

Supplementary information

Selective inhibition of sterol *O*-acyltransferase 1 isozyme by beauveriolide III in intact cells

Taichi Ohshiro¹, Keisuke Kobayashi¹, Mio Ohba¹, Daisuke Matsuda¹, Lawrence L. Rudel², Takashi Takahashi³, Takayuki Doi⁴, and Hiroshi Tomoda^{1,*}

¹Graduate School of Pharmaceutical Sciences, Kitasato University, Tokyo 108-8641, Japan;

²Section on Lipid Sciences, Department of Internal Medicine, Wake Forest School of Medicine, Winston-Salem, NC 27157, USA; ³Faculty of Pharmaceutical Sciences, Yokohama College of Pharmacy, Kanagawa 245-0062, Japan; ⁴Graduate School of Pharmaceutical Sciences, Tohoku University, Sendai 980-8578, Japan

* To whom correspondence should be addressed: Hiroshi Tomoda, Graduate School of Pharmaceutical Sciences, Kitasato University, 5-9-1 Shirokane, Minato-ku, Tokyo 108-8641, Japan, E-mail: tomodah@pharm.kitasato-u.ac.jp

Supplementary Figure S1

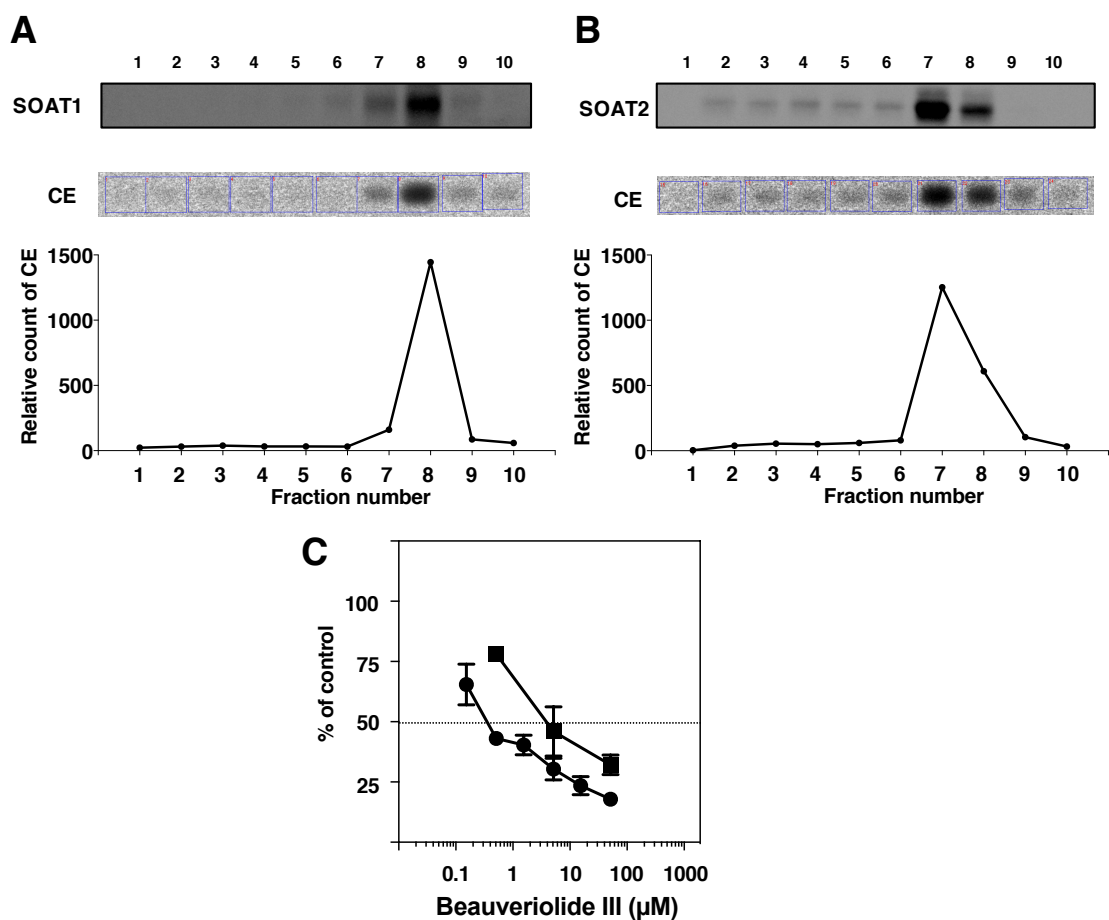
Inhibition of SOAT1 and SOAT2 by BeauIII in an enzyme-based assay using an intact ER fraction from SOAT1- and SOAT2-CHO cells sonicated for 10 min.

Supplementary Figure S2

Co-localization of beauveriolide and SOAT1 in mouse peritoneal macrophages.

Supplementary Figure S3

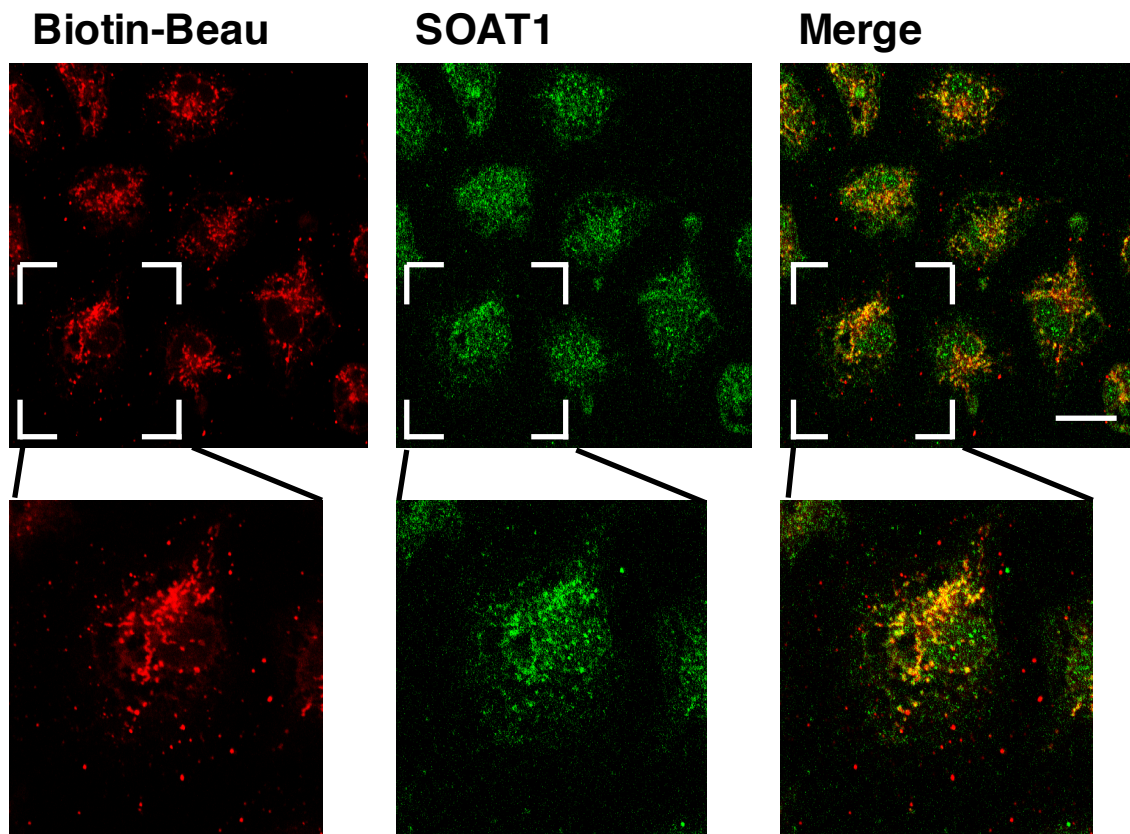
UFLC charts of BeauI, BeauIII and Biotin-Beau.



Supplementary Figure S1

Inhibition of SOAT1 and SOAT2 by BeauIII in an enzyme-based assay using an intact ER fraction from SOAT1- and SOAT2-CHO cells sonicated for 10 min.

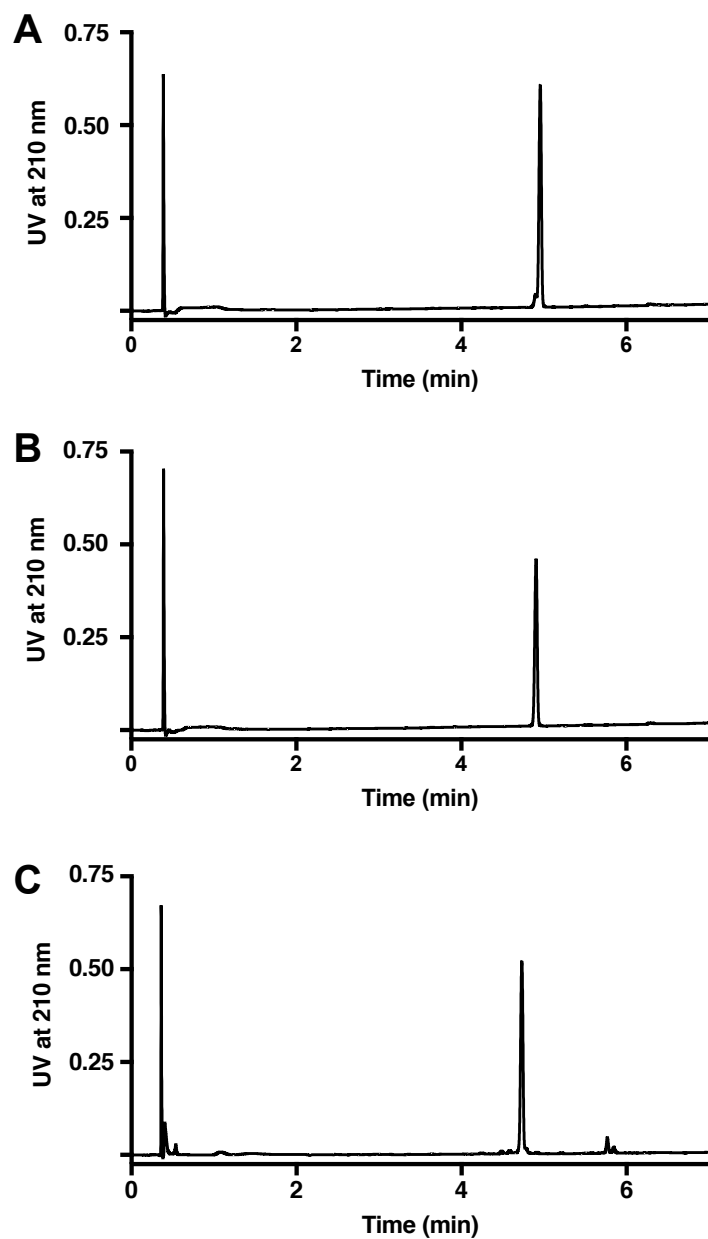
Each cell lysate in SOAT1-/SOAT2-CHO cells sonicated for 10 min was fractionated in 10 fractions with the Optiprep™ density gradient. In order to identify the ER fraction including SOAT proteins, SOAT protein levels and SOAT activities were measured using a Western blotting analysis and SOAT assay, respectively (A and B). Then, SOATA-rich ER fraction was then used as an enzyme sources for SOAT1 (●) and SOAT2 (■) assays (C). The results obtained were plotted as % of control (without drugs). Values represent means \pm SD (n=3~4).



Supplementary Figure S2

Co-localization of beauveriolide and SOAT1 in mouse peritoneal macrophages.

Macrophages were double-stained with anti-Biotin-Beau (red) and anti-SOAT1 (green). Inserts show a magnification of the area indicated by the squares. Scale bars, 20 μm .



Supplementary Figure S3

UFLC charts of BeauI, BeauIII and Biotin-Beau.

BeauI (A), BeauIII and Biotin-Beau (C) were analyzed by ultra fast liquid chromatography (UFLC)

Supplementary Experiment Methods

Materials

Goat anti-rabbit IgG conjugated to fluorescein isothiocyanate (FITC) was purchased from Sigma-Aldrich (St. Louis, MO). Hank's balanced salt solution was purchased from Nissui Seiyaku (Tokyo, Japan). Streptavidin conjugated to Alexa Fluor 546 was purchased from Thermo Fisher Scientific (Waltham, MA). A tissue culture chamber (LAB-TEK 8-chamber) was purchased from Nalge Nunc (Rochester, NY).

Immunofluorescence staining of SOAT1 and Biotin-Beau in mouse peritoneal macrophages

Mouse peritoneal macrophages from ICR mice (Japan SLC, Inc., Japan) were prepared as described previously¹. Briefly, primary mouse peritoneal macrophages (2.0×10^5 cells) with liposomes in each well of a tissue culture chamber (LAB-TEK 8-chamber) were incubated at 37°C in 5.0% CO₂. After a 14-h incubation, cells were washed three times with PBS, fixed with 3.7% formalin for 10 min, and permeabilized with PBS containing 0.10% Triton X-100 at room temperature for 20 min. After a brief rinse with PBS, cells were incubated with blocking buffer (3.0% BSA and 0.10% Triton X-100 in PBS) at room temperature for 1.0 h. Cells were then incubated with the anti-SOAT1 rabbit polyclonal antibody ($1.5 \mu\text{g/ml}$ in blocking buffer)² and Biotin-Beau ($9.8 \mu\text{M}$)³ at room temperature. After a 1-h incubation, cells were stained with the secondary antibody, a goat anti-rabbit IgG conjugated to FITC ($10 \mu\text{M}$) and streptavidin conjugated to Alexa Fluor 546 ($1.0 \mu\text{g/ml}$) in blocking buffer, for 1.0 h. Cells were washed three times after the incubation and analyzed with confocal laser scanning microscopy (LSM-510 META, Carl Zeiss, Germany). Images were analyzed with a Zeiss LSM Image Browser (Version 3.5.0.376)

UFLC analysis

BeauI, BeauIII and Biotin-Beau dissolved in methanol were analyzed by (UFLC) (Prominence, Shimadzu, Kyoto, Japan). under the following conditions: column, Shim Pack XR-ODS (Shimadzu), 2.0 x 75 mm; column temperature, 50°C; solvent, 6-min linear gradient from 5.0% acetonitrile in 0.10% phosphoric acid to 95% acetonitrile in 0.10% phosphoric acid; flow rate, 0.55 ml/min; detection, UV at 210 nm.

Supplementary References

- 1 Namatame, I., Tomoda, H., Arai, H., Inoue, K. & Omura, S. Complete inhibition of mouse macrophage-derived foam cell formation by triacsin C. *J Biochem* **125**, 319-327 (1999).
- 2 Lee, R. G., Willingham, M. C., Davis, M. A., Skinner, K. A. & Rudel, L. L. Differential expression of ACAT1 and ACAT2 among cells within liver, intestine, kidney, and adrenal of nonhuman primates. *J Lipid Res* **41**, 1991-2001 (2000).
- 3 Doi, T. *et al.* Conformationally restricted analog and biotin-labeled probe based on beauveriolide III. *Bioorg Med Chem Lett* **22**, 696-699 (2012).