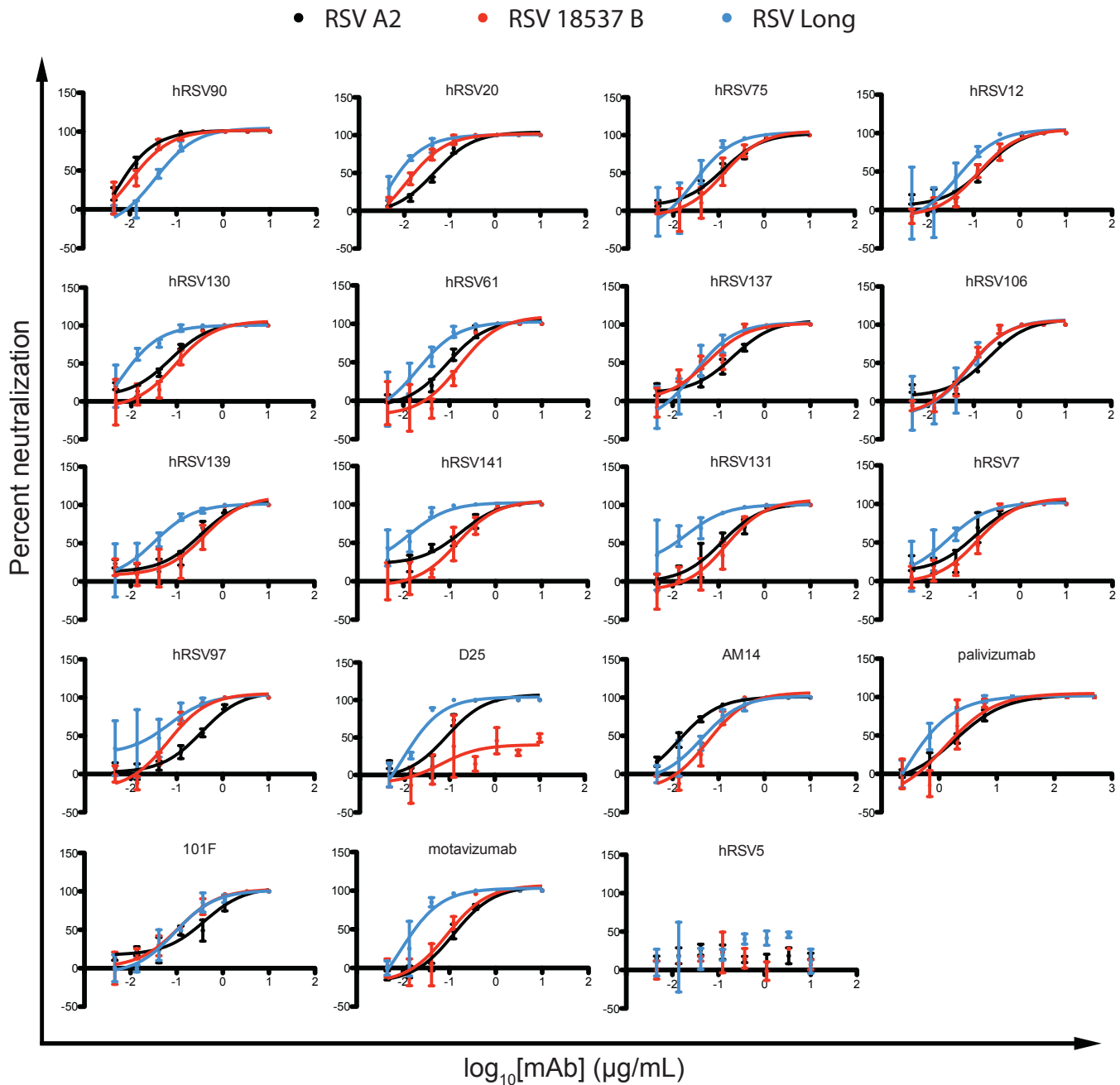


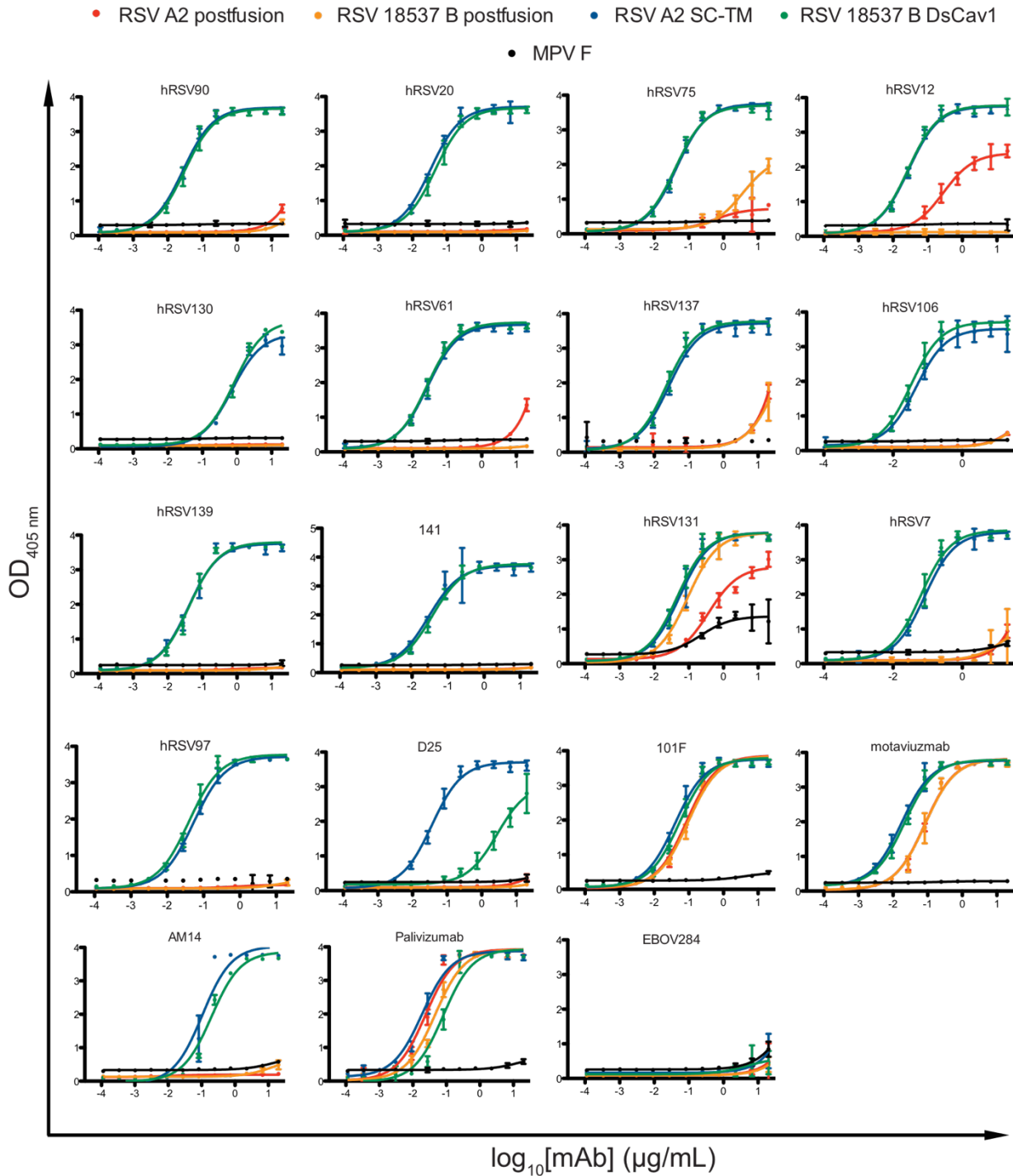
In the format provided by the authors and unedited.

# **A novel pre-fusion conformation-specific neutralizing epitope on the respiratory syncytial virus fusion protein**

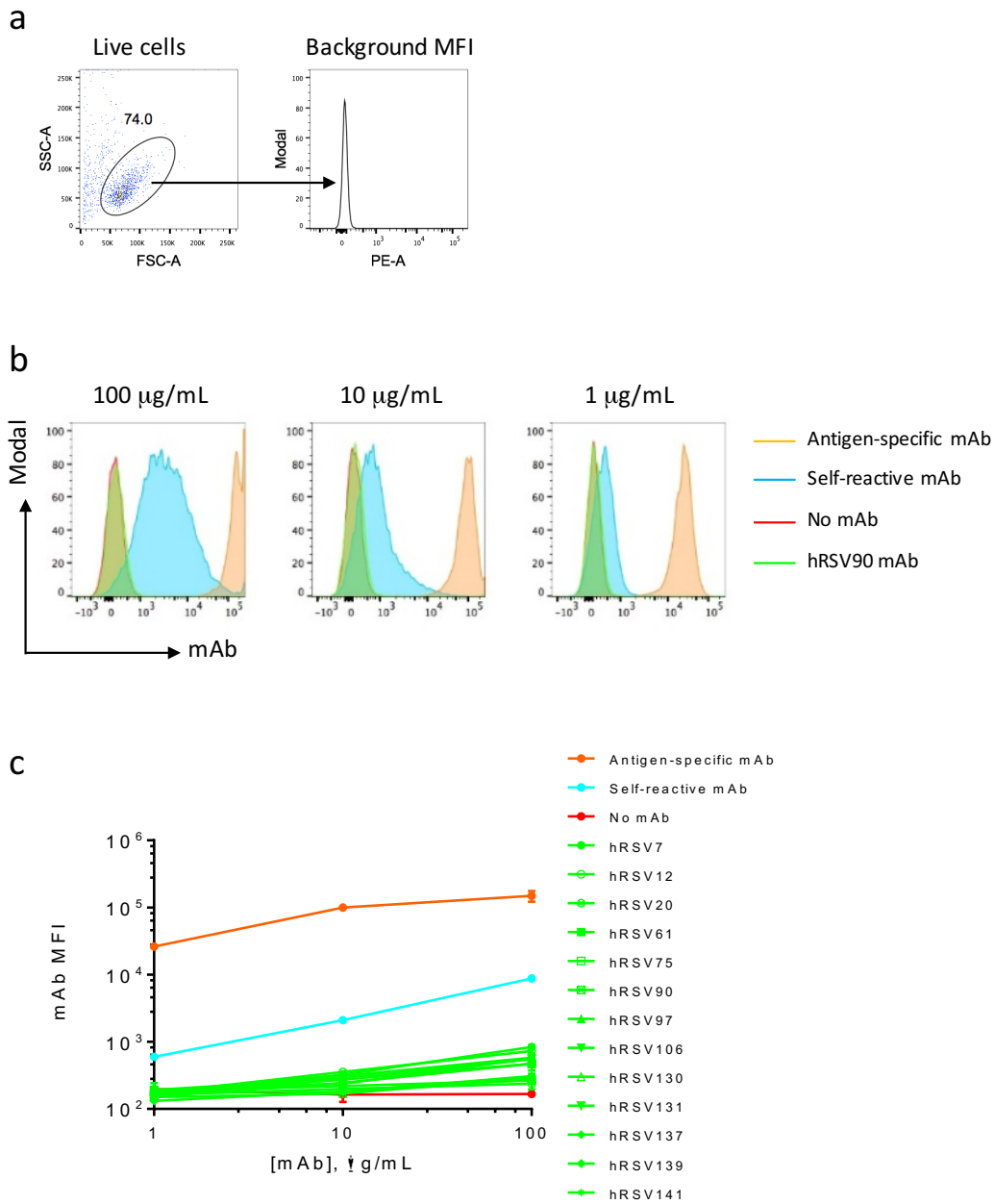
**Jarrold J. Mousa, Nurgun Kose, Pranathi Matta, Pavlo Gilchuk and James E. Crowe Jr**



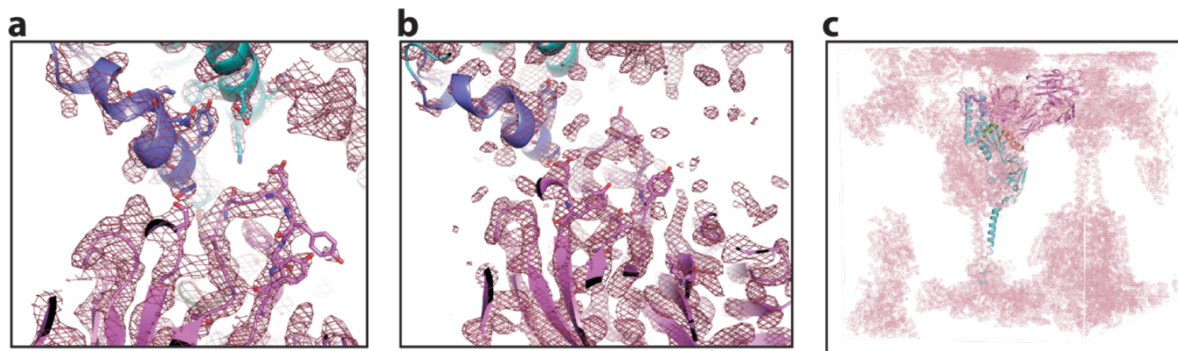
**Supplementary Figure 1. Neutralization curves for the newly isolated RSV F-specific mAbs.**  $\text{IC}_{50}$  values are displayed in Table 1. Error bars represent the standard deviation of three technical replicates from one of at least two independent experiments. A non-neutralizing RSV F specific mAb, hRSV5, was used as a negative control. MABs D25, 101F, AM14, motavizumab, and palivizumab were included as positive controls.



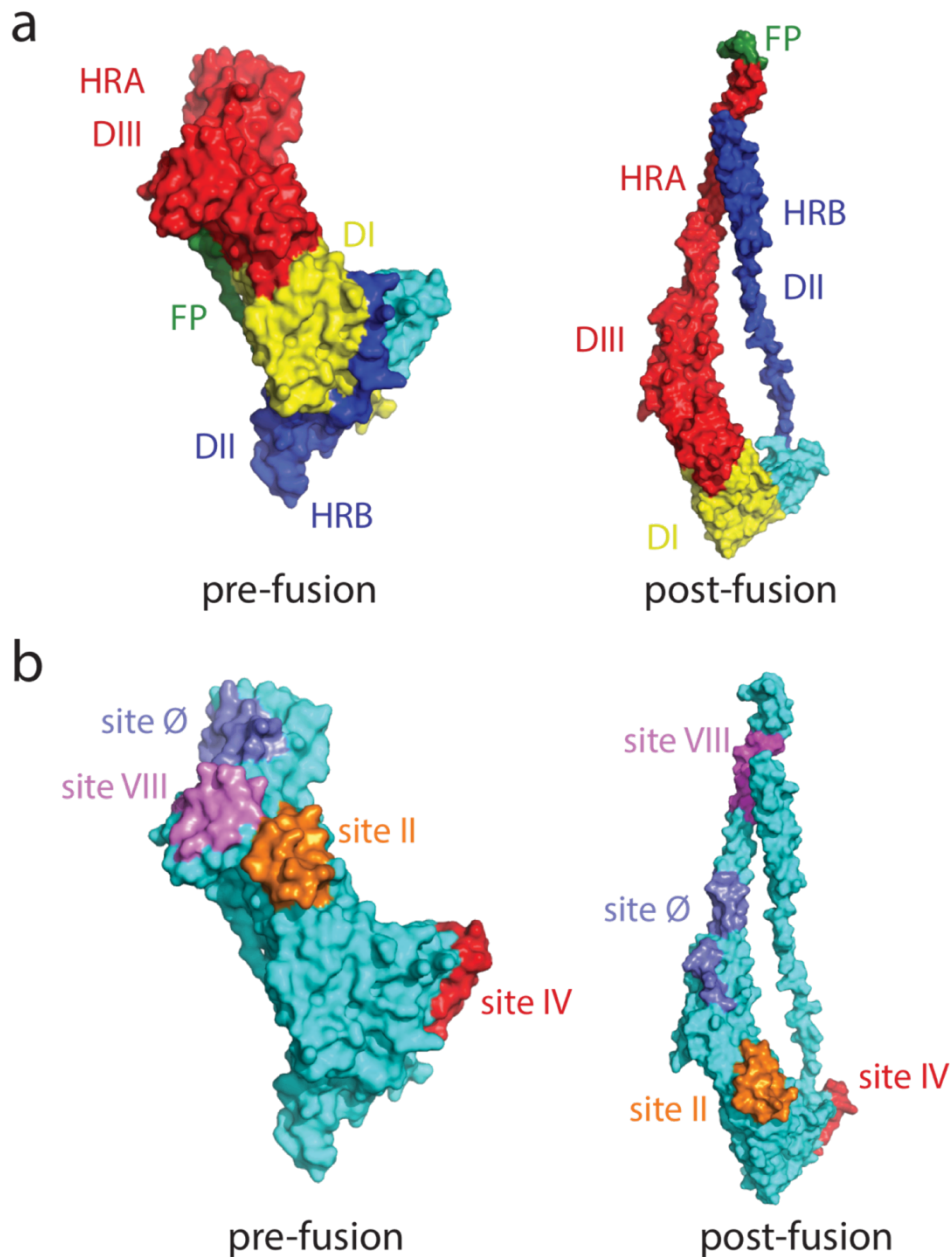
**Supplementary Figure 2. ELISA binding curves for the newly isolated mAbs and positive controls to RSV F protein strain and construct variants.** The metapneumovirus F protein was used as a negative binding control. An Ebola-virus specific mAb EBOV284 was used as a negative mAb control. Error bars indicate 95% confidence intervals of four technical replicates from one of at least two independent experiments. EC<sub>50</sub> values for these curves are displayed in Table 1.



**Supplementary Figure 3. Assessing self-reactivity of hRSV mAbs by flow cytometry.** Jurkat cell line was stained with individual mAbs followed by incubation with secondary phycoerythrin (PE)-conjugated Ab and flow cytometric analysis. (a) Gating strategy for measuring binding of mAb to Jurkat cells. (b) Representative flow cytometric histograms showing dose-dependent binding of antigen-specific, self-reactive, or hRSV90 mAbs to Jurkat cells. Binding of BDBV289 Ebola virus GP-specific mAb to transfected Jurkat cells that express Ebola virus GP on their surface served as positive control for antigen-specific mAb (orange histogram); a mAb with known self-reactivity (BDBV223) served as a control for self-reactivity (light blue histogram); Jurkat stained with secondary detection PE-conjugated Ab only served as a control for assay background (red histogram). (c) Dose-dependent binding of hRSV mAbs to Jurkat cells measured as mean fluorescence intensity (MFI). Error bars represent the SD of two technical replicates.

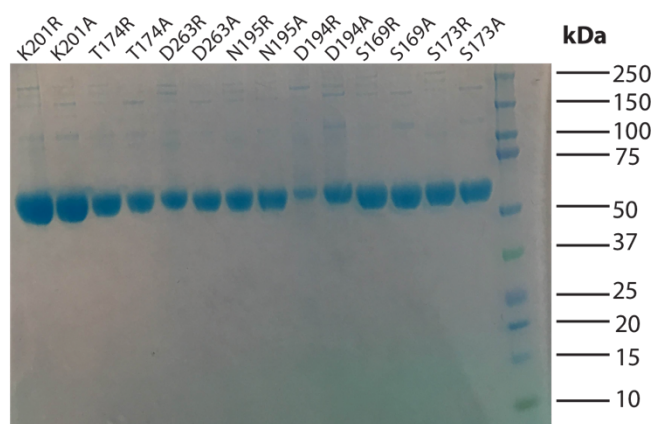


**Supplementary Figure 4. Density maps for the hRSV90-RSV F A2 SC-TM interface.** (a) 2F<sub>o</sub>-2F<sub>c</sub> density map. (b) Simulated annealing composite omit density map of the same interface. (c) 2F<sub>o</sub>-2F<sub>c</sub> Density map of the entire structure showing density for the foldon trimerization domain.



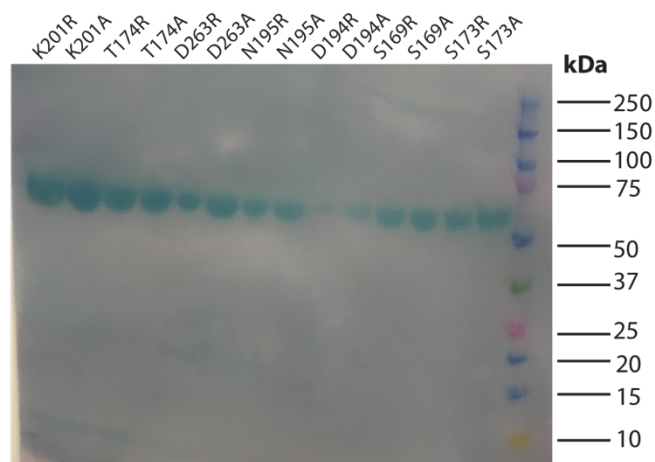
**Supplementary Figure 5. Structural and antigenic regions of the RSV F protein.** (a) Structural regions of the RSV F protein are shown with corresponding labels in both pre-fusion and post-fusion structures. HRA is heptad repeat A (red as part of DIII), HRB is heptad repeat B (blue as part of DII), FP is fusion peptide (green), and DI is shown in yellow. (b) Antigenic regions in the pre-fusion and post-fusion RSV F structures are colored. Site IV is red, site II is orange, site Ø is blue, and site VIII is magenta. Residues comprising antigenic sites Ø and VIII are rearranged in the post-fusion conformation, resulting in loss of mAb binding. Site VIII becomes part of the six-helix bundle of the post-fusion RSV F protein.

a



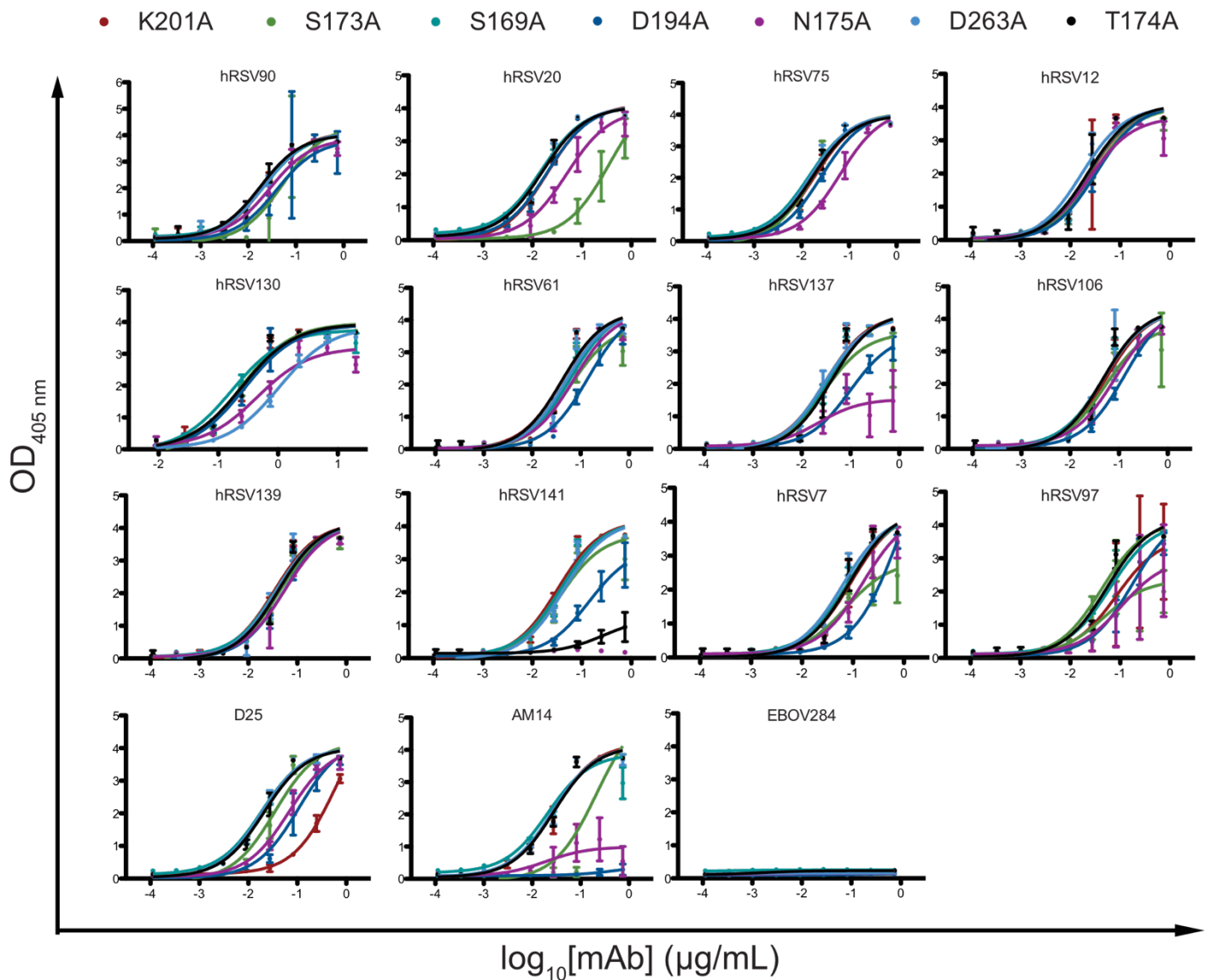
Coomassie-stained SDS-PAGE

b



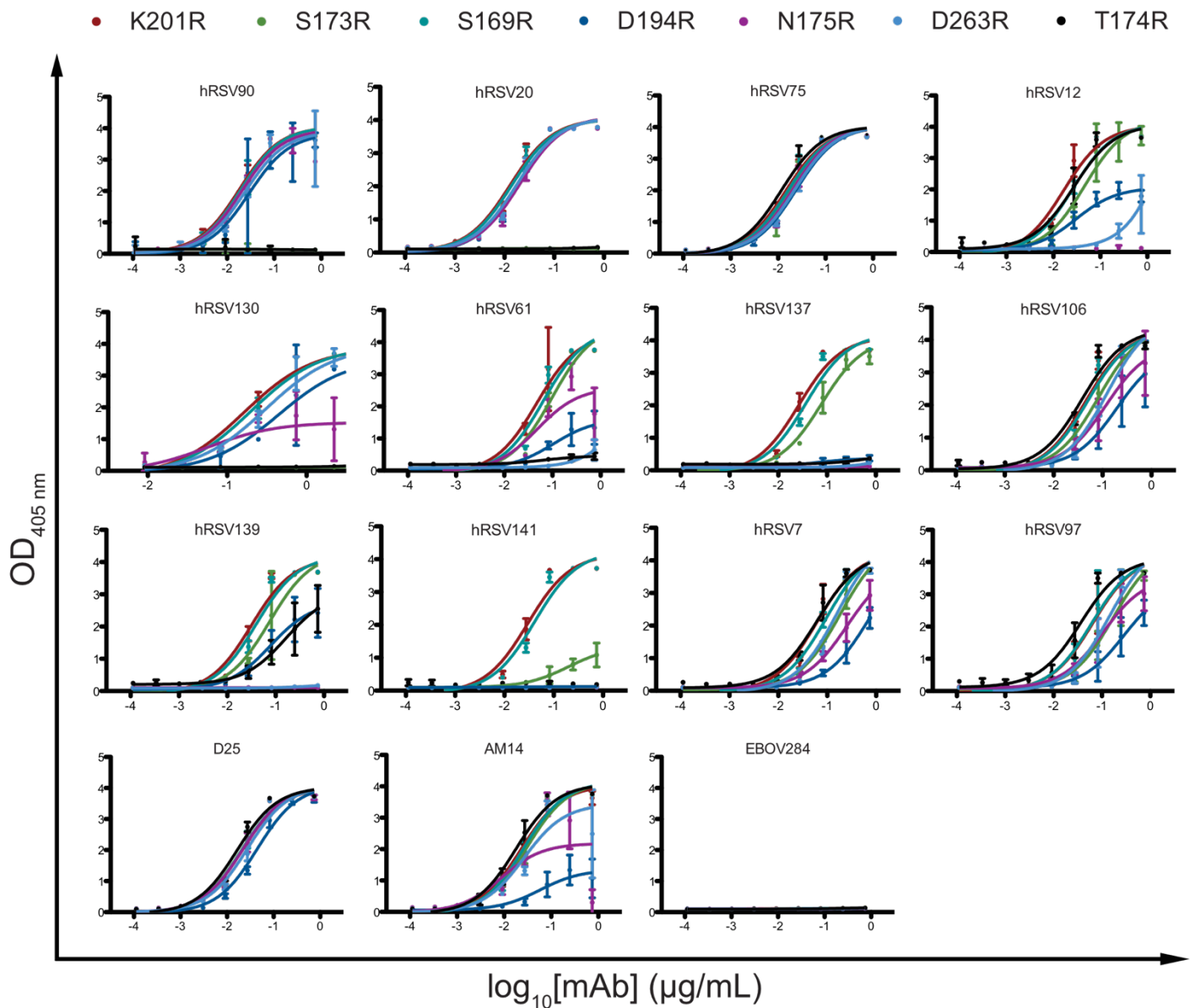
Anti-polyhistidine-alkaline phosphatase antibody  
BM purple chromogenic substrate

**Supplementary Figure 6. RSV F SC-TM mutants.** (a) A Coomassie-stained SDS-PAGE is displayed with bands displayed for each purified mutant protein. (b) A corresponding western blot is shown using a monoclonal anti-polyhistidine-alkaline phosphatase antibody with BM purple chromogenic substrate to visualize the RSV F mutants directly on the PVDF membrane. Purifications, SDS-PAGE, and western blot were conducted once.

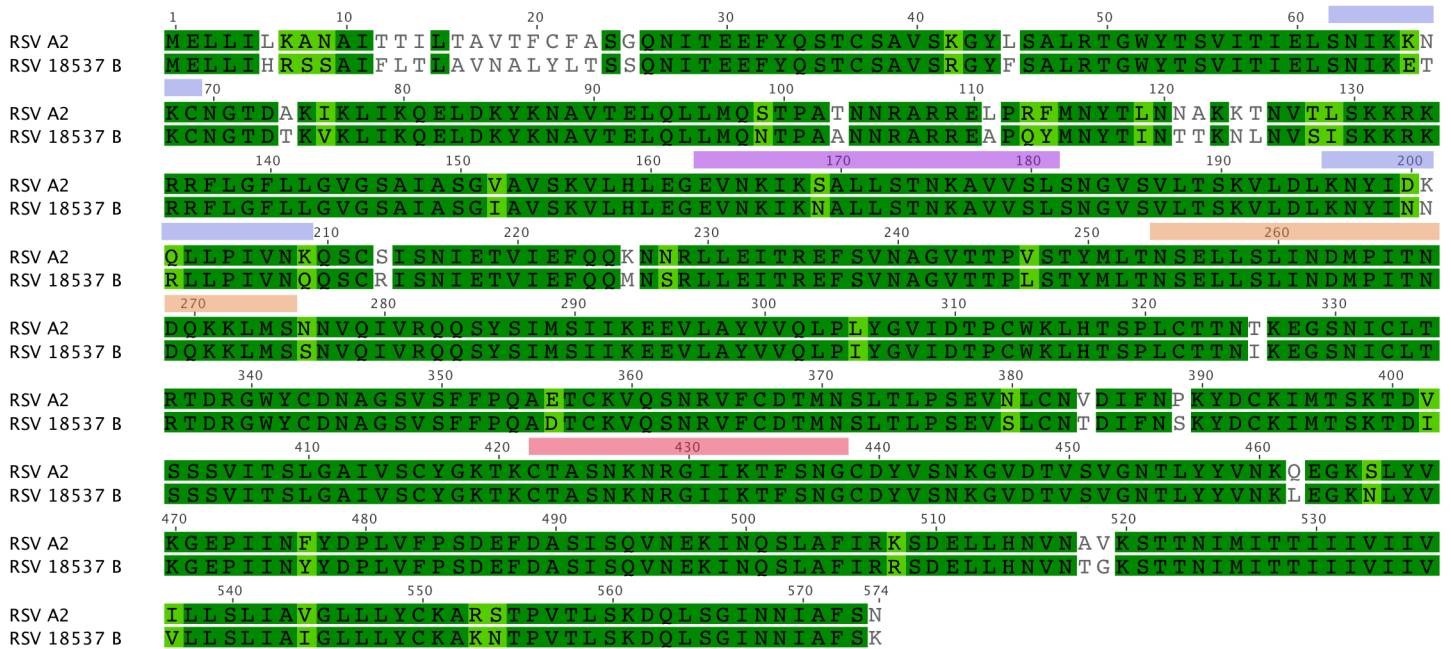


**Supplementary Figure 7. ELISA binding curves for RSV F SC-TM alanine mutations at the hRSV90 binding site.** Error bars indicate 95% confidence intervals of four technical replicates from one of at least two independent experiments.





**Supplementary Figure 8. ELISA binding curves for RSV F SC-TM arginine mutations at the hRSV90 binding site.** Error bars indicate 95% confidence intervals of four technical replicates from one of at least two independent experiments.



**Supplementary Figure 9. Alignment of RSV F proteins from subgroups A and B.** Amino acids in green are conserved among the two proteins, those in light green are semi-conserved, and those in white are not conserved. Antigenic sites are shaded above the corresponding sequences, with site Ø in blue, site VIII in magenta, site II in orange, and site IV in red.

**Supplementary Table 1. Data collection and refinement statistics**

<b>Data collection*</b>	hRSV90+RSV A2 F SC-TM	
Beamline	LS-CAT 21-ID-G	Anisotropy correction
Number of crystals	1	
Space group	R 3 2 H	
Cell dimensions		
<i>a</i> , <i>b</i> , <i>c</i> (Å)	148.2, 148.2, 538.2	
$\alpha$ , $\beta$ , $\gamma$ (°)	90, 90, 120	
Resolution (Å)	49.23 – 3.14 (3.26 – 3.14)	a= 3.6, b= 3.6, c= 3.1
<i>R</i> <sub>merge</sub>	0.385 (4.715)	0.266 (0.768)
<i>I</i> / $\sigma I$	5.2 (0.5)	6.7 (2.2)
Completeness (%)	99.9 (99.8)	75.2 (4.1)
Redundancy	5.9 (5.9)	4.4 (0.2)
<b>Refinement</b>		
Resolution (Å)		48.31 – 3.14
No. unique reflections		30527 (272)
<i>R</i> <sub>work</sub> / <i>R</i> <sub>free</sub>		0.2212 (0.2603)
No. atoms		
Protein		7077
<i>B</i> -factors		
Protein		71.68
R.m.s. deviations		
Bond lengths (Å)		0.012
Bond angles (°)		1.41
Ramachandran statistics		
Favored regions (%)		95
Allowed regions (%)		4.8
Outliers (%)		0.22

Values in parentheses are for the highest resolution data shell.