4, 9.7, 8.8, 6.4, 2.3 Hz, 1H), 1.65 (dddd, J = 12.4, 11.3, 9.5, 6.4 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 157.2, 147.1, 133.1, 129.0, 128.63, 128.59, 127.3, 126.9, 126.0, 61.4, 55.2, 54.8, 40.3, 31.5, 22.0; m/z (ESI-MS) $225.2 [M + H]^+$.

Alternate procedure for the preparation of (\pm) -5: Following general procedure H, 1a and proline were heated under reflux for 1h. Product (\pm) -5 was obtained as a yellow solid in 70% yield (78 mg). The characterization data matched those shown above.

2D-NMR Analysis for (±)-2j, Selected Interactions (in CDCl₃)



2D-NMR Analysis for (±)-2j', Selected Interactions (in CDCl₃)

 H_4



4.02

2D-NMR Analysis for (±)-2k, Selected Interactions (in CDCl₃)

GCOSY

NOESY





NOESY

Protons	Chemical Shifts (ppm)
H ₁	3.92
H ₂	3.51
Me	1.73

2D-NMR Analysis for (±)-2k', Selected Interactions (in CDCl₃)

GCOSY	NOESY
N He Me	N He Me
Protons	Chemical Shifts (ppm)
H ₁	4.02
H_2	3.74
Me	1.13

2D-NMR Analysis for (\pm) -2l, Selected Interactions (in CDCl₃)





Protons	Chemical Shifts (ppm)
H_1	4.53
H ₂	4.45
H ₃	3.65
H_4	3.18
Me	1.48

2D-NMR Analysis for (±)-2l', Selected Interactions (in CDCl₃)





NOESY

> >	5
Protons	Chemical Shifts (ppm)
H ₁	4.03
H ₂	3.98
H ₃	3.72
H ₄	3.30
Me	1.69

2D-NMR Analysis for (±)-20, Selected Interactions (in CDCl₃)

GCOSY	NOESY
H ₂ ,H ₁ N H ₄	H ₂ ,H ₁ N H ₄ H ₄
Protons	Chemical Shifts (ppm)
H_1, H_2	4.50, 3.97
H ₃ , H ₄	4.38, 4.31

2D-NMR Analysis for (\pm) -20', Selected Interactions (in CDCl₃)



2D-NMR Analysis for (\pm) -2p, Selected Interactions (in CDCl₃)

GCOSY

NOESY

N N	
N Me H	

N Ma Ha

Protons	Chemical Shifts (ppm)
H ₁	3.85-3.79
H ₂	3.26
Me	1.61

2D-NMR Analysis for (\pm) -2p', Selected Interactions (in CDCl₃)

GCOSY NOESY NH2 Me Protons Chemical Shifts (ppm)

Protons	Chemical Shifts (ppm)
H_1	3.90
H_2	3.49
Me	1.04

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¹H NMR of **11k** in CDCl₃









¹H NMR of **15b** in CDCl₃

COOEt N Bn



 $\frac{1.37}{1.36}$









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Me








$$\begin{array}{c} 10.31 \\ 10.31 \\ 8.63 \\ 8.17 \\ 8.15 \\ 8.17 \\ 8.15 \\ 8.17 \\ 8.17 \\ 8.17 \\ 8.17 \\ 8.17 \\ 8.17 \\ 8.17 \\ 8.17 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.28 \\ 17.$$

¹H NMR of **1j** in CDCl₃









¹H NMR of **1k** in CDCl₃













¹H NMR of **1o** in $CDCl_3$















¹H NMR of **2a** in CDCl₃

















¹H NMR of **2d** in CDCl₃









¹H NMR of **2e** in CDCl₃









¹H NMR of **2f** in CDCl₃





















¹H NMR of **2j** in CDCl₃













¹H NMR of **2j'** in CDCl₃




























¹H NMR of **2I** in CDCl₃























































¹H NMR of **2p'** in CDCl₃










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¹H NMR of **4a** in CDCl₃















8.00 8.00 7.98 7.79 7.73 7.77 7.77 7.77 7.77 7.77 7.77	7.63 7.66 7.66 7.66 7.66 7.66 7.66 7.66	2255 2255 2255 2555 2555 2555 2555 255	2.2.33 2.2.33 2.2.20 2.2.20 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.19 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118 2.2.118

¹H NMR of **5** in CDCl₃





