

**Title: Microbial Signatures Associated with Oropharyngeal and Oral Squamous Cell Carcinomas**

**Sagarika Banerjee, Tian Tian, Zhi Wei, Kristen N Peck, Natalie Shih, Ara A Chalian, Bert W O'Malley Jr, Gregory S Weinstein, Michael D Feldman, James Alwine, Erle S. Robertson**

**Supplemental figure legends.**

**Figure S1.** (Figure S1a-S1f). Probe capture sequencing alignments post MiSeq. The MiSeq reads from individual capture when aligned with the metagenome of PathoChip (Chip probes) was found to cluster mostly at the capture probe regions. The genomic location along with the number of MiSeq reads are mentioned in the figure. The alignment for O capture pool library is shown in Figure S1a; that of B library is shown Figure S1b & S1c; that of F library is shown in Figure S1d; that of P library is shown in Figure S1e & S1f.

Figure S1a

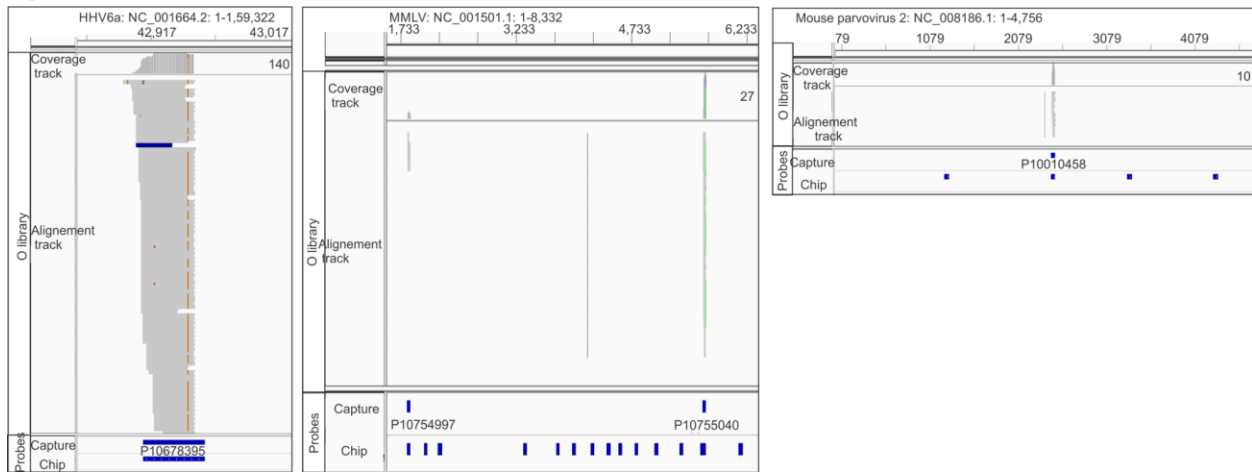


Figure S1b

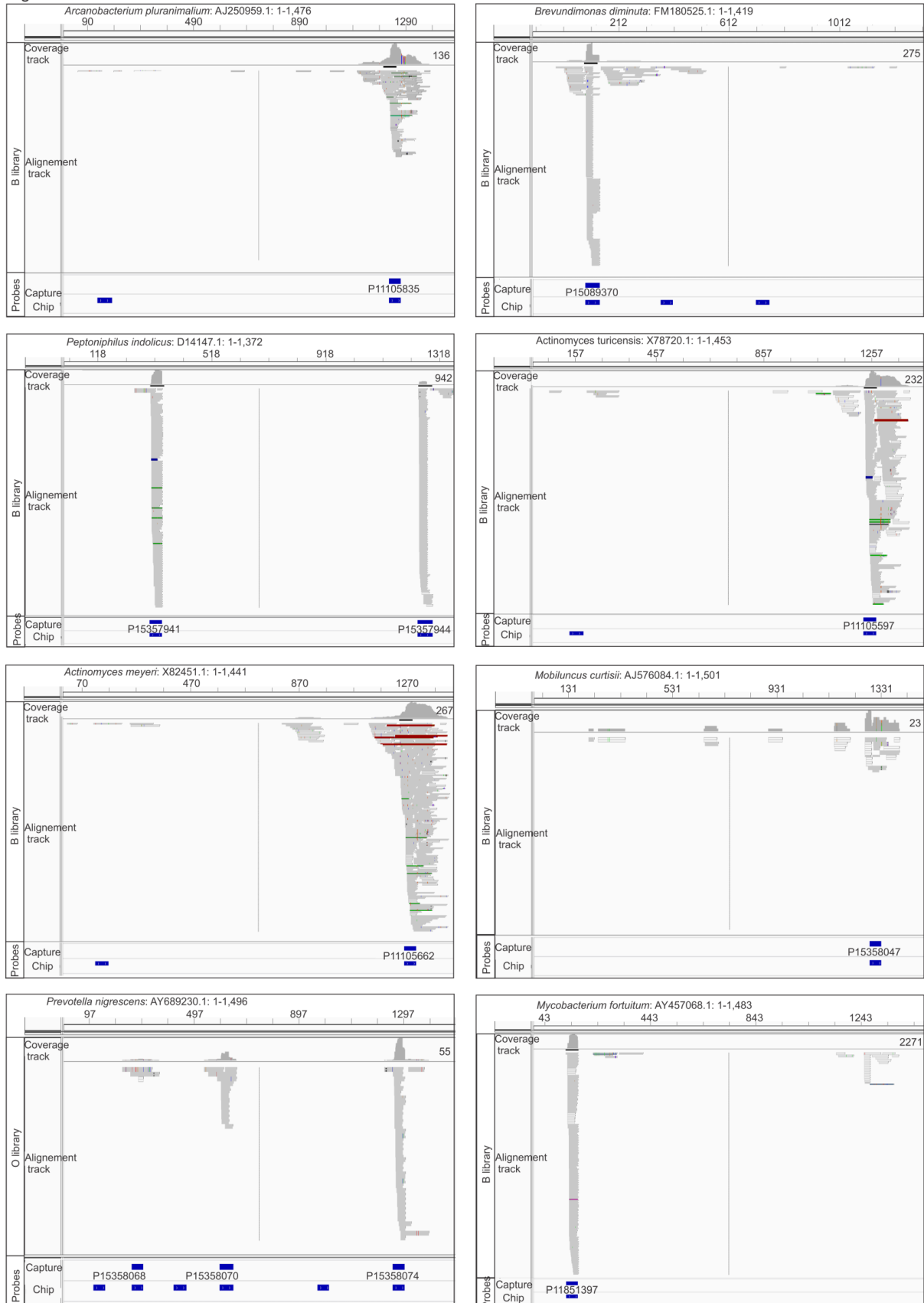


Figure S1c

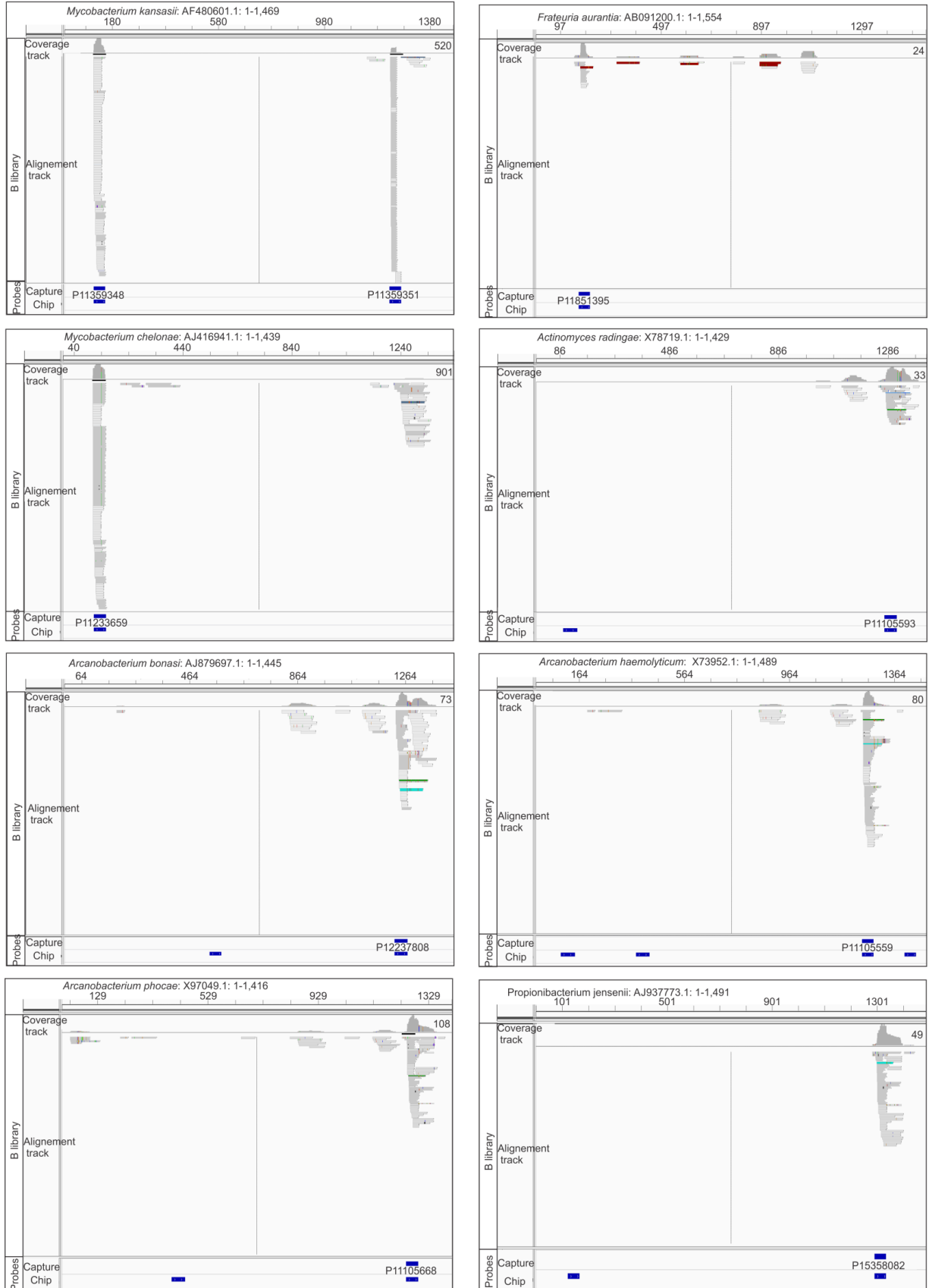


Figure S1d

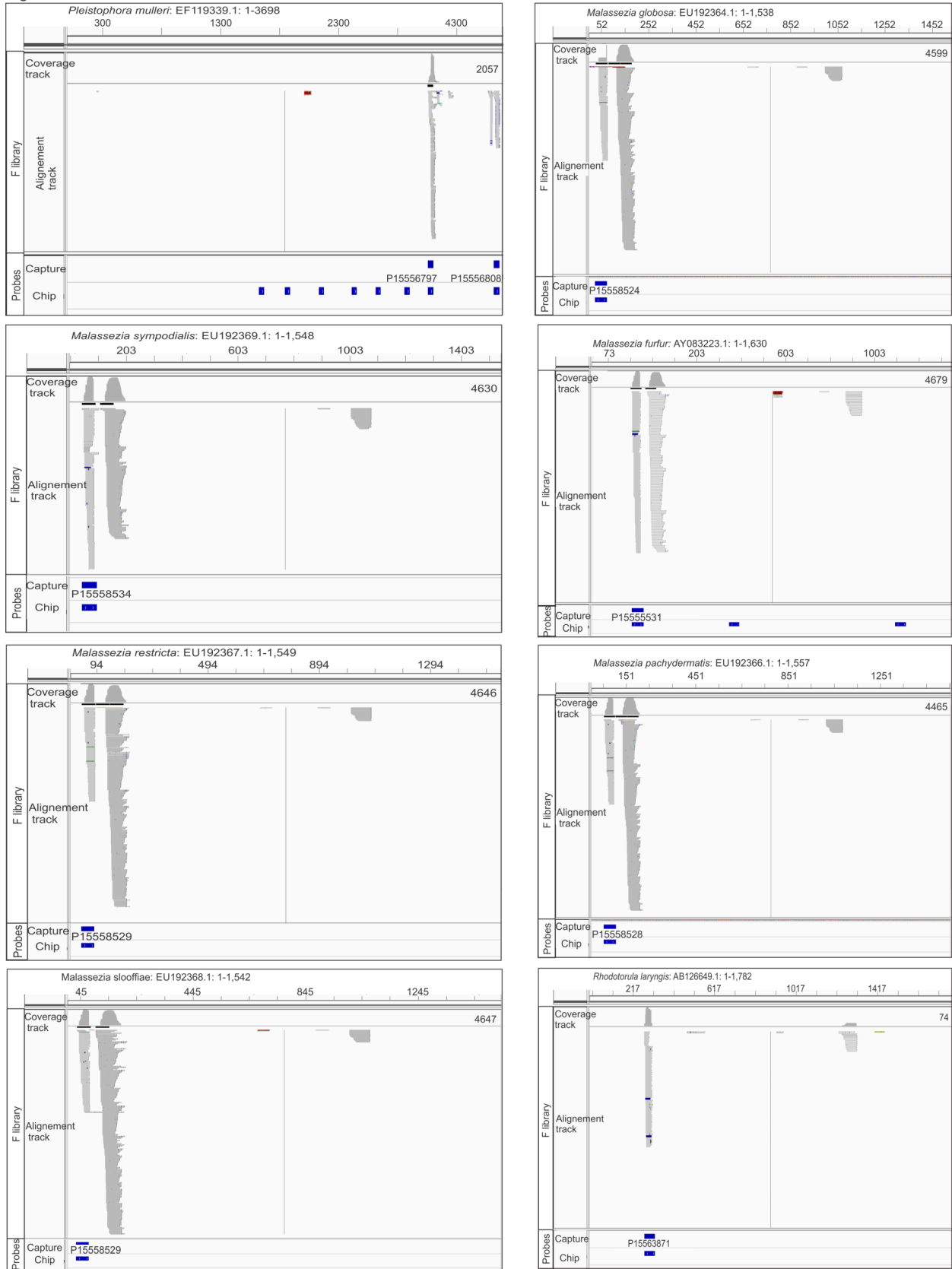


Figure S1e

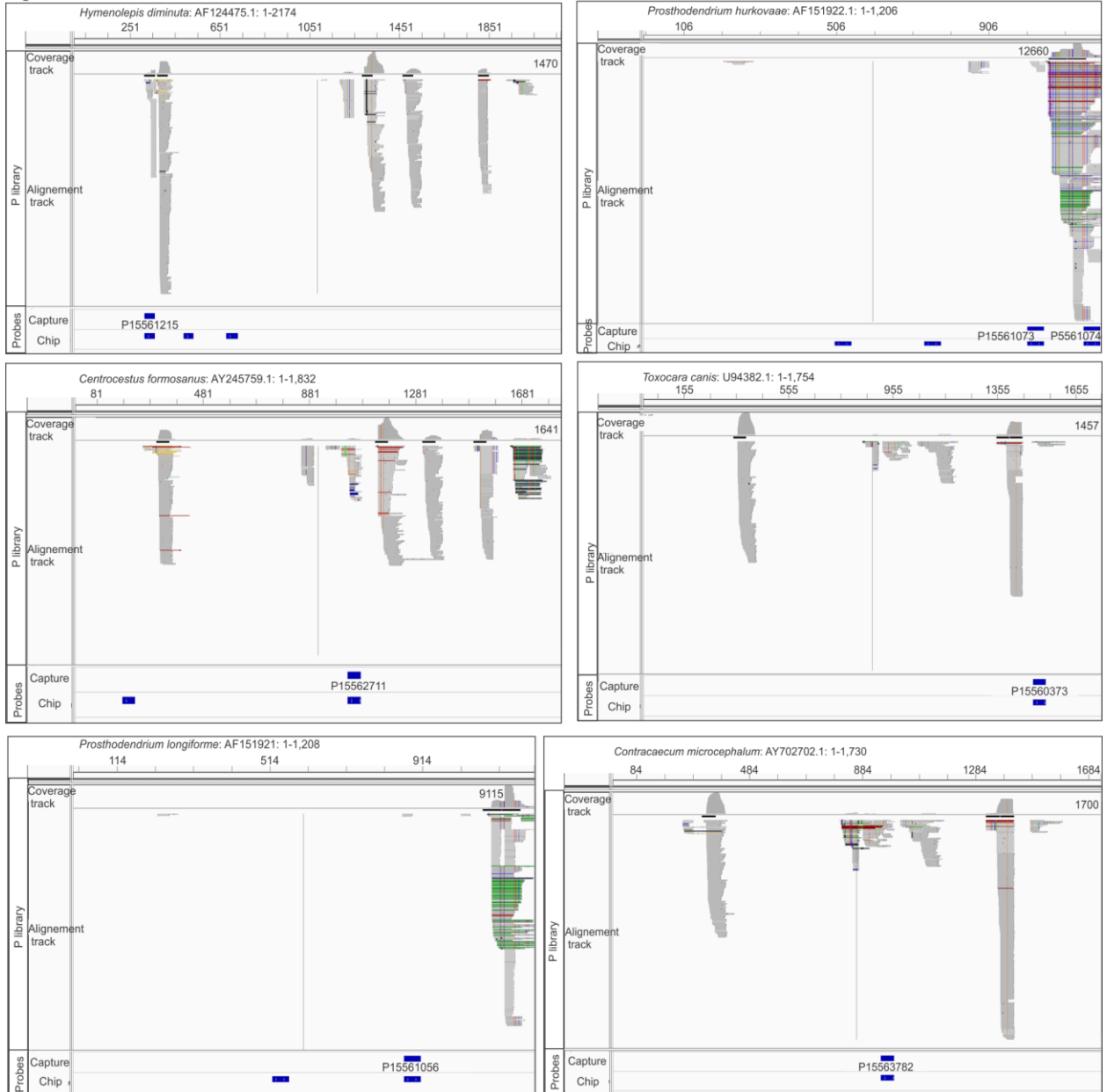
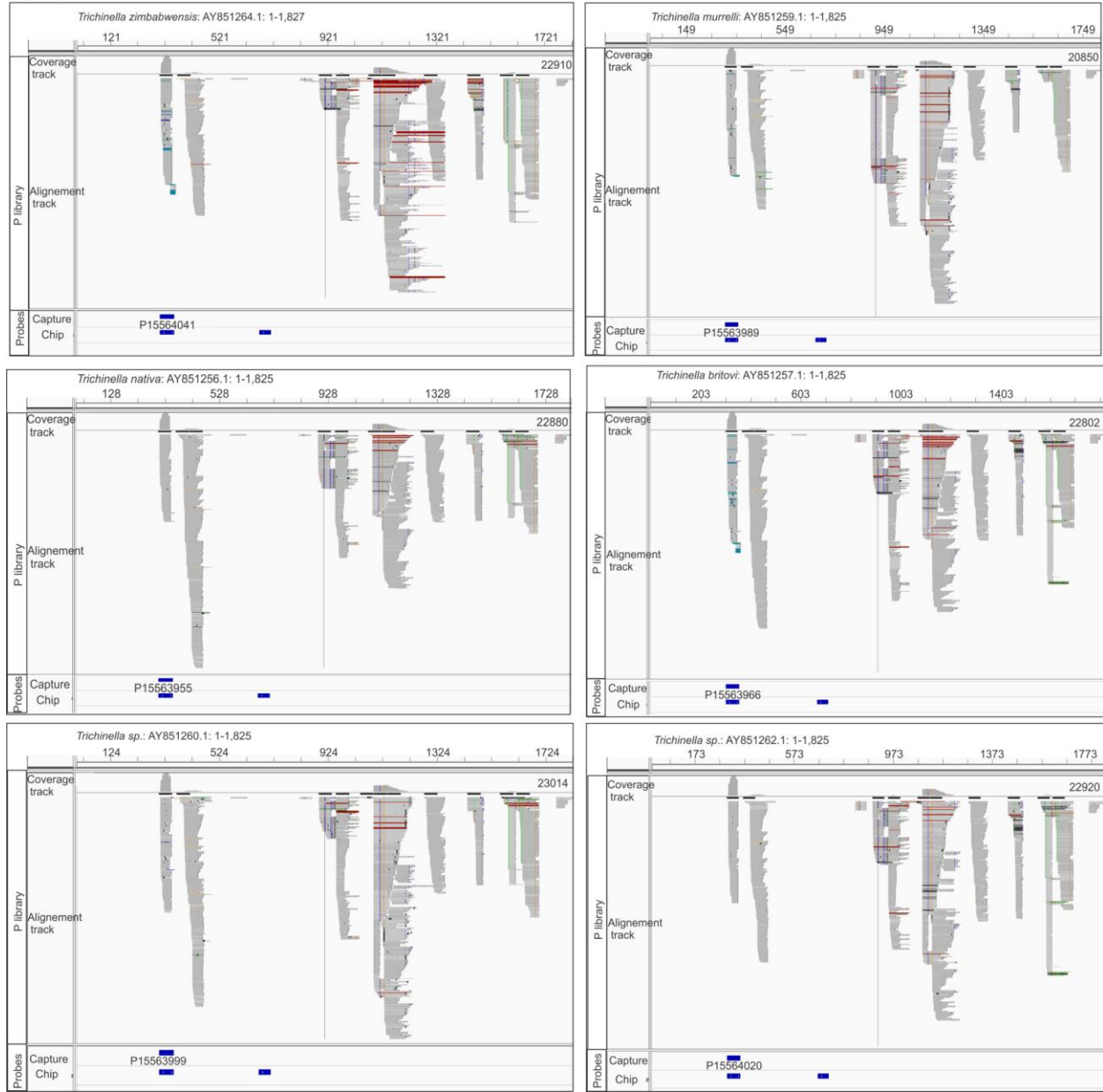
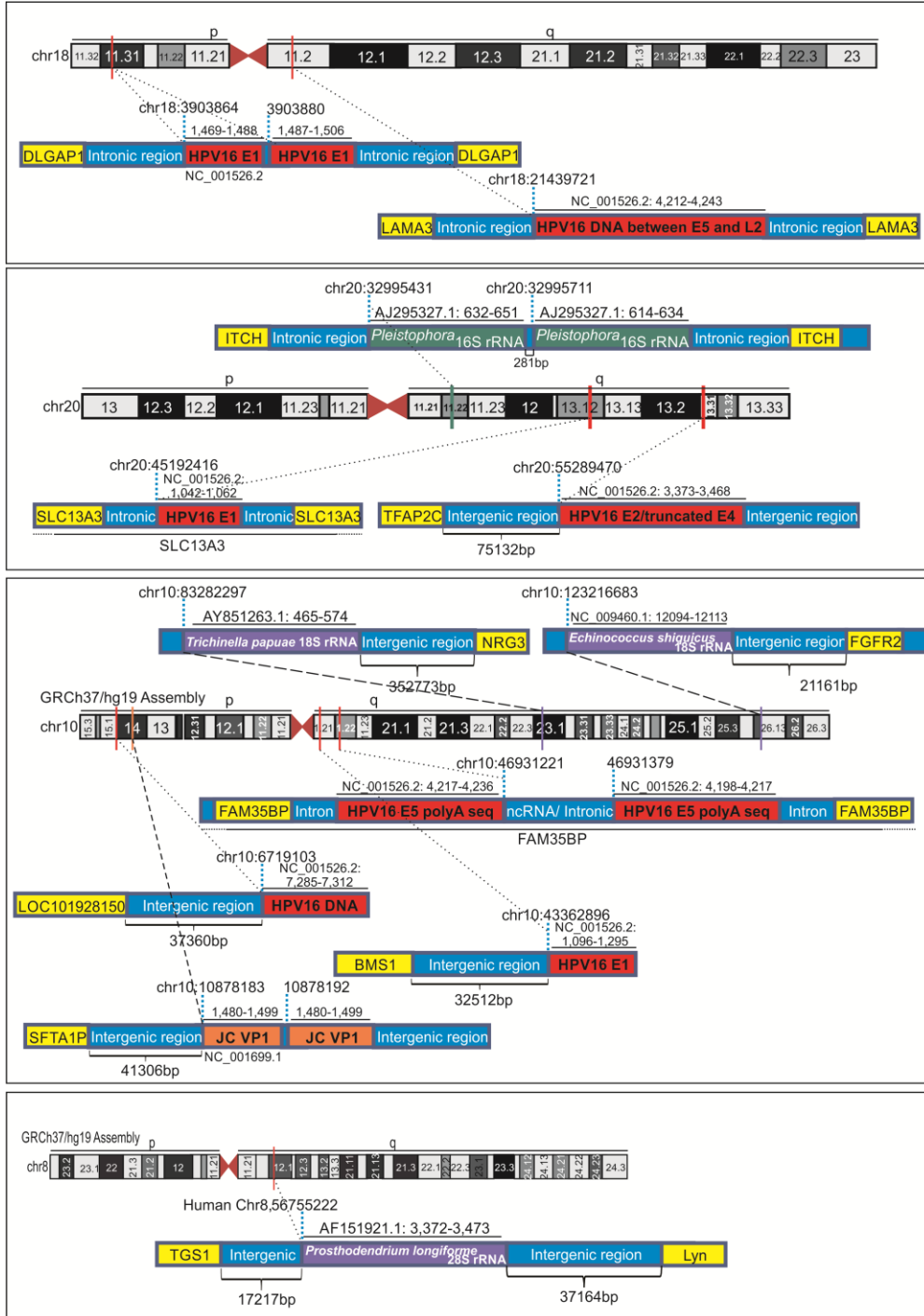


Figure S1f



**Figure S2.** Schematic representation of viral, bacterial, fungal and parasitic genomic insertional sites in human chromosomes. The genomic co-ordinates of the pathogens integrated and that of the host chromosome integration sites are mentioned. The co-ordinates for human chromosomes are from GRCh37/hf19 Assembly.

Figure S2



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**Supplemental table legends.**

**Table S1.** Tumor location and HPV16 positivity status of the OSCC samples screened by PathoChip.

**Table S2.** List of the capture probe sequences used for capture sequencing.

**Table S3.** Microbial signatures detected in OSCC and control samples.

**Table S4.** MiSeq reads aligning to capture probe locations. The total number of MiSeq reads per capture pool for each accessions are mentioned. For example Bcounts for each accession designates the no. of reads in B library that aligned to the particular accession, of which Bcounts.probe designates the no. of reads that aligned to the capture probe location of that particular accession and Bcounts.not.probe designates the no. of reads aligning outside the capture probe region for that particular accession. Reads are mentioned similarly for F, P, O and HPV16 library as well.

**Table S5.** Statistical significance between cluster 1 and cluster 2 oral cancer samples that are clustered by NBClust software. The significant differences between the clusters observed by these methods were determined using two-sided t-test.

**Table S6.** Statistical significance between 'group a', 'group b' and 'un-ungrouped' oral cancer samples that are obtained by topological-based data analyses using Ayasdi software. The significant differences between the clusters observed were determined using two-sided t-test.

**Table S7-S10.** Gene fusion results. List of viral (Table S7), bacterial (Table S8), fungal (Table S9) and parasitic (Table S10) genomic regions integrated either in intronic, exonic, downstream or upstream of certain human genomic regions. Point of integration with genomic co-ordinates, both for the pathogen and human genome are mentioned. The number of reads for such kind of fusion are also mentioned. The description of the gene fusion results are as follows: Column A-H: type of left/right element of fusion fragment (micro-organism or human), left/right chromosome name (pathochip genome or human), left/right start and end co-ordinate. Column I: number of reads supporting fusion; Column J: Gene\_region, indicating region (like intergenic, intronic, exon etc) of the fusion site in human genome; Column K: affected human gene name; Column L: Microbial genomic insertion position in human chromosome; Column M: Number of read support for human sequences; Column N: (Virus/bacteria/parasite/fungus).gene start coordinate of fusion organism in pathochip metagenome; Column O: (Virus/bacteria/parasite/fungus)\_ coordinate of fusion site in pathochip metagenome; Column P: integration site of (Virus/bacteria/parasite/fungus)\_coordinates at their genome; Column Q: (Virus/bacteria/parasite/fungus) region integrated; Column E: Accession number of microorganism integrated (description of accession).



Table S1

Study Code	Tumor location	Tumor type	p16 status
ENT01	tongue/tonsil	Squamous Cell Carcinoma (SCC)	Positive
ENT02	tongue/tonsil	SCC	Positive
ENT03	tonsil	SCC	Positive
ENT04	tonsil/base of tongue	SCC	Positive
ENT05	tonsil	SCC	Positive
ENT06	tonsil	SCC with sarcomatoid growth	Positive
ENT07	tonsil	SCC	Positive
ENT08	tonsil	SCC	Positive
ENT09	tonsil/tongue base	SCC	Positive
ENT10	floor of mouth/cheek	SCC	Negative
ENT11	tonsil/tongue base	SCC	Positive
ENT12	tonsil/tongue base	SCC	Positive
ENT13	tonsil	SCC	Positive
ENT14	oropharynx/palatine tonsil	SCC	Positive
ENT15	base of tongue	SCC	Negative
ENT16	tongue base	SCC	Positive
ENT17	tonsil/pharynx	SCC	Positive
ENT18	tonsil	SCC	Positive
ENT19	tonsil	SCC	Positive
ENT20	tonsil	SCC	Positive
ENT21	tongue base	SCC	Positive
ENT22	tonsil	SCC	Positive
ENT23	tonsil	SCC	Positive
ENT24	tongue base	SCC	Positive
ENT25	tongue base	SCC	Negative
ENT26	tonsil	SCC	Positive
ENT27	palatine tonsil	SCC	Positive
ENT28	oropharynx/base of tongue including lingual tonsil	SCC	Positive
ENT29	oropharynx	SCC	Positive
ENT30	Retromolar trigone	SCC with clear cell change	Positive
ENT31	oropharynx/palatine tonsil	SCC	Positive
ENT32	tongue/floor of mouth	SCC	Negative
ENT33	Tonsil	SCC	Positive
ENT34	tonsil/tongue base	SCC	Positive
ENT35	tonsil	Poorly differentiated carcinoma	Positive
ENT36	retromolar trigone/palate	SCC	Positive
ENT37	pharynx	SCC	Negative
ENT38	buccal mucosa	SCC	Negative
ENT39	tonsil	SCC	Positive
ENT40	tonsil	SCC	Positive
ENT41	base of tongue including lingual tonsil	SCC	Positive
ENT42	tonsil	SCC	Positive
ENT43	tonsil	SCC	Positive
ENT44	oropharynx, base of tongue including lingual tonsil	SCC	Positive
ENT45	oropharynx, palatine tonsil	SCC	Positive
ENT46	base of tongue, including lingual tonsil	SCC	Positive
ENT47	base of tongue including lingual tonsil	SCC	Negative
ENT48	tongue	SCC	Negative
ENT49	oropharynx/palatine tonsil, base of tongue including lingual tonsil	SCC	Positive
ENT50	palatine tonsil, base of tongue including lingual tonsil	SCC	Positive
ENT51	oropharynx/palatine tonsil, base of tongue including lingual tonsil	SCC	Positive
ENT52	tonsillar pillar	SCC	Negative
ENT53	oropharynx/palatine tonsil, base of tongue including lingual tonsil	SCC	Positive
ENT54	tongue/floor of mouth	SCC	Negative
ENT55	oropharynx/palatine tonsil	SCC	Positive
ENT56	oropharynx/soft palate	SCC	Positive
ENT57	tonsil	SCC	Positive
ENT58	oropharynx/base of tongue including lingual tonsil	SCC	Positive
ENT59	oropharynx/base of tongue including lingual tonsil	SCC	Positive
ENT60	oropharynx/palatine tonsil, base of tongue including lingual tonsil/pharyngeal wall (posterior)	SCC	Positive
ENT61	tonsil	SCC	Positive
ENT62	base of tongue including lingual tonsil	SCC	Positive
ENT63	oropharynx base of tongue, including lingual tonsil	SCC	Positive
ENT64	oropharynx base of tongue, including lingual tonsil	SCC	Positive
ENT65	oropharynx base of tongue, including lingual tonsil	SCC	Positive
ENT66	palatine tonsil	SCC	Positive
ENT67	oropharynx palatine tonsil base of tongue, including lingual tonsil	SCC	Positive
ENT68	tonsil, base of tongue	SCC	Positive
ENT69	hard palate	SCC	Positive
ENT70	tonsil	SCC	Positive
ENT71	tonsil	SCC	Positive
ENT72	tongue base	SCC	Positive
ENT73	tonsil, tongue base	SCC	Negative
ENT74	tonsil and tongue base	SCC	Positive
ENT75	tonsil	SCC	Positive
ENT76	oropharynx base of tongue, including lingual tonsil, pharyngeal wall (posterior)	SCC	Positive
ENT77	tonsil, tongue base, pharynx	SCC	Positive
ENT78	oropharynx palatine tonsil	SCC	Positive
ENT79	tonsil/tongue base	SCC	NA
ENT80	tongue base	SCC	NA
ENT81	tonsil/base of tongue	SCC	NA
ENT82	tongue base	SCC	Positive
ENT83	tonsil	SCC	Positive
ENT84	buccal mucosa	SCC	NA
ENT85	tongue	SCC	Negative
ENT86	oropharynx, tonsil	SCC	Positive
ENT87	oropharynx, tongue base	SCC	Positive
ENT88	tonsil/tongue base	SCC	Positive
ENT89	tonsil	SCC	Positive
ENT90	tonsillar pillar	SCC	Positive
ENT91	tongue base	SCC	Positive
ENT92	tongue base	SCC	Positive
ENT93	tonsil	SCC	Positive
ENT94	tonsil	SCC	NA
ENT95	left cheek and lip resection	SCC	NA
ENT96	Left radical tonsillectomy, tongue base resection	SCC	Positive
ENT97	Tongue, right	SCC	NA
ENT98	Left radical tonsillectomy, tongue base resection, limited pharyngectomy	SCC	Positive
ENT99	Left tonsil, left radical tonsillectomy, tongue base resection, limited pharyngectomy	SCC	Positive
ENT100	Right tonsillectomy, tongue base resection	SCC	NA

Table S2. Microbial signatures detected in OSCC and control samples.

	Cancer	MC	NC	Cancer/MC	Cancer/NC	Cancer/MC/NC	NC/MC
Viral signatures	<b>HPV16</b>	Coronaviridae	Togaviridae			Papillomaviridae	Adenoviridae
						Poxviridae	Picornaviridae
						Retroviridae	Flaviviridae
						Polyomaviridae	Hepadnaviridae
						Herpesviridae	Parvoviridae
						Reoviridae	Anelloviridae
						Orthomyxoviridae	
Bacterial signatures	<i>Eshcherichia</i>	<i>Campylobacter</i>	<i>Porphyromonas</i>	<i>Arcanobacterium</i>	<i>Mobiluncus</i>	<i>Mycobacterium</i>	
	<i>Rothia</i>	<i>Streptobacillus</i>		<i>Actinomyces</i>	<i>Propionibacterium</i>	<i>Sphingobacterium</i>	
	<i>Peptoniphilus</i>			<i>Aeromonas</i>	<i>Prevotella</i>	<i>Streptococcus</i>	
	<i>Brevundimonas</i>			<i>Bordetella</i>	<i>Citrobacter</i>		
	<i>Comamonas</i>			<i>Aerococcus</i>	<i>Sphingomonas</i>		
	<i>Alcaligenes</i>			<i>Pediococcus</i>	<i>Peptostreptococcus</i>		
	<i>Caulobacter</i>			<i>Acinetobacter</i>			
	<i>Cardiobacterium</i>			<i>Actinobacillus</i>			
	<i>Plesiomonas</i>			<i>Veillonella</i>			
	<i>Serratia</i>						
	<i>Edwardsiella</i>						
	<i>Haemophilus</i>						
<i>Frateuria</i>							
Fungal signatures	<i>Rhodotorula</i>	<i>Pseudallescheria</i>	<i>Piedraia</i>	<i>Fonsecaea</i>		<i>Malassezia</i>	<i>Microsporium</i>
	<i>Geotrichum</i>		<i>Trichosporon</i>			<i>Pleistophora</i>	<i>Alternaria</i>
	<i>Pneumocystis</i>		<i>Encephalitozoon</i>			<i>Absidia</i>	
			<i>Trichophyton</i>			<i>Phialophora</i>	
			<i>Paecilomyces</i>			<i>Cladophialophora</i>	
		<i>Madurella</i>			<i>Cladosporium</i>		
Parasitic signatures	<i>Hymenolepis</i>	<i>Echinococcus</i>	<i>Dipylidium</i>		<i>Prosthodendrium</i>	<i>Toxocara</i>	<i>Diphyllobothrium</i>
	<i>Centrocestus</i>		<i>Anisakis</i>		<i>Contracaecum</i>		<i>Strongyloides</i>
	<i>Trichinella</i>		<i>Echinostoma</i>				

Table S3

Capture Probe Library	Microorganisms	Probe sequences used for capture 5'-3'	
B (Bacterial pool)	A250959.1  <i>Arcanobacterium</i> sp.	/5Biosg/GAATCCCTTAAAGCCAGTCTCAGTTCGGATTGGGGTCTGCAACTC	
	X73952.1  <i>Arcanobacterium haemolyticum</i>	/5Biosg/GAATCCCTTAAAGCCAGTCTCAGTTCGGATTGGGGTCTGCAACTC	
	X97049.1  <i>Arcanobacterium phocae</i>	/5Biosg/GAATCCCTTAAAGCCAGTCTCAGTTCGGATTGGGGTCTGCAACTC	
	A879697.1  <i>Arcanobacterium</i> sp.	/5Biosg/CTAATCACTTAAAGTCTGCTCAGTTCGGATTGGGGTCTGCAACTCGAC	
	AJ576084.1  <i>Mobiluncus curtisii</i>	/5Biosg/CAACTCGACCCCATGAAGTTGGAGTCGTAGTAAATCGAGATCAG	
	J01859.1  <i>Escherichia coli</i>	/5Biosg/TAAACTCAATGAATTGACGGGGCCGCCACAAGCGGTGGAGCATGTGGTTTAAATTCGA	
	X78719.1  <i>Actinomyces radingae</i>	/5Biosg/ATCGGGTCTGCAACTCGACCTCGTGAAGGTGGAGTCTGAGTAAAT	
	X78720.1  <i>Actinomyces turicensis</i>	/5Biosg/ATATCGTAAGGTGGAGCGAATCACTTAAAGCCAGTCTCAGTTCGGATTG	
	X82451.1  <i>Actinomyces meyeri</i>	/5Biosg/GAATCCCTTAAAGCCAGTCTCAGTTCGGATTGGGGTCTGCAACTC	
	AJ171364.1  <i>Rothia dentocariosa</i>	/5Biosg/CTAATCCCTAAAGCCGGTCTCAGTTCGGATTGGGGTCTGCAACT	
	AJ937773.1  <i>Propionibacterium jensenii</i>	/5Biosg/GAATCTCAAAAAGCCAGTCTCAGTTCGGATTGGGGTCTGCAACTC	
	D14147.1  <i>Peptoniphilus indolicus</i>	/5Biosg/CCTGAGAGGGTGAACGGCCACATTTGAACTGAGAAACGGTCCAAA	
	D14147.1  <i>Peptoniphilus indolicus</i>	/5Biosg/GATAGCAAGCAATCTCAAAAAGCCTATCTCAGTTCGGATTGTTCTGCAACT	
	AF480601.1  <i>Mycobacterium kansasii</i>	/5Biosg/CGGGATAAGCCTGGGAACTGGGTCTAATACCGGATAGGACCACT	
	AF480601.1  <i>Mycobacterium kansasii</i>	/5Biosg/CGAGGTTAAGCGAATCTCTTTAAAGCCGGTCTCAGTTCGGATCGG	
	AJ416941.1  <i>Mycobacterium chelonae</i>	/5Biosg/ATAAGCCTGGGAACTGGGTCTAATACCGGATAGGACCACACTT	
	AY457068.1  <i>Mycobacterium fortuitum</i>	/5Biosg/CACITTTGGGATAAGCCTGGGAACTGGGTCTAATACCGAATAGGAC	
	FM180525.1  <i>Brevundimonas diminuta</i>	/5Biosg/GAGTTGCAGAGGACAATCCGAATGAGACAATTTTAAAGATTAAACCTCTGTAG	
	F (Fungal pool)	AB091200.1  <i>Frateuria aurantia</i>	/5Biosg/GAACTTACGCTAATACCCATACGCTACGGGAGAAGCAGG
		AB126649.1  <i>Rhodotorula laryngis</i>	/5Biosg/CTTCATTCAAATACCTGCCCTATCAACTTTCGATGGTAGAGGCCCTAC
DQ325447.1  <i>Geotrichum citri-aurantii</i>		/5Biosg/AGAGCTTTCATGATTTTGTGGTGGTGGTGCATGGCCGTCTTCTAGTGTGGAGTGT	
EU192364.1  <i>Malassezia globosa</i>		/5Biosg/GTCTGGCGATCTCTCAAATATCGCCCTATCAACTCTCACTGTCGATGGATG	
AY083223.1  <i>Malassezia furfur</i>		/5Biosg/TTTCATTCAAATATCGCCCTATCAACTCTGATGGTAGGATAGAGCCCTACCAT	
EU192366.1  <i>Malassezia pachydermatis</i>		/5Biosg/ATGCTTCATTCAAATATCGCCCTATCAACTCTGATGGTAGGATAGAGGCCCTA	
EU192367.1  <i>Malassezia restricta</i>		/5Biosg/CTGGCGGTCTCTCAAATATCGCCCTATCAACTCTCACTGTCGATGGATG	
EU192368.1  <i>Malassezia slooffiae</i>		/5Biosg/CGGGGTGCTTCATTCAAATATCGCCCTATCAACTCTGATGGTAG	
EU192369.1  <i>Malassezia symyodialis</i>		/5Biosg/ATGCTTCATTCAAATATCGCCCTATCAACTCTGATGGTAGGATAGAGGCCCTA	
EF119339.1  <i>Pleistophora mulleri</i>		/5Biosg/GAGGAAATCACTGTAAAGCAGCGGTTAACGGCGGAGTAACATGACT	
EF119339.1  <i>Pleistophora mulleri</i>		/5Biosg/GCAGAAAGCGACGGTCTTTTGTATTCTCGATTCGACTCTCTATCATG	
AF044392.1  <i>Pleistophora ehrenbaumi</i>		/5Biosg/AAAGCACAGGAAGGAGGGTCAAGAAACAGTCACTGATGCCCTAG	
P (Parasitic pool)	AB266392.1  <i>Pneumocystis jirovecii</i>	/5Biosg/CTGTGCTGGCGATGGTTCATTCAAATATCGCCCTATCAACTCTGATG	
	AF124475.1  <i>Hymenolepis diminuta</i>	/5Biosg/CTTCAAATGTCTGCCCTATCAACTTTCGATGGTAGGTCACCTGCCCTAC	
	AY245759.1  <i>Centrocestus formosanus</i>	/5Biosg/AAAGTCAGAGGTTGCAAGAGCAGTCACTCCGCTAGTTCGACCAATAAC	
	AF151921.1  <i>Prosthodendrium longiforme</i>	/5Biosg/GTCTTGAACACCGGACCAAGGAGAGTAACATGCGCGCAGTCACT	
	AF151922.1  <i>Prosthodendrium hurkovaee</i>	/5Biosg/GAGCGGTAGGCGCATCACCGCCCGTCCATGACAATTGTTCTTG	
	AF151922.1  <i>Prosthodendrium hurkovaee</i>	/5Biosg/CCGAAGCGGTTCTGACGTGCAAACTGATCGCCAGACATGAGTATA	
	AY851256.1  <i>Trichinella nativa</i>	/5Biosg/ACATCACTTTCAAGTGTCTGCCCTATCAACTCTCGATGGTAGTCTATGCGACTA	
	AY851257.1  <i>Trichinella britovi</i>	/5Biosg/ACATCACTTTCAAGTGTCTGCCCTATCAACTCTCGATGGTAGTCTATGCGACTA	
	AY851259.1  <i>Trichinella murrelli</i>	/5Biosg/ACATCACTTTCAAGTGTCTGCCCTATCAACTCTCGATGGTAGTCTATGCGACTA	
	AY851260.1  <i>Trichinella</i> sp.	/5Biosg/ACATCACTTTCAAGTGTCTGCCCTATCAACTCTCGATGGTAGTCTATGCGACTA	
	AY851262.1  <i>Trichinella</i> sp.	/5Biosg/ACATCACTTTCAAGTGTCTGCCCTATCAACTCTCGATGGTAGTCTATGCGACTA	
	AY851263.1  <i>Trichinella papuae</i>	/5Biosg/ACATCACTTTCAAGTGTCTGCCCTATCAACTCTCGATGGTAGTCTATGCGACTA	
O (Other pool)	AY851264.1  <i>Trichinella zimbabwensis</i>	/5Biosg/ACATCACTTTCAAGTGTCTGCCCTATCAACTCTCGATGGTAGTCTATGCGACTA	
	AY702702.1  <i>Contraecaecum microcephalum</i>	/5Biosg/GATCAGATACCCCTAGTTCGACCGTAACACGATCAACCACTAGCGTT	
	U94382.1  <i>Toxocara canis</i>	/5Biosg/TAACCCGTTGAAATCCTCGTGTGCGGATCGGGAATGCAATTAATTT	
	NC_008186.1  Mouse parvovirus 2	/5Biosg/AAAGACGCTTAAAGACTGGGGAGGCAAGGTTGGTCACTCTTTTAGAA	
	NC_001664.2  Human herpesvirus 6A	/5Biosg/TATAGCAGCGCCGATGCCAACCGCAGAGGTTGATCAGTACATAGAT	
	NC_001664.2  Human herpesvirus 6A	/5Biosg/CAATACAGAGTGTGTTTGGCAAATTTAAGCAGCAATTTGTTGATCTAAATTAAGG	
	NC_006879.1  Simian adenovirus 1	/5Biosg/TCTTGATCAGTCTTCCACCGGAACTCTGCTTCCCTGTCT	
	NC_006879.1  Simian adenovirus 1	/5Biosg/AGAGGCCAGAGAGGAGCAACTGGCAAAGCAGCTGAACAGCATC	
	NC_002657.1  Classical swine fever virus	/5Biosg/ATTGGGGAACAAGCCAGAGAAAGGGGGAGATTGGGAGAGTAAAG	
	NC_001699.1  JC polyomavirus	/5Biosg/TAACAGCCAGTAAACAAAGCACAAGGGGAAGTGGAAAGCAGCCAA	
	NC_001699.1  JC polyomavirus	/5Biosg/AAAGGGAGGGAACCTATATTTCTTTGGCCACTCATACCCAAAGTATAGATGATG	
	NC_001699.1  JC polyomavirus	/5Biosg/TTATAGGGGTGACAAGTTTGTATGATGTGCACTCTAATGGGCAAGCACTCAT	
HPV16 pool	NC_001664.2  Human herpesvirus 6A	/5Biosg/CTAAAACCGTCTTGCCTCTCAAATCTTTAATGAGAAAAACGTTACGGGGTAAAC	
	NC_001664.2  Human herpesvirus 6A	/5Biosg/ACATTAATACAGGCAAGGACACAATTCACGGCATTCTTGAAGCCGTGAC	
	NC_001664.2  Human herpesvirus 6A	/5Biosg/CTATCAGAATTACGAACGCCCGATAAACTAATCCGCGCATCAAAATTAAT	
	NC_001664.2  Human herpesvirus 6A	/5Biosg/AAACTTACTCATGTTTCTGCAGTCTCACTTTCGAAGACATTCAGTTCACG	
	NC_001664.2  Human herpesvirus 6A	/5Biosg/AAGTGGTAACTTAAATCGGAGTGGTGTGTCGGCGTGGTGAATTTT	
	NC_001664.2  Human herpesvirus 6A	/5Biosg/CGATGACCGCTAAATTTGATAAATGATCTGCTCCACTCCGATTCAAAGAAA	
	NC_001664.2  Human herpesvirus 6A	/5Biosg/TTAGCAATTTTAAAGTTTCTAACCCGCTGGGTAGGAAAGACCTCAACCAC	
	NC_001501.1  Moloney murine leukemia virus	/5Biosg/GATAGACAGGAGGAGAACGAAGGAGTCCCAACTCGATCGCGAC	
	NC_001501.1  Moloney murine leukemia virus	/5Biosg/GTAAACCAATGGAGATCGGGAGACGGTATGGGCAACTCTGGCAAC	
	NC_014480.2  Torque teno virus 2	/5Biosg/AAAACACAGAATCACAGTCAGAATAGGAGCCCTAAACCTTTGAGGACAAG	
	NC_014480.2  Torque teno virus 2	/5Biosg/GAGACAGGAGACAGCCATAAACACCAACGCTAGACTGACACTAATATGC	
	NC_001583.1  Human papillomavirus type 26	/5Biosg/CCTATGATGTTATTAATGTGCCAGAAACCAATTAATTAAGCAACCAAAATTCGG	
HPV16 pool	NC_001583.1  Human papillomavirus type 26	/5Biosg/TGTTGCAATGGGCGTTCGATCATGACATAACAGATGATAGTGAATTTGCAATATG	
	NC_015961.1  Tomato chlorotic leaf distortion virus	/5Biosg/TATGGCTCTCTCTCTGTCACCTGGCCGATTATAGACCATGGTCTAAATAA	
	NC_010729.1  <i>Porphyromonas gingivalis</i>	/5Biosg/GGGGAGAGGTAAGAGGACTAAGGAGCAGAACTTATGAATAGAAAATAGTATGCA	
	AY689230.1  <i>Prevotella nigrescens</i>	/5Biosg/CAACGGCCCAACGAAGCCAGCAGTCACTAGGGGTTCTGAGAGGAAG	
	AY689230.1  <i>Prevotella nigrescens</i>	/5Biosg/TGTTGTGAATTTAGGTGCTCAACATTTAACTTGCAGCGCAACTGTCAAGCTT	
	AY689230.1  <i>Prevotella nigrescens</i>	/5Biosg/TAATGCAAGTGTCACTCAACTTGAATCCGGTCCAGTTCGGACT	
	NC_001526.2  Human papillomavirus type 16	/5Biosg/TGCAAGGCAAGTGTGCAAAAATTTAAAGAAATACGGGGTGGATTTTCAGAAAT	
	NC_001526.2  Human papillomavirus type 16	/5Biosg/CAACGAAGTATCTCTCTGAAATTTAATAGCAGCACTTGCCCAACAC	
	NC_001526.2  Human papillomavirus type 16	/5Biosg/CCTTGGGACCCGAAAGAACACAGACGACTTCCAGCGACCAAGAT	
	NC_001526.2  Human papillomavirus type 16	/5Biosg/GGAATTTTGGTCTACAACCTCCCGAGGAGGACACTAGAAGATA	

