

## Supplementary Figure 1. *Ndfip1*-deficient $T_{reg}$ cells have normal thymic numbers and peripheral suppression.

(**a-b**) *Ndfip1*<sup>+/+</sup>*Foxp3*-Cre (WT) only or WT/ *Ndfip1*<sup>fl/fl</sup> *Foxp3*-Cre mixed bone marrow chimeras were analyzed 6-8 weeks after reconstitution. (a) Percentages of thymic T<sub>reg</sub> cells for WT only or WT/ *Ndfip1*<sup>fl/fl</sup> *Foxp3*-Cre mixed chimeras. Gated on singlets, live cells, FSCa by SSCa lymphocytes. (b) Normalized congenic contributions to total  $T_{reg}$  cells in the thymi of mixed chimera animals. Normalization to account for reconstitution bias was done by dividing the percentage of CD45.1<sup>+</sup> WT cells or CD45.2<sup>+</sup> Ndfip1-deficient cells within the pool of total  $T_{reg}$  cells (TCR $\beta$ +CD4+CD8<sup>-</sup> Foxp3<sup>+</sup>) by the percentages within the pool of total CD4+ single positive cells (TCRβ+CD4+CD8- Foxp3-). (**c-e**) *Ndfip1*-deficient regulatory T cells have normal *in vitro* and *in vivo* suppression of wild type T<sub>conv</sub> cells. (c-d) In vitro suppression of congenically marked (CD45.1), CFSE-labeled, WT CD4<sup>+</sup> T<sub>conv</sub> cells by congenic WT T<sub>reg</sub> cells or *Ndfip1*<sup>fl/fl</sup> *Foxp3*-Cre T<sub>reg</sub> cells (c) CFSE dilution of  $T_{conv}$  cells after 4 days in culture (**d**) Quantification of the percentage of T<sub>conv</sub> cells that diluted CFSE in culture (e) Weight changes during an *in vivo* suppression assay using CD45.1 CD4+ T<sub>conv</sub> cells and CD45.2 WT or Ndfip1fl/fl Foxp3-Cre T<sub>reg</sub> cells. Error bars indicate mean ± SEM. Each dot represents an individual mouse. Male donors were used for mixed bone marrow chimeras (a-b). These chimeras are also shown in Figure 4. (c-e) represents data from at least 3 male animals of each genotype. (e) Represents one experiment. P values were determined by a paired t test (**a-b**) and student's t test (**c-e**) \* p < 0.05 \*\* p < 0.01



Supplementary Figure 2. *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre<sup>+/-</sup> female animals develop **disease**. (a) Inflammation index calculated by spleen weight/body weight for male *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre<sup>+</sup> animals, female *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre<sup>+/-</sup> animals, compared to male *Ndfip1*<sup>+/+</sup>*Foxp3*-Cre<sup>+</sup> WT controls and female *Ndfip1*<sup>+/+</sup>*Foxp3*-Cre<sup>+/-</sup>WT controls all between 8-16 weeks old. Male data from Figure 1 is added here for comparison. (b) Ex vivo PMA/ionomycin/golgi plug restimulated lung homogenates were analyzed by flow cytometry for IFN<sub>γ</sub>, IL-4 and IL-17A cytokine production from WT *Ndfip1*<sup>+/+</sup>*Foxp3*-Cre<sup>+</sup> male, *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre male, and *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre<sup>+/-</sup> female animals. (c-f) Analysis of ICOS, GITR, CD25 and PD-1 MFI are shown for the *Ndfip1*-sufficient (YFP<sup>-</sup>) T<sub>reg</sub> cells in unstimulated lung homogenates from *Ndfip1*<sup>+/+</sup>*Foxp3*-Cre<sup>+</sup> WT female or *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre<sup>+/-</sup> female animals. (g) Summary of T<sub>reg</sub> cell proteins whose expression is increased by inflammation versus those which are increased directly due to loss of Ndfip1. Error bars indicate mean ± SEM. Each dot represents an individual mouse. All experiments were performed on at least two independent occasions using at least 3 female animals of each genotype. *P* value calculated by one way ANOVA (**a**,**b**) or unpaired *t* test (**c**-**f**). \* *p*<0.05 \*\* *p*<0.01 \*\*\* *p*<0.001 \*\*\*\**p*<0.0001.

#### **Supplementary Figure 3**

• WT Foxp3-Cre+/- Female (day 13) • Ndfip1fl/fl Foxp3-Cre+/- Female (day 13)



Supplementary Figure 3. *Ndfip1*-deficient  $T_{reg}$  cells have altered phenotypes prior to inflammation onset. (a) Spleen weights and (b) Inflammation index calculated by spleen weight/body weight for 13 day old female *Ndfip1*<sup>fl/fl</sup> *Foxp3*-Cre +/- neonates compared to age-matched *Ndfip1*<sup>+/+</sup>*Foxp3*-Cre<sup>+</sup> WT controls. (c-d) Lung homogenates were analysed by flow cytometry for total Foxp3<sup>+</sup> cells (c) or stimulated *ex vivo* with PMA/ionomycin/golgi plug and analysed for total cytokineproducing CD4<sup>+</sup> T cells (d). (e-f) Percentages of (e) Foxp3<sup>+</sup> cells or (f) CD44<sup>+</sup> eT<sub>reg</sub> cells are shown as a ratio of YFP<sup>+</sup>: YFP<sup>-</sup> cells to normalize for effects of Cre expression (g-h) Analysis of ICOS (g) and GITR (h) MFI on unstimulated lung homogenates. Error bars indicate mean ± SEM. Each dot represents an individual mouse. All experiments were performed on at least two independent occasions. P value calculated by unpaired *t* test (a-c, e-h) or one-way ANOVA (d). \* *p*<0.05 \*\* *p*<0.01 \*\*\* *p*<0.001



Supplementary Figure 4. *Ndfip1*-deficient  $T_{reg}$  cells show normal *Foxp3* mRNA levels but lose Foxp3 protein *in vivo*. (a) Analysis of the methylation status of the CNS2 region in *Foxp3*<sup>-</sup> (Cre negative) cells isolated from WT male animals. (b) Representative alleles showing methylation status of the *Foxp3* promoter region from  $eT_{reg}$  cells isolated from 9-12 week old male WT or *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre mice. (c) Expression of *Foxp3*, as measured by qPCR analysis in unstimulated or  $\alpha$ CD3/CD28-bead-stimulated  $T_{reg}$  cells. *Foxp3* mRNA expressed as 2<sup>ΔCT</sup>, relative to *Actb*. (d-e) *In vitro* percentages (d) and MFI (e) of sorted YFP+ Foxp3 cells are increased in response to IL-2 and decreased in response to IL-4 and anti-IL-2. *P* value calculated by one way ANOVA \* *p*<0.05 \*\* *p*<0.01 \*\*\* *p*<0.001 \*\*\*\**p*<0.0001.





Supplementary Figure 5. Ndfip1-deficient T<sub>reg</sub> cells are more likely than controls to lose Foxp3 in vivo. (a-g) CD45.1 CD4+ Tconv cells and CD45.2 WT or *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre YFP<sup>+</sup> T<sub>reg</sub> cells were co-injected via IP at a 5 T<sub>conv</sub>:1 T<sub>reg</sub> ratio into *Rag1<sup>-/-</sup>* animals, as described in Figure 5. (a) Graph shows percentage change in weight over the course of the experiment. (b) Lung homogenates were sorted for T<sub>reg</sub> cells that remained Foxp3<sup>+</sup> (current T<sub>reg</sub> cells), those that had become Foxp3<sup>-</sup> following transfer (former T<sub>reg</sub> cells), and for CD45.1 T<sub>conv</sub> cells, restimulated, and analyzed for *Ndfip1* mRNA by qPCR. (**c-e**) IFNy production by current (**c**) and former (d)  $T_{reg}$  cells and from the co-transferred  $T_{conv}$  cells (e). (f-g) Absolute numbers of CD45.2<sup>+</sup> current and former  $T_{reg}$  cells (f) or of total CD45.2<sup>+</sup> cells (g) in recipients of WT or *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre T<sub>reg</sub> cells. All mice were analyzed 13 weeks after the cotransfer. Error bars indicate mean ± SEM. Each dot represents an individual recipient mouse. All experiments were performed on at least two independent occasions using at least 2 animals of each genotype. P value was calculated by unpaired *t* tests (b-e,g) or by one-way ANOVA (f). \**p*<0.05 \*\**p*<0.01 \*\*\**p*<0.001 \*\*\*\*p<0.0001

#### **Supplementary Figure 6.**



Supplementary Figure 6. *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre<sup>+/-</sup> T<sub>reg</sub> cells do not have decreased Foxp3 MFL. (a-c) Lung homogenates, from 8-16 week old hemizygous female *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre<sup>+/-</sup> mice and *Ndfip1*<sup>+/+</sup>*Foxp3*-Cre<sup>+/-</sup> controls, were analyzed *ex vivo* by flow cytometry as described in Figure 3. Data is shown for the ratio of Foxp3 MFI on: YFP<sup>+</sup> to YFP<sup>-</sup> Foxp3<sup>+</sup> cells (a) YFP<sup>+</sup> to YFP<sup>-</sup> eT<sub>reg</sub> cells (b), or on YFP<sup>+</sup> to YFP<sup>-</sup> cT<sub>reg</sub> cells (c) in *Ndfip1*<sup>+/+</sup>*Foxp3*-Cre<sup>+/-</sup> controls versus *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre<sup>+/-</sup> mice. *P* values determined by student's *t* test. \**p*<0.05 \*\**p*<0.01 \*\*\**p*<0.001 \*\*\*\**p*<0.0001. Each dot shows data acquired from a single female mouse. Graphs show mean ± SEM. All experiments were performed on at least two independent occasions.

#### Supplementary Figure 7.



# Supplementary Figure 7. IL-4 does not drive the *in vivo* fitness of *Ndfip1*deficient $T_{reg}$ cells. (a-g) Regulatory T cells from the lung of *IL-4* KO and *Ndfip1 IL-4* DKO mice were analyzed by flow cytometry *ex vivo*. (a) Representative flow plots of Foxp3<sup>+</sup> $T_{reg}$ cells from lung. (b-c) Percentage of (b) total lung $T_{reg}$ cells or (c) total lung Ki-67<sup>+</sup> $T_{reg}$ cells. (d) Representative flow plots of eT\_{reg} and cT\_{reg} cells, identified by CD44 and CD62L staining of lung homogenate. Previously gated on Foxp3. (e) Summary of percentages of $eT_{reg}$ cells from (d). (f-g) CD44<sup>+</sup> lung Foxp3<sup>+</sup> $eT_{reg}$ cells were analyzed for expression of the proliferation marker, Ki-67 (f) or ICOS (g). Error bars indicate mean ± SEM. N=2-4 (age-matched male or female) animals per genotype in one experiment. P values determined by student's *t* test \* *p*<0.05 \*\* *p*<0.01.

#### **Supplementary Figure 8.**



Supplementary Figure 8. *Ndfip1*-deficient  $T_{reg}$  cells do not show increased IL-2induced STAT5 phosphorylation. Sorted YFP<sup>+</sup> total  $T_{reg}$  cells were rested overnight without TCR or cytokines. The MFI for STAT5 phosphorylation as measured by flow cytometry, is shown for the  $T_{reg}$  cells with or without addition of 119 U/ml (~50ng/ml) of rhIL-2 for 30minutes in a 37°C incubator. Each dot represents data acquired from a single mouse (male or female, between 9-16 weeks old). Graphs show mean ± SEM and represent data from at least two independent occasions.

### **Supplementary Figure 9**

a Gating to look at surface markers or at intracellular proteins on eTreg versus cTreg YFP+ versus YFP- Treg cells



Supplementary Figure 9. Flow cytometry gating schemes. (a) Gating scheme for analyzing T<sub>reg</sub> cell surface proteins and intracellular proteins in female *Ndfip1*<sup>+/+</sup>*Foxp3*-Cre<sup>+/-</sup> controls and *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre<sup>+/-</sup> mice shown in Figures 3, Supplementary Figure 2, and Supplementary Figure 3. Male animals were gated similarly, omitting the GFP by Foxp3 step, since all Foxp3<sup>+</sup> T<sub>reg</sub> cells in male animals are YFP<sup>+</sup>. YFP was detected using an anti YFP/GFP antibody. The gating schema for males was used for Figure 1e-f, Figure 2, Figure 4, Figure 5, and Figure 8. (b) Gating scheme for sorting total CD4<sup>+</sup> YFP<sup>+</sup> Treg cells. Total T<sub>reg</sub> cells were sorted for Figure 1a, Figure 4c, Figure 7, Figure 8, Supplementary Figure 4c, and Supplementary Figure 5b. To sort CD62L<sup>lo</sup> CD44<sup>+</sup> eT<sub>reg</sub> and CD62L<sup>hi</sup> CD44<sup>-</sup> cT<sub>reg</sub> cells, CD4<sup>+</sup> YFP<sup>+</sup> cells were further separated by CD44 and CD62L staining as shown in (a). cT<sub>reg</sub> versus eT<sub>reg</sub> YFP<sup>+</sup> Foxp3<sup>+</sup> cells were sorted for Figure 5a-b, Figure 6, Figure 7, and Figure 8. (c) Gating schema for mixed chimera experiments and adoptive transfer experiments. In the mixed chimera experiments in Figure 4, congenically marked *Ndfip1*<sup>+/+</sup>*Foxp3*-Cre WT cells (CD45.1<sup>+</sup> or CD45.1<sup>+</sup>/CD45.2<sup>+</sup>) were compared to CD45.2<sup>+</sup> *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre cells. In the adoptive transfer experiments for Figure 5 and Supplementary Figure 5, WT T<sub>conv</sub> cells (CD45.1<sup>+</sup>/CD45.2<sup>+</sup>) were co-transferred and compared to either CD45.2+ Ndfip1+/+Foxp3-Cre WT Treg cells or CD45.2+ *Ndfip1*<sup>fl/fl</sup>*Foxp3*-Cre Treg cells.