THE LANCET Diabetes & Endocrinology

Supplementary appendix

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Supplementary Methods

Biochemical Analyses

In PROMIS, non-fasting blood samples (with the time since last meal recorded) have been drawn by phlebotomists from each participant and centrifuged within 45 minutes of venepuncture. Serum, plasma and whole blood samples were stored at -70°C within 45 minutes of venepuncture¹. All samples were transported on dry-ice to the central laboratory at the Center for Non-Communicable Diseases (CNCD), Pakistan, where serum and plasma samples were aliquoted in different storage vials. Samples from cases and controls were randomly assorted in storage boxes (each box containing 81 samples) and stored at -70°C for any subsequent laboratory analyses. All biochemical assays were conducted in automated auto-analysers. At CNCD Pakistan, measurements for total-cholesterol, HDL-C, LDL-C, triglycerides, creatinine were conducted in serum samples using enzymatic assays; whereas levels of HbA1c were measured using a turbidemetric assay in whole-blood samples (all using kits manufactured by Roche Diagnostics). For further measurements, aliquots of serum and plasma samples were transported on dry ice to the Translational Research Center (TRC), University of Pennsylvania, USA, where following biochemical assays were conducted: apolipoproteins (apoA-I, apoA-II, apoB, apoC-III, apoE) and non-esterified fatty acids were measured through immunoturbidometric assays using kits by Roche Diagnostics or Kamiya: lipoprotein (a) levels were determined through a turbidimetric assay using reagents and calibrators from Denka Seiken (Niigata, Japan); LpPLA2 mass and activity levels were determined using immunoassays manufactured by diaDexus; measurements for insulin, leptin and adiponectin were made using radio-immunoassays by LINCO; levels of adhesion molecules (ICAM-1, VCAM-1, P- and E-Selectin) were determined through enzymatic assays by R&D; and measurements for CRP, ALT, AST, cystatin-C, ferritin, ceruloplasmin, thyroid stimulating hormone, alkaline phosphatase, sodium, potassium, choloride, phosphate, sex-harmone binding globulin were made using enzymatic assays manufactured by Abott Diagnostics. Glomerular filtration rate (eGFR) was estimated from serum creatinine levels using the MDRD equation².

Statistical Methods

Genome-wide analyses Participants were genotyped on the Illumina 660W-Quad or the Illumina OmniExpress Bead chip. Quality control (QC) criteria on the genotyped data included, removal of participants or single nucleotide polymorphisms (SNP's) that had a missing rate >5%. SNPs with a MAF <1% and a P-value of <10⁻⁷ for the Hardy-Weinberg equilibrium test were also excluded from the analyses. Further QC included removal of participants with discrepancy between their reported sex and genetic sex determined from the X chromosome. To identify sample duplications, unintentional use of related samples (cryptic relatedness) and sample contamination (individuals who seem to be related to nearly everyone in the sample), identity-by-descent (IBD) analyses were

conducted in PLINK⁴. To account for population substructure in Pakistanis, principal component analyses were conducted on the pruned dataset as described previously¹. After implementing QC, imputations were conducted using the global reference panel in the 1000-Genomes project (March 2012 (v3) release) using IMPUTE-2. All analyses were conducted in SNPTEST, adjusted for the first 10 principal components, age and sex.

Genetic Risk Score Analysis We utilized a two-sample MR method⁵ to estimate effects for a multi-SNP genetic instrument by using summary statistics from the CARDIoGRAMplusC4D consortium. This method has been previously validated to infer causal effects (odds ratio) and associated standard error⁶.

Supplementary References

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eFigure 1a. Cross-sectional associations of LPA KIV2 repeats in PROMIS controls participants

eFigure 1b. Cross-sectional association of *LPA* KIV2 repeats with apo(a) protein isoforms in PROMIS controls participants



Partial correlation coefficients (r) adjusted for age and sex have been calculated using all data from the study control participants.



eFigure 2. Cross-sectional associations of apo(a) protein isoform in PROMIS controls participants



eFigure 3. Cross-sectional associations of Lp(a) concentration in PROMIS control participants

eFigure 4. Correlation of Lp(a) levels measured by the Denka Seiken immunoturbidemetric method and the Northwest Lipid Metabolism and Diabetes Research Laboratories (NWRL) ELISA method (n = 2617)



Partial correlation coefficients adjusted for age and sex were calculated to assess correlations between the two traits. Association of the two traits was further characterized by using linear regression analyses adjusted for age and sex. From each fitted regression model, overall adjusted mean values and 95 percent confidence intervals for Lp(a) (nmol/L) measured by NWRL within quintiles of Lp(a) (nmol/L) measured by Denka Seiken, were obtained.

eFigure 5. Association of rs10455872 and rs3798220 variants with Lp(a) levels measured by the Denka Seiken immunoturbidemetric method and the Northwest Lipid Metabolism and Diabetes Research Laboratories (NWRL) ELISA method (n = 2617)



Per-SD change in log-Lpa levels (nmol/L)

eFigure 6: Association of the genetic risk score, comprising of rs10455872 and rs3798220 variants, with LPA KIV2 repeats, Lp(a) levels, apo(a) isoform

size and other traits



SD change in trait per allele increase in the risk score (95% CI)

eFigure 7a-b. Genetic determinants of apo(a) isoform size and KIV2 repeats



(b) LPA KIV2 repeats





The following SNPs were independently associated with apo(a) protein isoform size on step-wise conditional analyses in PROMIS: rs12525588 rs7761293 rs7770628 rs1367211 rs13213129 rs2665356 rs3798950 rs142231215. These SNPs were further used to create a genetic risk score to assess association with CHD risk.

eFigure 9. Genetic determinants of Lp(a) levels







The following SNPs were independently associated with lp(a) levels on step-wise conditional analyses in PROMIS: rs73028918, rs9456488, rs77153348, rs8191871, rs2297362, rs73023700, rs3737088, rs148868952, rs316024, rs17589858, rs4708867, rs58432601, rs6919346, rs62441903, rs41272114, rs7765803, rs4708876, rs41269876, rs1367210, rs783144, rs2064712, rs7756550, rs12174748, rs35533223, rs79067311. These SNPs were further used to create a genetic risk score to assess association with CHD risk.

eTable 1. Analyses of rs3777392 and rs2457564 variants with various traits in international consortia

	P-values		
	rs3777392	rs2457564	
ICBP (n = 134,433)			
Systolic blood pressure	0.78	0.74	
Diastolic blood pressure	0.12	0.02	
Pulse Pressure	0.46	0.09	
Mean arterial pressure	0.80	0.41	
GLGC (n = 134,433)			
Total Cholesterol	0.12	0.02	
LDL-C	0.13	0.13	
HDL-C	0.92	0.02	
Log-triglycerides	0.25	0.92	
MAGIC (n = 46,368)			
Post-load glucose	0.84	0.33	
Fasting glucose	0.93	0.66	
Hba1c	0.43	0.45	
Fasting insulin	0.24	0.28	
Fasting pro-insulin	0.7	0.73	
HOMA-B	0.28	0.43	
HOMA-IR	0.17	0.70	
GIANT (n=183,727)			
Waist-to-hip ratio	0.14	0.03	
Height	0.41	0.91	
BMI	0.64	0.12	

	1-SD higher LPA KIV2 repeats (qPCR)		1-SD higher log-Lp(a) levels	
	(6020 MI cases and 6686 controls)		(5878 MI cases and 6886 controls)	
Model	OR (95% CI)	P-value	OR (95% CI)	P-value
Age, gender, recruitment center and ethnicity	0.93 (0.90 – 0.97)	<0.0001	1.15 (1.11 - 1.20)	<0.0001
plus tobacco use, history of diabetes, and hypertension				
and waist-to-hip ratio	0.92 (0.89 - 0.96)	<0.0001	1.16 (1.12 - 1.21)	<0.0001
plus apo(a1) and apo(b)	0.92 (0.88 - 0.96)	<0.0001	1.11 (1.06 - 1.15)	<0.0001
plus log-Lp(a) levels / LPA				
KIV2 repeats (qPCR)	0.93 (0.90 -0.97)	0.001	1.10 (1.05 - 1.14)	<0.0001





Interaction *P-value* was calculated by fitting an interaction term (x) in the logistic regression model for MI as an outcome while treating *LPA* KIV2 repeats on a continuous scale

eFigure 12. Association of the *LPA* locus with OxPL-apoB (a) unadjusted, (b) adjusted for *LPA* KIV2 repeats, (c) adjusted for Lp(a) levels and (d) adjusted for KIV2 repeats and Lp(a) levels



(c) Adjusted for log-Lp(a) levels



(d) Adjusted for LPA KIV2 repeats and log-Lp(a) levels

