

Supplementary Information

A subset of cerebrovascular pericytes originates from mature macrophages in the very early phase of vascular development in CNS

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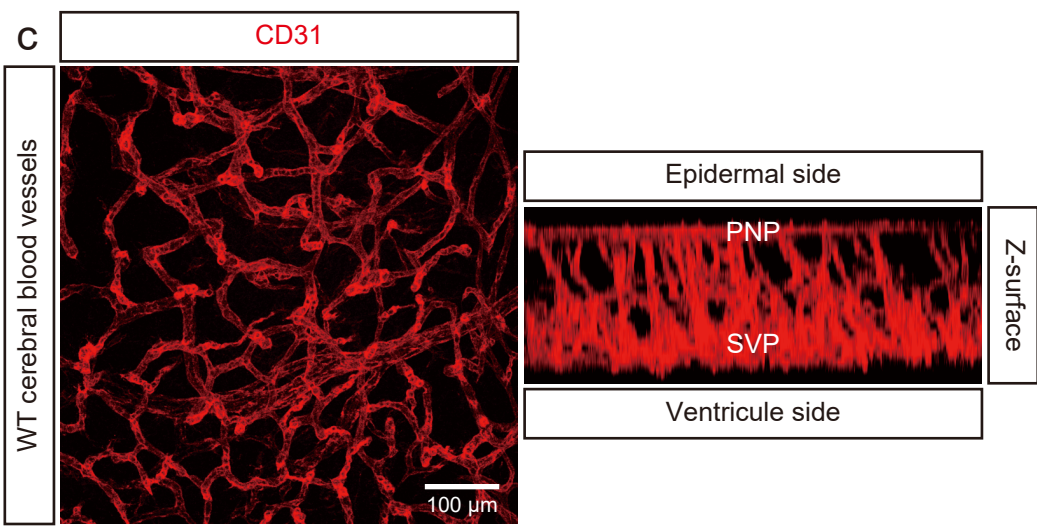
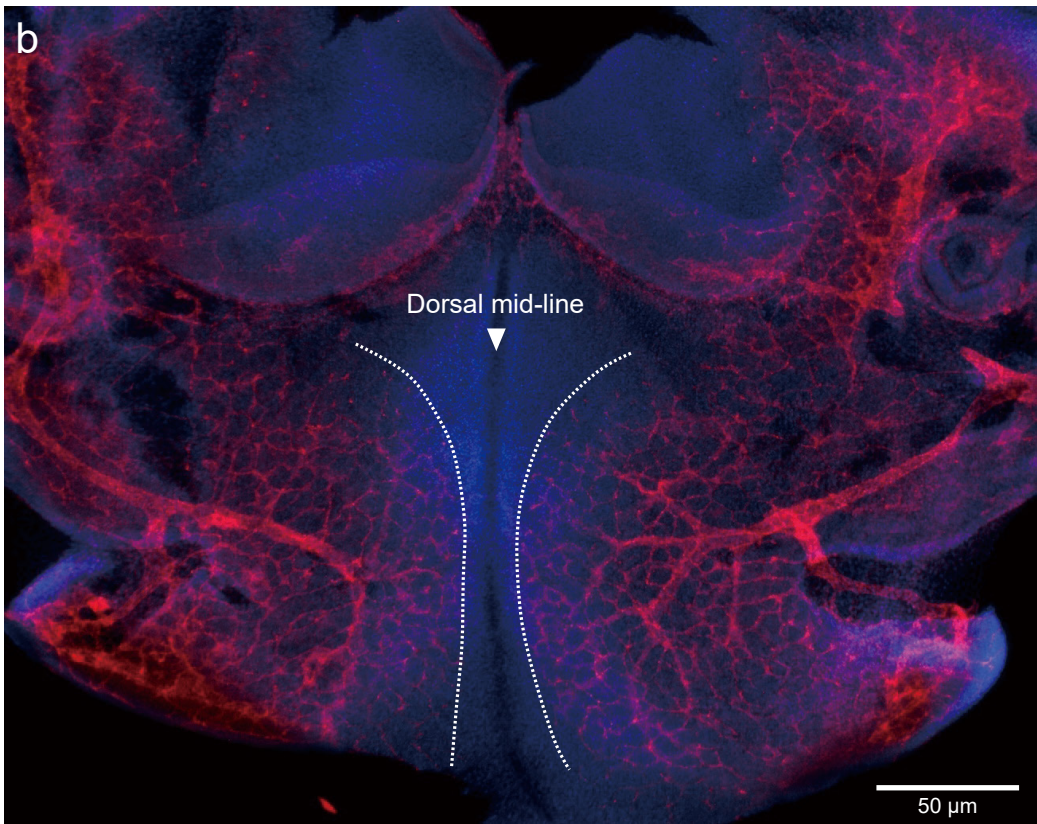
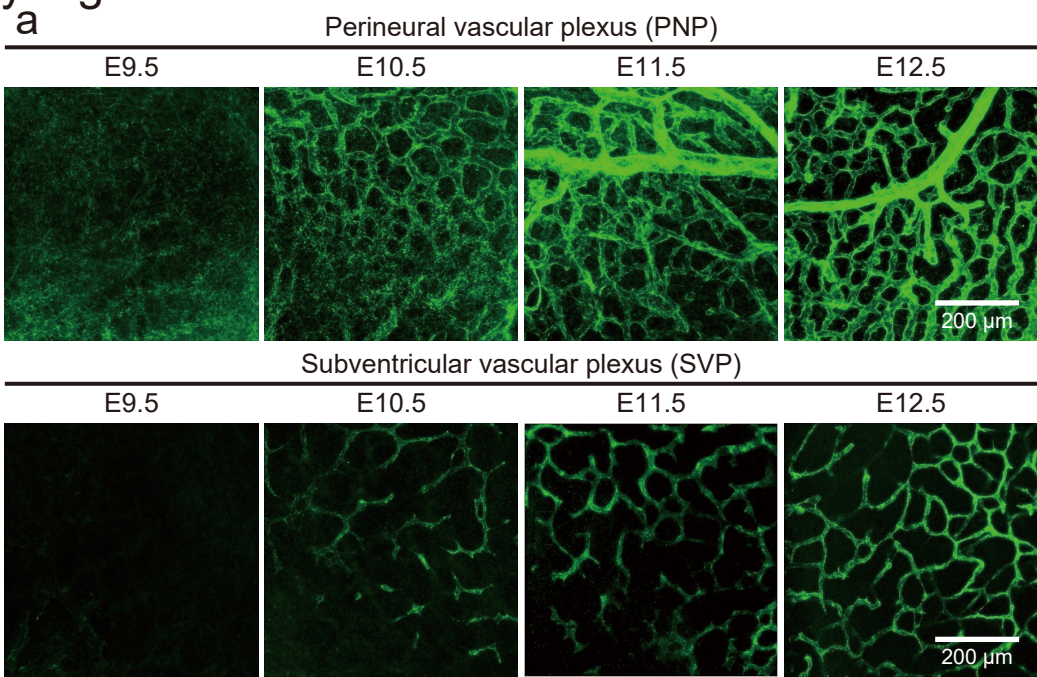
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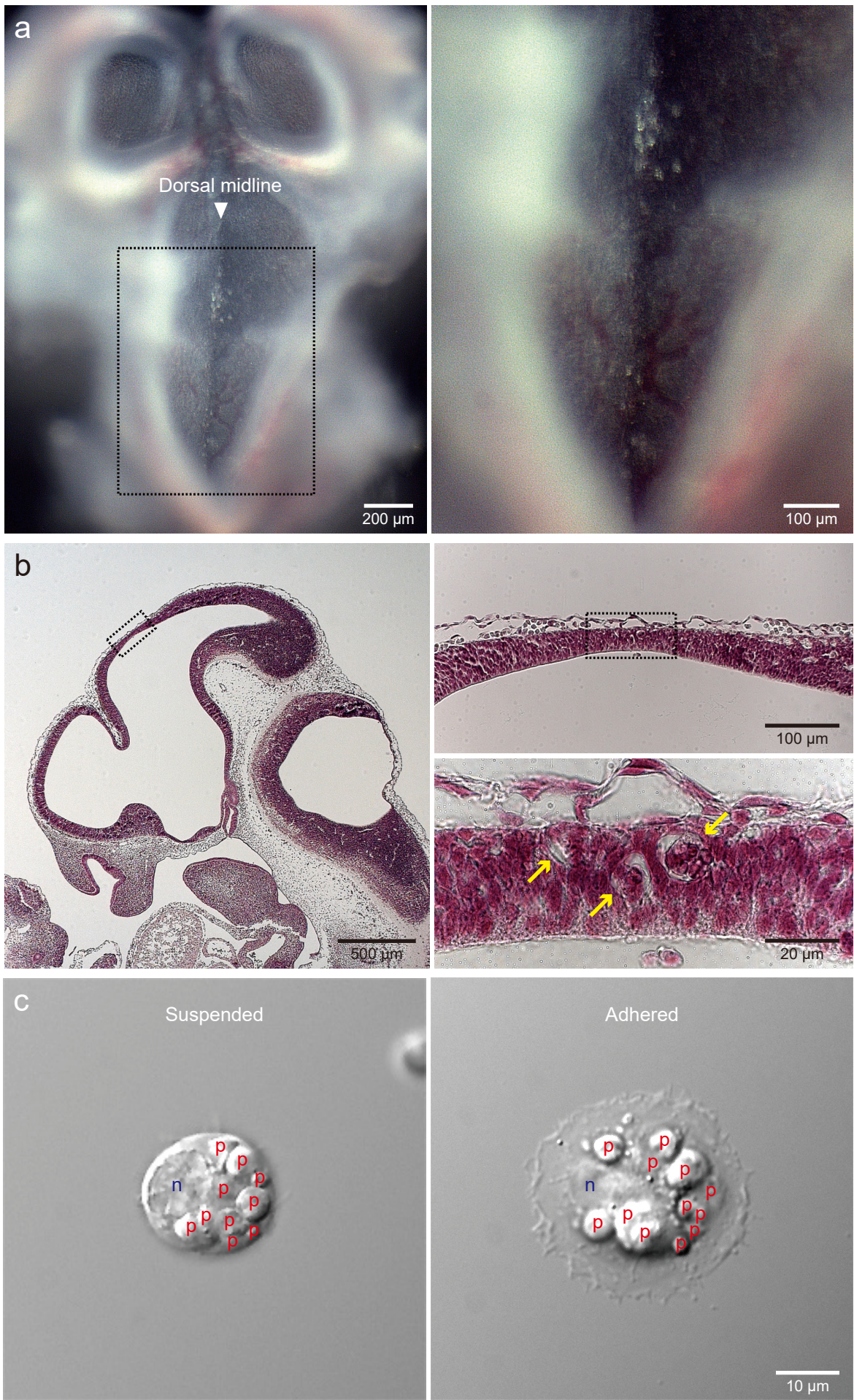
Supplementary Figure 1



Supplementary Figure 1. PNP and SVP development from E9.5 to E12.5 in mouse embryos.

(a) PNP development of the midbrain occurs from E9.5 to E12.5. Blood vessels are delineated by collagen type IV immunostaining (green). SVP development also occurs from E9.5 to E12.5 through angiogenesis. On E9.5, a few sprouts are observed in the neuroepithelium. SVP develops significantly throughout the entire avascular area of the neuroepithelium during this period. (b) Midbrain cerebral vascular formation in the E10.5 mouse embryo. CD31 staining concurrently delineates the formation of the SVP and PNP (red). Dotted lines indicate the vascular front of the SVP. In the SVP at E10.5, the dorsal midline area (arrowhead) is still avascular. Nuclei are counterstained with Hoechst (blue). (c) Rendered Z-stack confocal image showing dense vascular networks (CD31, red). Reconstituted Z-surface image depicting SVP and PNP.

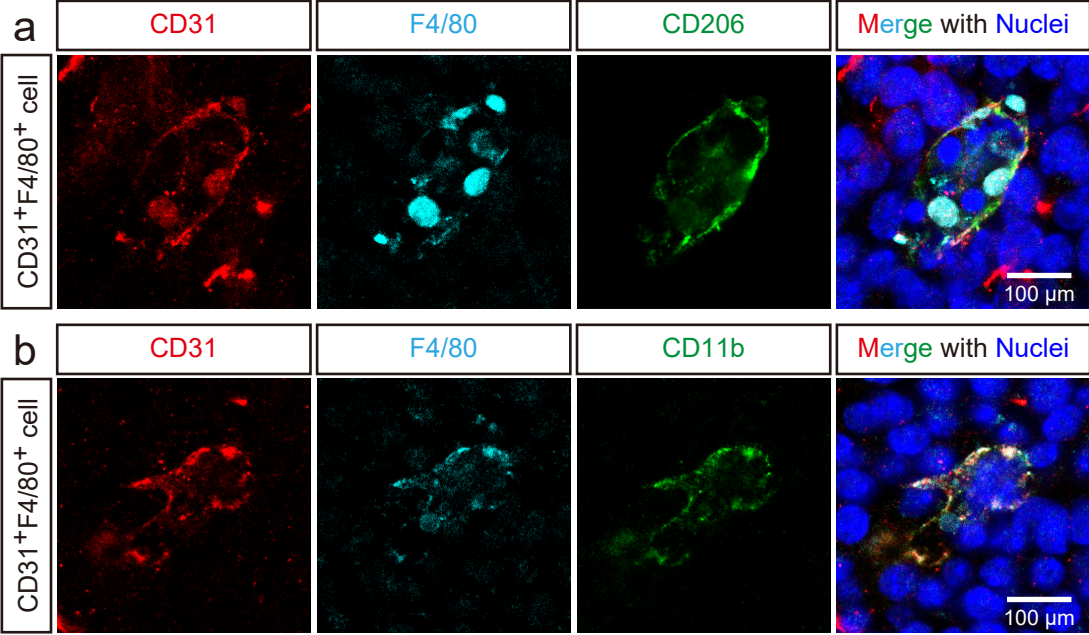
Supplementary Figure 2



Supplementary Figure 2. The phagocytes infiltrating the dorsal midline area are equivalent to the CD31⁺F4/80⁺ cells.

(a) Ventricle side view of the E10.5 mouse brain. Relatively large, light-reflecting, complex cells are observed in the dorsal midline area. The right panel shows a high magnification image of the boxed area in the left panel. The localization, morphology, and number of phagocytes correspond to the CD31⁺F4/80⁺ cells observed in Figures 1c-f, 4a, 5a, 7a. (b) Hematoxylin and eosin stained parasagittal section of the E10.5 dorsal midline area. The boxed area is enlarged in the upper right panel and further magnified on the lower right. Arrows indicate phagocytes. (c) The phagocytes can be isolated under the microscope based on their distinct tissue localization and unique phagocyte morphology (see Figure 3a). Phagocytosed apoptotic cells are observed in the isolated phagocytes when they are either suspended (left) or adhered (right). p (red), phagocytosed apoptotic cells; n (blue), nuclei.

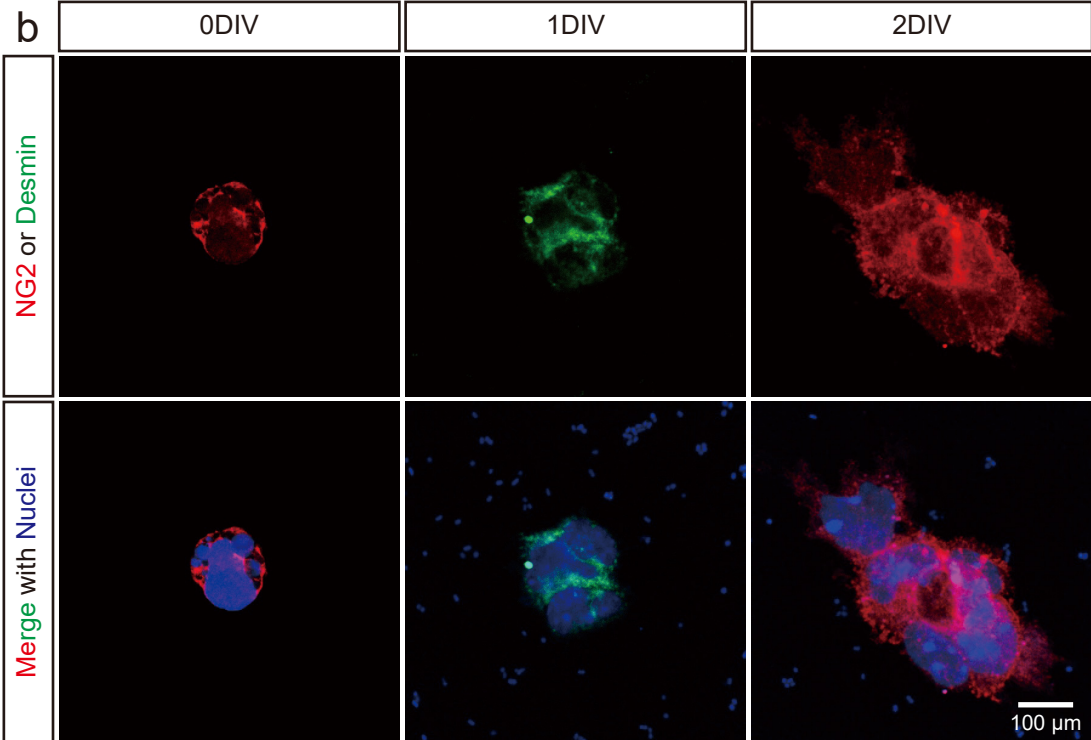
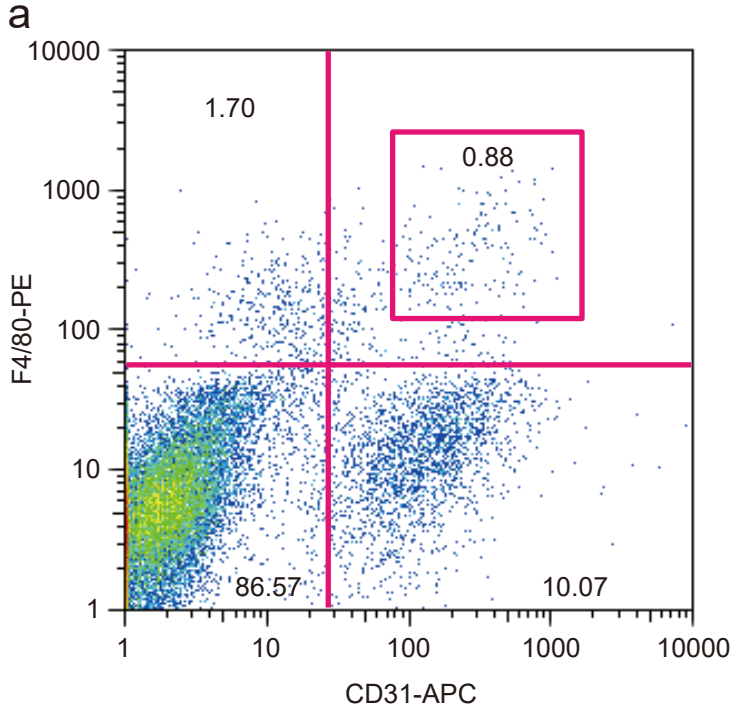
Supplementary Figure 3



Supplementary Figure 3. CD31⁺F4/80⁺ cells express multiple macrophage surface markers.

CD31⁺F4/80⁺ cells, which infiltrate the dorsal midline at E10.5, express multiple macrophage markers. (a) CD31⁺F4/80⁺ cells express CD206, an M2 macrophage marker. (b) CD31⁺F4/80⁺ cells express CD11b, a pan-marker for monocytes/macrophages. CD31 (red), F4/80 (cyan), CD206/CD11b (green), and nuclei (blue).

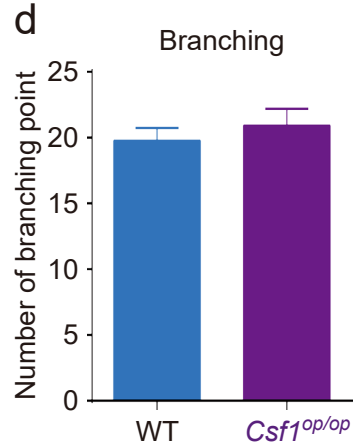
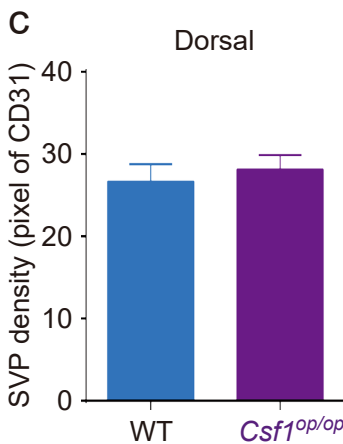
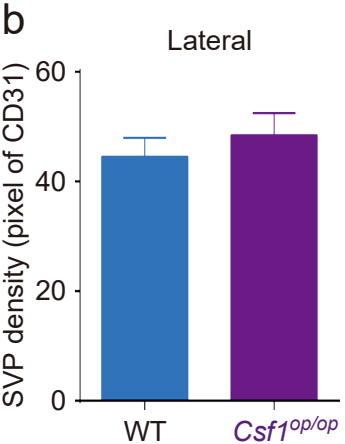
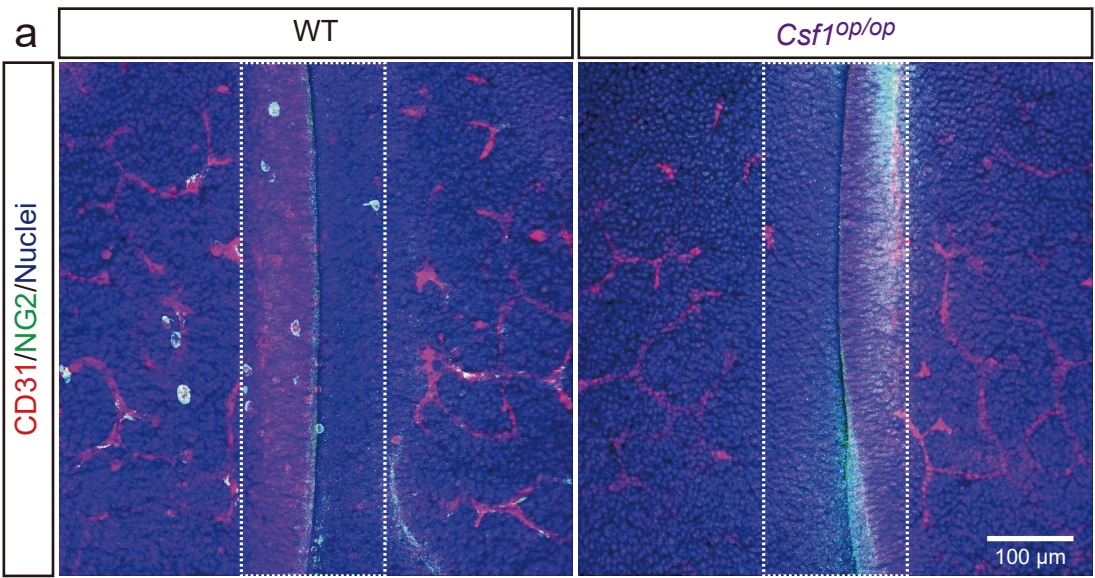
Supplementary Figure 4



Supplementary Figure 4. Fractionation of phagocytes by flow cytometry using their CD31 and F4/80 surface markers and subsequent *in vitro* culture.

(a) CD31⁺F4/80⁺ cells fractionated from the E10.5 dorsal midline area by flow cytometry. The double positive fraction is less than 1%. (b) Cultured CD31⁺F4/80⁺ cells express the pericyte markers NG2 (red) and desmin (green). On 0DIV, fractionated CD31⁺F4/80⁺ cells exhibit a phagocytic morphology. On 1DIV, the CD31⁺F4/80⁺ cells proliferate and express desmin (green). On 2DIV, a couple of daughter cells migrate and express NG2 (red). Nuclei are counterstained with TO-PRO-3 (blue).

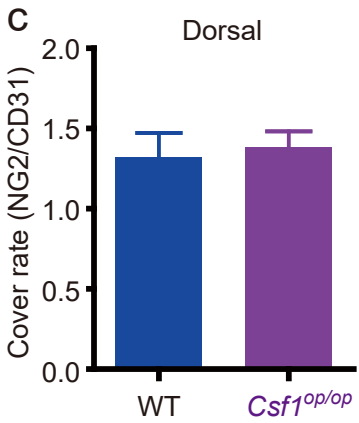
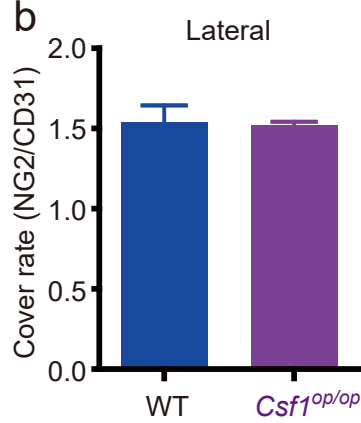
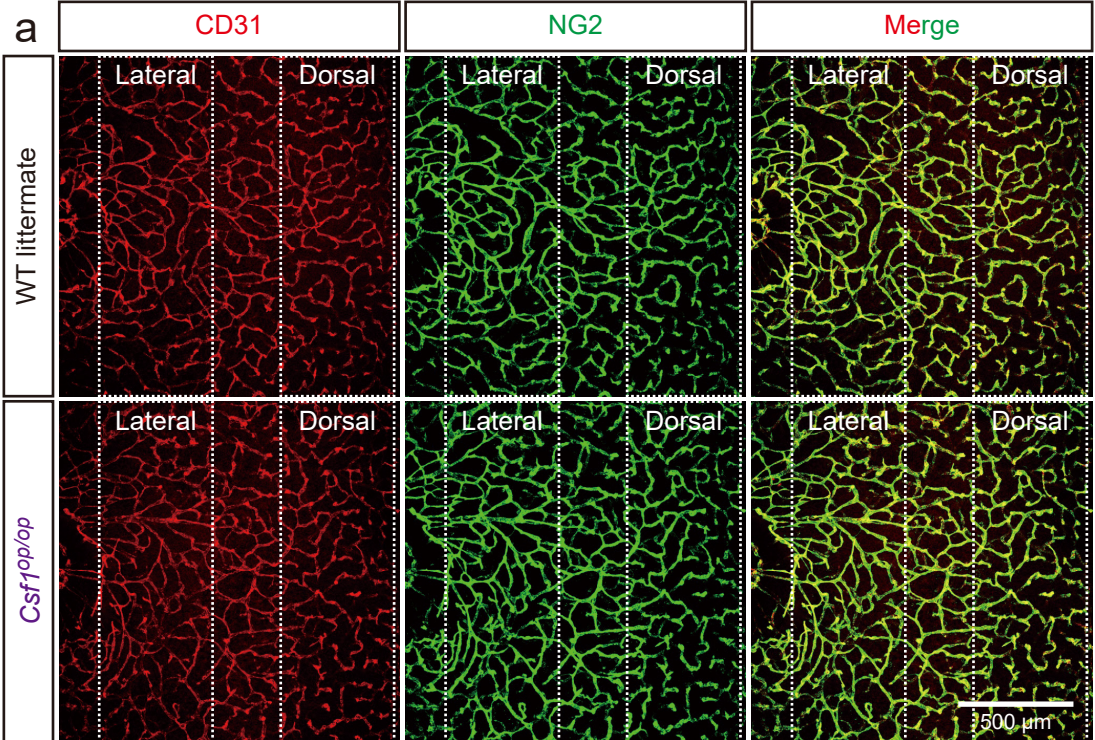
Supplementary Figure 5



Supplementary Figure 5. E10.5 *Csf1^{op/op}* mouse embryos do not display NG2⁺ cells at the dorsal midline area.

(a) In WT littermates, NG2⁺ cells (green, equivalent to the CD31⁺F4/80⁺ cells) are observed in the dorsal midline area (dotted area) at E10.5. In contrast, no NG2⁺ cells are observed in the *Csf1^{op/op}* mouse embryos. Nuclei are counterstained with Hoechst (blue). (b-d) CD31⁺F4/80⁺ cells may not be involved in SVP angiogenesis. There are no significant difference in SVP density on the lateral side (b) or dorsal side (c), or SVP branching point frequency (d) between the two genotypes (WT = 8; *Csf1^{op/op}* = 7; 200 × 200 μm). All error bars indicate the mean ± s.e.m.

Supplementary Figure 6



Supplementary Figure 6. E12.5 *Csf1^{op/op}* embryos show no change in pericyte coverage of the SVP.

(a) There is no significant difference in NG2⁺ pericyte coverage in the dorsal midline area on E12.5 between *Csf1^{op/op}* mice and WT littermates. Statistical analysis further confirms no significant difference in the lateral (b) or dorsal (c) areas (ns, WT = 5, *Csf1^{op/op}* = 5, 400 × 400 μm). All error bars indicate the mean ± s.e.m.

Supplementary Figure 7

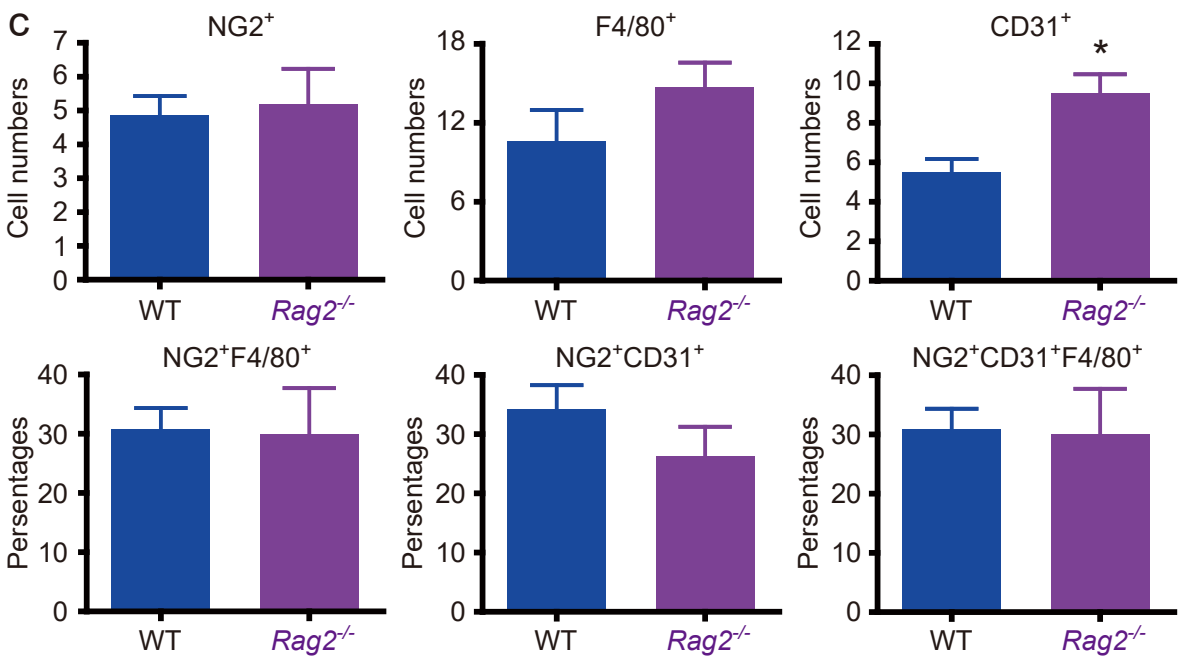
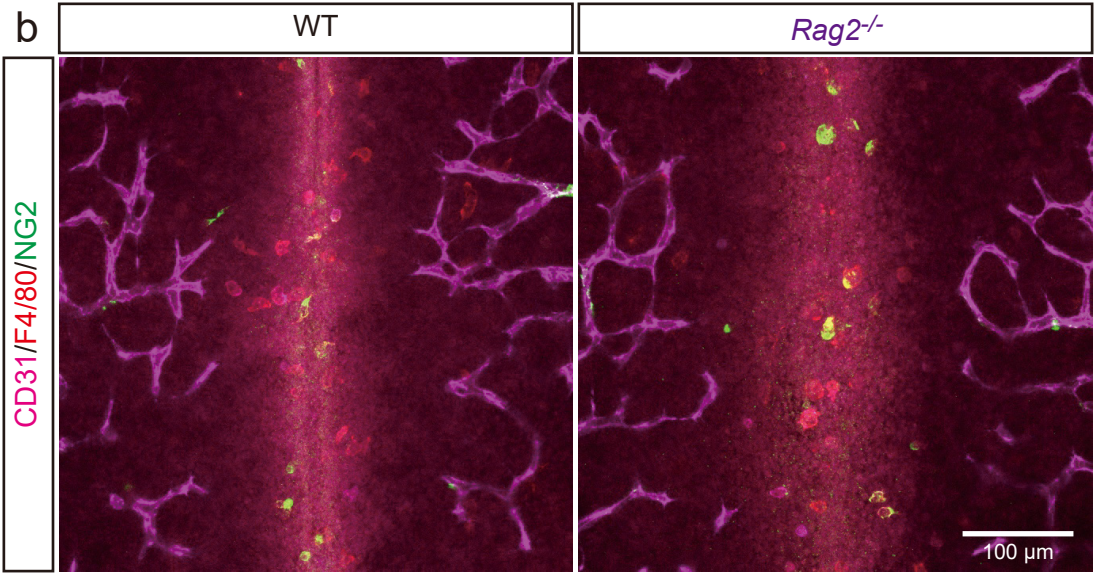
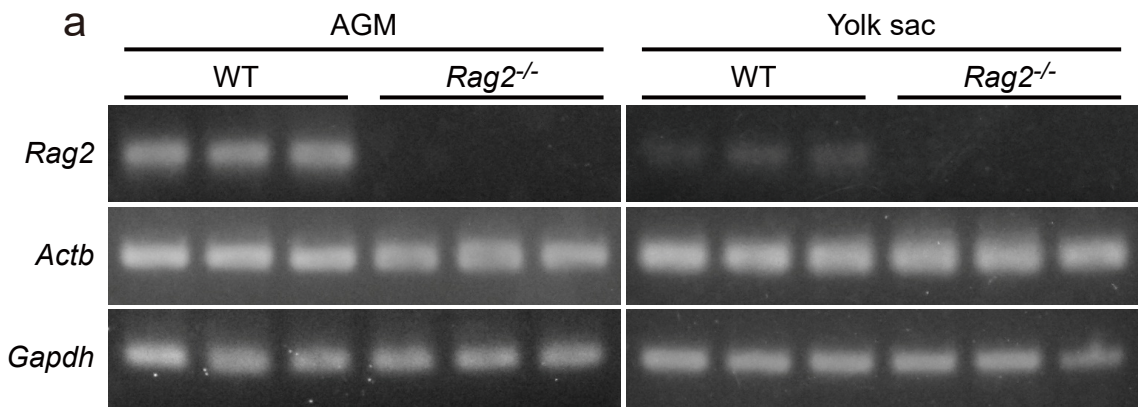
Gene Symbol	OP/WT ratio	Entrez Gene
630663	23.033	107849
<i>Hand1</i>	6.171	15110
<i>Fgf8</i>	5.667	14179
<i>Fgf2</i>	4.650	14173
<i>Shh</i>	4.543	20423
<i>Gbx2</i>	3.715	14472
<i>Prl7d1</i>	3.707	18814
<i>Thy1</i>	3.556	21838
<i>Prl2c2</i>	3.466	18811
<i>Fgf18</i>	2.931	14172
<i>Sema5a</i>	2.068	20356
<i>Angpt2</i>	1.768	11601
<i>Scg2</i>	1.766	20254
<i>Adra2b</i>	1.537	11552
<i>Runx1</i>	1.531	12394
<i>Wisp3</i>	1.526	327743
<i>Serpinf1</i>	1.510	20317
<i>Foxc2</i>	1.498	14234
<i>Cyr61</i>	1.486	16007
<i>Cx3cl1</i>	1.401	20312
<i>Shc1</i>	1.375	20416
<i>Col4a3</i>	1.321	12828
<i>Lect1</i>	1.313	16840
<i>Tbx1</i>	1.310	21380
<i>Pitx2</i>	1.305	18741
<i>Tbx20</i>	1.293	57246
<i>Bmp4</i>	1.271	12159
<i>Thbs1</i>	1.269	21825
<i>Eng</i>	1.254	13805
<i>Il1b</i>	1.253	16176
<i>Tie1</i>	1.234	21846
<i>Meox2</i>	1.228	17286
<i>Ctgf</i>	1.214	14219
<i>Klf5</i>	1.214	12224
<i>Nos3</i>	1.214	18127
<i>Pnpla6</i>	1.212	50767
LOC640441	1.209	21825
<i>Gna13</i>	1.204	14674
<i>Fgf9</i>	1.193	14180
<i>Epas1</i>	1.192	100048537
<i>C1galt1</i>	1.189	94192
<i>Id1</i>	1.163	15901
<i>Meis1</i>	1.152	17268
<i>B4galt1</i>	1.151	14595
<i>Sox18</i>	1.147	20672
<i>Ednra</i>	1.147	13617
<i>Vhl</i>	1.130	22346
<i>Notch4</i>	1.126	18132
<i>S1pr1</i>	1.124	13609
<i>Ubp1</i>	1.114	22221
<i>Amot</i>	1.107	27494
<i>Nrp1</i>	1.107	18186
<i>Hbegf</i>	1.102	15200
<i>Edn1</i>	1.085	13614
<i>Wasf2</i>	1.059	242687
<i>Erap1</i>	1.043	80898
<i>Col18a1</i>	1.042	12822
<i>Cxcr4</i>	1.037	12767
<i>Casp8</i>	1.035	12370
<i>Ptk2</i>	1.030	14083
<i>Nf1</i>	1.028	18015
<i>Elk3</i>	1.016	13713
<i>Vegfa</i>	1.011	22339

Gene Symbol	OP/WT ratio	Entrez Gene
<i>Acvr1l1</i>	0.999	11482
<i>Hs6st1</i>	0.998	100047260
<i>Fgfr1</i>	0.995	14182
<i>Rbm15</i>	0.994	229700
<i>Tgfa</i>	0.993	21802
<i>Sox17</i>	0.990	20671
<i>Pten</i>	0.985	19211
<i>Map3k7</i>	0.985	26409
<i>Adamts1</i>	0.984	11504
<i>Dicer1</i>	0.984	192119
<i>Hif1a</i>	0.984	15251
<i>Mapk7</i>	0.980	23939
<i>ErbB2</i>	0.979	13866
<i>Pofut1</i>	0.976	140484
<i>Rhob</i>	0.971	11852
<i>VeZF1</i>	0.971	22344
<i>Flt1</i>	0.969	14254
<i>Pml</i>	0.967	18854
<i>Cxcl12</i>	0.967	20315
<i>Rtn4</i>	0.959	68585
<i>Acvr1</i>	0.958	11477
<i>Gpx1</i>	0.956	14775
<i>Pdgfa</i>	0.952	18590
<i>Rbpj</i>	0.950	19664
<i>Dll4</i>	0.948	54485
<i>Crhr2</i>	0.945	12922
<i>Epas1</i>	0.935	13819
<i>Vash1</i>	0.930	238328
<i>Pknox1</i>	0.911	18771
<i>Plxnd1</i>	0.909	67784
<i>Anxa2</i>	0.905	12306
<i>Plcd1</i>	0.901	18799
<i>Tgfr2</i>	0.896	21813
<i>Bai1</i>	0.891	107831
<i>Hhex</i>	0.890	15242
<i>Ctnnb1</i>	0.877	12387
<i>Gata2</i>	0.860	14461
<i>Mapk14</i>	0.856	26416
<i>Fgfr2</i>	0.826	14183
<i>Tek</i>	0.818	21687
<i>Htatip2</i>	0.802	53415
<i>Plcd3</i>	0.799	72469
<i>Ihh</i>	0.765	16147
<i>Notch1</i>	0.759	18128
<i>Il18</i>	0.667	16173
<i>Cx3cr1</i>	0.478	100047704
<i>Ang</i>	0.381	11727
<i>Plg</i>	0.367	18815
<i>Ovol2</i>	0.278	107586
<i>Enpep</i>	0.207	13809
<i>Serpine1</i>	0.195	18787
<i>Tbx4</i>	0.126	21387
<i>Il1a</i>	0.119	16175

Supplementary Figure 7. Microarray data regarding angiogenesis-related genes.

Microarray data showing the ratio of angiogenesis-related genes expressed in the dorsal midline area of the *Csf1^{op/op}* mouse embryo as compared with the WT littermate. Some genes are down-regulated; however, the expression levels of angiogenesis-related genes basically appear of a similar level to the WT littermate.

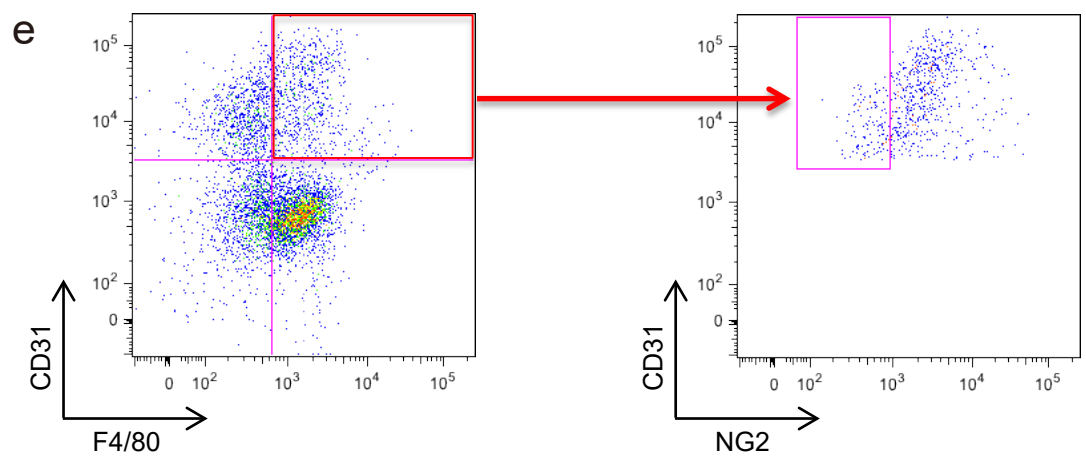
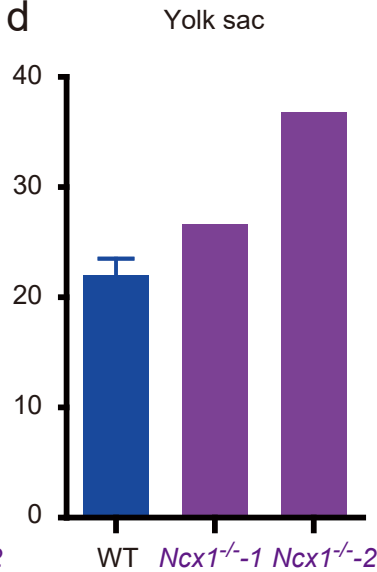
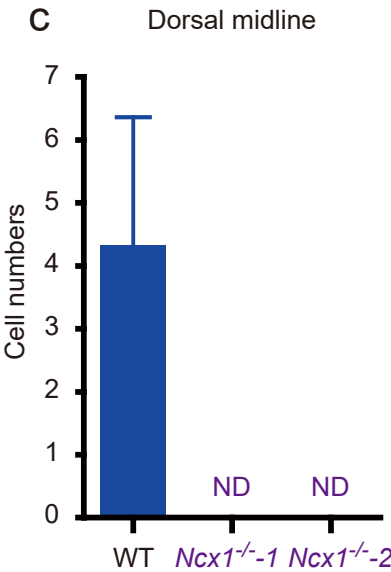
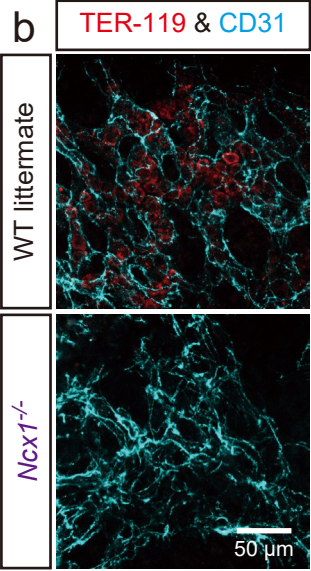
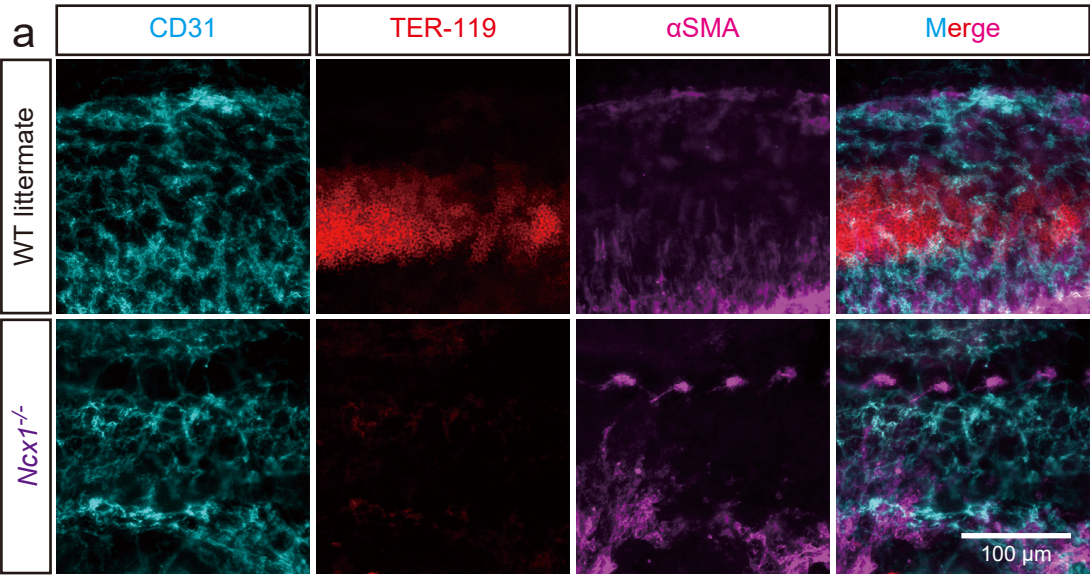
Supplementary Figure 8



Supplementary Figure 8. *Rag2*^{-/-} mice exhibit no alteration in NG2⁺ cell recruitment to the dorsal midline area.

(a) AGM and yolk sac *Rag2* mRNA expression levels are confirmed by RT-PCR. (b) Immunofluorescence on E10.5 *Rag2*^{-/-} mouse embryos. Compared to their WT littermates, *Rag2*^{-/-} mice showed no significant difference in NG2⁺ cells (green, equivalent to the CD31⁺F4/80⁺ cells). (c) Statistical analysis of the cells infiltrating the dorsal midline area (WT = 4, *Rag2*^{-/-} = 5, 200 × 200 μm). Cell surface marker analysis-based immunostaining shows essentially no difference between the 2 genotypes, with the exception of the single positive CD31⁺ cell population (*, *P* < 0.05). All error bars indicate the mean ± s.e.m.

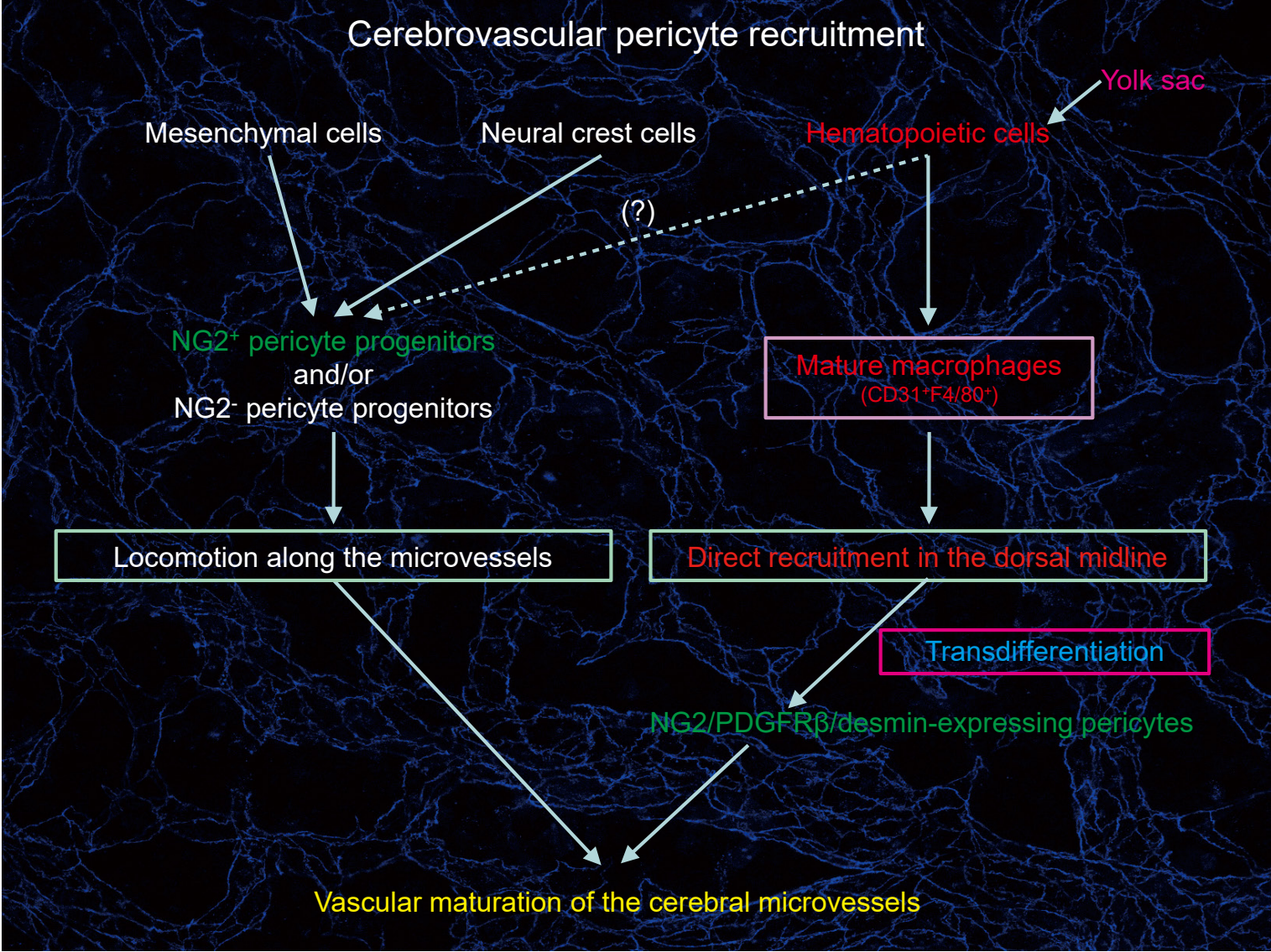
Supplementary Figure 9



Supplementary Figure 9. NG2⁺ cells are not observed in the dorsal midline of *Ncx1*^{-/-} mice and CD31⁺F4/80⁺NG2⁻ cells can be fractionated from yolk sac.

- (a) Although TER-119⁺ embryonic erythrocytes (red) are recruited to the dorsal aorta region (cyan) encircled by α SMA⁺ cells (magenta) in the WT littermates, this event is not observed in *Ncx1*^{-/-} mouse embryos on E10.5. (b) Compared to the embryos of WT littermates, *Ncx1*^{-/-} mouse embryos also show no TER-119⁺ embryonic erythrocytes (red) in the cutaneous microvasculature (cyan). (c) In *Ncx1*^{-/-} mice, no recruitment of the NG2⁺ cells (equivalent to the CD31⁺F4/80⁺ cells) can be observed in the dorsal midline (WT = 3, *Ncx1*^{-/-} = 2, 200 \times 200 μ m). (d) In contrast, yolk sac hematopoiesis, as surveyed by the major hematopoietic population, shows relatively increased TER-119⁺ erythrocyte numbers (WT = 3, *Ncx1*^{-/-} = 2, 50 \times 50 μ m). (e) CD31⁺F4/80⁺NG2⁻ cells are fractionated from yolk sacs of mCherry⁺ embryos by flow cytometry.

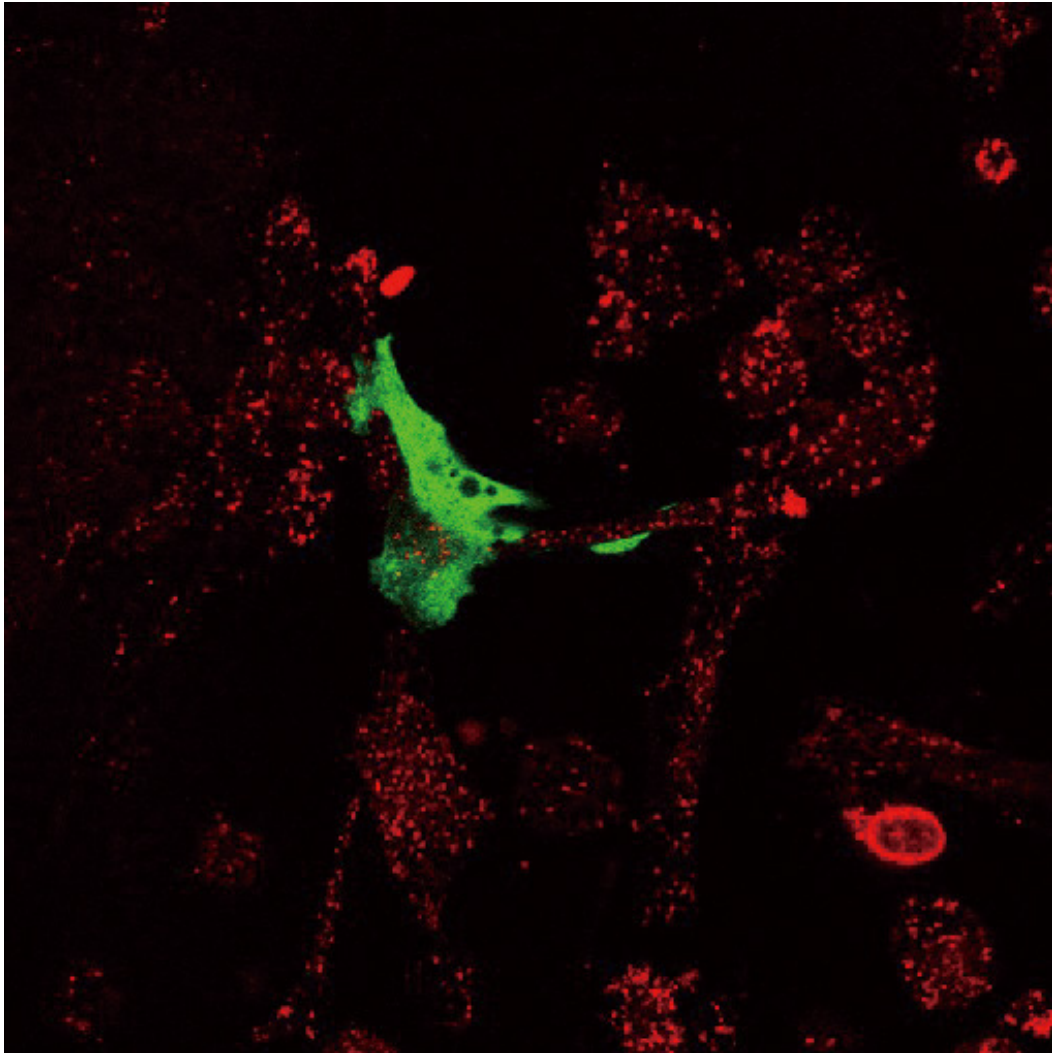
Supplementary Figure 10



Supplementary Figure 10. Two cerebrovascular pericyte recruitment mechanisms.

Direct recruitment to the dorsal midline (right scheme), which was demonstrated in this study *ad initium*, and locomotion along the microvessels (left scheme), which is well established in the vascular biology field, are suggested as two methods of pericyte recruitment.

Supplementary Movie 1



Supplementary Movie 1. Single-cell-tracing time-lapse analysis of target cell transdifferentiation.

The single-cell-tracing time-lapse study shows that EGFP reporter cells proliferate and transdifferentiate to pericytes *in vitro*.

Supplementary References

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