

## Supplementary Figure 1. Snail1 and Twist1 non-cell autonomously increase invasion of HMLER-Ctrl cells without altering EMT markers

(a) Representative image of HMLER-Ctrl cells co-cultured with HMLER-Snail1/Twist1 cells at 1:1 and 10:1 ratio (day 11 of 14).

(b) Representative Western blot analyses of whole cell lysates (WCLs) from GFP<sup>+</sup> or tRFP<sup>+</sup> sorted from co-culture experiments at day 14. (HDAC1 – loading control), n≥3.

Lanes: (1) GFP<sup>+</sup>HMLER-Ctrl cells, (2) tRFP<sup>+</sup> HMLER-Snail1 cells, (3, 4) GFP<sup>+</sup>HMLER-Ctrl cells from (3) 1:1 and (4) 10:1 co-culture with HMLER-Snail1 cells, (5) tRFP<sup>+</sup> HMLER-Twist1 cells, (6, 7) GFP<sup>+</sup>HMLER-Ctrl cells from (6) 1:1 and (7) 10:1 co-culture with HMLER-Twist1 cells.

(c,e) Representative 18hr invasion assays of HMLER-Snail1/Twist1 and HMLER-Ctrl cells in indicated CM.

 $(\mathbf{d},\mathbf{f})$  Quantitation of invasion assays in panel  $\mathbf{c}$  and  $\mathbf{e}$ .

(g,h) Representative proliferation assay by MTS (g) and CellTiter-Glo (h) for HMLER-Ctrl cells in indicated CM. Scale bar-100 $\mu$ m; S.E.M shown, One way ANOVA with Tukey post test in d,f; n≥6; RM Two-way ANOVA in g,h; ns- not significant, \*\*p value<0.001, \*\*\*p value<0.0001



### Supplementary Figure 2. Six1 is upregulated downstream of Snail1 and Twist1 in HMLER cells and only Six1 is overexpressed in MCF7 cells

(a) qRT-PCR analyses of HMLER-Ctrl, Snail1 and Twist1 cells show that Six1 is upregulated downstream of Snail1 and Twist1 and is knocked down in cells transfected with 150nM of siSix1 compared to a non-targeting siRNA pool (siNT).
 (b,c) Representative 18hr invasion assay of HMLER-Ctrl cells in CM from HMLER-Snail1 or Twist1 cells +/- siSix1.
 (d,e) Quantitation of cell invasion in panel b and c respectively.

(f) qRT-PCR analyses of Six1 levels in HMLER-Ctrl cells cultured in CM from HMLER-Ctrl or HMLER-Snail1/Twist1 +/siSix1.

(g-i) qRT-PCR analyses of 3 clones each of MCF7-Ctrl and MCF7-Six1 cells (combined data shown for each group). Gene expression normalized to 18s, GAPDH or PP1B.

(j) 7hr migration assay of HMLER-Ctrl cells in CM from MCF7-Ctrl and Six1 cells (combined data shown for >3 experiments).

S.E.M shown, One way ANOVA with Tukey post test on compiled experiments n≥3 in **a,d-f;** two tailed unpaired t-test in **g-j**, n=3; ns- not significant, \*p<0.05, \*\*p<0.001, \*\*\*p<0.001



### Supplementary Figure 3. Hh ligand levels are differentially regulated downstream of Snail1, Twist1 and Six1

(**a**,**b**) qRT-PCR analyses in HMLER-Ctrl, Snail1 and Twist1 cells transfected with 150nM of siNT or siSix1. (**c**,**d**) qRT-PCR analyses in combination of 3 clones each of MCF7-Ctrl and MCF7-Six1 cells.

(e) qRT-PCR analyses in A2780 cells transfected with 100nM of siNT or siSix1.

(f) qRT-PCR analyses in HMLER-Ctrl cells transfected with 1µg of Six1 and/or Eya2. EV-Empty vector.

(g) Western blot analyses of whole cell lysates from HMLER and MCF7 cells showing endogenous levels of Eya2.

Gene expression normalized to PP1B. S.E.M shown, compiled experiments n≥3. One way ANOVA with Tukey post test in (**a,b,f**), two tailed unpaired t-test in (**c-e**). ns – not significant; \*p value<0.05, \*\*p<0.001, \*\*\*p<0.0001



#### Supplementary Figure 4. Hh/Gli signaling is specifically activated in the presence of CM from cells expressing EMT-TFs

(a,b) mutant (m)-Gli1-GFP reporter assay in HMLER cells cultured in CM from HMLER-Ctrl and (a) HMLER-Snail1+/siSix1 cells or (b) HMLER-Twist1+/-siSix1 cells.

(c) qRT-PCR analyses of Hh pathway target genes in HMLER-Ctrl cells in different CM; gene expression normalized to PP1B.

(d) m-Gli1-GFP reporter assay in MCF7-Ctrl cells cultured in MCF7-Ctrl/Six1 CM.

(e) 7-Gli1 and m-Gli1-GFP reporter assays in MCF7-Ctrl cells with addition of 1µg/ml of rhSHH to media.

(f,g) m-Gli1-GFP reporter assay in (f) HMLER cells in CM from HMLER-Ctrl, Snail1 or Twist1 cells, and in (g) MCF7-Ctrl cells in MCF7-Ctrl/Six1 CM treated with indicated drug/antibodies.

All Gli1-GFP reporter assays are represented as %GFP<sup>+</sup> cells.

Veh

Cyclo.

61

S.E.M shown, compiled experiments n≥2 (c),n=3 (with different sets of CM). One way ANOVA with Tukey post test in all cases. ns - not significant; \*p value<0.05, \*\*\*p value<0.0001







MCF7 cells







## Supplementary Figure 5. Hh pathway inhibition via upstream vs downstream inhibitors differentially regulates non-cell autonomous phenotypes dependent on context.

(a,b) 16-18hr invasion assays of HMLER-Ctrl cells cultured in HMLER-Snail1/Twist1 CM treated with (a) cyclopamine or (b) GANT61 or corresponding vehicle control.

(c) Representative ICC of E-cadherin (green) in MCF7-Ctrl cells cultured in CM from MCF7-Ctrl and Six1 cells, treated with 5E1 and control NS-1 antibody for 48hrs (Dapi, blue), scale bar – 20µm.

(d) Quantitation of % membranous E-cadherin in panel c, cell counted per condition  $\geq$  45-60.

(e) Anoikis resistance assay in MCF7-Ctrl cells cultured in indicated CM +/- cyclopamine or 5E1 for 24hrs.

S.E.M shown, n≥3. One way ANOVA with Tukey post test in all cases. ns – not significant; \*p<0.05, \*\*p<0.001, \*\*\*p<0.0001



# Supplementary Figure 6. Six1-expressing EMT cells benefit from inter-clonal cooperation with non-EMT non-Six1 expressing cells.

Weeks

(a) Representative fluorescent images of NOG/SCID mice at same time point (corresponding to similar tumor volumes) with MCF7-Six1-tRFP "singly" injected tumors and GANT61 or vehicle treated mixed injection groups.
(b) Quantitation of red fluorescent signal (EMT cells) from primary tumor and distant sites (in lymph nodes/lungs - yellow boxed region in a representing MCF7-Six1 cells) in MCF7-Six1-tRFP and mixed tumor groups represented as c/s, counts per second.

(c,e) Quantitation of fluorescent signal (representing MCF7-Six1 EMT cells) from (c) distant sites and (e) primary tumor in mixed tumors groups treated with vehicle or GANT61, represented as fold change over pre-treatment signal.

(d,f) Normalized fluorescent signal (MCF7-Six1 cells) from (d) distant sites and (f) primary tumors of individual mice pre- and post-treatment.

(g) Overall tumor volumes of mice that received vehicle or GANT61 treatment over the course of the experiment, measured by calipers.

S.E.M shown, two tailed unpaired t-test for **a-f**; Two-way ANOVA followed by Bonferroni post test for **g**; ns- not significant; \*p value<0.05, \*\*p value<0.001



Supplementary Figure 7. Uncropped blots for western blots analyses in the main figures



Supplementary Fig. 7 continued



B – MCF7-Ctrl clones A – MCF7-Six1 clones



Blot is reversed in paper





B - MCF7-Ctrl A - MCF7-Six1 B+B - Ctrl cells in Ctrl CM B+A - Ctrl cells in Six1 CM A+A - Six1 cells in Six1 CM Cyc - cyclopamine Ab - 5E1





B – MCF7-Ctrl A – MCF7-Six1 Blot is reversed in paper



-150

### Supplementary Fig 1b.





G- HMLER-Ctrl S – HMLER-Snail1 T- HMLER-Twist1 1:1, 1:10 – ratio of G to S/T cells





Blot is reversed in paper

Supplementary Fig 3g.



Blot is reversed in paper

#### Supplementary Table 1.

Combination	Prognosis	Subtype/Grade	p value
Six1, Gli1	DMFS	Luminal A	0.0121
	DMFS	ER+ LN negative tumors	0.01923
	DMFS	LN negative tumors	0.00919
	DMFS mixed	PAM50-Luminal A	0.03606
	DMFS mixed	LN negative	0.00968
	DMFS mixed	ER+ LN negative tumors	0.01692
	DMFS mixed	Luminal B	0.02931
	RFS	Grade 3	0.04818
Snai1, Gli1	DMFS mixed	Grade 1	0.01909
	DMFS mixed	Luminal A	0.02437
	DMFS mixed	PAM50-Luminal A	0.0384
	RFS	Grade 1	0.01812
	RFS	-	0.041
Twist1, Gli1	DMFS	Basal	0.01376
	DMFS mixed	Basal	0.0268
	RFS	LN negative	0.05203

High expression of only both EMT-TFs *and GLI1* correlate with worsened prognosis across multiple breast subtypes and grades, while high expression of either the EMT-TF or *GLI1* alone does not. DMFS – Distant metastasis-free survival, RFS – Relapse-free survival, DMFS mixed – DMFS mixed with RFS data. Data obtained from GOBO or KM Plotter (in red).

#### Supplementary Table 2. Primer sequences for qRT-PCR analyses

Primer	Forward Primer (5'-3')	Reverse Primer (5'-3')
Six1	TGCGCCGAAAATTTCCA	TTGAAGCAGTAGCTGGTCTCC
Twist1	GGAGTCCGCAGTCTTACGAG	TCTGGAGGACCTGGTAGAGG
Snai1	GAAAGGCCTTCAACTGCAAA	TGACATCTGAGTGGGTCTGG
Gli1	GATGACCCCACCACCAATCA	AGAAAAGAGTGGGCCCTCGG
Ptch1	CTGGCTTCAGGGACTTCAGG	CGGTTTGCACCAGGAGTTTG
Ptch2	CCAGAGATCCTGAGTCCACC	GGCTGGATGGATGTAGGCAC
VEGF-A	AAGAAAATCCCTGTGGGCCTT	TTTCCTGGTGAGAGATCTGCA
Bcl-2	GGAGGCTGGGATGCCTTTGT	TTCACTTGTGGCCCAGATAGG
BMP-4	ACCGAATGCTGATGGTCGTT	CAGAAGTGTCGCCTCGAAGT
FoxF1	TCTCGCTCAACGAGTGCTTC	GTTCATCATGCTGTACATGGGC
CyclinD1	CTGCTCCTGGTGAACAAGC	TGTGGCACAGAGGGCAAC
MTSS1	AAGAACGTGGCCGATTCTGT	TGGACTTTCTGGACATGGTGG
SHH	GAAGAGGAGGCACCCCAAAA	CCCTTCATACCTTCCGCTGG
ІНН	CCGCGACCGCAATAAGTATG	CGAGTGCTCGGACTTGACG
DHH	ACATCACTACGTCTGACCGC	ACCGCCAGTGAGTTATCAGC
GAPDH	CATCACCATCTTCCAGGAGC	ATGCCAGTGAGCTTCCCGTC
PP1B	GGAGATGGCACAGGAGGAAA	CGTAGTGCTTCAGTTTGAAGT
18S	GTAACCCGTTGAACCCCATT	CCATCCAATCGGTAGTAGCG

#### Supplementary Table 3. Antibodies used for Western blot analyses and ICC

Antibody	Company	Catalog Number	Dilution Used
SHH	Cell signaling	2207	1:1000 – WB
E-Cadherin	BD Biosciences	610181	1:2500 – WB 1:300 – ICC
E-Cadherin	Cell signaling	3195S	1:300 – ICC
Vimentin	BD Biosciences	550513	1:2500 – WB
Cytokeratin-18	Abcam	ab82254	1:10,000 – WB
Fibronectin	BD Biosciences	610077	1:500 – WB
Eya2	Sigma-Aldrich	HPA027024	1:1000
HDAC1	Santa Cruz	sc-7872	1:1000 – WB
β-tubulin	Sigma-Aldrich	T8328	1:5000 – WB
GAPDH	Cell signaling	2118S	1:1000 – WB
AlexaFluor 488 α- rabbit IgG	Invitrogen	A11008	1:300 – ICC
AlexaFluor 594 α- mouse IgG	Invitrogen	A11005	1:300 – ICC
HRP-anti-mouse IgG	Sigma-Aldrich	A9044	1:10,000 – WB
HRP-anti-rabbit IgG	Sigma-Aldrich	A9169	1:10,000 – WB

WB – Western blot analysis, ICC - Immunocytochemistry