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### Supplementary Materials for

## Differential intron retention in *Jumonji* chromatin modifier genes is implicated in reptile temperature-dependent sex determination

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### The PDF file includes:

- fig. S1. Comparison of gene expression in male and female dragons.
- fig. S2. Comparison of gene expression in normal and sex-reversed female dragons.
- fig. S3. Differential JARID2 IR in normal and sex-reversed dragons.
- fig. S4. Temporal dynamics of *JARID2/JMJD3* expression and splicing in alligator and turtle embryo.
- fig. S5. Expression and splicing of *JMJD3* in the brain and gonad from normal and sex-reversed dragons.
- fig. S6. Differentially retained introns in *JARID2/JMJD3* are nonparalogous.

**Other Supplementary Material for this manuscript includes the following:** (available at advances.sciencemag.org/cgi/content/full/3/6/e1700731/DC1)

- table S1 (Microsoft Excel format). RNA sequencing libraries for *P. vitticeps*.
- table S2 (Microsoft Excel format). RNA sequencing libraries for *A. mississippiensis*.
- table S3 (Microsoft Excel format). RNA sequencing libraries for *T. scripta*.



**fig. S1. Comparison of gene expression in male and female dragons.** Gene expression profiles for normal female (ZWf) and male (ZZm) dragons were compared in order to identify sex-specific differences. (**A** and **B**) Volcano plots show average gene expression fold-change between adult brain from ZZm and ZWf relative to average normalized expression level (mean; n = 2). Expression measurements were made separately using Kallisto alignment-free quantification (A) and STAR-RSEM alignment-based quantification (B). Under each method, genes that exceeded a differential expression threshold of FDR < 0.01 are highlighted (red dots). (**C**) Venn diagrams show the number of genes that were found to be differentially expressed (FDR < 0.01) under either or both methods of quantification. Only genes concordant between the two methods were reported as differentially expressed. (**D**) We found eight genes that were significantly over-expressed in ZZm relative to ZWf brain. All eight are involved in canonical muscle processes, including the eminent muscle genes *Actin (ACTA1), Myosin (MYH1), Troponin (TNNI2)* and *Creatine kinase (CKM)*. The plot shows the average normalized expression (transcripts per million; mean +/- SD; n = 2) recorded in adult brain from

ZWf, ZZm and sex-reversed female (ZZf) dragons for the eight genes that were differentially expressed between ZZm and ZWf. Importantly, ZZf dragons exhibited normal female expression for all sex-biased genes. (E) Gene ontology terms that were enriched (GOrilla) among the top 500 genes upregulated in ZZm, relative to ZWf, brains included 'muscle system process' as well as related sub-terms like 'filament sliding' and 'electron transport chain'. Collectively, these data reveal a previously unreported male-bias in the expression of canonical muscle genes in dragon brain.

#### ZZ-female vs ZW-female



fig. S2. Comparison of gene expression in normal and sex-reversed female dragons. Gene expression profiles of normal female (ZWf) and sex-reversed female (ZZf) dragons were compared in order to identify sex-reversal specific transcriptome features. (A and B) Volcano plots show average gene expression foldchanges between adult brain from ZZf and ZWf relative to average normalized expression level (mean; n = 2). Expression measurements were made separately using Kallisto alignment-free quantification (A) and STAR-RSEM alignment-based quantification (B). Under each method, genes that exceeded a differential expression threshold of FDR < 0.01 are highlighted (red dots). (C) Venn diagrams show the number of genes that were found to be differentially expressed (FDR < 0.01) under either or both methods of quantification. Only genes concordant between the two methods were reported as differentially expressed. (D) 17 genes were classified as differentially expressed between ZZf and ZWf, of which 14 were down-regulated in ZZf. Expression of prominent immune genes, such as IgM and IRF1, was reduced in ZZf dragons compared to ZWf or ZZm, while the circadian regulator, CIART, was over-expressed. The immune and circadian systems are known to be intertwined with stress. (E) Annotated gene model for the predicted ortholog of *Proopiomelanocortin (POMC)* in dragon. Normalized coverage by mapped RNA sequencing reads shows expression of POMC in a single replicate of ZWf (blue), ZZm (yellow) and ZZf brain (red). (F) Top 5 nonredundant gene ontology terms (GOrilla) that were enriched among the top 500 genes down-regulated (above) or up-regulated (below) in ZZf, relative to ZWf, individuals.



**fig. S3. Differential JARID2 IR in normal and sex-reversed dragons.** Annotated gene model for the predicted ortholog of *JARID2* in dragon. Normalized coverage by mapped RNA sequencing reads (gray) and density of spliced-read junctions (colored) spanning annotated introns are shown for a single replicate from adult tissues for normal female (ZWf; blue), male (ZZm; yellow) and sex-reversed female (ZZf; red).



0

S12

S15

S16

0. D0 D3 D6



IR-IR+ FPT MPT

IR-IR+ FPT MPT

S16

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0

S12

fig. S4. Temporal dynamics of JARID2/JMJD3 expression and splicing in alligator and turtle embryo. (A) Alligator eggs were incubated under FPT (30°C) until Ferguson developmental stage 19, a period in which the gonads are still bipotential and morphologically indistinguishable. At stage 19, a subset of eggs was shifted to MPT (33.5°C) while the remaining eggs were maintained at FPT for the subsequent incubation period. Tissues comprising the developing gonad were sampled at FPT of day 0, then at FPT and MPT at multiple time points after stage 19. Full procedure is described in reference (4). Turtle eggs were separately incubated at FPT (31°C) or MPT (26°C) from the day of laying. Whole embryos were sampled at FPT and MPT at development stage 12, before the initiation of gonad development. Gonads were then sampled at FPT and MPT from stage 15-21, fully encompassing the temperature sensitive period of development. Full procedure is described in reference (5). (B) Normalized coverage by mapped RNA sequencing reads (gray) and density of spliced read junctions spanning annotated introns are shown for a single replicate of: (left) alligator embryonic gonad at FPT day 0 (stage 19; blue), FPT day 3 (blue) and MPT day 3 (yellow); (right) turtle whole stage 12 embryos at FPT (blue) and MPT (yellow) and embryonic gonad at stage 15 FPT (blue) and MPT (yellow). Note that the section of zero-coverage in the center of the retained intron for turtle is a string of undefined (N) bases in the genome, to which reads cannot be mapped. (C) Average normalized expression (transcripts per million; mean +/- SD; n = 3) for spliced (IR-) and intron retaining (IR+) isoforms of JARID2 and JMJD3 in alligator and turtle. Measurements shown are from alligator gonad at FPT day 0 and FPT/MPT day 3 and 6 and turtle FPT/MPT whole stage 12 embryos, and embryonic gonads at FPT/MPT stage 15 and 16.







**fig. S6. Differentially retained introns in JARID2/JMJD3 are nonparalogous.** (A to B) This schematic shows the relationship between orthologous introns, which fall between orthologous coding exons, and non-paralogous introns, which do not fall between paralogous coding exons. (C to E) Alignment (MUSLCE) of peptide sequences for dragon and aligator *JARID2* (C), dragon and aligator *JMJD3* (D) and dragon *JARID2/JMJD3* (E), with exon-intron architecture indicated. Alignments demonstrate that the differentially retained introns identified in dragon and aligator *for JARID2/JARID2* and *JMJD3/JMJD3* are orthologous but the differentially retained introns in *JARID2/JMJD3* are not paralagous to one and other, suggesting that their capacity for differential retention is not underpinned by a shared conserved sequence element. The same relationships were true for *JARID2/JMJD3* differentially retained introns between dragon/turtle and alligator/turtle (not shown).