SUPPLEMENTARY INFORMATION



Supplementary Figure 1: CCR6 and CCR2 expression by resting $\gamma\delta$ **T17 cells. a)** *Ccr6* and *Ccr2* mRNA in sorted CD3⁺CD4⁺CD44^{lo}CD25⁻ naïve CD4 T cells and CD90⁺CD3⁺TCR- $\gamma\delta^+$ IL-17A-YFP⁺ $\gamma\delta$ T17 and YFP⁻ IL-17⁻ $\gamma\delta$ T cells from skin-draining lymph nodes (sLNs) and spleen of naïve *Il17a^{Cre}×Rosa26^{eYFP}* mice (pooled from n=5). N.D.: not detected. **b)** Representative flow cytometry of CCR6/CCR2 expression by V $\gamma4^+$ and V $\gamma4^-$ (V $\gamma6^+$) $\gamma\delta$ T17 cells from sLNs and dermis of naïve *Il17a^{Cre}×Rosa26^{eYFP}* mice (n=3). **c)** Representative flow cytometry of CCR6/CCR2 and V $\gamma4^+$ and CD3^{bright} (V $\gamma6^+$) $\gamma\delta$ T17 cells from sLNs of naïve *Il17a^{Cre}×Rosa26^{eYFP}* mice (n=3). **b** c) Representative of two experiments.



Supplementary Figure 2: Downregulation of CCR6 in activated $\gamma\delta$ T17 cells. a) Representative flow cytometry of CCR6/CCR2 expression by subsets of $\gamma\delta$ T cells distinguished by CD44 and CD27 expression in skin-draining lymph nodes of wildtype (WT) mice either naïve or at experimental autoimmune encephalomyelitis (EAE) onset (n=3/group). b) Representative flow cytometry of CCR6/CCR2 expression by $\gamma\delta$ T17 cells from *Il17a^{Cre}xRosa26^{eYFP}* lymphocytes cultured with indicated stimuli for 72 hr (n=3). c) Representative flow cytometry of activation marker expression by $\gamma\delta$ T17 cells from *Il17a^{Cre}xRosa26^{eYFP}* lymphocytes either unstimulated or IL-23/IL-1 β stimulated for 72 hr (n=3). d) Representative flow cytometry of CCR6/CCR2 expression by splenic conventional (V γ 4⁺) and CD3^{bright} (V γ 6⁺) $\gamma\delta$ T17 cells (gated as in Supplementary Figure 1c) from *Il17a^{Cre}xRosa26^{eYFP}* mice either unstimulated or IL-23/IL-1 β stimulated for 72 hr (n=3). a, d) Representative of two experiments.



Supplementary Figure 3: B16 melanoma and EAE progression in chemokine receptor-deficient mice. a) Average mass (per mouse) of B16 melanomas 7d post-challenge in wildtype (WT) (n=19), $Ccr6^{-/-}$ (n=18), $Ccr2^{-/-}$ (n=19) and $Ccr2^{-/-}Ccr6^{-/-}$ mice (n=13). b) Clinical disease scores of WT, $Ccr6^{-/-}$, $Ccr2^{-/-}$ and $Ccr2^{-/-}Ccr6^{-/-}$ mice given experimental autoimmune encephalomyelitis (EAE) (n=17/group). Mean±SEM. a-b) Pooled from two experiments. a) One-way ANOVA with Bonferroni's multiple comparisons test. ** p < 0.01, **** p < 0.0001.



Supplementary Figure 4: *In vitro* expansion of $\gamma\delta$ T17 cells. a) Schematic of culture protocol. b) Representative flow cytometry and number and frequency of *in vitro*-expanded $\gamma\delta$ T17 cells at different stages of culture (n=3). c) Flow cytometry of CCR2 expression by *in vitro*-expanded $\gamma\delta$ T17 cells relative to isotype (grey). d) Transwell chemotaxis of *in vitro*-expanded $\gamma\delta$ T17 cells from wildtype (WT) and *Ccr2^{-/-}* mice to CCL2. e) Flow cytometry of V γ 4 expression by *in vitro*-expanded $\gamma\delta$ T17 cells. b) Mean±SEM, d) Mean±SD. a-e) Representative of two experiments.



Supplementary Figure 5: CCR6 regulates homeostatic $\gamma\delta$ T17 cell positioning in the dermis. a) Representative flow cytometry of IL-17A-YFP and IL-17A expression by dermal $\gamma\delta$ T^{lo} cells from ears of *Il17a*^{Cre}*xRosa26*^{eYFP} mice (n=3). b) Number of $\gamma\delta$ T17 cells in organs of unimmunized wildtype (WT), *Ccr6*^{-/-} and *Ccr2*^{-/-} mice (n=4-12/group). iLN: inguinal lymph node; mLN: mesenteric lymph node; PEC: peritoneal exudate cells. Mean±SEM. a) Representative of two experiments, b) pooled from two experiments. b) One-way ANOVA with Bonferroni's multiple comparisons test. * p < 0.05, ** p < 0.01.



Supplementary Figure 6: IRF8 and Blimp1 do not regulate CCR6 expression in $\gamma\delta$ T17 cells. a) *Ccr2* and b) transcription factor mRNA in sorted $\gamma\delta$ T17 cells from *Il17a^{Cre}xRosa26^{eYFP}* lymphocytes fresh *ex vivo* or cultured with IL-23/IL-1 β for indicated times (pooled from 5-7 mice). N.D.: not detected. c) Frequency of total and CCR6⁺ $\gamma\delta$ T17 cells in spleens of wildtype (WT) and transcription factor-deficient mice (n=3/group). d) Splenocytes from Ly5.1 and *Irf8^{-/-}* or *Lck^{Cre}Prdm1^{II/I}* mice were 670 dye-labelled, mixed 50:50 and stimulated with IL-23/IL-1 β for 72 hr. Representative flow cytometry and quantitation of CCR6 expression and proliferation in CD45.1⁺ or CD45.2⁺ $\gamma\delta$ T17 cells (n=3/group). a-b) Mean±SD, c) Mean±SEM. a-b, d) Representative of two similar experiments. c) One-way ANOVA with Dunnett's multiple comparisons test relative to WT control, d) paired two-tailed Student's *t*-test. * p < 0.05.



Supplementary Figure 7: IRF4 and BATF promote CCR6 downregulation in $\gamma\delta$ T17 cells. a) Representative flow cytometry of IL-23R and IL-1R1 expression by splenic CD44^{hi}CD27⁻ $\gamma\delta$ T cells (gated as in Supplementary Figure 2a) from wildtype (WT), $Irf4^{-/-}$ and $Batf^{-/-}$ mice (n=3/group). b) IRF4 and BATF ChIP-Seq data at the *Ccr6* locus from CD8⁺ T cells and T helper 17 (Th17) cells from published datasets. Line indicates binding site consistently detected for both transcription factors in both cell types. Datasets are from Kurachi *et al.* 2014 (BATF CD8⁺ T cells), Man *et al.* 2013 (IRF4 CD8⁺ T cells) and Ciofani *et al.* 2012 (IRF4/BATF Th17 cells) as referenced in results section.



Supplementary Figure 8: Flow cytometry gating strategies. Each panel denotes pre-gates to flow cytometry plots and/or data points in indicated figures. **a)** Lymphocyte gating, single cell discrimination and viability gating serves as pre-gate for all flow cytometry data and sorting strategies. **b)** Gating of $\gamma\delta$ T17 cells by IL-17A protein expression. **c)** Gating of $\gamma\delta$ T17 cells in *Il17a*^{Cre}×*Rosa26*^{eYFP} mice. **d)** Gating of neutrophils from nasal wash. **e)** Gating of donor CD45.1^{+/-}CD45.2⁺ *in vitro*-expanded $\gamma\delta$ T17 cells in co-transfer trafficking experiments, including V γ 4/V γ 6 distinction by CD3^{bright} staining. **f)** Gating of dermal $\gamma\delta$ T^{lo} cells. **g)** Gating of donor CD45.2⁺ $\gamma\delta$ T17 cells in skin-homing transfer experiments. **h)** Gating of CD45.1⁺ and CD45.2⁺ $\gamma\delta$ T17 cells in co-culture experiments.



Supplementary Figure 9: Flow sorting strategies. a) Sorting of $\gamma\delta$ T17 cells and IL-17 $\gamma\delta$ T cells from pooled splenocytes and lymph node cells from *Ill7a*^{Cre}×*Rosa26*^{eYFP} mice. **b)** Sorting of naïve CD4⁺ T cells from wildtype (WT) splenocytes. **c)** Sorting of indicated epidermal keratinocyte and dermal stromal populations from skin of WT mice. All strategies first gated on live single cells as in Supplementary Figure 8a.

Primary antibodies for flow cytometry, ELISA and cell culture					
Antigen	Conjugate	Clone	Company	Final	
				concentration	
BrdU	FITC	B44	BD	15µL/well	
CCL2	Biotin (ELISA)	Polyclonal	R&D	200ng/ml	
	Purified (ELISA)		R&D	200ng/ml	
CCR2	Purified	MC21	In house	5.6µg/ml	
CCR6	Purified	MAB590	R&D	8.3µg/ml	
	PE	140706	R&D	7µL/well	
CD3€	Biotin	17A2	eBioscience	2µg/ml	
	BV510	145-2C11	BD	0.833µg/ml	
	FITC		BD	2µg/ml	
	PECy7		eBioscience	2µg/ml	
CD4	Alexa 647	RM4-5	BD	0.67µg/ml	
CD11b	PECy7	M1/70	BD	0.67µg/ml	
CD25	Biotin	7D4	BD	4.17µg/ml	
	PE			$0.83 \mu g/ml$	
CD27	PECv7	LG.3A10	Biolegend	0.67µg/ml	
CD31	FITC	MEC 13.3	BD	4.17µg/ml	
CD38	APC	90	eBioscience	0.83µg/ml	
CD44	Biotin	IM7	BD	1.67µg/ml	
CDTT	FITC	11117	BD	$0.67 \mu g/ml$	
	V450		BD	$1.67 \mu g/ml$	
CD45	APC	30-F11	BD	0.67µg/ml	
02.10	FITC	00111	BD	$1.67 \mu g/ml$	
	PE		BD	$0.83 \mu g/ml$	
CD45 1	APC	A20	eBioscience	$0.67 \mu g/ml$	
02.011	Biotin		Biolegend	$1.67 \mu g/ml$	
	PerCPCv5.5		eBioscience	$1.67 \mu g/ml$	
CD45.2	Biotin	104	BD	1.67µg/ml	
	FITC		BD	$1.67 \mu g/ml$	
	PE		BD	$0.67 \mu g/ml$	
	PECv7		Biolegend	$0.83 \mu g/ml$	
	PerCPCv5.5		eBioscience	1.67µg/ml	
CD69	PECv7	HI 2F3	BD	0.83µg/ml	
CD90 2		53-2.1	BD	0.05µg/ml	
$\frac{CD}{121}\alpha (II - 1R1)$	PE	JAMA 147	Biolegend	3.33µg/ml	
$\frac{\text{CD1210}(\text{IL-IKI})}{\text{CD140a}}$	PV421		PD	1.11µg/ml	
En CAM	Dv421	C 8 8	BD RD	$2.08\mu g/ml$	
ap ²⁸	Piotin	00.0	aBioscience	2.08µg/ml	
	Biotin	260	PD	2.78µg/ml	
I-A/I-E	Biotini Derrificad (aplitante)	209 VMC1.2	DD DieVCell	2.08µg/III	
ΙΓΙΝ-γ	FUTC	AMG1.2	BIOACEII	$10\mu g/ml$	
II 17A	FIIC DV421	TC11	Dialagand	2.78µg/mi	
IL-I/A	BV421 DV510	1011-	Biolegend	$0.28 \mu g/ml$	
		18010.1	Diolegena	$1.55 \mu g/ml$	
н ээр	PE ADC	752217		1.11μg/IIII 20.11/mg/IIII	
1L-23K	Art	/3331/	DD	20µL/Well	
Lyou DOD:4	FIIU DerCDeFL = 710		DD	1.0/µg/ml	
κυκγι	PerCPeFluor/10	B2D	eBioscience	1.6/µg/ml	
Sca-I	FIIC D' c'		eBioscience	2.08µg/ml	
ΤСК-γδ	Biotin	GL3	eBioscience	$1.6/\mu g/ml$	
	BV421		Biolegend	$0.83 \mu g/ml$	
	PECy/		Biolegend	$0.83 \mu g/ml$	
X7 4	Purified (cell culture)		Biolegend	1µg/ml	
νγ4	PECy/	UC3-10A6	eBioscience	0.6/μg/ml	

Supplementary Table 1: Monoclonal antibodies and related reagents

Supplementary Table 1 (continued)						
Other flow cytometry reagents						
Reagent	Conjugate	Clone	Company	Final dilution		
α-rat IgG	Alexa 647	Polyclonal	Life Technologies	10µg/ml		
Proliferation dye	eFluor670	-	eBioscience	5μΜ		
Live/Dead fixable	Near infrared	-	Life Technologies	1/1000		
dye						
Streptavidin	Alexa 647	-	Jackson	2.5µg/ml		
	BV510		ImmunoResearch	0.33µg/ml		
	PerCPCy5.5		BD	0.67µg/ml		

Gene	Forward primer $(5' \rightarrow 3')$	Reverse primer $(5' \rightarrow 3')$
Rplp0	TGC AGA TCG GGT ACC CAA CT	ACG CGC TTG TAC CCA TTG A
Ccr2	GTT CAT CCA CGG CAT ACT ATC AAC	GCC CCT TCA TCA AGC TCT TG
Ccr6	CCT GGG CAA CAT TAT GGT GGT	CAG AAC GGT AGG GTG AGG ACA
Rorc	CAG CCA ACA TGT GGA AAA GCT	GGG AAG GCG GCT TGG A
Irf4	CGG GCA AGC AGG ACT ACA AT	ACA ATG CCC AAG CCT TGA TG
Irf8	GCT GAT CTG GGA AAA TGA TGA GA	CAC CTC CTG ATT GTA ATC CTG CTT
Prdml	TGG CAG AGA CTG GGA TCA TG	CTC GGC CTC TGT CCA CAA A
Batf	GTT CTG TTT CTC CAG GTC C	GAA GAA TCG CAT CGC TGC
Tbx21	GCC AGG GAA CCG CTT ATA TG	AAC TTC CTG GCG CAT CCA
Eomes	TGA GCT TCA ACA TAA ACG GAC TCA	CGG CCA GAA CCA CTT CCA

Supplementary Table 2: Primers used in qPCR