1	Role of methionine adenosyltransferase 2A in bovine preimplantation development and its
2	associated genomic regions
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11 Supplementary figure legends

Figure S1. Nuclear localization of MAT2A in bovine blastocyst cells. (A) Immunofluorescence image of MAT2A protein in blastocyst cells. Nuclei showing more abundant MAT2A compared with the cytoplasm are indicated by arrowheads. (B) Nuclear counterstaining with propidium iodide. (C) Merged image of (A) and (B). Scale bar represents 50 µm.

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17 Figure S2. Identification of MAT2A-ChIP specific peaks in duplicated ChIP-seq (Rep 1 and Rep 2).

- (A) 39 peaks (intersection) are specific for MAT2A-ChIP samples against Input in both replicates.
 (B) 59 peaks (intersection) are specific for MAT2A-ChIP samples against IgG control in both replicates. (C) The union of these peaks makes 76 MAT2A-ChIP specific peaks with 22 overlapping
- 21 peaks.
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23Figure S3. Hierarchical tree graph of overrepresented GO terms in biological process generated by 24Singular Enrichment Analysis (SEA) of agriGO (http://bioinfo.cau.edu.cn/agriGO/). The first line in 25each significant box represents the GO term ID with the adjusted P-value in parentheses, followed 26by the lines indicating the term definition. In the last line of the significant box, the first pair of 27numerals represents the number of genes associated with the GO term and that of all genes in the 28query list and the second pair of numerals represents those in the Bos taurus database. The degree of 29colour saturation of the boxes is positively correlated with the enrichment level of the term: yellow, 30 <0.05; orange, <5e-03 - <5e-09; and red, <5e-10. Solid, dashed, and dotted lines represent two, one and zero enriched terms at both ends connected by the line, respectively. 31

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Figure S1



Figure S2



Figure S3

Table S3. The primer pairs used for validation of ChIP-seq peaks.

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Peak ID	Primer sequences		
Dook 168	Forward	GGGTAGTTCCCCAAAGGCTG	
reak 400	Reverse	CAAGGGCTGGGGCATGTAAT	
Deak 1939	Forward	CAGCAGTCAGGCACCACTC	
reak 1252	Reverse	CTGCCCAGCTTCTAGCGT	
Dook 1485	Forward	AAGTCAACACGAAGGGGCA	
reak 140J	Reverse	GGTTGAGGCATGGAACTCCG	
Deak 1630	Forward	TTTGGTCTCTCTGTGCAGCTT	
reak 1000	Reverse	CCTGCTGTCTTATGGAGGGC	
Deak 1723	Forward	TTCTGCCCTCTGACATGATTC	
	Reverse	CAGATTGGACTGAAAGGACCA	
Dook 2012	Forward	TAAGCACCTGGTGAACTGAG	
reak 2045	Reverse	TATTACGTGGCGTGTCAGG	
CAPDH promotor *	Forward	TGTTATATCCTTGCGGCAGCTT	
	Reverse	AGCACTGCGGGAGAGTAGTAAACT	
* Herrmann D et al., Epigenetics 2013 8(3): 281-289 doi:10.4161/epi.23899			