

Supplementary data

Table 1 Antibodies used in this study

Vender	Conjugated Antibodies	Clone
Tonbo Biosciences (San Diego, CA. USA)	CD3 PE-Cy7	UCHT1
	CD4 PE	OKT4
	CD8 APC	OKT8
	CD19 PE-Cy7	HIB19
Biolegend (San Diego, CA USA)	CD45RA APC-Cy7	H100
	CD45RO Pacific Blue	UCHL1
	CCR7 APC	G043H7
	CXCR3 Brilliant Violet 421™	G025H7
	CD161-FITC	HP-3G10
	CCR6 Per CP-Cy5.5	G034E37
	CD25 Alexa Fluor® 488	BC96
	CD56 Brilliant Violet 421™	HCD56
	CD69 Brilliant Violet 510™	FN30
	CD4Brilliant Violet 510™	OKT4
	CD4 Pacific Blue	OKT4
BD Biosciences (San Diego, CA. USA)	IL-4 APC	8D4-8
	CD14 APC-Cy7	MφP9
	TCR _{αβ} FITC	T10B9.1A-31
	CCR3 Horizon™ BV510	5E8
eBiosciences (San Diego, CA. USA)	CCR6 PE	11A9 (RUO)
	Fox P3 PE	PCH101
	Ki67 eFluor® 450	20Raj1
	IL-17A PE	SHLR17
Thermo Fisher/Invitrogen (Waltham, MA. USA)	IL-17F PE	eBio64CAP17
	IFN _γ FITC	B27

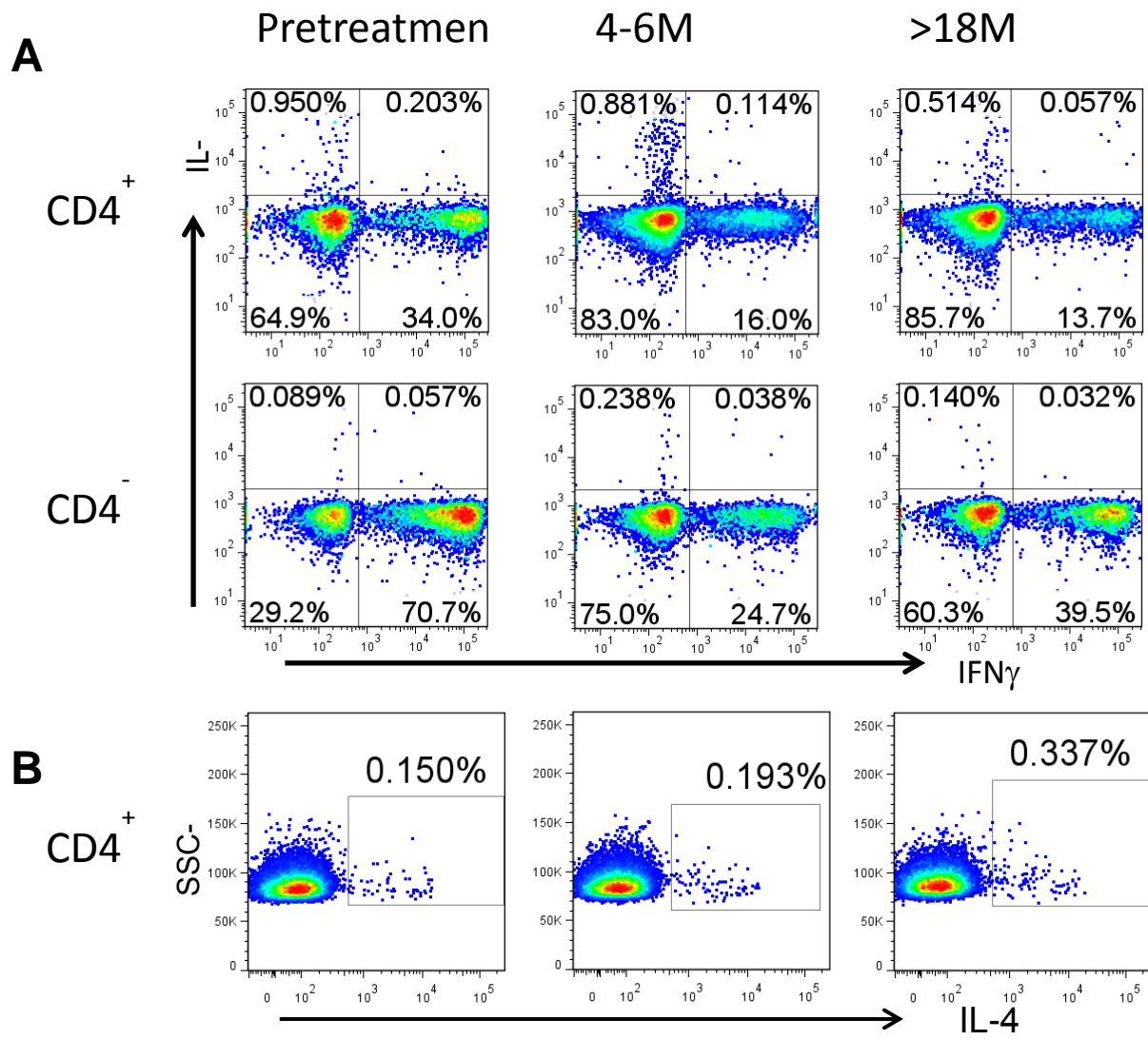


Figure 1: DMF effect on ex vivo IFN γ , IL-17 and IL-4 production by T cells (A) Representative FACS profile of IFN γ and IL-17 (IL-17A and IL-17F) expression on 6hr PMA/ ionomycin-activated CD4⁺ (upper) and CD4⁻ (lower) T cells from patients before (left hand), 6 month (middle) or 24 month after DMF treatment (right hand). **(B)** Representative FACS profile of IL-4 expression on ex vivo CD4⁺ T cells from a MS patient before (left hand), 6 month (middle) and 26 month after DMF treatment (right hand).

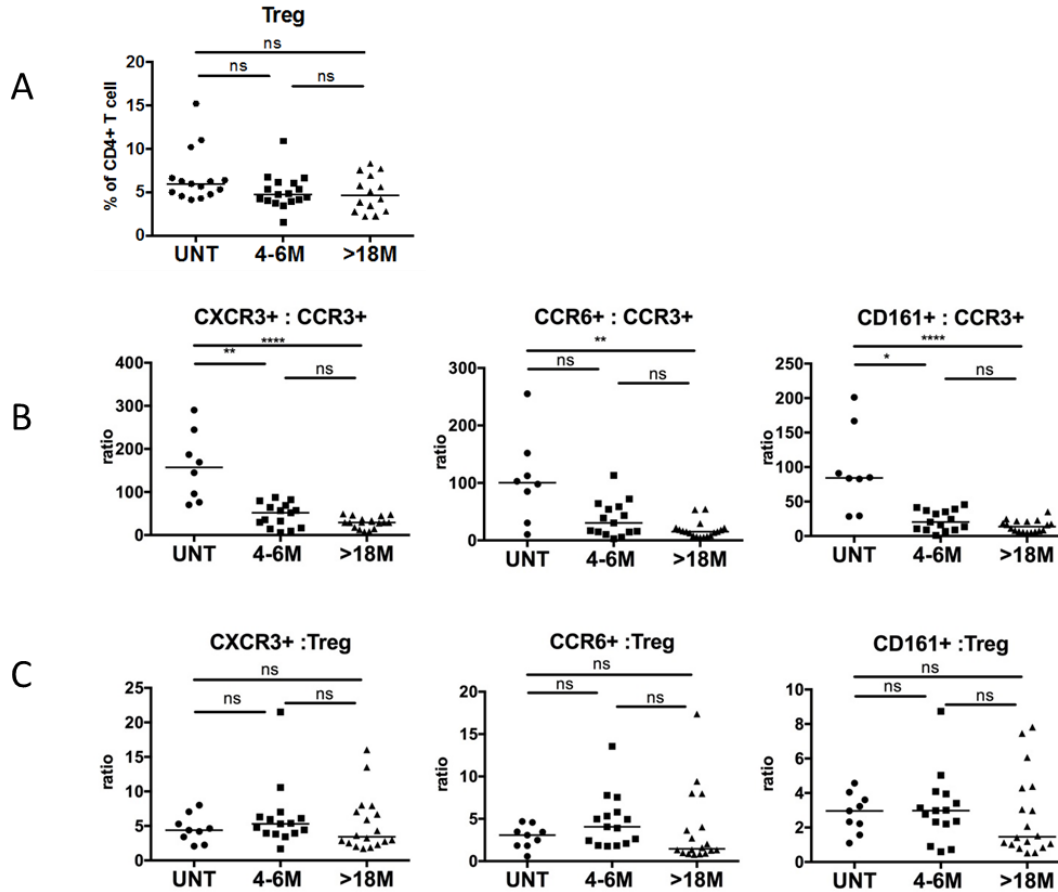


Figure 2: DMF effect on Th and Treg subsets and ratios. (A) DMF effect on Treg PBMC derived from untreated (UNT), 4-6 month DMF-treated (4-6M) and 18-26M DMF-treated (>18M) RRMS patients were stained with fluorochrome-conjugated antibodies against CD3, CD4, and CD25 followed by intracellular staining of fluorochrome-conjugated anti-human FOXP3 and analyzed by flow cytometry. The Treg (CD25⁺FOXP3⁺) of total CD4⁺ T cells were analyzed at three time point (UNT: n=15, 4-6M: n=17, >18M: n=14). (B) DMF effect on Th1/Th17:Th2 ratios: PBMC derived from untreated (UNT), 4-6 month DMF-treated (4-6M) and 18-26M DMF-treated (>18M) RRMS patients were stained with fluorochrome-conjugated antibodies against CD3, CD4, CXCR3, CD161, CCR6 and CCR3 and analyzed by flow cytometry. Th1:Th2 ratio was calculated using CXCR3⁺% divided by CCR3⁺% in CD4⁺ T cells (left panel); Th17:Th2 ratio was calculated using CCR6⁺% divided by CCR3⁺% in CD4⁺ T cells (middle panel). The ratio of CD161⁺:Th2 was also calculated (right panel). Each dot represents the ratio from an individual patient. Upper panel is the cross-sectional analysis (UNT: n=8; 4-6M: n=15 and >18M: n=18). (C) DMF effect on Th1/Th17 : Treg cells ratio Th1:Treg ratio was calculated using CXCR3⁺% divided by CD25⁺FOXP3⁺ Treg% in CD4⁺ T cells (left lower panel); Th17: Treg ratio was calculated using CCR6⁺% divided by Treg% in CD4⁺ T cells (middle lower panel). The ratio of CD161⁺:Treg was also calculated (right lower panel). Each dot represents the ratio from an individual patient. Upper panel is the cross-sectional analysis (UNT: n=9; 4-6M: n=15 and >18M: n=18). *P*-values from Kruskal-Wallis ANOVA with Dunn's multiple comparison tests are shown above the data for each cell type group. **P* < 0.05; ***P* < 0.01; ****P* < 0.001; ns, not significant.

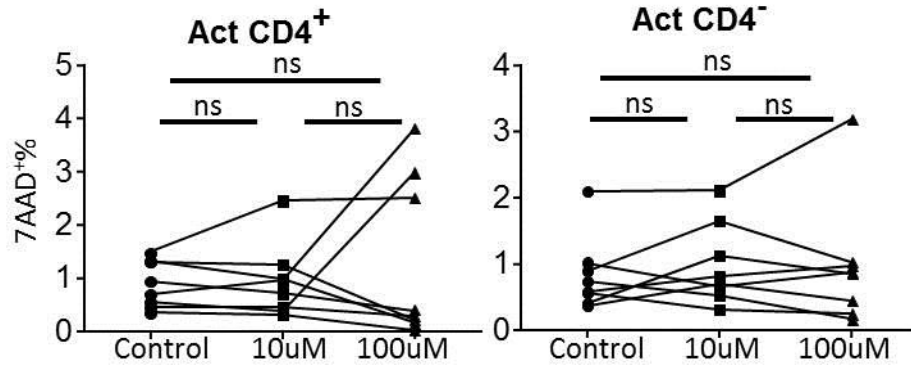


Figure 3: Cell Death of activated T cells *in vitro* PBMCs from 8 healthy donors were activated by anti-plate- coated anti-CD3 and soluble anti-CD28 in complete RPMI with or without 10 μ M or 100 μ M of DMF for 48 hours. Cells were harvested, washed and surface stained with fluorochrome-conjugated antibodies against CD3, CD8, CD4, as well as 7AAD and Annexin V and analyzed by flow cytometry. Large (FSC-A^{hi}) CD3⁺ T cell blasts were gated as activated T cells. The percentage of dead cells measured by positive staining of 7AAD in activated CD4⁺ T cells (left panel) and CD4⁻ T cells (right panel) populations.