

Supplementary Figure 1

EAE induced by several immunization methods and phenotypes on cell migration, demyelination and nociceptive sensitivity in mice with Type-A and Type-B EAE.

(a) EAE severity in NIrp3^{-/-} mice was evaluated. EAE was induced with 6 different methods, as indicated in Online Methods. (b) AUC between 0 and 20-dpi. Method 1 (n=4), Method 2-6 (n=5). P(Method 1 vs 2)=0.0006, t(7)=5.98, P(Method 1 vs 3)<0.0001, t(7)=12.54, P(Method 1 vs 2)=0.0006, t(7)=5.98, P(Method 1 vs 3)<0.0001, t(7)=12.54, P(Method 1 vs 2)=0.0006, t(7)=5.98, P(Method 1 vs 3)<0.0001, t(7)=12.54, P(Method 1 vs 2)=0.0006, t(7)=5.98, P(Method 1 vs 3)<0.0001, t(7)=12.54, P(Method 1 vs 2)=0.0006, t(7)=5.98, P(Method 1 vs 3)<0.0001, t(7)=12.54, P(Method 1 vs 2)=0.0006, t(7)=5.98, P(Method 1 vs 3)<0.0001, t(7)=12.54, P(Method 1 vs 2)=0.0006, t(7)=5.98, P(Method 1 vs 3)<0.0001, t(7)=12.54, P(Method 1 vs 2)=0.0006, t(7)=5.98, P(Method 1 vs 3)<0.0001, t(7)=12.54, P(Method 1 vs 2)=0.0006, t(7)=5.98, P(Method 1 vs 3)<0.0001, t(7)=12.54, P(Method 1 vs 2)=0.0006, t(7)=5.98, P(Method 1 vs 3)<0.0001, t(7)=12.54, P(Method 1 vs 3)=0.0001, t(7)=10.0001, t(7)=10001, t(7)=10.0001, t(7)=10 4)=0.0027, t(7)=4.54, P_(Method 1 vs 5) <0.0001, t(7)=8.15, P_(Method 1 vs 6) <0.0001, t(7)=10.63. (c) Numbers of total immune cells were evaluated in the brains and spinal cords (S.C.) of mice with Type-B EAE at 17-dpi. n=4. Brain: $P_{(WT vs Nirp3-\ell)}=0.8159$, t(6)=0.2433, $P_{(WT)}=0.2433$, $P_{(WT)}=0.8159$, t(6)=0.2433, $P_{(WT)}=0.8159$, t(6)=0.2433, t(6)=0.vs Asc-/-)=0.5685, t(6)=0.6031. Spinal cord: P(WT vs Nlrp3-/-)=0.5226, t(6)=0.6787, P(WT vs Asc-/-)=0.8984, t(6)=0.1332. (d) IFNβ treatment on *NIrp*3^{-/-} mice with Type-B EAE induced with Method 6. IFN β (3x10⁴ unit/mouse) were *i.p.* injected every other day from day 0 to 8 as previously performed. (e) Time course on body weight change in Type-A and Type-B EAE. (f) II1b mRNA levels in splenic DC from naïve, Type-A, and Type-B EAE mice at 9 dpi. P_(Naive vs Type-A)=0.0059, t(4)=5.344, P_(Naive vs Type-B)<0.0001, t(4)=18.55, P_(Type-A vs Type-A) _B=0.0004, t(4)=10.83. (g,h) Levels of extracellular IL-1β (f) and p20 caspase-1 (g) in 24h splenocyte culture supernatants (n=4 for Type-A, n=5 for Type-B). P=0.0261, t(6)=2.724 (g). P=0.0020, t(6)=4.794 (h). (i) Infiltrated cell numbers in the brain and spinal cord (n=7). Brain: $P_{(Total)}=0.0028$, t(12)=3.747, $P_{(CD4)}=0.018$, t(12)=2.737, $P_{(Th17)}=0.0167$, t(12)=2.78, $P_{(Th1)}=0.0165$, t(12)=2.784. $P_{(CD8)}=0.0329, t(12)=2.41, P_{(B)}=0.0013, t(12)=4.165, P_{(DC)}=0.0003, t(12)=5.008, P_{(PMN)}=0.0004, t(12)=4.912, P_{(Mac)}=0.0027, t(12)=3.759$ Spinal cord: $P_{(Total)}=0.0035$, t(12)=3.621, $P_{(CD4)}=0.0002$, t(12)=5.372, $P_{(Th17)}=0.0003$, t(12)=4.997, $P_{(Th1)}=0.00465$, t(12)=2.221 $P_{(CD8)}=0.0104, t(12)=3.032, P_{(B)}=0.0918, t(12)=1.833, P_{(DC)}=0.0429, t(12)=2.263, P_{(PMN)}=0.0011, t(12)=4.29, P_{(Mac)}=0.089, t(12)=1.849$ (j) Representative LFB-stained images of brains at 17-dpi. Red arrows indicate reduced LFB intensity, *i.e.*, reduced myelin. Scale bars. 200 µm. (k) T2 FLAIR MRI analysis of spinal cords obtained from mice at 18-dpi. Yellow arrows indicate areas of potential myelin loss. (I) Thermal sensitivity evaluated by a hot-plate test in 9-dpi mice, which did not show any EAE symptoms and motor dysfunction. n=8. P(Naive vs Type-A)=0.1524, t(14)=1.514, P(Naive vs Type-B)=0.0013, t(14)=4.026, P(Type-A vs Type-B)=0.0217, t(14)=2.582. *; p<0.05. All statistical analyses in this figure were performed by two-tailed unpaired Student's t-test. All the experimental data and images are representatives from at least 2 similar experiments for each.



9-dpi. (b) Percentages of LTa⁺ (*i.e.*, mLT⁺) macrophages, determined by flow cytometry, in DLNs of naïve mice or mice with EAE at 9dpi. Naïve (*n*=4), Type-A (n=3), Type-B (n=4). $P_{(Naive vs Type-A)}$ =0.1340, *t*(5)=1.7887, $P_{(Naive vs Type-B)}$ =0.0024, *t*(6)=45.021, $P_{(Type-A vs Type-B)}$ =0.0268, *t*(5)=3.101, by two-tailed unpaired Student's *t*-test. (c) Evaluating mLT expression on DCs from mice received CFA injection alone without MOG. One group received 200 µg *Mtb* in CFA (*Mtb* dosage for Type-A EAE), and another group had 400 µg *Mtb* in CFA twice (*Mtb* dosage for Type-B)=0.0040, *t*(6)=4.519, $P_{(Type-A vs Type-B)}$ =0.0095, *t*(6)=3.748, by two-tailed unpaired Student's *t*-test. (d) Methylation analysis was carried out by bisulfite conversion on the *Lta* promoter in DCs from naïve mice. Methylated and unmethylated CpG were shown with black and gray boxes, respectively





Comparison of IFN β -responders and non-responders in mouse EAE and human RRMS.

(a) Numbers of indicated cell types obtained from DLNs and spleen on 17-dpi were evaluated between Type-A and Type-B EAE. n=4. DLNs: $P_{(Total)}=0.2073$, t(6)=1.413, $P_{(CD3)}=0.8466$, t(6)=0.202, $P_{(CD4)}=0.9285$, t(6)=0.09353, $P_{(CD8)}=0.7413$, t(6)=0.3459, $P_{(B)}=0.4141$, t(6)=0.8773, $P_{(Th17)}=0.9724$, t(6)=0.03605, $P_{(Th1)}=0.9017$, t(6)=0.1289, $P_{(Treg)}=0.3556$, t(6)=1.001. Spleen: $P_{(Total)}=0.0072$, t(6)=3.99, $P_{(CD3)}=0.2537, t(6)=1.262, P_{(CD4)}=0.1781, t(6)=1.525, P_{(CD8)}=0.3999, t(6)=0.906, P_{(B)}=0.1371, t(6)=1.715.$ (b, c) Representative flow charts (b) and proportion (c) for GM-CSF-producing Th1 and Th17 cells from Type-A and Type-B EAE at 9-dpi. n=6. Th17: P=0.6568. t(10)=0.4579, Th1: P=0.3954, t(10)=0.8879. (d) Proportion for IFNγ-, GM-CSF-, IL-22-producing and CD5L-expressing Th17 cells in spinal cords from Type-A and Type-B EAE mice at 17-dpi. Type-A (*n*=4), Type-B (n=5). *P*_(/FNv)=0.3493, *t*(7)=1.0037, *P*_(GM-CSF)=0.1760, t(7)=1.505, $P_{(II-22)}$ =0.2859, t(7)=1.155, $P_{(CD5L)}$ =0.2641, t(7)=1.214. (e) Proportions and MFI of CXCR2⁺ neutrophils and macrophages obtained from naïve mice and mice with either Type-A or Type-B EAE at 9-dpi. n=4. PMN %: P(Naive vs Type-A)=0.0102, t(6)=3.695, P(Naive vs туре-в)=0.0004, t(6)= 7.001, P_(Туре-А vs Туре-В)=0.0022, t(6)=5.131. PMN MFI: P_(Naive vs Type-A)=0.0777, t(6)=2.125, P_(Naive vs Type-B)=0.0014, t(6)= 5.588, P_(Type-A vs Type-B)=0.0090, t(6)=3.81. Macrophage %: P_(Naive vs Type-A)=0.0005, t(6)=6.774, P_(Naive vs Type-B)=0.0006, t(6)= 6.48, P(Type-A vs Type-B)=0.0161, t(6)=3.314. Macrophage MFI: P(Naive vs Type-A)=0.33, t(6)=1.06, P(Naive vs Type-B)=0.0001, t(6)= 11.11, P(Type-A vs Type-A)=0.0001, t(6)= 11.11, P(Type-A vs Type-A)=0.0001, t(6)= 11.00, $_{B}$ =0.0001, t(6)=10.89. (f) Flow cytometry results showing CXCR2 protein expression on the CD4⁺ T surface in each group (naïve mice, Type-A, or Type-B EAE mice at 9-dpi). (g) Comparison of relative gene expression levels between Ltbr and Cxcr1, Cxcr1 and Cxcr2, *Ltbr* and *Cxcr2*, normalized to *Vcam1* expression. Total PBMCs from IFNβ-responder and non-responder RRMS patients were compared. All statistical analyses in this figure were performed by two-tailed unpaired Student's t-test. All the experimental data sets. except for (g), are representatives from at least 2 similar experiments for each.



Supplementary Figure 5

CXCL1 expression in spinal cords.

Shown are representative images of typical CXCL1 staining in spinal cords from naïve mice and mice with either Type-A or Type-B EAE at 9-dpi. Scale bars, 200 μm. Images are representatives from 3 similar experiments.



Histology of spinal cords in mice with Type-A or Type-B EAE.

(a-e) Shown are representative images of typical staining from multiple mice. (a) H/E staining in the spinal cord of Type-A and Type-B EAE mice at 70-dpi. (b) IGF-1 staining in spinal cord of naïve mice and mice with either Type-A or Type-B EAE at 22-dpi. (c) Bielschowsky neuron staining in spinal cords of naïve mice (e), and Type-A and Type-B EAE at 30-dpi. Red arrows indicate area showing reduced staining intensity in the spinal cord. (d,e) Golgi's silver staining in spinal cord of naïve mice (d), and Type-A and Type-B at 22-dpi (e). All scale bars in this figure except magnified figure in (e), 200 μm. All scale bars in magnified figure in (e), 50 μm. All images are representatives from 3 similar experiments.





than Type-A EAE induction, and is NLRP3 inflammasome-independent and IFN β -resistant. Type-B EAE can also be induced with Type-A EAE induction methods with MHV68 infection or with rLT (rLT $\alpha 2\beta 1$) injection. In Type-B EAE, *Lta* gene expression is epigenetically induced in DCs and the expression of membrane-bound LT (mLT) on DCs are enhanced. mLT stimulate LT β R on CD4⁺ T cells, resulting in the upregulation of CXCR2 on CD4⁺ T cells. Blockade of LT β R (with LT β R-Fc) and CXCR2 (with SB225002) selectively inhibits the Type-B EAE progression. mLT is also involved in the induction of Sema6B in T cells. Sema6B causes neural damages, and this may be a reason for the prolonged and minimal remission in Type-B EAE.