

Type of file: PDF

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Title of file for HTML: Supplementary Information

Description: Supplementary Figures and Supplementary Table.

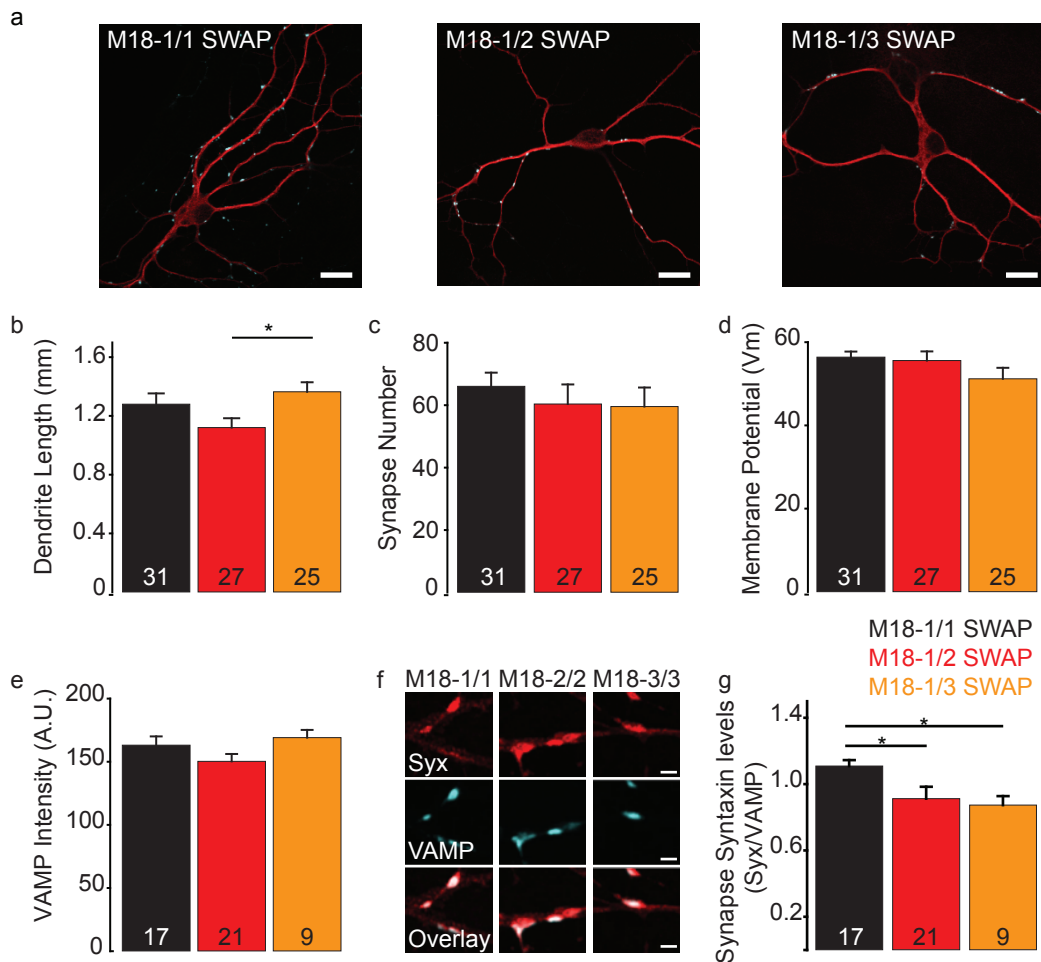
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## Supplementary Figure. 1



### Supplementary Figure1

#### Viral expression of Munc18-2 or -3 rescues munc18-1 null mutant neurons and restores neuronal viability and morphology

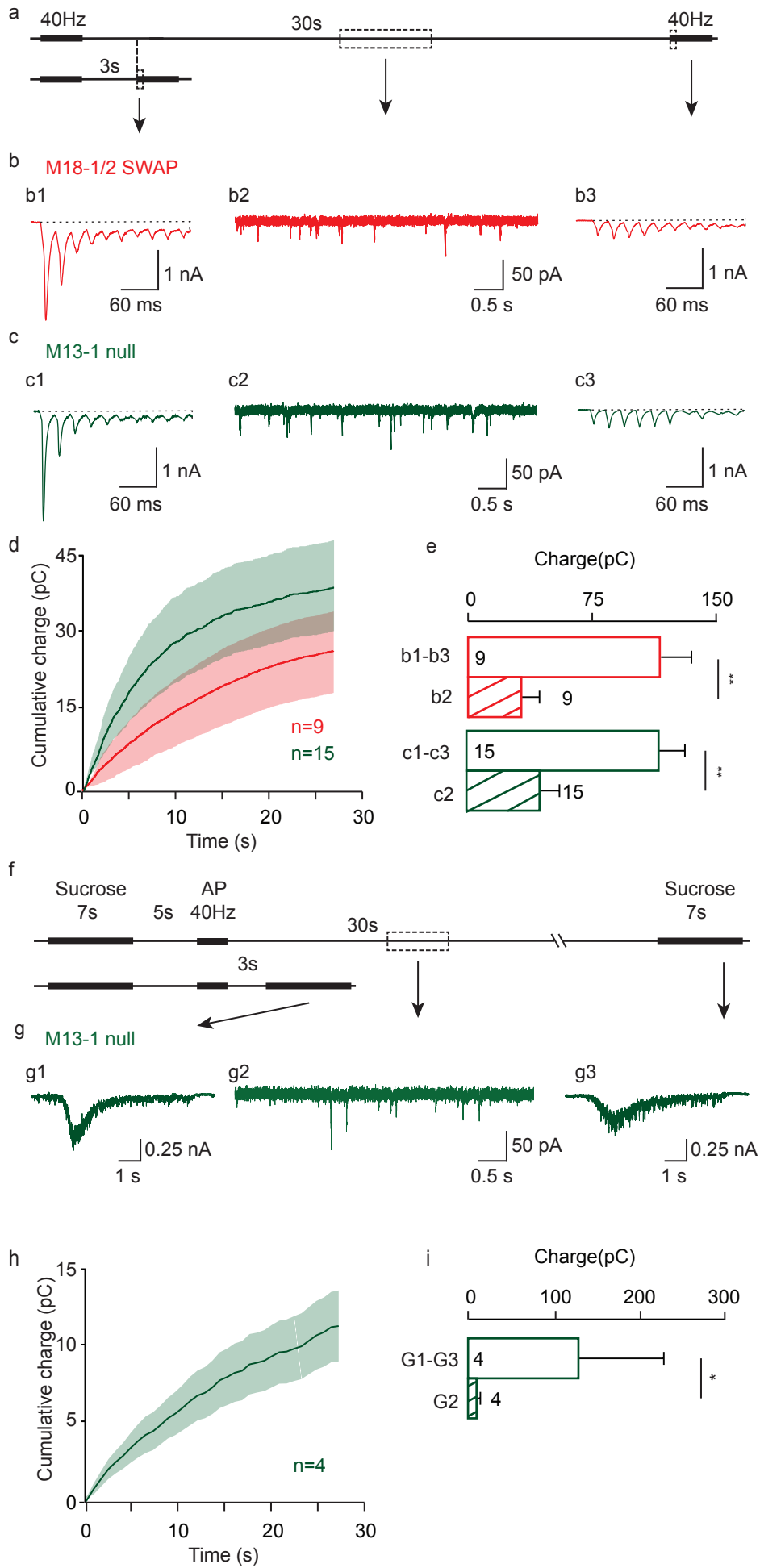
(a) Typical examples of null mutant neurons infected with M18-1, -2, -3 viral particles at DIV1 and stained for MAP2 (dendritic maker) and synapsin (presynaptic terminal marker) at DIV14. Scale bar represents 50  $\mu$ m.

(b, c) Total dendrite length (b, synapse number (c) and membrane potential (d) of autaptic null mutant neurons expressing M18-1, -2, -3.

(e, f, g) VAMP/synaptobrevin expression level (e), example of syntaxin and VAMP co-staining (f) and syntaxin levels compared to VAMP in autaptic null mutant neurons expressing M18-1, -2, -3. Scale bar represents 2  $\mu$ m.

All data in this figure are means  $\pm$  SEMs; \* $p$  < 0.05, \*\* $p$  < 0.01, \*\*\* $p$  < 0.001 as determined by ANOVA. See Table S1 for all values, SEMs and n-numbers plotted in this figure.

# Supplementary Figure. 2



## Supplementary Figure2

### Primed vesicles rapidly de-prime in the absence of Munc13-1

(a) Paradigm to assess de-priming by dual 40Hz train stimulation (see methods for details).

(b, c) sample traces of dual 100 stimuli at 40Hz with 3s or 30s interval in *munc18-1/2SWAP* (b) and *munc13-1 null* (c) synapses.

(d) Spontaneous fusion events were quantified for the 3-30s intervals.

(e) To quantify de-priming, the total charge of the first 10 responses (b1, b3; c1, c3) of each dual 100 stimuli at 40Hz was quantified. The spontaneous fusion of vesicles in the 3-30s intervals (b2; c2) cannot explain the loss of fusion-competent vesicles after 30s, defined as the difference in total charge between 3s and 30s intervals (b1-b3; c1-c3).

(f) Paradigm to assess de-priming by dual 500 mM hypertonic sucrose application.

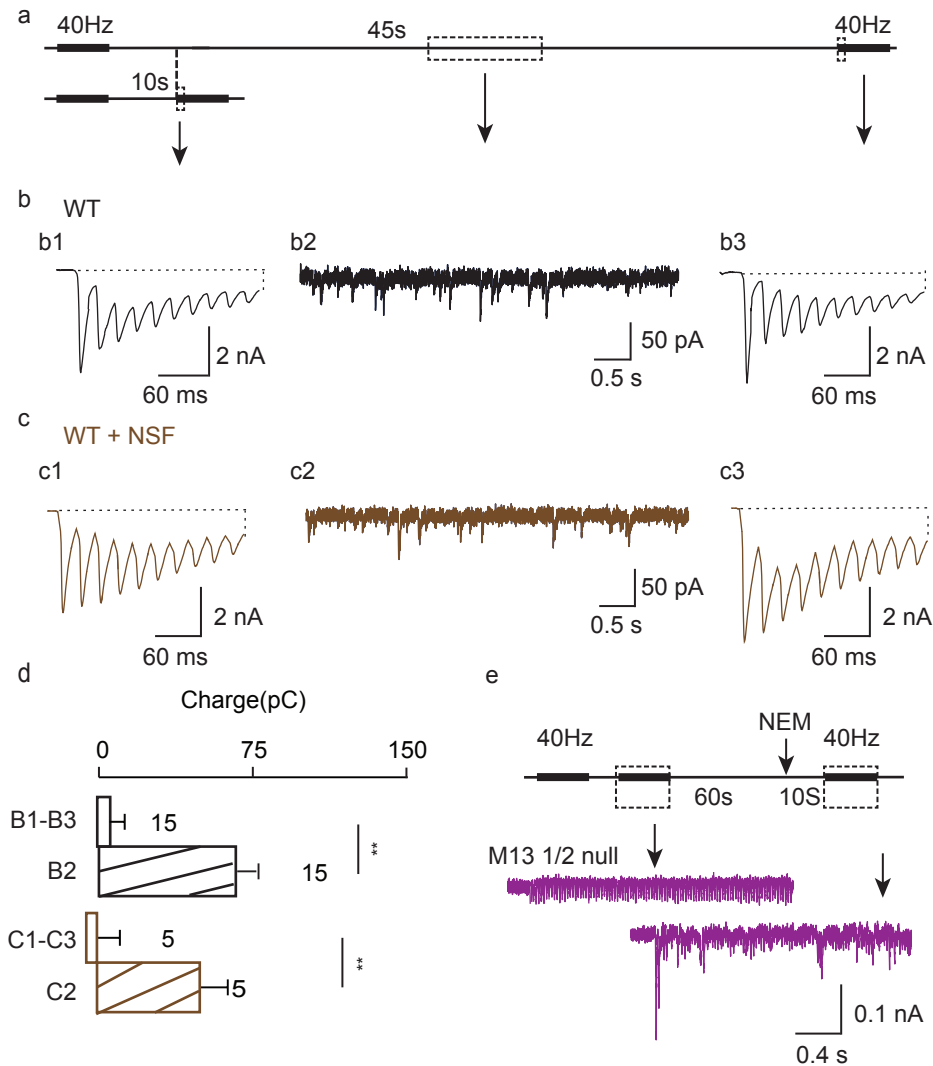
(g) Sample traces of dual sucrose application with 3s or 30s interval in *munc13-1 null* synapses. (h) Spontaneous fusion events were quantified for the 3-30s intervals.

(h) Spontaneous fusion events were quantified for the 3 - 30s intervals.

(i) To quantify de-priming, the total charge of the sucrose responses (g1, g3) after 100 stimuli at 40Hz was quantified. The spontaneous vesicle fusion in the 3-30s intervals (g2) cannot explain the loss of fusion-competent vesicles after 30s, defined as the difference in total charge between 3s and 30s intervals (g1-g3).

All data in this figure are means  $\pm$  SEMs; p value determined by Wilcoxon signed rank test. See supplementary Table 1 for all values, SEMs and n-numbers plotted in this figure.

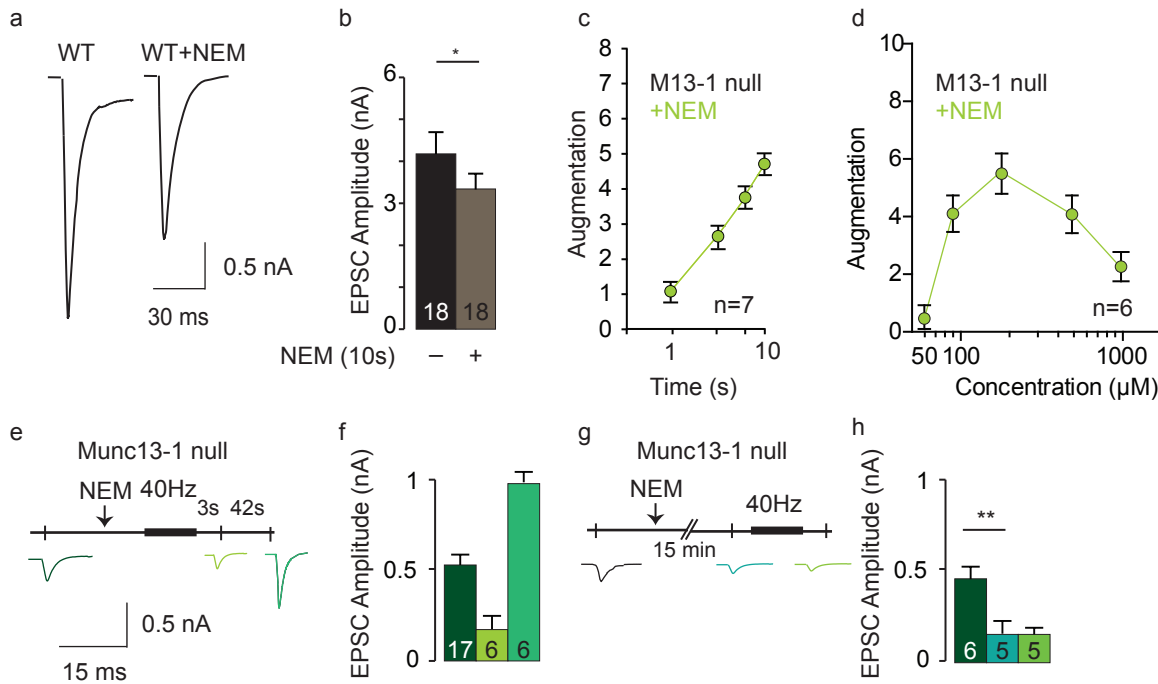
## Supplementary Figure. 3



### Supplementary Figure3 de-priming is undetected in the WT and NSF over-expressed WT neurons

- (a) Paradigm to assess de-priming by dual 40Hz train stimulation (see methods for details).
- (b, c) sample traces of dual 100 stimuli at 40Hz with 10s or 45s interval in WT (b) and NSF over-expressed WT (c) synapses.
- (d) To quantify de-priming, the total charge of the first 10 responses (b1, b3; c1, c3) of each dual 100 stimuli at 40Hz was quantified. The spontaneous fusion of vesicles in the 10-45s intervals (b2; c2) cannot explain the loss of fusion-competent vesicles after 45s, defined as the difference in total charge between 10s and 45s intervals (b1-b3; c1-c3).
- (e) Paradigm and sample traces of munc13-1/2 null with train stimulation and NEM application.
- All data in this figure are means  $\pm$  SEMs; p value determined by Wilcoxon signed rank test. See Table S1 for all values, SEMs and n-numbers plotted in this figure.

## Supplementary Figure. 4



## Supplementary Figure 4

### NEM effect on evoked EPSC in WT and munc13-1 null autaptic neurons

(a, b) Sample traces (a) and quantification (b) of EPSCs with or without 10 s NEM application.

(c) The effect of NEM on munc13-1 null neurons EPSC builds up during the 1<sup>st</sup> second after application and reaches highest level within 10s.

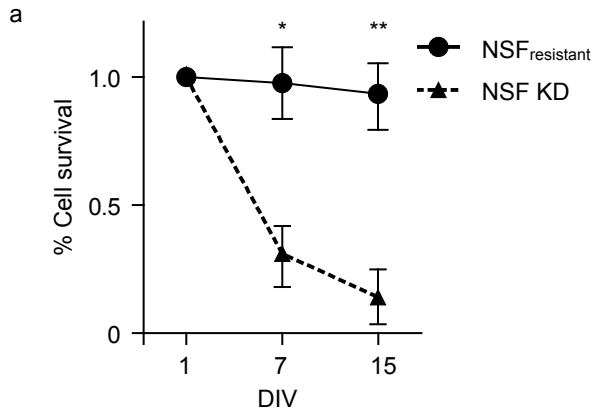
(d) NEM potentiates EPSC in a dose-dependent manner.

(e, f) After NEM application, EPSC potentiation recovered in 45 second after 100 stimuli with 40Hz; Paradigm and sample trace (e), and quantification (f).

(g, h) After NEM application, EPSC potentiation vanished after 15 min; 100 stimuli at 40Hz cannot re-potentiate EPSC ; Paradigm and sample trace (g), and quantification (h).

p values determined by paired t-test. See Table S1 for all values, SEMs and n-numbers plotted in this figure.

## Supplementary Figure. 5



### Supplementary Figure 5 NSF knock down by shRNA causes neuronal death

(a) Neuron viability was measured at various time after infection at DIV0. Black circle: neurons with NSF knock down by short hairpins and rescued by shRNA resistant NSF (n=10); Black triangle: neurons with NSF knock down by short hairpins only (n=10). All data in this figure are means  $\pm$  SEMs;

\* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$  as determined by student t-test.

**Supplementary Table 1**

	M18 -1/1SWAP	M18 -1/2SWAP	M18 -1/3SWAP	M13 -1 null	CAPS -1/2 null	M13 -1/2 null	WT
<i>number of docked vesicles</i>	6.38±0.26, n=56, fig.1G	4.6±0.26, n=54, fig.1G	2.69±0.23, n=65, fig.1G				
<i>active zone size</i>	284.5±15.9 nm, n=56, fig.1H	283.6±17.9 nm, n=54, fig.1H	312.9±16.3 nm, n=65, fig.1H				
<i>total number of vesicles</i>	79.3±4.7, n=56, fig.1I	72.4±5.2, n=54, fig.1I	72.8±4.8, n=65, fig.1I				
<i>EPSC amplitude</i>	3.7±0.1 nA, n=73, fig.1B 3.1±0.3 nA, n=9, *	0.42±0.07 nA, n=90, fig.1B 0.36±0.13 nA, n=48, fig.3F	0.1±0.02 nA, n=49, fig.1 0.07±0.03 nA, n=6, *	0.4±0.08 nA, n=119, fig.3B 0.51±0.14 nA, n=6, s fig.3F	0.75±0.19 nA, n=68, fig.3H 3.4±0.3 nA, n=68, fig.3H	0.003±0.003 nA, n=5, fig.3K 3.8±0.3 nA, n=51, *	4.1±0.8 nA, n=18, s fig.3B
<i>EPSC amplitude after 40 Hz</i>		1.8±0.3 nA, n=48, fig.3F		2.1±0.1 nA, n=119, fig.3B	3.4±0.3 nA, n=68, fig.3H		3.4±0.5 nA, n=51, *
<i>EPSC amplitude in presence of 100 μM NEM &lt;10s</i>	2.7±0.3 nA, n=10, *	2.2±0.2 nA, n=23, fig.3F	0.07±0.03 nA, n=6, *	2.0±0.2 nA, n=41, fig.3B	0.71±0.24 nA, n=17, fig.3H	0.15±0.07 nA, n=19, fig.3K	3.3±0.6 nA, n=18, s fig.3B
<i>EPSC amplitude in presence of 100 μM NEM &gt;15min</i>				0.6±0.1 nA, n=6, s fig.3F			0.2±0.05 nA, n=5, *
<i>mEPSC amplitude</i>	25.4±2.0 pA, n=51, fig.1E 21.3±2.5 pA, n=15, *	21.7±2.2 pA, n=68, fig.1E 18.9±3.4 pA, n=20, *	19.1±0.9 pA, n=27, fig.1E	26.1±2.8 pA, n=29, fig.3D	23.8±3.1 pA, n=17, *		22.4±5.1 pA, n=35, * 29.1±6.9 pA, n=16, *
<i>mEPSC amplitude in presence of 100 μM NEM &lt;10s</i>	24.5±6.2 pA, n=15, *	20±5.5 pA, n=20, *		25.4±3.2 pA, n=29, fig.3D	20.1±2.7 pA, n=17, *		27.6±7.2 pA, n=16, *
<i>mEPSC frequency</i>	17±1.6 Hz, n=51, fig.1D 16.5±4.1 Hz, n=15, *	2.1±0.9 Hz, n=68, fig.1D 2.9±1.2 Hz, n=20, *	0.1±0.04 Hz, n=27, fig.1D	1.6±0.4 Hz, n=29, fig.3D	2.1±0.7 Hz, n=17, *		10.3±4.6 Hz, n=35, * 8.7±3.9 Hz, n=16, *
<i>mEPSC frequency in presence of 100 μM NEM &lt;10s</i>	12.9±3.3 Hz, n=15, *	7.1±1.5 Hz, n=20, *		4.6±0.6 Hz, n=29, fig.3D	1.2±0.4 Hz, n=17, *		6.1±2.4 Hz, n=16, *
<i>mEPSC charge in 45s-10s</i>		23.9±3.4 pC, n=30, fig.2		20.4±2.8 pC, n=47, fig.2			
<i>first 10 EPSCs charge in second 40 Hz train (45s interval)</i>		51.8±9.0 pC, n=30, fig.2		83.2±8.4 pC, n=47, fig.2			
<i>first 10 EPSCs charge in second 40 Hz train (10s interval)</i>		126.4±11.4 pC, n=30, fig.2		158.5±15.4 pC, n=47, fig.2			
<i>mEPSC charge in 30s-3s</i>		27.9±8.4 pC, n=9, s fig.2		36.1±9.1 pC, n=15, s fig.2			
<i>first 10 EPSCs charge in second 40 Hz train (30s interval)</i>		49.2±11.6 pC, n=9, s fig.2		105.2±12.5 pC, n=15, s fig.2			
<i>first 10 EPSCs charge in second 40 Hz train (3s interval)</i>		155.1±18.8 pC, n=9, s fig.2		199.5±22.7 pC, n=15, s fig.2			
<i>mEPSC charge in 30s-3s</i>				11.7±2.8 pC, n=4, s fig.2			
<i>sucrose charge after 40 Hz train (30s interval)</i>				289.9±42.1 pC, n=4, s fig.2			
<i>sucrose charge after 40 Hz train (3s interval)</i>				371.5±51.4 pC, n=4, s fig.2			

\* not shown in figures