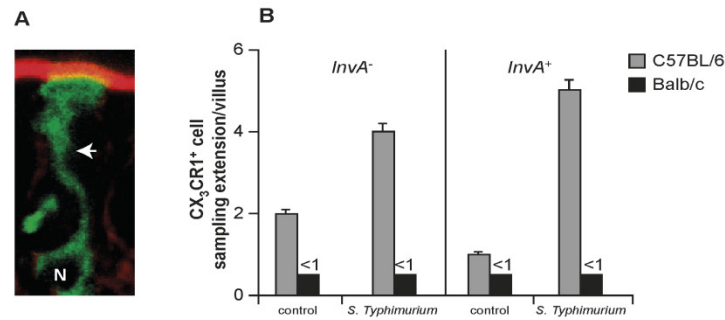


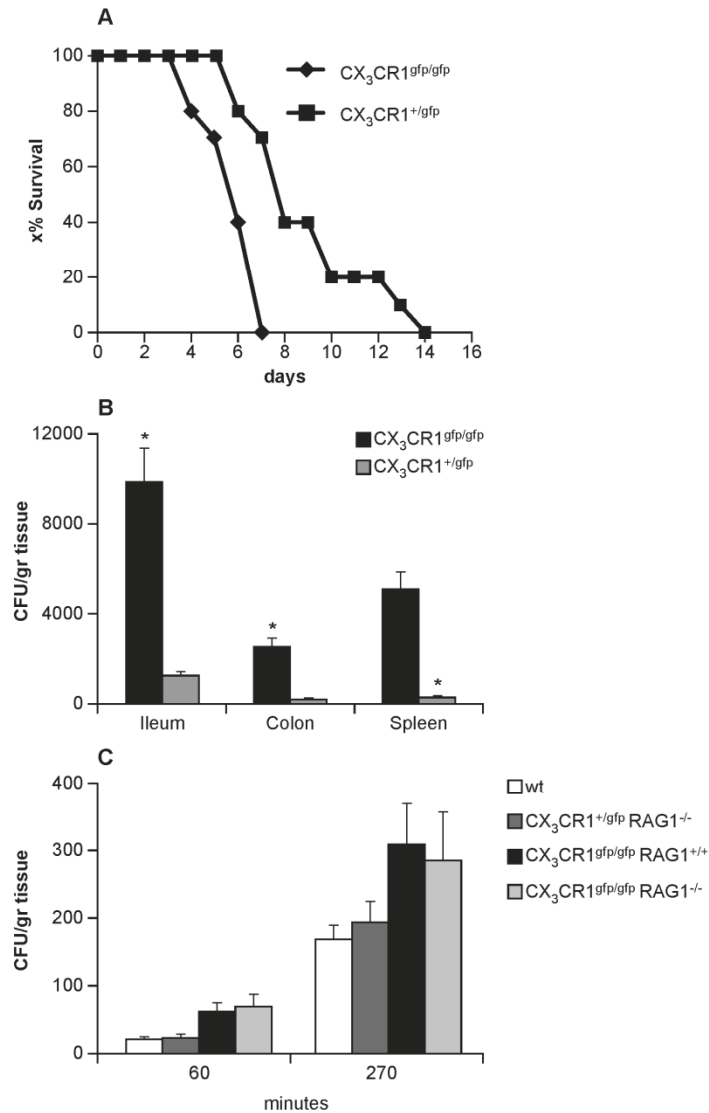
## Supplemental Materials

Supplemental Fig 1



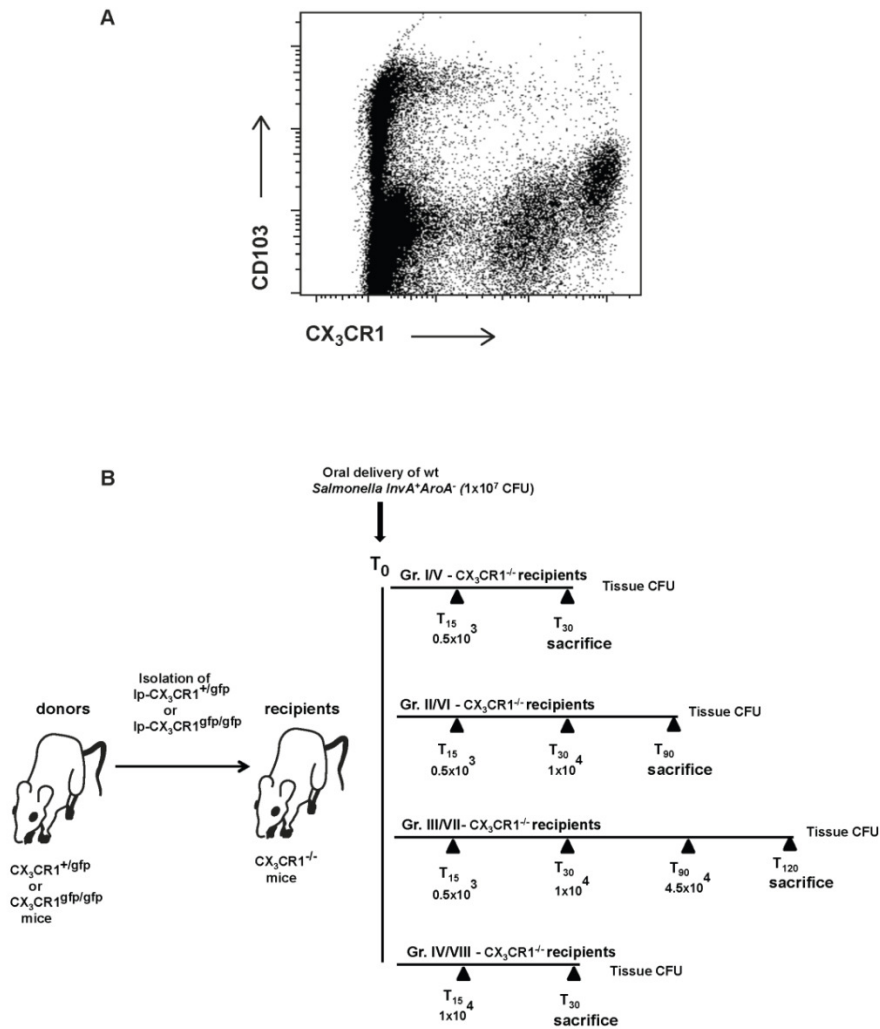
**Supplemental Fig 1:** *CX<sub>3</sub>CR1*-mediated antigen-sampling in mice of different genetic background. Confocal microscopy was used to enumerate cellular extension, or dendrites (arrow head) considered being sampling devices of *CX<sub>3</sub>CR1*<sup>+gfp</sup> cells (1A) in the epithelium of the ileum of both C57BL/6 and Balb/c. The number of dendrites/villus increased significantly in C57BL/6, but not in Balb/c mice following challenge with both non-invasive (*InvA*<sup>-</sup>) and invasive (*InvA*<sup>+</sup>) *Salmonella*. As previously reported (8), the ability of *S. Typhimurium* to invade did not have a significant impact on the number of dendrites produced by *CX<sub>3</sub>CR1*<sup>+gfp</sup> cells.

Supplemental Fig 2



**Supplemental Fig 2.** Role of CX<sub>3</sub>CR1 and RAG<sup>-/-</sup> background on susceptibility to salmonella infection and invasion of *S. Typhimurium*. CX<sub>3</sub>CR1<sup>gfp/gfp</sup> mice succumbed within 7 days from infection at a faster rate compared to CX<sub>3</sub>CR1<sup>+/gfp</sup> mice (A) (12 mice/group). It appeared that in spite of lacking the ability to sample *S. Typhimurium* via the indirect route CX<sub>3</sub>CR1<sup>gfp/gfp</sup> mice showed significantly higher bacterial load both locally (ileum and colon) and systemically (spleen) (B). Furthermore, increased susceptibility of these mice to infection could not be attributed to reduced antibody responses to *S. Typhimurium* as shown in Figures 6. Furthermore, to rule out that the impact of CX<sub>3</sub>CR1 deficiency in the number of *S. Typhimurium* traversing the epithelium could be related to altered activation of adaptive immune responses we assessed bacterial translocation in CX<sub>3</sub>CR1<sup>+/-</sup> and CX<sub>3</sub>CR1<sup>-/-</sup> on RAG<sup>-/-</sup> background (C). We observed that the RAG<sup>-/-</sup> background did not affect bacterial translocation

Supplemental Fig 3



**Supplemental Fig 3. Protocol for *Salmonella*-exclusion assay.** Flow cytometry analysis was carried out to identify CX<sub>3</sub>CR1<sup>-</sup>CD103<sup>-</sup>, CX<sub>3</sub>CR1<sup>-</sup>CD103<sup>+</sup> DC and CX<sub>3</sub>CR1<sup>+</sup>CD103<sup>-</sup> cells in the lamina propria of CX<sub>3</sub>CR1<sup>+/gfp</sup> mice (A). The latter population was introduced directly into the intestinal lumen at various intervals and increasing number after *Salmonella* infection as detailed in the pathogen exclusion assay protocol (B). Four groups of mice (6 mice/group) (I-IV) received a single oral delivery of  $1 \times 10^7$  invasive/non-replicating *InvA<sup>+</sup>AroA<sup>-</sup>* *Salmonella* at T<sub>0</sub>; this was followed by the intraluminal injection of CX<sub>3</sub>CR1<sup>+/gfp</sup> cells. The number of CX<sub>3</sub>CR1<sup>+/gfp</sup> cells for the adoptive transfer of intraluminal cells in Gr I, II and III at any given time (T<sub>15</sub>, T<sub>30</sub> and T<sub>90</sub>) was established according to the results of the time course experiment illustrated in Figure 3C. Group IV received a number of CX<sub>3</sub>CR1<sup>+</sup> cells ( $1 \times 10^4$ ) that far exceeded the number of cells observed in the lumen 15 minutes (T<sub>15</sub>) after infection. At the end of the experiments mice were sacrificed and CFU gr/tissue determined. The same protocol was repeated in four additional groups (V-VIII) to test the ability of CX<sub>3</sub>CR1<sup>gfp/gfp</sup> cells to prevent *S. Typhimurium* from traversing the epithelial barrier.