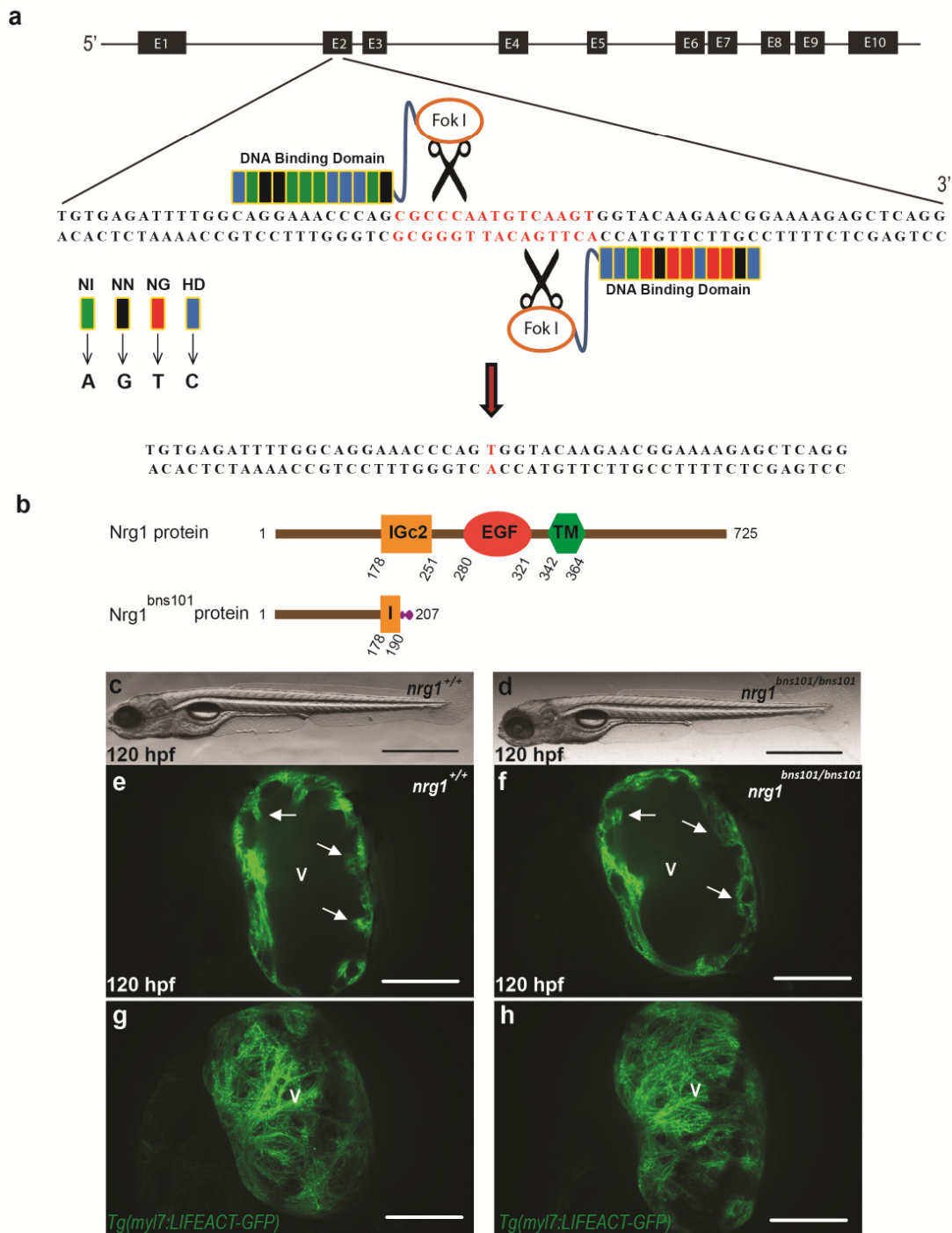


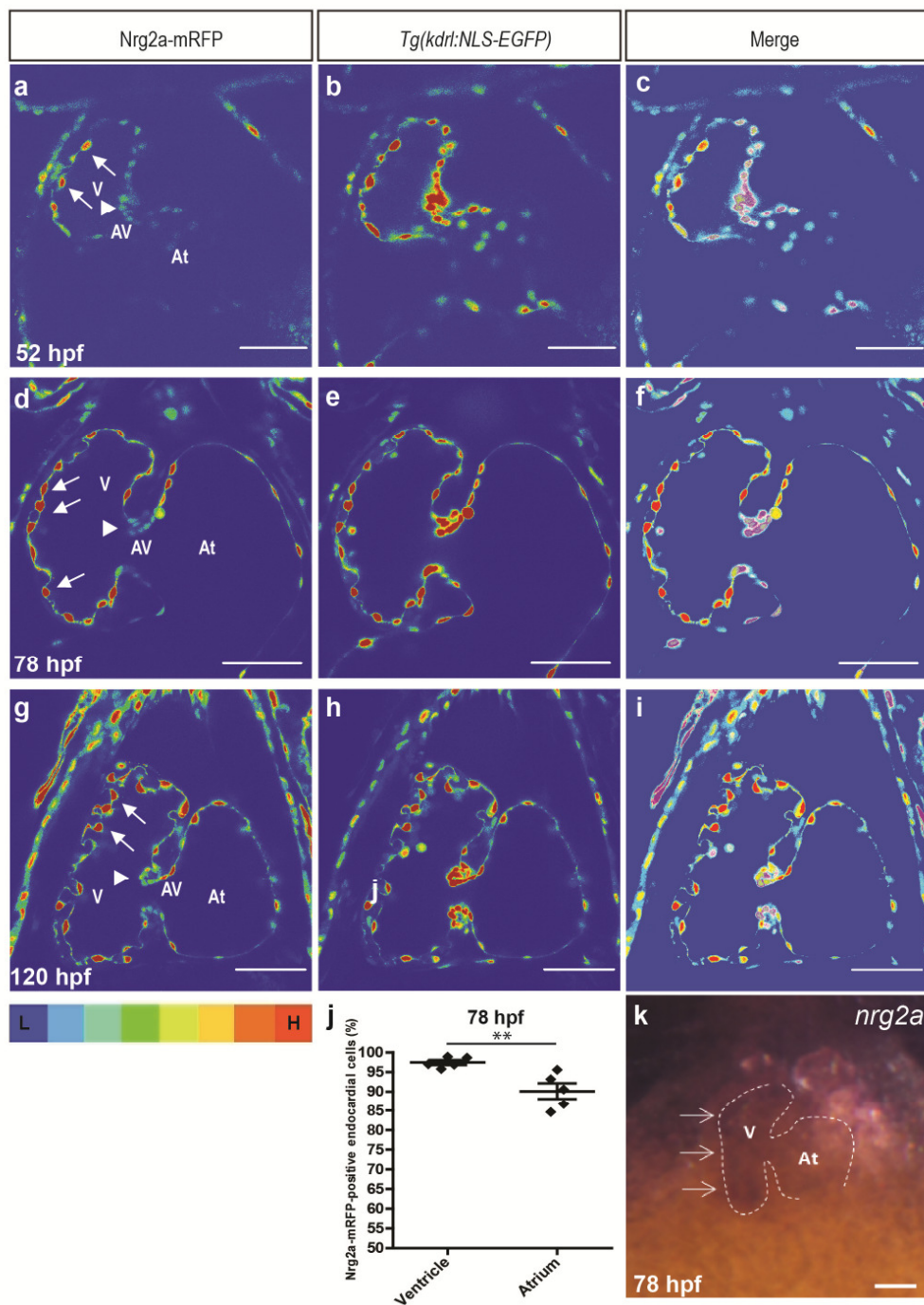
Supplementary Figures



Supplementary Figure 1. *nrg1*^{bns101/bns101} embryos develop a functional heart and survive to adulthood

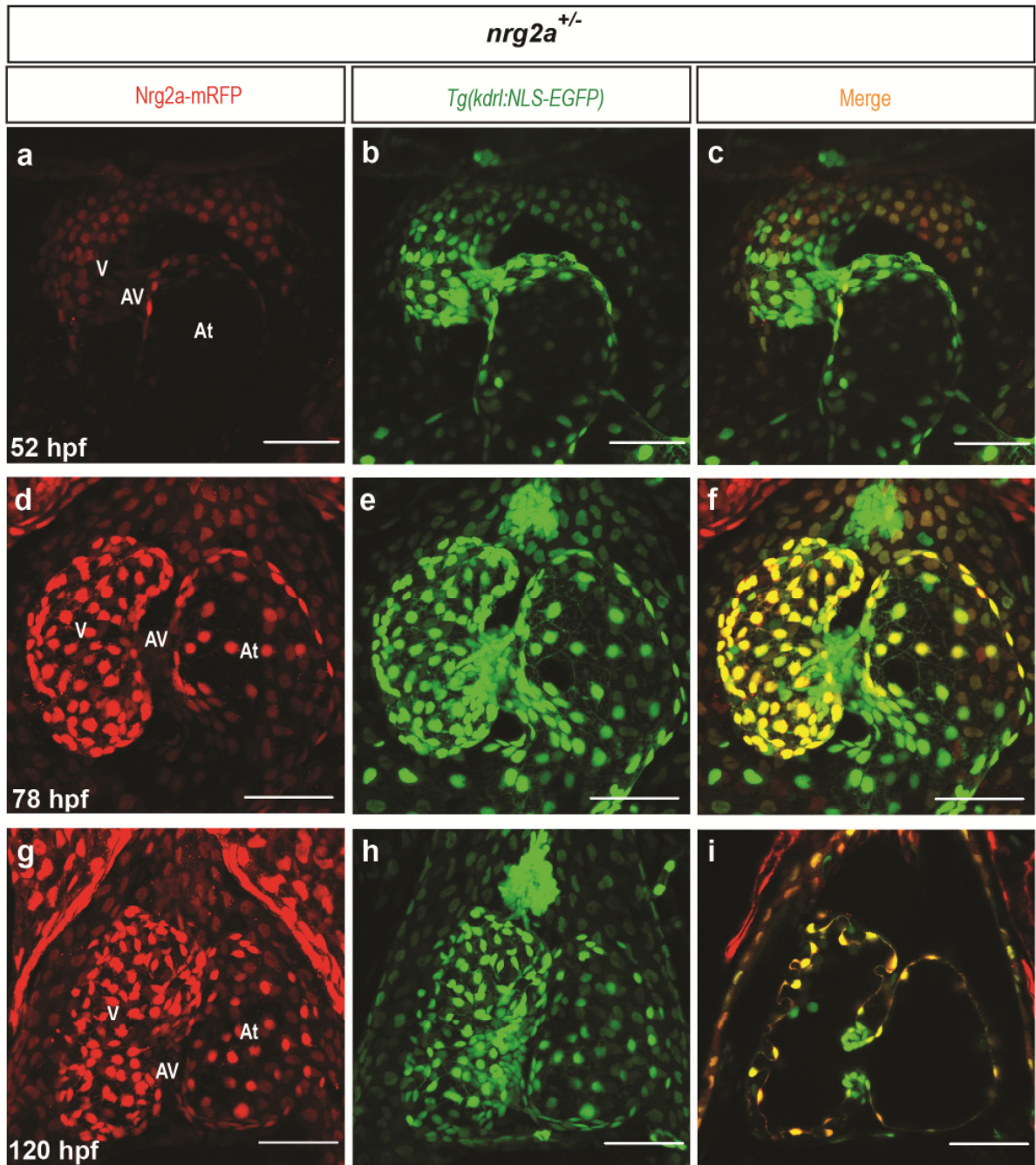
(a-b) Cartoon of Talen-induced *nrg1* mutation with a 14-base-pair deletion in exon 2 (a); this deletion causes a frame shift leading to a premature stop codon in exon 2 resulting in a

predicted truncated protein **(b)**. Notably, the predicted truncated Nrg1 protein in *nrg1^{bns101/bns101}* animals lacks the Ig-like, EGF-like and TM domains that have all been reported to be essential for mouse cardiac trabeculation ^{1,2,3}. If the first ATG downstream of the 14 bp deletion is used to reinitiate translation, a protein consisting of part of the TM domain and the carboxy terminal domain will be made. However, the type IIIa Nrg1 isoform ⁴ is not predicted to be affected by the *bns101* mutation. **(c-d)** *nrg1^{bns101/bns101}* larvae do not exhibit any gross morphological defects as compared to wild-type; lateral views, anterior to the left; scale bars, 0.5 mm. **(e-h)** Trabeculation appears unaffected in *nrg1^{bns101}* mutants. 120 hpf larvae from *Tg(myI7:LIFEACT-GFP); nrg1^{bns101/+}* incrosses imaged by spinning disk confocal microscopy; 2D views (mid-sagittal sections) **(e and f)**, and 3D maximum intensity z-projections **(g and h)**; V: ventricle; arrows point to trabecular cardiomyocytes; scale bars, 50 μ m.



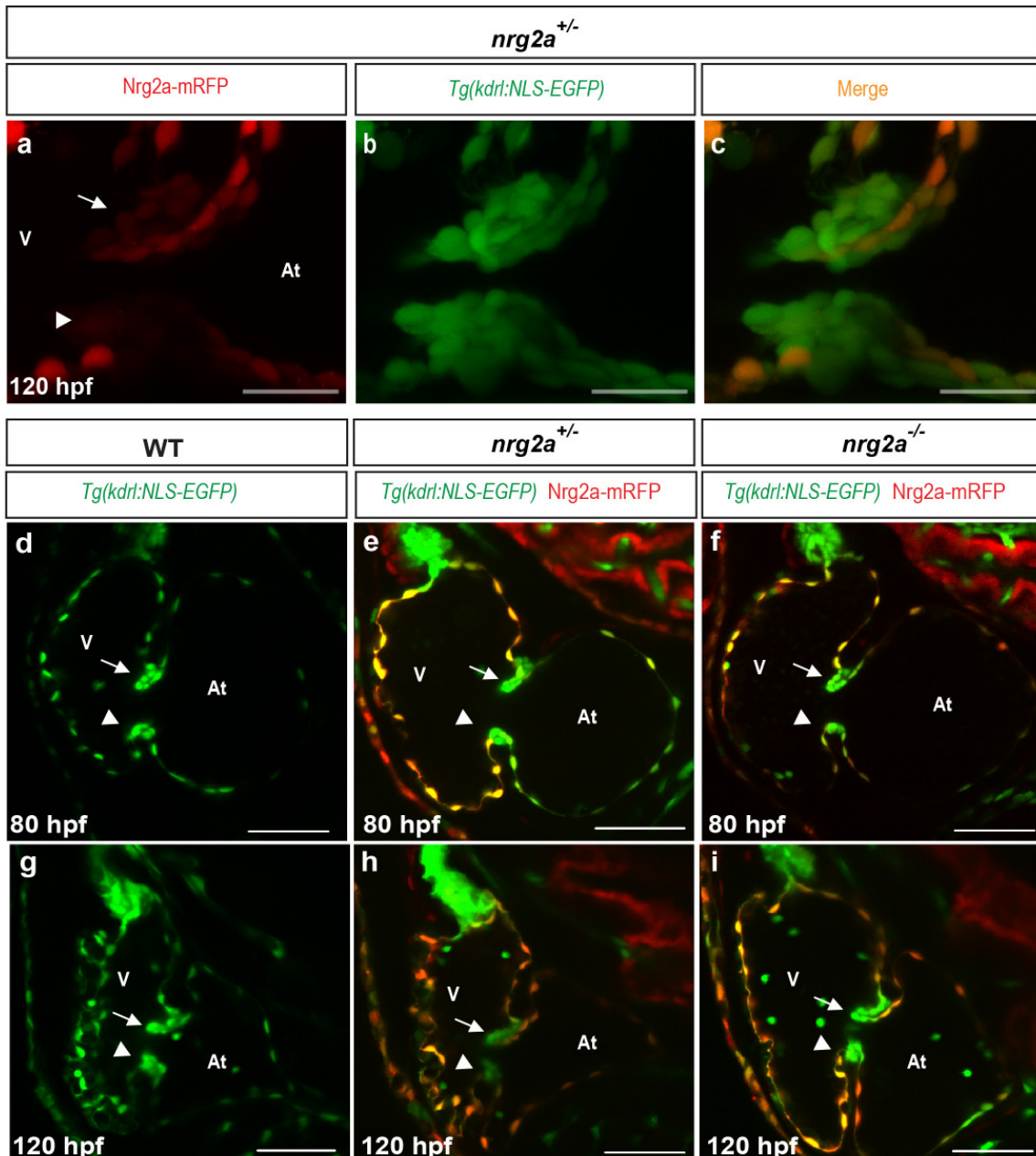
Supplementary Figure 2. Cardiac Nrg2a-mRFP expression during embryonic and larval development

(a-i) Mid-sagittal confocal images of *Tg(kdrl:NLS-EGFP);nrg2a^{+/-}* hearts; heat map; low intensity (blue) to high intensity (red) at 52 (a-c), 78 (d-f) and 120 (g-i) hpf; arrows and arrowheads point to endocardial cells in ventricular outer curvature and superior AV valve leaflet, respectively. (j) Graph showing that on average there is a higher percentage of endocardial cells positive for Nrg2a-mRFP in the ventricle than in the atrium at 78 hpf; dots in this graph represent individual hearts; N=5 hearts; values represent means \pm SEM; ** $P \leq 0.01$ by Student's *t*-test. (k) *In situ* hybridization for *nrg2a* expression in 78 hpf heart. V: ventricle, AV: atrioventricular canal, At: atrium; scale bars, 50 μ m.



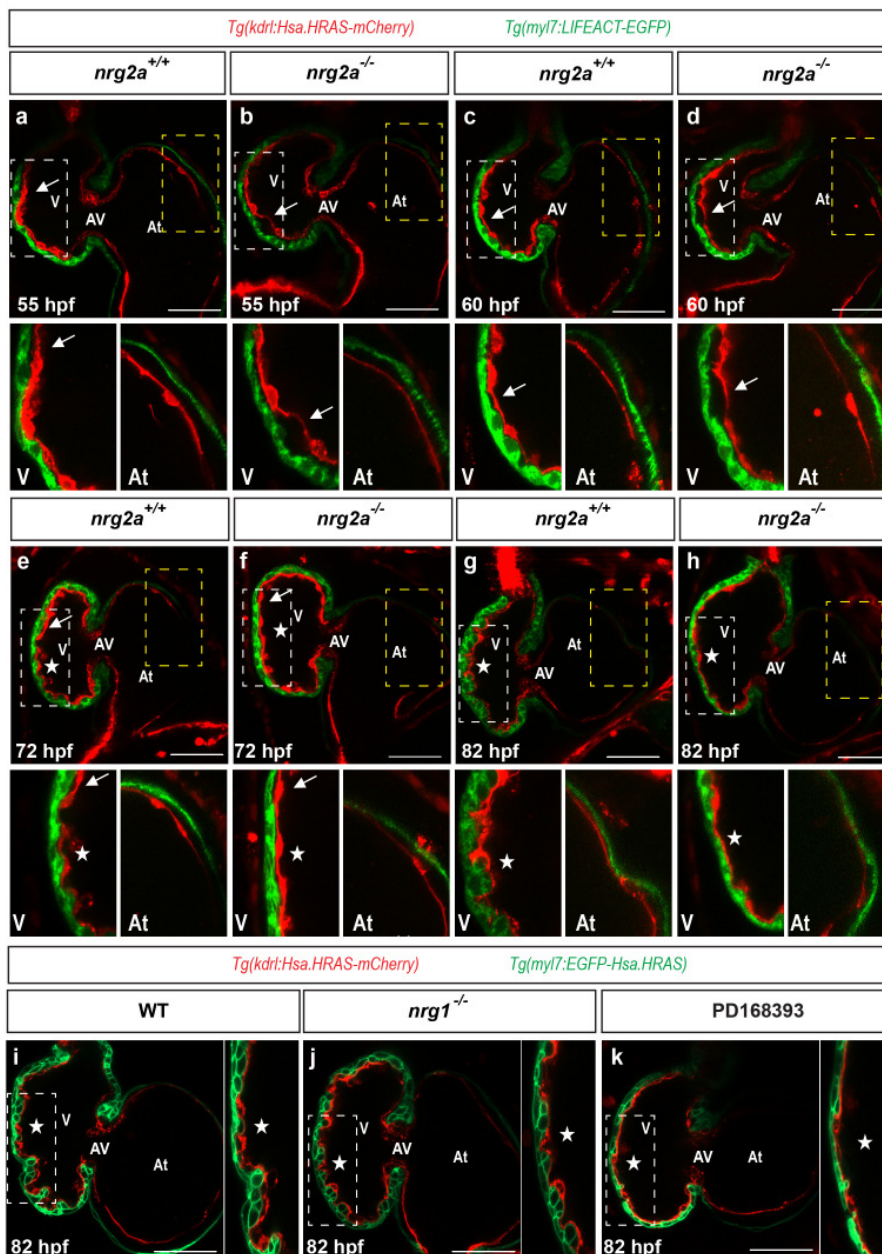
Supplementary Figure 3. Spatiotemporal pattern of cardiac Nrg2a-mRFP expression during embryonic and larval development

(a-i) 3D views (maximum intensity z-projections) of *Tg(kdrl:NLS-EGFP);nrg2a*^{+/-} hearts at 52 (a-c), 78 (d-f) and 120 (g-i) hpf; Nrg2a-mRFP expression appears stronger in the ventricle than in the atrium or AV canal; V: ventricle, AV: atrioventricular canal, At: atrium; scale bars, 50 μ m.



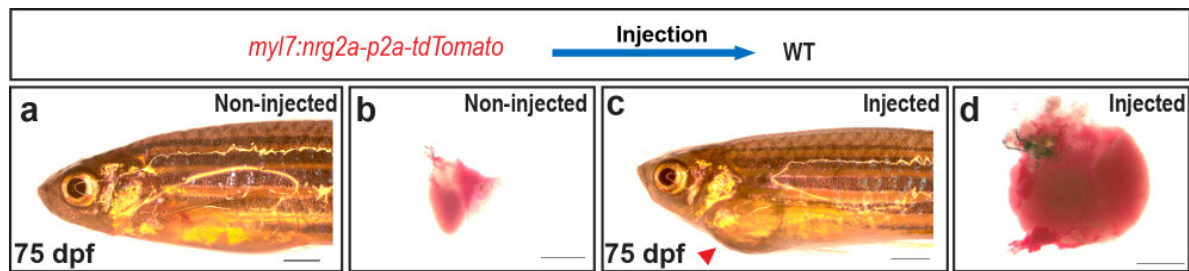
Supplementary Figure 4. *nrg2a*^{-/-} hearts develop functional atrioventricular valves

(a-c) Maximum intensity z-projections (25 z-stacks, sagittal sections) of 120 hpf *Tg(kdrl:NLS-EGFP);nrg2a*^{+/-} AV canal showing that Nrg2a-mRFP is weakly expressed in the AV valve leaflets. **(d-i)** Mid-sagittal confocal images of hearts from *Tg(kdrl:NLS-EGFP);nrg2a*^{+/-} incrosses at 80 **(d-f)** and 120 **(g-i)** hpf; arrows and arrowheads point to superior and inferior valve leaflets, respectively; V: ventricle, AV: atrioventricular canal, At: atrium; scale bars, 50 μm.



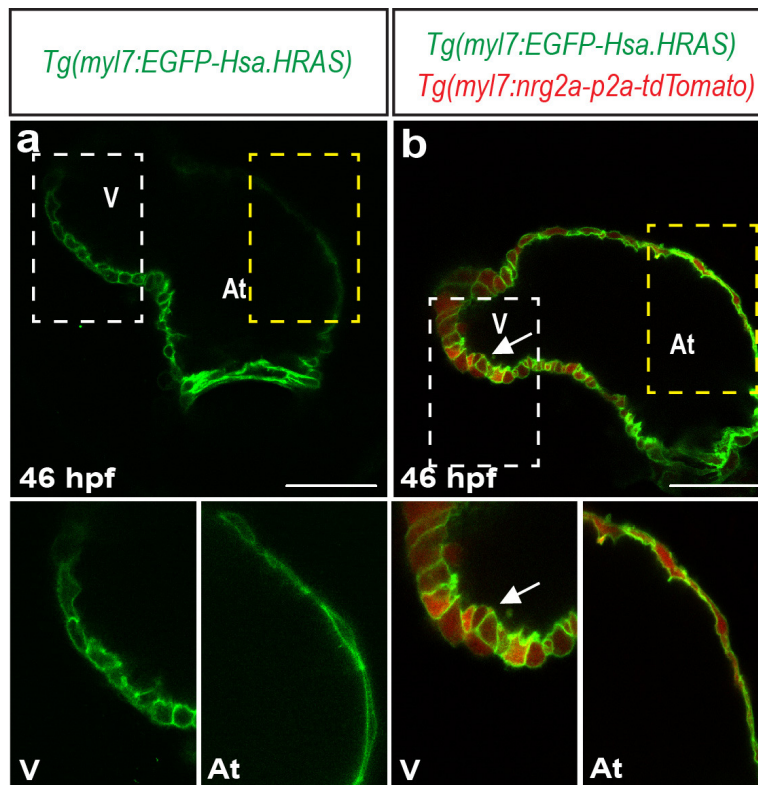
Supplementary Figure 5. Developmental analysis of cardiac jelly thickness in the presence or absence of Nrg/ErbB2 signaling

(a-h) Mid-sagittal views of wild-type and *nrg2a*^{-/-} hearts from *Tg(kdrl:Hsa.HRAS-mCherry);Tg(myI7:LIFEACT-GFP);nrg2a*^{+/+} incrosses at 55 **(a-b)**, 60 **(c-d)**, 72 **(e-f)** and 80 **(g-h)** hpf; the myocardium and endocardium are labeled in green and red, respectively; magnified images of dashed boxes are shown below each time point. The thickness of the cardiac jelly progressively diminishes in wild-type and *nrg2a*^{-/-} hearts. **(i-k)** Confocal sagittal sections of 82 hpf *Tg(kdrl:Hsa.HRAS-mCherry);Tg(myI7:EGFP-Hsa.HRAS)* hearts from wild-type **(i)**, *nrg1*^{-/-} mutant **(j)** or ErbB2 inhibitor (PD168363) treated animals showing that cardiac jelly reduction also appears to be an Nrg1/ErbB2 independent process; magnified views of dashed boxes in **i-h** are shown on the right; arrows and asterisks indicate the presence and absence of cardiac jelly, respectively; V: ventricle, AV: atrioventricular canal, At: atrium; scale bars, 50 μm.



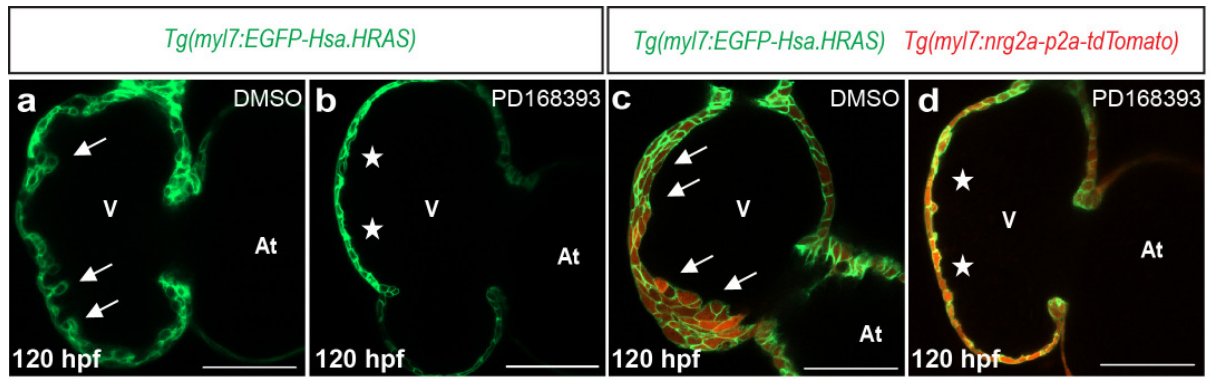
Supplementary Figure 6. Myocardial *nrg2a* overexpression causes cardiomegaly in wild-type fish

(a-d) A number of injected F0 wild-type fish (6 out of 31) exhibited pericardial edema at juvenile and adult stages as compared to non-injected ones; lateral views of non-injected **(a)** and injected **(c)** zebrafish imaged by brightfield microscopy at 75 dpf; red arrowhead in **c** points to pericardial edema; scale bars, 1mm. 75 dpf extracted hearts from non-injected **(b)** and injected **(d)** fish; scale bars, 200 μ m.



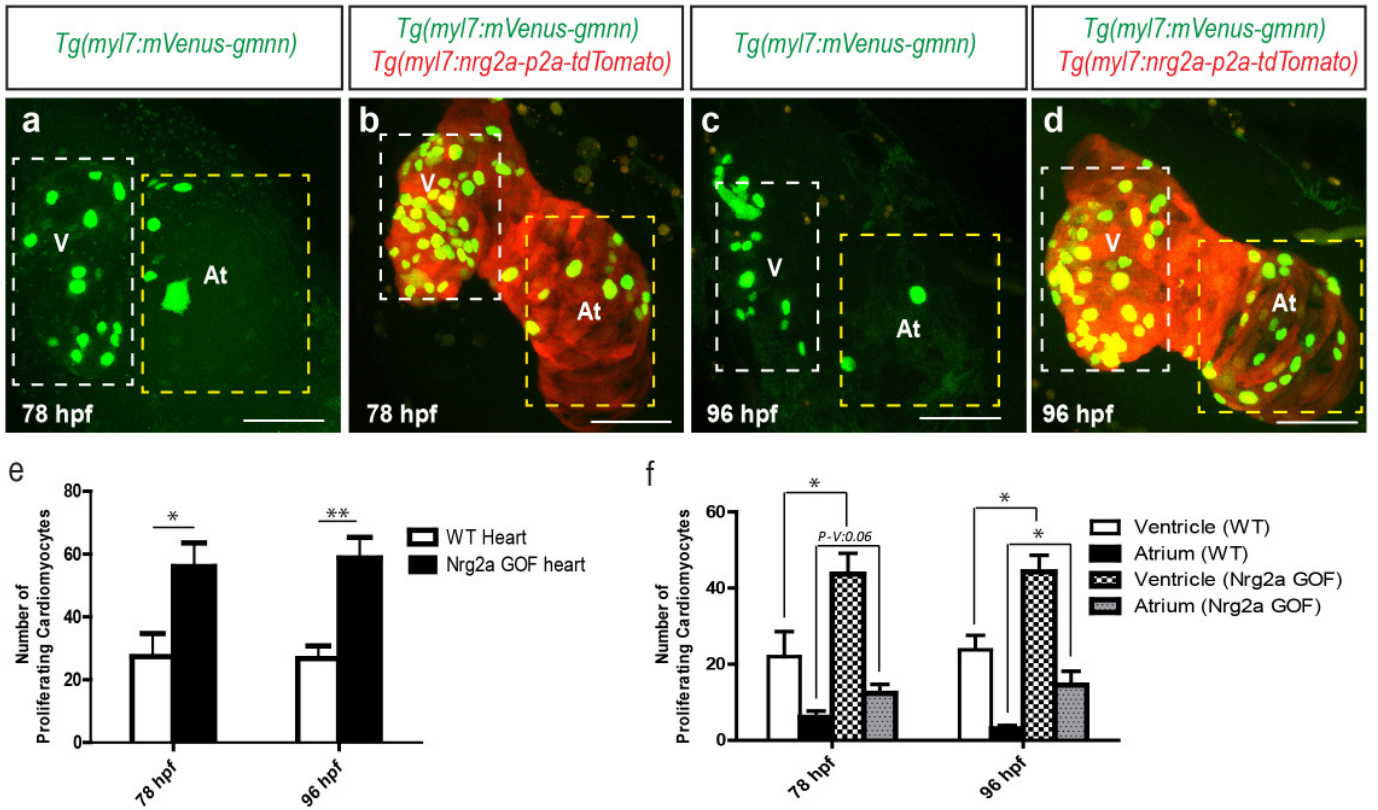
Supplementary Figure 7. Overexpression of *nrg2a* under control of a myocardial specific promoter induces cardiomyocyte bilayering in the ventricle by 46 hpf

(a-b) 2D ventral views (mid-sagittal sections) of 46 hpf *Tg(myI7:EGFP-Hsa.HRAS)* **(a)** or *Tg(myI7:EGFP-Hsa.HRAS);Tg(myI7:nrg2a-p2a-tdTomato)* **(b)** hearts; Magnified images of dashed boxes are shown below **a** and **b**; arrows point to bilayered wall; V: ventricle, At: atrium; scale bars, 50 μ m.



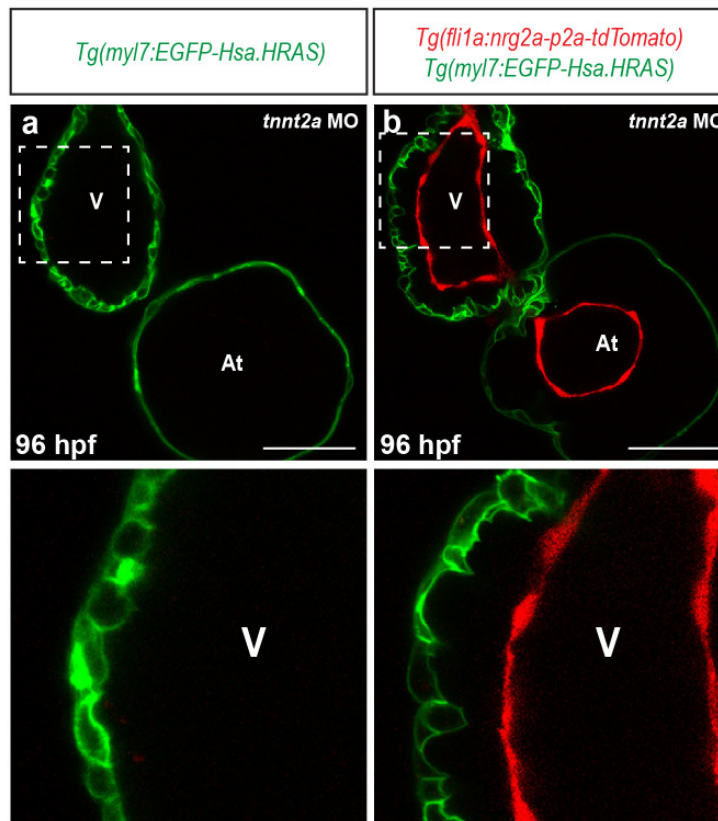
Supplementary Figure 8. Ectopic *nrg2a* overexpression in cardiomyocytes requires *ErbB2* function

(a-d) Confocal sagittal sections of *Tg(myI7:EGFP-Hsa.HRAS)* (**a-b**) or *Tg(myI7:EGFP-Hsa.HRAS);Tg(myI7:nrg2a-p2a-tdTomato)* (**c-d**) hearts, treated with DMSO (**a and c**) or the *ErbB2* inhibitor PD168363 (**b and d**) showing that *nrg2a* overexpression-induced cardiomyocyte multilayering requires *ErbB2* function; asterisks and arrows indicate single-layered and multilayered ventricular walls, respectively; V: ventricle, At: atrium; scale bars, 50 μm .



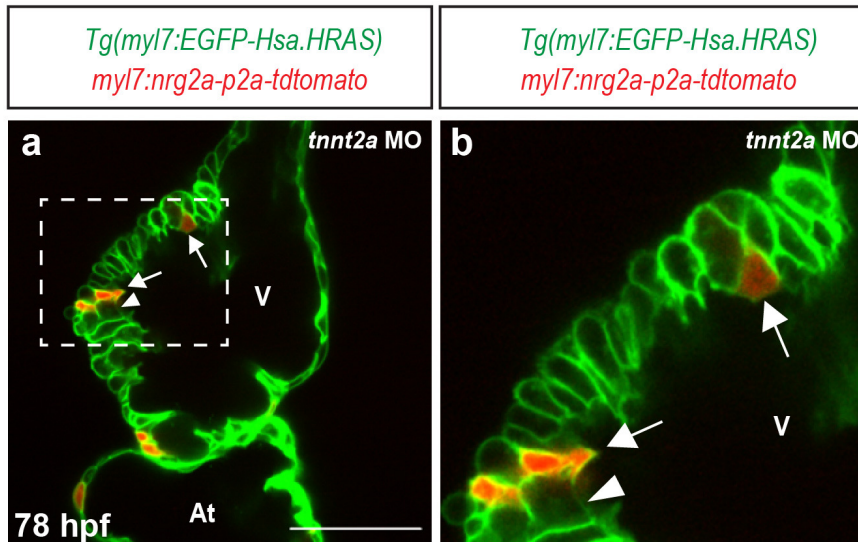
Supplementary Figure 9. Myocardial specific *nrg2a* overexpression leads to an increase in cardiomyocyte proliferation

(a-d) 3D views (maximum intensity z-projections) of *Tg(myl7:mVenus-gmnn)* (a and c) or *Tg(myl7:mVenus-gmnn);Tg(myl7:nrg2a-p2a-tdTomato)* (b and d) hearts at 78 (a-b) and 96 (c-d) hpf. Proliferating cardiomyocytes are labeled in green; white and yellow dashed boxes outline the ventricular and atrial chambers, respectively; V: ventricle, At: atrium; scale bars, 50 μ m. (e-f) Graphs showing the average number of proliferating cardiomyocytes per heart (e) and per ventricular or atrial (f) chamber at 78 and 96 hpf; N=9 hearts; values represent means \pm SEM; * P \leq 0.05,** P \leq 0.01 by Student's *t*-test.



Supplementary Figure 10. Endocardial specific overexpression of *nrg2a* does not induce cardiomyocyte multilayering in *tnnt2a* morphants

(a-b) 2D ventral views (mid-sagittal sections) of 96 hpf *Tg(myI7:EGFP-Hsa.HRAS)* (**a**) or *Tg(fli1a:nrg2a-p2a-tdTomato);Tg(myI7:EGFP-Hsa.HRAS)* (**b**) hearts from *tnnt2a* MO (0.5 ng) injected animals; magnified images of dashed boxes are shown below **a** and **b**; myocardium and endocardium are labeled in green and red, respectively; V: ventricle, At: atrium; scale bars, 50 μ m.



Supplementary Figure 11. Nrg2a can affect cardiomyocyte behavior via both autocrine and paracrine signals

(a-b) 2D ventral views (mid-sagittal sections) of 78 hpf *Tg(myI7:EGFP-Hsa.HRAS)* hearts from animals injected with *tnnt2a* MO (0.5 ng) and *myI7:nrg2a-p2a-tdTomato* DNA (5-10 pg). This low amount of DNA allows one to get single cardiomyocytes to overexpress the *nrg2a* transgene; arrows and arrowhead point to *nrg2a-p2a-tdTomato* expressing and non-expressing cardiomyocytes, respectively; higher magnification of dashed box in **a** is shown in **b**; V: ventricle, At: atrium; scale bar, 50 μ m.

Supplementary References

1. Erickson SL, *et al.* ErbB3 is required for normal cerebellar and cardiac development: a comparison with ErbB2-and heregulin-deficient mice. *Development* **124**, 4999-5011 (1997).
2. Kramer R, Bucay N, Kane DJ, Martin LE, Tarpley JE, Theill LE. Neuregulins with an Ig-like domain are essential for mouse myocardial and neuronal development. *Proceedings of the National Academy of Sciences* **93**, 4833-4838 (1996).
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4. Honjo Y, Kniss J, Eisen JS. Neuregulin-mediated ErbB3 signaling is required for formation of zebrafish dorsal root ganglion neurons. *Development* **135**, 2615-2625 (2008).