

SUPPLEMENTARY INFORMATION

Heterologous expression of cytotoxic sesquiterpenoids from the medicinal mushroom

Lignosus rhinocerotis in yeast

Hui-Yeng Yeannie Yap^{1,3}, Mariano Jordi Muria-Gonzalez^{2†}, Boon-Hong Kong¹, Keith A. Stubbs³, Chon-Seng Tan⁴, Szu-Ting Ng⁴, Nget-Hong Tan¹, Peter S. Solomon², Shin-Yee Fung^{1*}, and Yit-Heng Chooi^{2,3*}

¹Department of Molecular Medicine, Faculty of Medicine, University of Malaya, 50603 Kuala Lumpur, Malaysia; ²Research School of Biology, The Australian National University, Canberra, Australian Capital Territory 2601, Australia; ³School of Molecular Sciences, University of Western Australia, Crawley, Western Australia 6009, Australia; ⁴Ligno Biotech, 43300 Balakong Jaya, Selangor, Malaysia

*Corresponding authors:

Y-H Chooi*, yitheng.chooi@uwa.edu.au; S-Y Fung*, syfung@um.edu.my

Co-authors' email addresses:

H-YY Yap, yea_ny_nie@yahoo.com; MJ Muria Gonzalez, jordi.muria@curtin.edu;

B-H Kong, bh_kong84@yahoo.com; KA Stubbs, keith.stubbs@uwa.edu.au;

C-S Tan, tanchonseng@gmail.com; S-T Ng, szuting@ligno.com.my;

N-H Tan, tanngethong@yahoo.com.sg; PS Solomon, peter.solomon@anu.edu.au;

Present address: [†]Centre for Crop and Disease Management, Curtin University, Perth, WA 6102, Australia.

Table S1. Coding sequences of *GME3634*, *GME3638*, and *GME9210*

Gene ID	Coding sequence
<i>GME3634</i>	ATGGCTGTAACCTCTGCAAACGTCGCCTCCCCGACTCGCAAAGAGATCGTGCTCAAATCCCCGATTTTCATCTCTCCCATCCCCCTACCCCTTCGCTGCCATGTGCAGGAGCGAGAAGTGTACGCCAATCCGAGGAATGGCTCCTCAGCATGGCCAACTTCTCCGAGAAACAGCGCAGCAAGTTCCTTACCCTCAACGGTGGTCTTCTTAGCGGCATGTGCTACATCGACTGCACGTTTACGAACTGCGGGTCTGCACCGACTTCATGAACTTCTGTTCACACTCGACGACTGGACGGACGAGTTCGACACGACCCGGCACGCGGGTCTCGCAGAGTGCATGAACACGCTTACTGGCCGCACAGCTACCAGGCAGACACCGCCGCGCACCGCTTGACAAAGTCTTCTGGGTCCGCATGAAGCAGACGGCAGGCCCGGGTGTGACGACGCGCTCATGTCGACGCTCGACACGTAATCCAGGCCATCATGCAACAGGCCGCCGACCGTGGTTCACATAACATTCCGGAGTTGGAGGAGTACATCTTGTCCGCCGAGACACGAGCGGGTGCAAGATCGGTTTCGCGTTCATCGAATACGCAGCCAACATTGATCTGCCCGACGACGTCATTGAGCACCCAATCATCAAGGCCATGGCCGATGCGACCAATGATCTCGTTCGTTGGGCGAATGATGTCCTTTCGTACAACGCTGAACAGTACGCGGAGACACGCACAACCTCGTTTGTGTTCTCATGGCTCAGAACGGCCTCGACCCGGCAGGGTGCATCGAGCTGGCGGGGAGCTATGGGAGAAGACCCTCCACTTGTCTTCGAGTCCCGAAGAACGTCCCTTCGTGGGGTCCGAGATCGACCGTCCGCTCGTCTGTACATCCAGGGGCTCGAAGACTGGATTATCGCGAACGCCGAGTGGAGCTTCGAGACGGAGCGCTACTTTGGCAAGGACGGGCATCTCGTGAAGAAGACGCGGCAGGTTACACTGCTTCCGGTCCGTACAGCTGCG
<i>GME3638</i>	ATGCGCGCTCGATCGTTTCTTCCCGACCTCGTGTGCGGATTGCCCATATACGCTCAGGTGCAACTCCAATTGCGAAGCTGTTGCGCGCGCTTCAGAGCCTGGATGCTCGAAGACGCGAATCTCTACCGAAACGTGTGACGCCTTCTGCGTCTGCGGGGGGAGAACTCACGGCAGCGTGCTACCCCGATACGGACGAGGCCTGCCGAGTCGCCGCGGACTTCTCAATTTCTGTTAGTTGGACGACTGGTTCGGACGAGTTCAGTATGGAGACACTTGCAGGCTCGCGCAGTGCATGTGTGTGCTCCATGATCCTGATGACTTCCAGACTGAAAAGGCTGCTGGCAAGCTCGCCAAAAGCTTTTTCAATCGGTTCCGGCAGACGGCGGGGCCGAGGTGTACTCGTCGATTCATCGATAGTATGGACCTTCTTTTCATGCGATTGCACAGCAAGCCAGGACCGCGCGTCCGGTCTGCTCCCTCACTCGAAGAATATGTGGCCCTCCGCGAGGACACGAGCGGGTGCAAGCCCTGCTTCGCTCTCATCGAATACGCCGCGGTATGGACCTTCCAGACCAGTCGCCCATCATCAACAATCACCGCCCTCGAGCGGGAAGCCAACGCGTGTATATCATGGTCGAACGATCTTCTCGTACAACGTCGAGCAAGCGCGGGTGACACACACAACATGATTGCGGTGATCATGCGCGAGGACGGGCGCAGCTTCAAGAAGCCGTTGAATACTTGGGCGCTCTGCAAGCTCTGCATAGTGCACTTCGAGGAAAACCGAGCCATGCTGCCATCGTGGGGTCCGGAGATTGACGGGGAAGTCGACAGGTATGTGCTCGGCCTCCAGGACTGGATGGTTCGGTCCCTCCATTGGAGCTTCGACACCGCACGTTATTTCCGGGATGAAGGCCCTGCGATCAAGAAGCACGGTGTCTGACTACTACCGCGGAAGTCTTCATCG
<i>GME9210</i>	ATGCCTTTGTCTTCGTCTGTTGTCGCTTCCGCCTTCTGACACGCTTGGGTGCTGGCCATGGCGACGCTGCCTGAACACGCATTATGTGGAGGCCAAACAGGATTCCGCATCCTGGCTGGAGTCATTCCACCCGTTCCGGCCCAAGGCACAAAGGGCCTTCAACAAGTGCAGCTTCAACCTTCTCGCTCCCTAGCGTATCCGGTGGCCAGCAAAGACCAACTGCGTGCAGGGTGTGACCTGATGAACGCTTCTTTCGTCTTCGACGAGTACTCTGATGTGAGAACGAGAAGACGGTCCAGCAGCTCGCGGACATCATGACGCACTGCGAAACCCTCACAAGCCTCGTCTTCGAGGAGAATCTCTCGTGGGCGAAATCACTAGGCAGTTCTGGGCGCGCACGATCAAGGTAGCGAGCGAACCCTCGCAGCGCCGATTTCATCGAGACCTTTGACGACTACTGCCAGTCCGTAACAGGCTGCGGACCGGTTCGAGAACCCTTGCCTGACGTCGAGACTACTTGGAGAACCAGGAGGAGAATCGGGCCAAAGCCGTCGTTTGCAGTCTCGAGTTGGACATGAACCTCCCCGACGAGGTGATCGAGCACCCGACCATCGTCAACCTGACGACATGGGCCATTGACATGATCATCTGGGAAACGACATTGTCTCGTACAACGTTGGAGCAGGCGCGTGGCGACGACGGACACAACGTCGTGACCATCGTCATGCATCCTACAACGTCGACGTGCAGGGTGCCATGGACCGCATCGCCGAGTGGACCAGAGGCTGGCGGACCAGTTCCTCACCAACTACAACAAGCTGCGTCTGGGGACGCGAGATTGACGCGCAAGTTGAGCGGTACATCCAGGGCATCGAAACTGGGTACGCGCGAACGACGATGGAGTTTCGAGAGCGAGAGGTAATTTGGATTGAACGGACGCGAGATCGAGCAGAGTCTGTTGGGTAACCCTACTCCCTCGGGTCTCCGCGGAGAAGCCTGCGGTTGTG

Table S2. PCR primers information

Gene ID	Primer name	Sequence	Product size (bp)
<i>GME3634</i>	XW-GME3634-F	ATCAACTATCAACTATTA ACTATATCGTAAT ACCAATGGCTGTA ACTCCTGCAAAC	410
	GME3634-fg1-r	GCTTCATGCGGACCCAGAAGGACTTTGTCAA GCGGTGCG	
	GME3634-fg2-f	CGCACCGCTTGACAAAGTCCTTCTGGGTCCG CATGAAGC	289
	GME3634-fg2-SOE-r	GTCTCCGCGTGACTGTT CAGCGTTGTACGAA AGGACATCATT CGCCCACGAAACGAGATC	
	GME3634-fg3-SOE-f	GATGTCCTTTTCGTACAACGCTGAACAGTCAC GCGGAGACACGCACAACCTCGTTTGTGT	297
	XW-GME3634-R	TGTCATTTAAATTAGTGATGGTGATGGTGAT GCACCGCAGCTGTACGGACCG	
<i>GME3638</i>	XW-GME3638-F	ATCAACTATCAACTATTA ACTATATCGTAAT ACCAATGCGCGCTCGATCGTT	374
	GME3638-fg1-r	GCCGGAACCGATTGAAAAGCTTTTGGCGAG CTTGCCAGC	
	GME3638-fg2-f	GCTGGCAAGCTCGCCAAAAGCTTTTCAATC GGTTCCGGC	289
	GME3638-fg2-r	ACGTTGTACGAGAAGAGATC- GTTTCGACCATGATATACACGCG	
	GME3638-fg3-f	CGTGTATATCATGGTTCGAACGATCTCTTCTCG TACAACGTCGA	336
	XW-GME3638-R	TGTCATTTAAATTAGTGATGGTGATGGTGAT GCACCGATGAAGACTTCCGCGGTA	
<i>GME9210</i>	XW-GME9210-F	ATCAACTATCAACTATTA ACTATATCGTAAT ACCAATGCCTTTGTCTTCGTCTGTTG	178
	GME9210-fg1-r	CTTTGCTGGCCACCGGATACGCTAGGGACGC GAGAAGGTTGAAGTCGCACTTGTTGAAGG	
	GME9210-fg2-f	ACCTTCTCGCGTCCCTAGCGTATCCGGTGGCC AGCAAAGACCAACTGCGTGCAGGGT	177
	GME9210-fg2-r	CTTGATCGTGCGCGCCAGA ACTGCCTAGTG ATTTGCCCCA	
	GME9210-fg3-f	GGGCGAAATCACTAGGCAGTTCTGGGCGCGC ACGATCAA	282
	GME9210-fg3-r	CACGTTGTACGAGACAATGTCGTTTCCCAGG ATGATCATGTCA	
	GME9210-fg4-f	CATGATCATCCTGGGAAACGACATTGTCTCG TACAACGTGGAG	353
	XW-GME9210-R	TGTCATTTAAATTAGTGATGGTGATGGTGAT GCACCACAACCGCAGGCTTCTCCG	

Table S3. ^{13}C (150.903 MHz) and ^1H (600.130 MHz) NMR spectral data for torreyol and α -cadinol (using CDCl_3 as internal standard, 77.16 ppm)

Carbon*	Torreyol (1)		α -Cadinol (2)	
	δC (ppm)	δH (ppm)	δC (ppm)	δH (ppm)
1	45.55 d	1.60 m	50.15 d	1.21
2	18.52 t	1.89 m, 1.58 m	22.09 t	1.60, 1.14
3	31.14 t	1.99	31.07 t	1.98
4	134.37 s	---	135.12 s	---
5	124.61 d	5.51 m	122.46 d	5.50 s
6	36.79 d	2.02 d	40.00 d	1.72
7	44.10 d	1.3	46.83 d	1.04
8	21.54 t	1.51, 1.01	22.81 t	1.27, 2.00
9	35.34 t	1.56 t, 1.52 t	42.33 t	1.82, 1.80
10	72.58 s	---	72.62 s	---
11	23.64 q	1.66	23.99 q	1.67 s
12	26.42 d	1.97 m	26.12 d	2.15 m
13	21.69 q	0.89 d	21.67 q	0.92 d
14	15.33 q	0.81 d	15.26 q	0.77 d
15	27.97 q	1.29	20.91 q	1.10 s

*Carbon signal assignments are based on Van Eijk et al. [1].

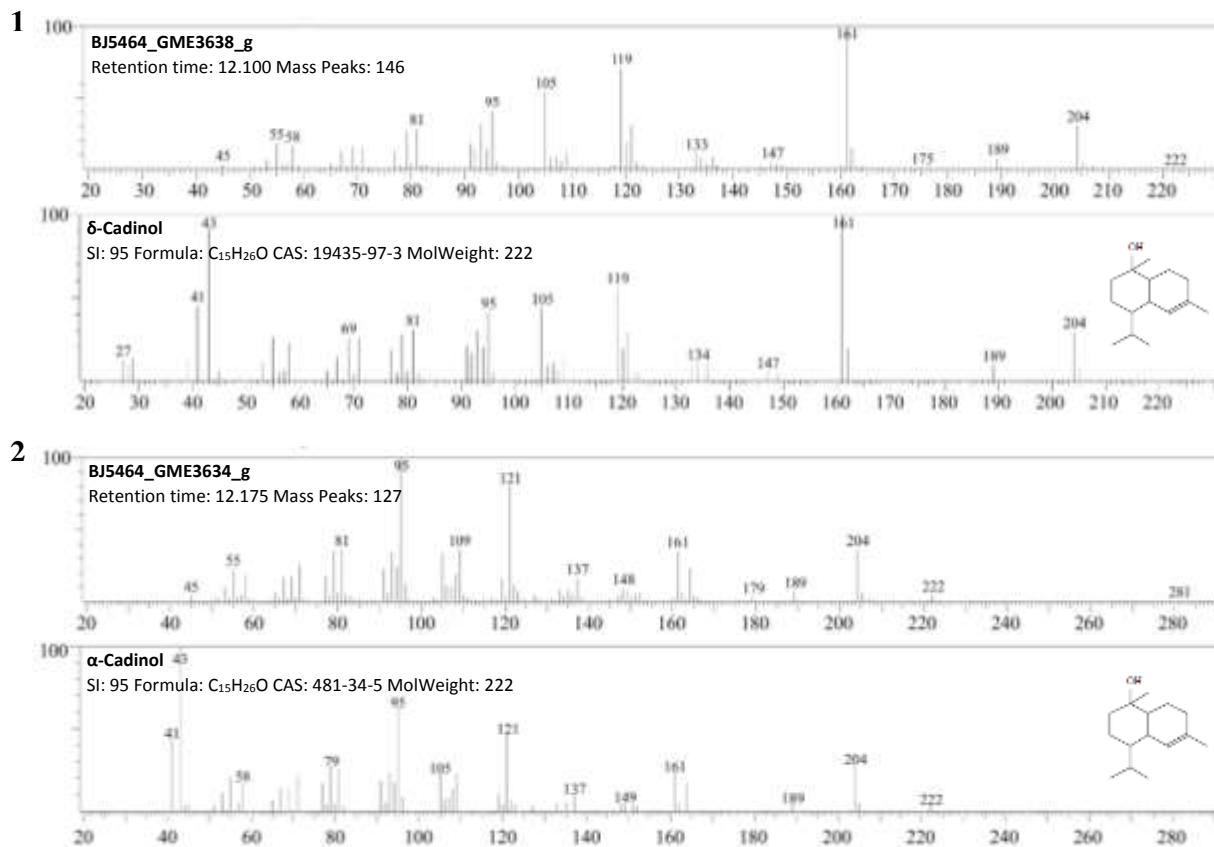


Figure S1. Mass spectrums of putative torreyol (1) and α -cadinol (2) to that of known compounds in NIST 05 library.

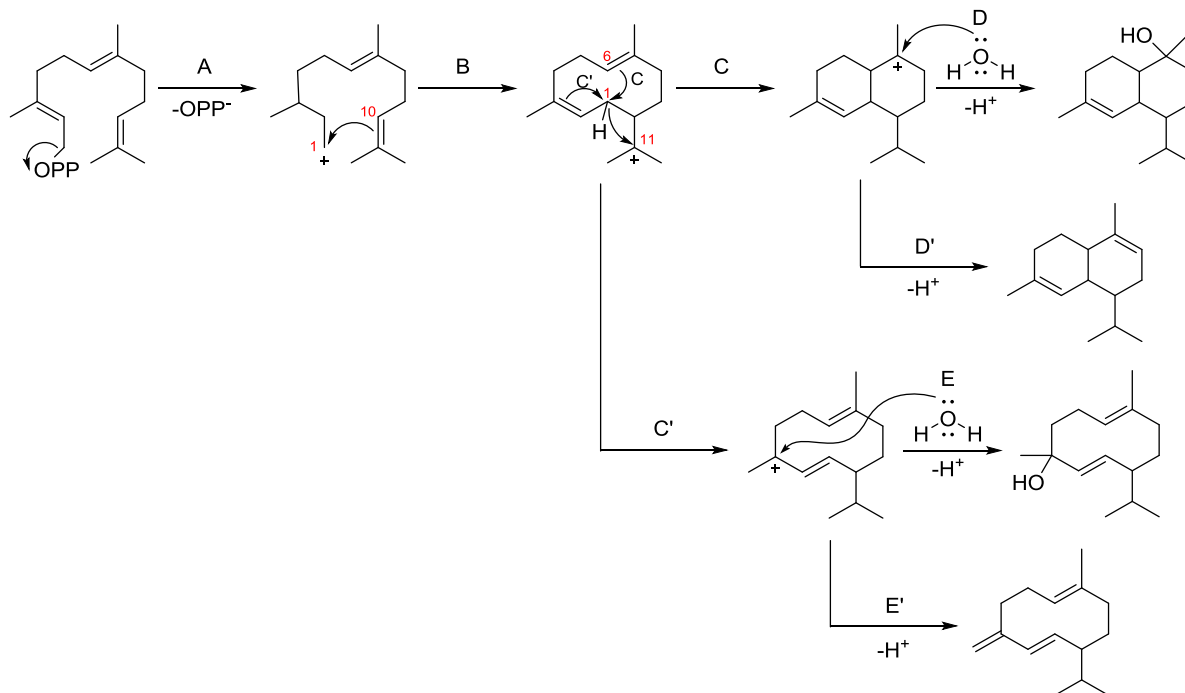


Figure S2. Proposed general mechanism for the biosynthesis of torreyol and α -cadinol from farnesyl pyrophosphate. The excision of the pyrophosphate group is catalysed by the enzyme producing the farnesyl cation (A: farnesyl carbocation formation). This is followed by the nucleophilic attack of carbon 10 (C-10) to the cation in C-1 that produces the first cyclisation reaction leading to germacranyl cation (B: cyclisation leading to germacranyl cation). At this point, the germacranyl cation can be quenched (C': hydride shift) by deprotonation or hydrolysis to form germacrene (E: carbocation quenching by hydrolysis producing germacrene D-ol) and germacrene-4-ol [E': hydride shift leading to a germacrene (germacrene D is presented here)] respectively, or continue the second cyclisation reactions via a Wagner-Meerwein rearrangement followed by the nucleophilic attack of C-6 over C-1. Depending on which face the hydride shift occurs, this leads to either the muurolenyl or cadinenyl cation (C: hydride shift). Finally, the resulted cation is quenched by either deprotonation [D': hydride shift resulting in a cadinene (α -cadinene is shown here)] or hydrolysis (D: carbocation quenching by hydrolysis leading to torreyol and α -cadinol) to yield cadinene/muurolene or cadinols/muurolols, respectively. Note: The numbering in here follows the farnesyl numbering, not the final product numbering. In the case of GME3634 and GME3638, the deprotonation is a competing reaction that also yields sesquiterpene products that can be seen in Table 1; however, hydrolysis appears to be the preferred mechanism.

Reference

1. Van Eijk GW, Roeijmans HJ, Verwiel PEJ. Isolation and identification of the sesquiterpenoid (+)-torreyol from *Xylobolus frustulatus*. *Exp Mycol.* 1984;8:273-5.