

Supplement Table 1. Donor Characteristics and Clinical Information ^A

Disease State ^B	Sample ^C (n)	Sex		Age	Time ^D	Cause of Death (n) ^E
		M	F	Mean ± SD	Mean ± SD	
No AMD	31	14	17	65 ± 11	19 ± 5	ACE (2), ALS (1), ABI (3), Cancer (8), CHF(2), CVA (2), Heart Failure (3) ICH/ICB (2), Organ Failure (2), Pneumonia (2), PE (1), Sepsis (3)
AMD	49	30	19	75 ± 11	20 ± 3	ACE (3), CABG (1),Cancer (10), CHF (4), COPD (1),CVA (4),Heart Failure (7), Organ Failure(3), Pneumonia (4), PE (1), Respiratory Failure (1), Sepsis (9), Unknown (1)

ABI = Anoxic Brain Injury; ACE = acute myocardial event; ALS=amyotrophic lateral sclerosis; CABG= coronary artery bypass grafting; CHF = congestive heart failure. COPD = chronic obstructive pulmonary disease; CVA = cerebrovascular accident (stroke); Heart failure = Myocardial infarction or cardiac arrest; ICH/ ICB = Intracerebral hemorrhage/ Intracerebral bleed; Organ failure=kidney and/or multiple organ failure; PE= pulmonary embolism.

^A Information supplied by Minnesota Lions Eye Bank.

^B Minnesota Grading System (MGS) was used to evaluate the presence or absence of AMD in eye bank eyes (Olsen 2004).

^C Sample number indicates the total donors used in the current study for each MGS stage.

^D The time from death to freezing is provided in hours.

^E The number of donors for each cause of death is indicated in parentheses.

Supplement Table 2. Antibodies used for Immunohistochemistry and Western Immunoblotting.

<u>Antibody</u>	<u>Type</u>	<u>Assay</u>	<u>Dilution</u>	<u>Company</u>
RPE65	P	W	1:2000	Dr. T. Michael Redmond, NEI
Bestrophin	M	W, I	1:1000, 1:500	Novus Biologicals, Littleton, CO
CRABP	M	I	1:250	Sigma-Aldrich, St. Louis, MO
CRALBP	M	W, I	1:1000, 1:500	Novus Biologicals, Littleton, CO
NaK ATPase	M	W, I	1:1000, 1:250	Novus Biologicals, Littleton, CO
MCT3	P	W	1:200	Santa Cruz Biotechnology, Dallas, TX
Ezrin	P	W	1:1000	Cell Signaling, Danvers, MA
ZO-1	M	I	1:100	Life Technologies, Grand Island, NY
Otx2	P	I	1:500	EMD Millipore, Darmstadt, Germany
MitF	M	I	1:200	Abcam, Cambridge, MA
CD31	M	W	1:1000	Cell Signaling, Danvers, MA
CD45	M	W	1:1000	Cell Signaling, Danvers, MA
Mast Cell Tryptase	M	W	1:1000	Novus Biologicals, Littleton, CO
B-Actin	M	W	1:5000	Santa Cruz Biotechnology, Dallas, TX
SRXN1	P	W	1:1000	Abcam, Cambridge, MA
GPX1	M	W	1:1000	Cell Signaling, Danvers, MA
PRDX3	P	W	1:1000	Abcam, Cambridge, MA
GST π	M	W	1:5000	Novus Biologicals, Littleton, CO
Catalase	M	W	1:2000	Sigma-Aldrich, St. Louis, MO
PGC1 α	M	W	1:1000	EMD Millipore, Darmstadt, Germany
<u>PPARα</u>	<u>M</u>	<u>W</u>	<u>1:1000</u>	<u>Abcam, Cambridge, MA</u>
<u>CuZnSOD (SOD1)</u>	<u>P</u>	<u>W</u>	<u>1:1000</u>	<u>Stressgen, Victoria, BC Canada</u>
<u>MnSOD (SOD2)</u>	<u>P</u>	<u>W</u>	<u>1:1000</u>	<u>Abcam, Cambridge, MA</u>

Monoclonal (M), polyclonal (P), Immunohistochemistry (I), Western Immunoblotting (W).

Supplement Table 3. Primers for qPCR Analysis

Gene	Gene ID		Sequence 5'-3'	Amplicon (bp)	Annealing Temp
Cell Characterization					
MITF	4286	Forward	CCCAGTTCATGCAACAGAGAG	79	59
		Reverse	GCAGAGGGAAGGGTGGTG		
RPE65	6121	Forward	CGTATGGACTTGGCTTGAATC	129	59
		Reverse	CTGGGTGAGAAACAAAGATGG		
PMEL17	6490	Forward	TGATGGCTGTGGTCCTTGC	94	59
		Reverse	CAGTGACTGCTGCTATGTGG		
PEDF	5176	Forward	TATCACCTTAACCAGCCTTTC	83	59
		Reverse	GGGTCCAGAATCTTGCCAATG		
TYRP1	7306	Forward	GATTCCACTCTAATAAGCCCAAA	198	59
		Reverse	TTCCAAGCACTGAGCGACAT		
BEST1	7439	Forward	TAGAACCATCAGCGCCGTC	261	59
		Reverse	TGAGTGTAGTGTGTATGTTGG		
RBP1	5947	Forward	CAGGTTGGGAAGGAGTTTGA	168	59
		Reverse	TGGTGGCGTGTGACTGTAAT		
GAPDH	2597	Forward	GAGTCAACGGATTTGGTCGT	185	59
		Reverse	GACAAGCTTCCCCTTCTCAG		
mtDNA Content					
Total mtDNA Set 1	4550	Forward	CCCCACAAACCCCACTACTAAACCCA	222	55
		Reverse	TTTCATCATGCGGAGATGTTGGATGG		
Total mtDNA Set 2	4519	Forward	CAGTGAAATTGACCTGCCCGTGAA	191	55
		Reverse	TCTTAGCATGTACTGCTCGGAGGT		
β-globin	3043	Forward	CTTGGGTTTCTGATAGGCAC	147	60
		Reverse	CTTAGGGTTGCCATAACAG		
Oxidative Stress Response					
ARBP	6175	Forward	CGACCTGGAAGTCCAACACTAC	109	58
		Reverse	ATCTGCTGCATCTGCTTG		
HO-1	3162	Forward	CCAGCGGGCCACAACAAAGT	262	58
		Reverse	GCCTTCAGTGCCACGGTAAGG		
NQO1	1728	Forward	AAAGGACCCTTCCGGAGTAA	222	58
		Reverse	CCATCCTTCCAGGATTTGAA		
SOD2	6648	Forward	GTGTGGGAGCAGCCTTACTA	153	58
		Reverse	AGAGCTTAACATACTCAGCATAACG		
CAT	847	Forward	GATAGCCTTCGACCCAAGCAAC	201	58
		Reverse	TGATTGTCCTGCATGCACATCG		
SOD1	6647	Forward	GTGCAGGTCTCACTTTAATCCTC	212	58
		Reverse	ACCTTTGCCCAAGTGATCTGC		
NRF2	4780	Forward	TCCAGTCAGAAACCAGTGGAT	107	58
		Reverse	GAATGTCTGCGCCAAAAGCTG		
PGC1α	10891	Forward	GTCACCACCCAAATCCTTAT	131	59
		Reverse	ATCTACTGCCTGGAGACCTT		
PPARα	5465	Forward	ATCGAATGTAGAATCTGCGGG	116	60
		Reverse	TCGCACTTGTGCATACACCAG		
CYTC	54205	Forward	CTTCGGAGCGGGAGTGTTCCG	229	59
		Reverse	CAGATGATGCCTTTGTTCTTATTG		
PPARγ	5468	Forward	AGGCGAGGGCGATCTTGACAG	150	59
		Reverse	GATGCGGATGGCCACCTCTTT		
GPX1	2876	Forward	CCAGTCGGTGTATGCCTTCTC	107	61
		Reverse	CGAGAATGTGGCGTCCCTC		