

Supplemental Figure Legends

Figure S1. Related to Figures 1 and 3: Circadian study design for (A) in vivo mouse, and (B) Cell-autonomous U2 OS cell line (C) Cycling of the top 12 metabolites in mouse liver with JTK q-values $< 4 \times 10^{-10}$. The y-axis is the intensity following run-order correction as described in the methods. Data was derived from two biological replicates and two analytical replicates. Error bars represent standard error of the mean. D) Temporal acrophases of glycogen (from literature, see text), glucose, citrate, and malate. E) Juxtaposition of metabolites indicative of oxidative status and methylation. Numbers in red indicate acrophase in CT. Data for all time-series plots was derived from two biological replicates and two analytical replicates. Error bars represent standard error of the mean.

Figure S2. Related to Figure 4: Effect of oscillations following knockdown of circadian clock components in U2 OS cells. U2 OS cells were transfected with siRNAs and relative expression is normalized to GAPDH. (A) Relative expression of *BMAL1* (left panel), *CRY1* (middle panel), *CRY2* (right panel). (B) Distribution of periods with each metabolite plotted as points. Size indicates JTK BH.Q $-\log$ value, and the line indicates density of periods. (C) *BMAL1* knockdown dampens oxidative cycling as measured by the NAD⁺/NADH raw ion count ratio (top panel) as compared to *CRY1* (middle) and *CRY2* (bottom). (D) Cycling of the top 13 metabolites under control conditions in the cell-autonomous model with JTK q-values < 0.05 . The y-axis is the intensity following run-order correction as described in the methods. Data for all cycling plots was derived from two analytical replicates, and error bars represent standard error of the mean.

Figure S3. Related to Figure 4: Metabolic oscillations and pathway analysis under *CRY1* and *CRY2* knockdown conditions in U2OS cells. *CRY1* (left panel) and *CRY2* (right panel) knockdown increases the number of cycling metabolites, eliminates circadian oscillations. 69 metabolites (~50%) in *CRY1* knock down cells and 64 metabolites (47%) in *CRY2* knockdown cells show cycling.

Figure S4. Related to Figure 5: A) Differential analysis between U2 OS cells with *Cry1* KD (green bar) or scrambled control (blue bar). Samples were considered from all time points

to understand global Cry1 knockdown effect on metabolism. B) Frozen liver samples from mice of the indicated genotype (WT or CRY1^{-/-}) and circadian timepoint (ZT 10 and ZT 22) were suspended in lysis buffer, lysed by Tissue Lyser II from Qiagen and further homogenized by passage through a Qias shredder column. Lysates were processed for expression by immunoblot for Cry1 and mTOR pathway proteins, with tubulin serving as a loading control.

Figure S5. Related to Figure 7: A) Cultured hepatocytes were synchronized with 0.1 μ M dexamethasone, and mRNA was collected at the indicated timepoints after synchronization. Expression of *Cry1* and *Cry2* were determined by qPCR, normalized to expression of 36B4 (*Rplp0*) as has been described (Gréchez-Cassiau, 2008) previously. Data are from two biological replicates (ie, *Cry1-1* is the first replicate, *Cry1-2* is the second replicate). (B) Heatmap of significantly cycling metabolite from primary hepatocyte metabolite profiling.

Table S1. Related to Figures 1 and 3: Compilation of characteristics of published circadian metabolomics studies.

Table S2. Related to STAR methods: Oligonucleotides used in this study.

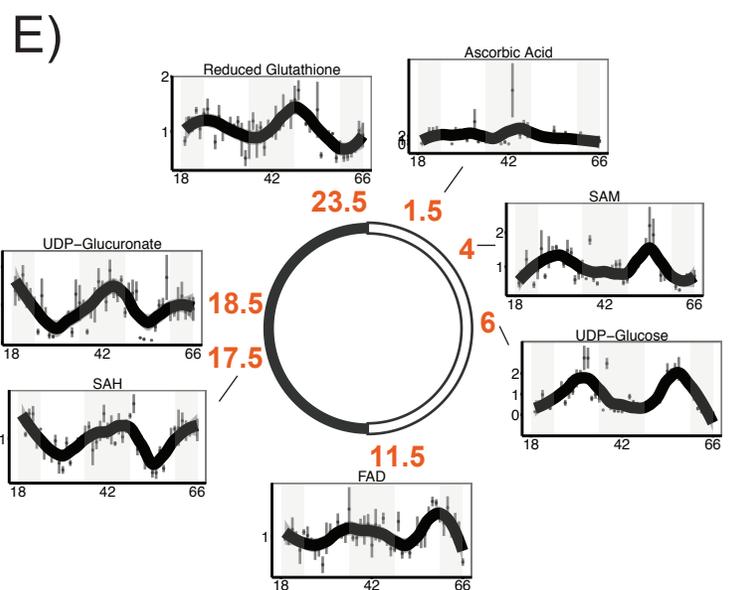
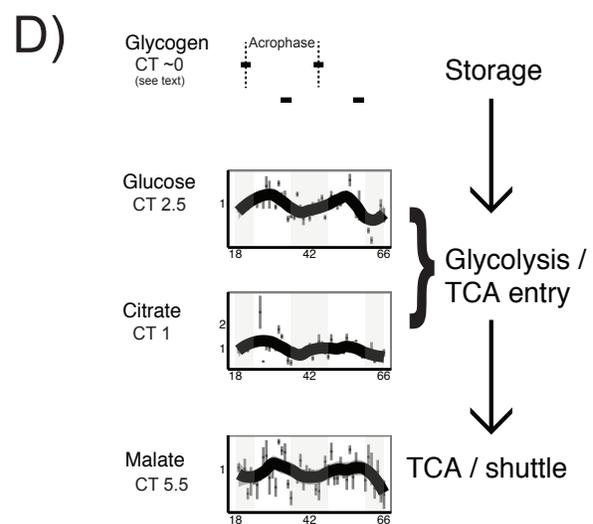
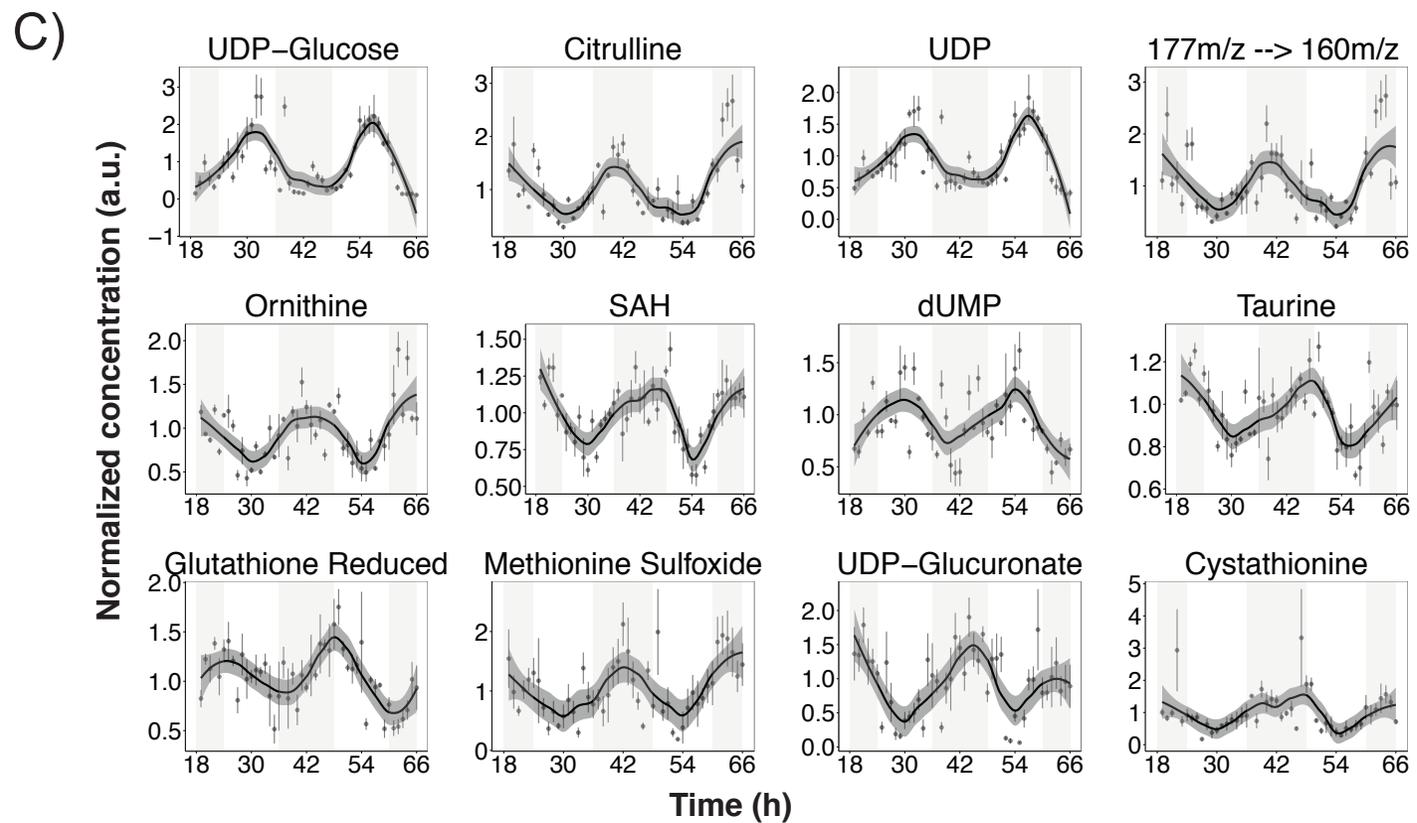
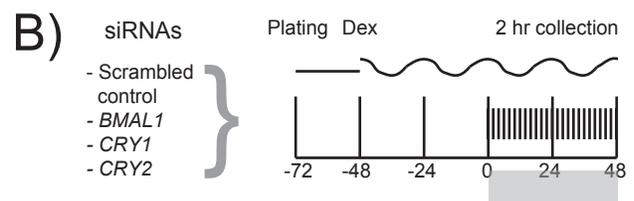
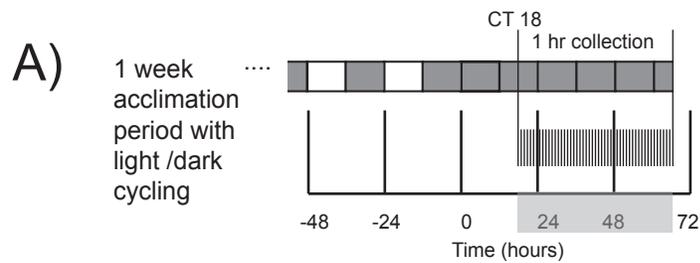


Figure S1

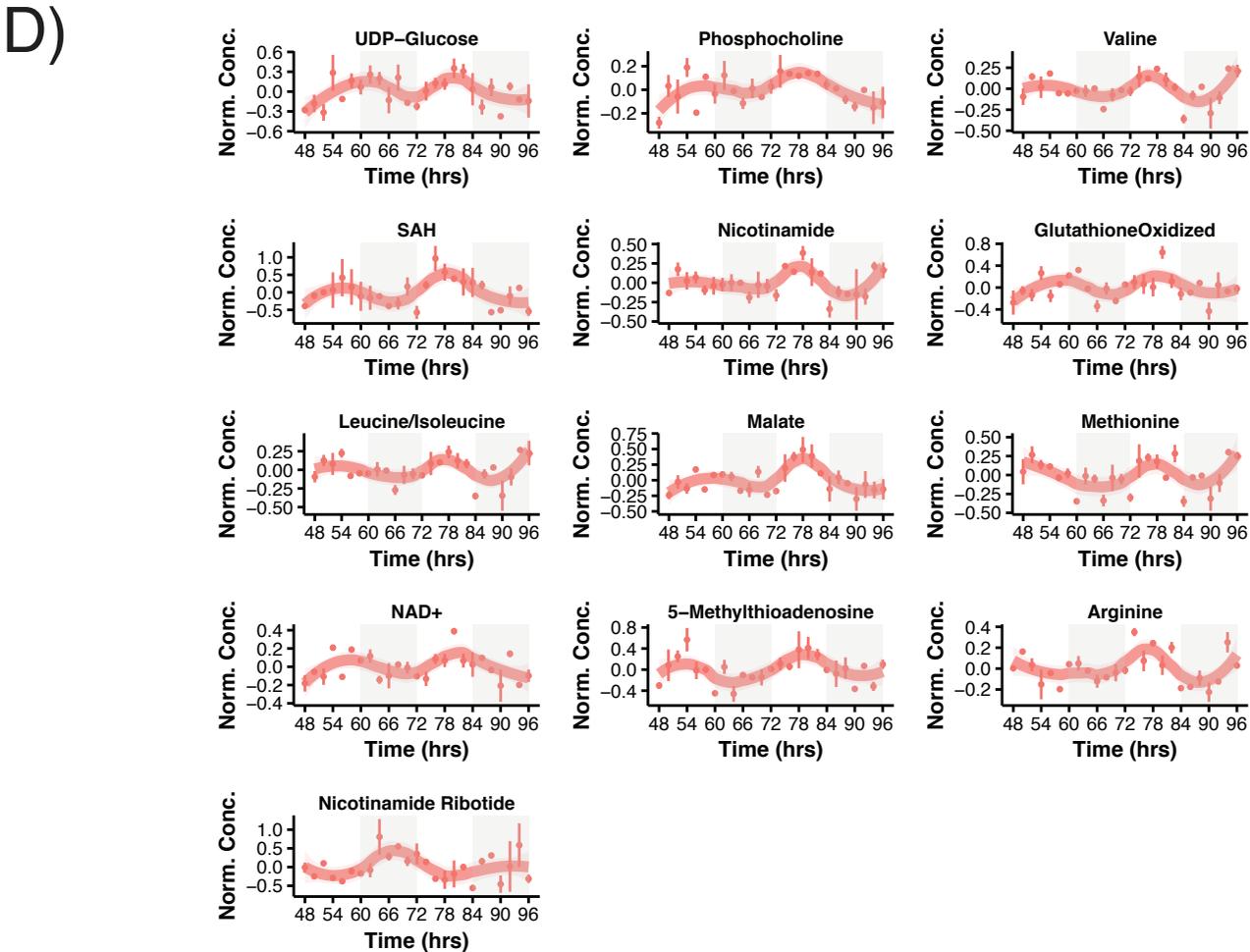
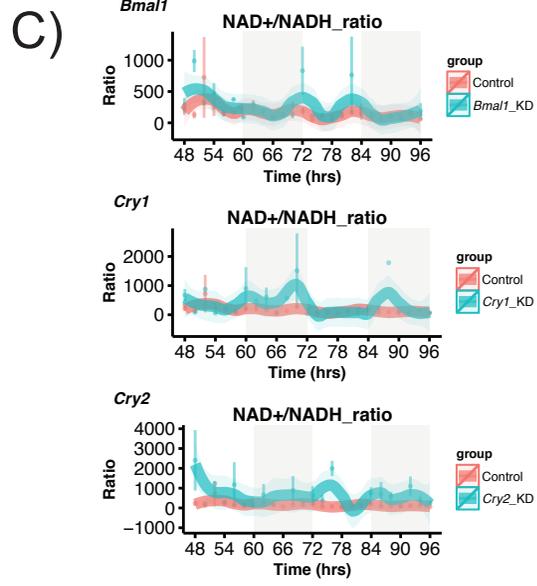
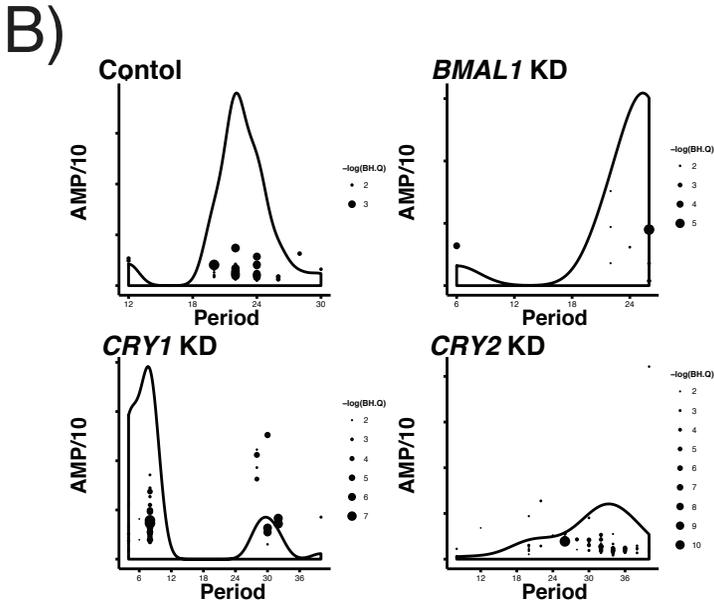
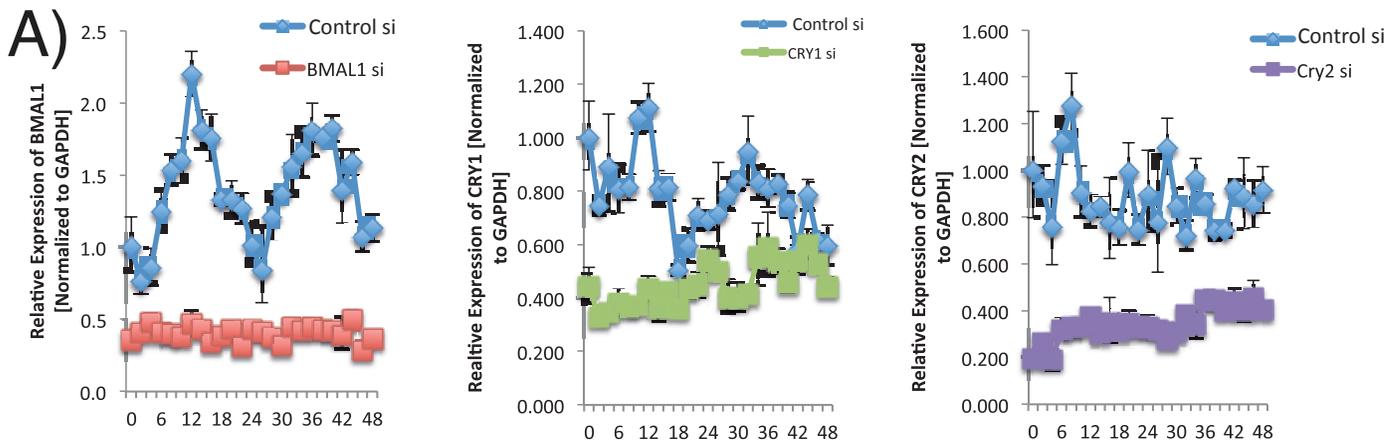


Figure S2

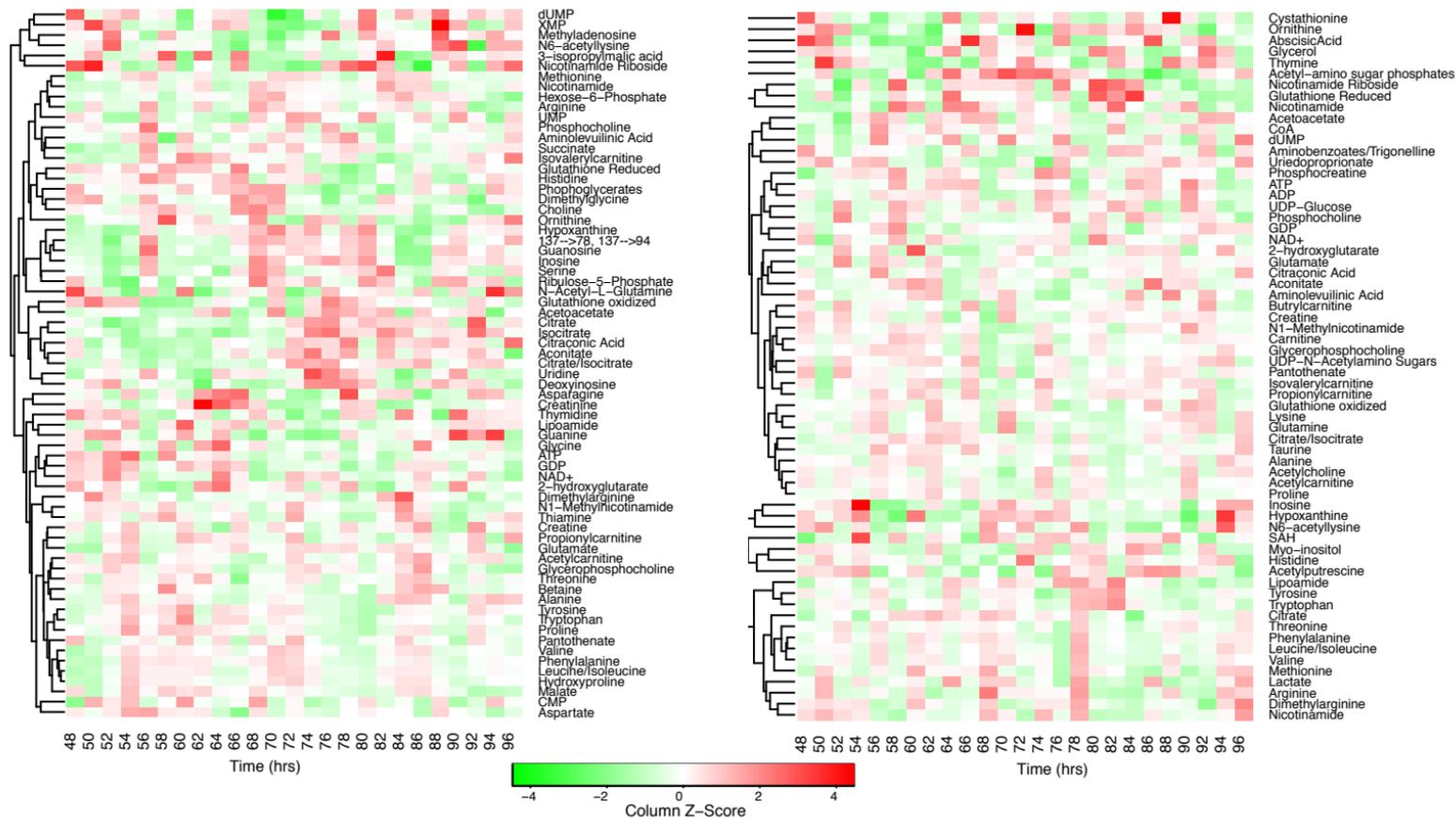


Figure S3

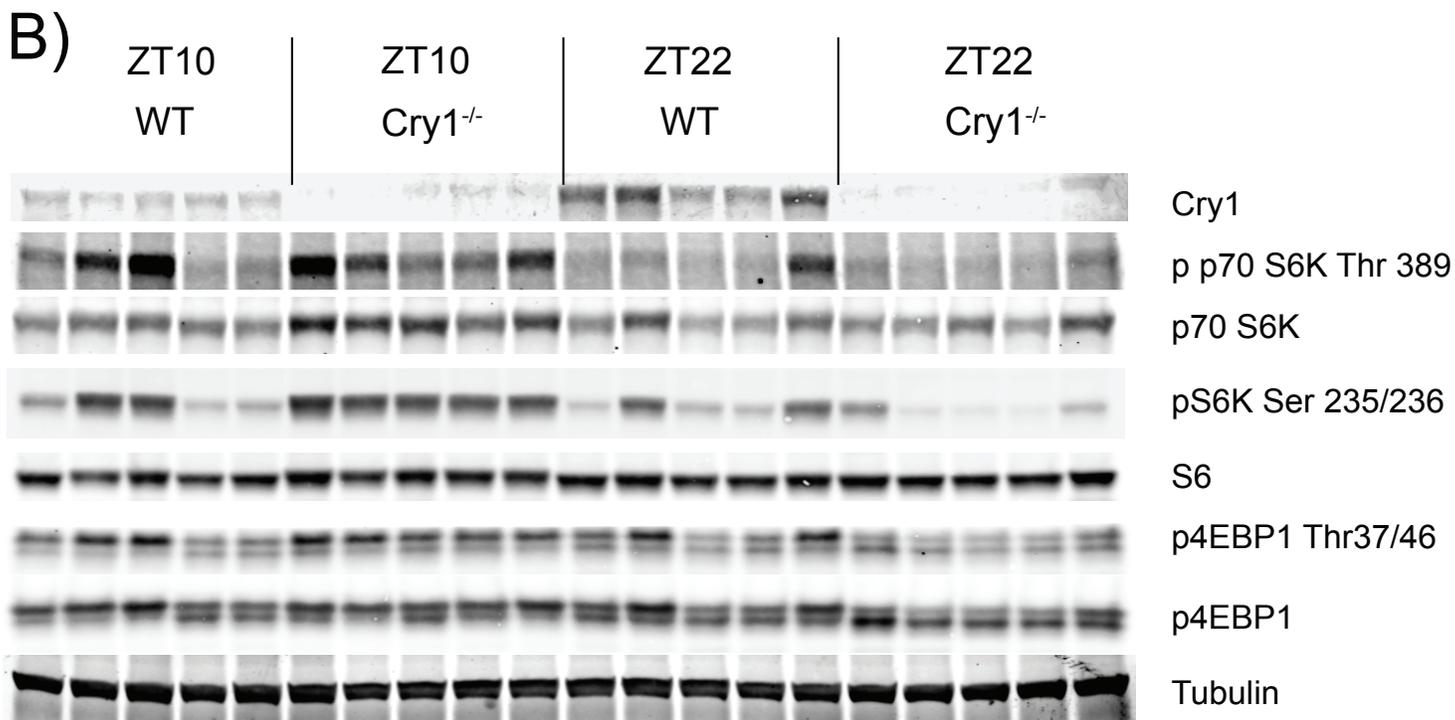
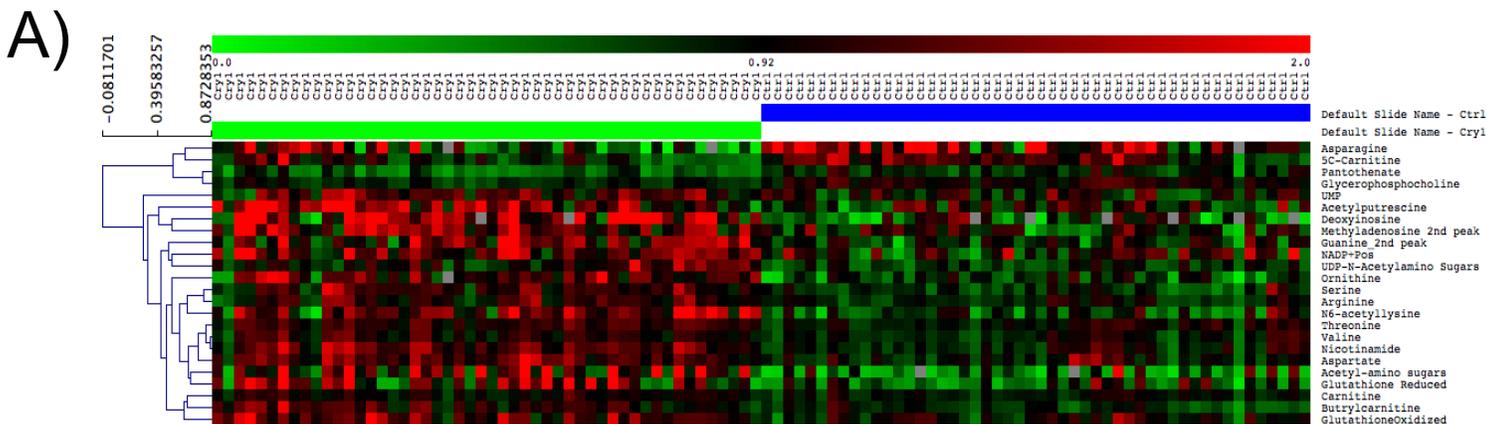
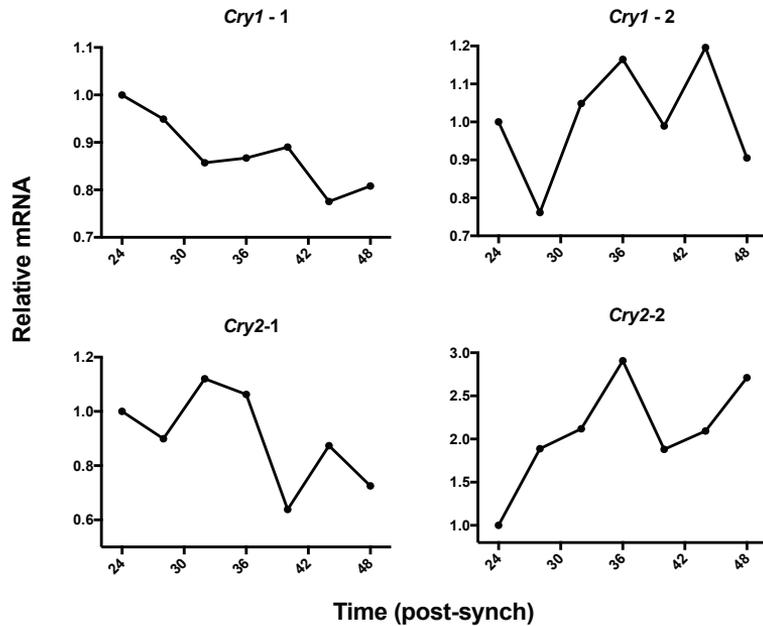


Figure S4

A)



B)

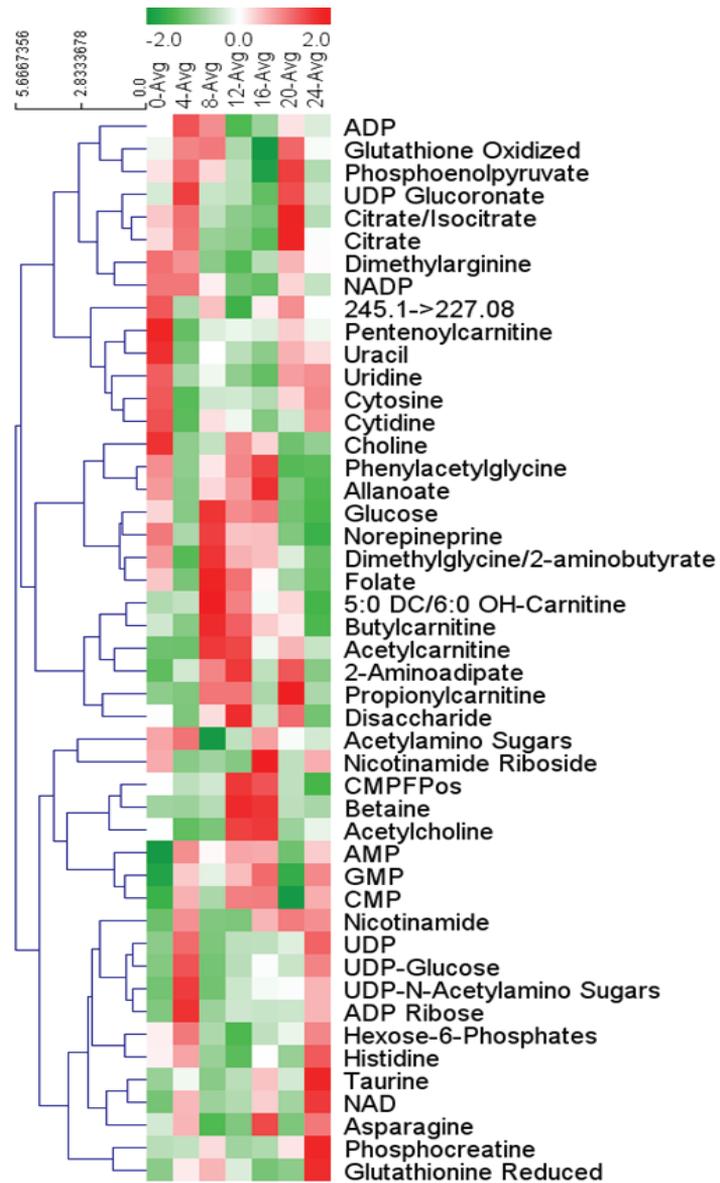


Figure S5

Table S1

	Organism	Circadian / Diurnal	Sample studied	Total Length	Resolution	Replicates? (Number)	Total samples	Metabolites measured
Current study	Mouse	Dark / Dark	Liver	48 h	1 h	Y (2)	96	179
	Human	N/A	U2 OS	48 h	2 h	N	24	137
			Primary Hepatocytes	24 h	4 h	Y (2-3)	16	147
(Thaiss et al., 2016)	Mouse	Light / Dark	Feces Serum	48 h	6 h	Y (18)	162	ND
(Tran et al., 2016)	Mouse	Light / Dark	Liver Serum	24 h	6 h		4	110 80
(Masri et al., 2016)	Mouse	Light / Dark	Liver	24 h	4 h	Y (5)	30	ND
(Abbondante et al., 2015)	Mouse	Light / Dark	Liver Serum	24 h	4 h	Y (5)	30	306 362
(Gogna et al., 2015)	Fly	Light / Dark & Temperature	Whole body	24 h	2 h	Y (5)	12	14 metabolites / NMR
(Li et al., 2015)	Zebrafish	Light / Dark	Whole larvae	24 h	6 h	Y (3)	12	73 assigned signals by NMR
(Zwighaft et al., 2015)	Mouse	Light / Dark	Liver	24 h	4 h	Y (4)	24	Polyamines / HPLC
(Giskeødegård et al., 2015)	Human	Dim light, constant routine, 24 hour wake/sleep, 24 hour wakefulness	Urine	24 / 24	2-8 h	Y (15)	105	NMR / binning
(Castro et al., 2015; Paschos et al., 2012)	Mouse	Light / Dark	White and Brown Adipose Tissues	24 h	6 h	Y (3)	12	- Fatty acids - Intact lipids - NMR polar
(Davies et al., 2014)	Human	Dim light, constant routine, 24 hour wake/sleep, 24 hour wakefulness	plasma	24 / 24	2 h	Y (12)	144 / 144	171
(Masri et al., 2014)	Mouse	Light / Dark	Liver	24 h	4 h	ND	ND	ND
(Dyar et al., 2014)	Mouse	Light / Dark	Muscle	24 h	4 h	Y (5)	30	277 (only sugars reported)
(Chaix et al., 2014)	Mouse	Light / Dark	Serum	24 h	4 h	N	6	278
(Shostak et al., 2013)	Mouse	Light / Dark and Dark / Dark	Blood / adipose	24 h	6 h	Y (3-5)	12-20	Selected fatty acids / lipid compounds
(Eckel-Mahan et al., 2013)	Mouse	Light / Dark	Liver	24 h	4 h	Y (5)	30	306 total
(Chua et al., 2013)	Human	Dim light, Constant lab conditions, wakefulness	plasma	28 h	4 h	Y (20)	140	LCMS / 263 lipids
(Hatori et al., 2012)	Mouse	Light / Dark	Liver	24 h	3 h	Y (3-4)	24-32	324
(Eckel-Mahan et al., 2012)	Mouse	Light / Dark	Liver	24 h	6 h	Y (5)	20	309 named (Metabolon)
(Dallmann et al., 2012)	Human (pooled)	Dim light / constant protocol / sleep deprivation	Blood plasma	40 h	4 h	Y (10)	100	258 total, 219 across two individuals (Metabolon)
(Fustin et al., 2012)	Mouse	Light / Dark and Dark / Dark	Liver	24 h	4 h	Y (3)	18	CE-TOF-MS; only nucleic acid metabolites reported

Table S2

REAGENT or RESOURCE	SOURCE	IDENTIFIER
36B4 (<i>Rplp0</i>)	IDT	TTA TAA CCC TGA AGT GCT CGA C, CGC TTG TAC CCA TTG ATG ATG
REV-ERBα (<i>Nr1d1</i>)	Taqman Gene Expression Assay	Mm00520708_m1
REV-ERBβ (<i>Nr1d2</i>)	IDT	Mm.PT.51.12747673
BMAL1 (<i>Arntl</i>)	Taqman Gene Expression Assay	Mm00500226_m1
<i>Cry1</i>	Taqman Gene Expression Assay	Mm00514392_m1
<i>Cry2</i>	Taqman Gene Expression Assay	Mm01331543_g1
<i>GAPDH</i>	Taqman Gene Expression Assay	Endogenous Human control
<i>ARNTL</i>	Taqman Gene Expression Assay	Hs00154147_m1
<i>CRY1</i>	Taqman Gene Expression Assay	Hs00172734_m1
<i>CRY2</i>	Taqman Gene Expression Assay	Hs00391360_m1
siRNA targeting human <i>CRY1</i>	Qiagen	GS1407
siRNA targeting human <i>CRY2</i>	Qiagen	GS1408
siRNA targeting human <i>ARNTL</i>	Qiagen	GS406
AllStars Negative Control siRNA	Qiagen	SI03650318