Parkin regulates translesion DNA synthesis in response to UV radiation

Supplementary Materials



Supplementary Figure 1: K164R mutation doesn't affect the interaction between Parkin and PCNA. HEK293T cells transfected with Flag-PCNA or Flag-PCNA^{K164R} were lysed and incubated with protein A/G agarose conjugated with normal rabbit IgG or anti-Parkin antibody for immunoprecipitation followed by blotting with the indicated antibodies.



Supplementary Figure 2: Interaction between Parkin and RPA. (A) HEK293T cells were lysed and incubated with protein A/G agarose conjugated with normal rabbit IgG or anti-Parkin antibody for immunoprecipitation followed by blotting with the indicated antibodies. (B) Endogenous immunoprecipitation was performed in HEK293T cell lysates containing EB or not as in (A), followed by blotting with the indicated antibodies.



Supplementary Figure 3: The association between Parkin and NBS1 is independent of RPA complex. HEK293T cells were transfected with siNC, siRPA32 or siRPA70. 48 h later cells were transfected with Myc-NBS1. Then cells were lysed and incubated with protein A/G agarose conjugated with normal rabbit IgG or anti-Parkin antibody for immunoprecipitation followed by blotting with the indicated antibodies.



Supplementary Figure 4: Expression of NBS1 in WT and Parkin-/- cells. WT and Parkin-/- cells were collected and examined by western-blot with an anti-NBS1 antibody. β -actin: loading control.



Supplementary Figure 5: The function of Parkin in TLS regulation is independent of its E3 ligase activity. (A) WT, Parkin-/-, and Parkin-/- cells stably expressing Flag-Parkin or Flag-Parkin^{C431S} were irradiated with 15 J/m² UV and further incubated for 2 h. The chromatin fractions were harvested and separated by SDS-PAGE. The levels of RPA32 and mUb-PCNA were detected by blotting with the indicated antibodies. The levels of RPA32 and PCNA in soluble fraction were also detected. (B) WT, Parkin-/-, and Parkin-/- cells stably expressing Flag-Parkin or Flag-Parkin^{C431S} were irradiated with 15 J/m² UV and further incubated for 2 h. Cells were pre-extracted with 0.5% Triton for 10 min and immunostained with anti-NBS1 antibody and Hoechst-stained. The percentage of cells with NBS1 foci was determined. Error bars represent SD. *t*-test, n = 2. (C) U2OS cells transfected with empty vector, Parkin or Parkin^{C431S} were irradiated with 15 J/m² UV and recovered for 3 h. Cells were pre-extracted with 0.5% Triton for 10 min and then fixed, immunostained with anti-Rad18 antibody and Hoechst-stained. Quantification of the percentage of cells with more than 20 Rad18 foci. Error bars represent SD. *t*-test, n = 3. (**D**) Viability of cells after exposure to UV radiation. WT, Parkin-/-, and Parkin-/- cells complemented with Flag-Parkin or flag-Parkin flag-Parkin or flag-Parkin or flag-Parkin flag-Parkin or flag-Parkin flag-Parkin flag-Parkin or flag-Park



GFP-Polη Laser microirradiation

Supplementary Figure 6: Parkin deficiency attenuates laser microirradiation-induced Polq recruitment. WT or Parkin-/- cells transfected with GFP-Polq were microirradiated. Dynamic redistribution of GFP-Polq was monitored at the indicated time points.