

### **Supporting Information**

Fluoroacetamide Moieties as NMR Spectroscopy Probes for the Molecular Recognition of GlcNAc-Containing Sugars: Modulation of the CH- $\pi$  Stacking Interactions by Different Fluorination Patterns

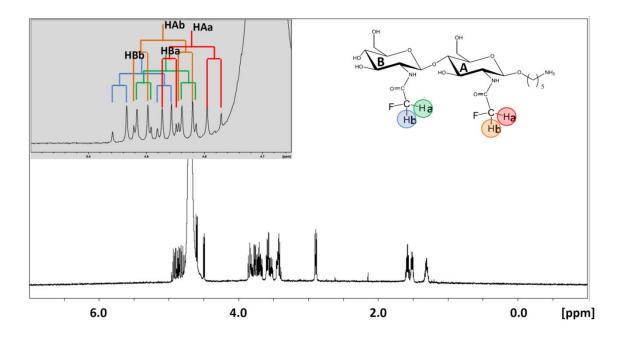
Luca Unione,<sup>[a]</sup> Maria Alcalá,<sup>[b]</sup> Begoña Echeverria,<sup>[b]</sup> Sonia Serna,<sup>[b]</sup> Ana Ardá,<sup>[a]</sup> Antonio Franconetti,<sup>[c]</sup> F. Javier Cañada,<sup>[d]</sup> Tammo Diercks,<sup>[a]</sup> Niels Reichardt,\*<sup>[b, e]</sup> and Jesús Jiménez-Barbero\*<sup>[a, f, g]</sup>

chem\_201605573\_sm\_miscellaneous\_information.pdf

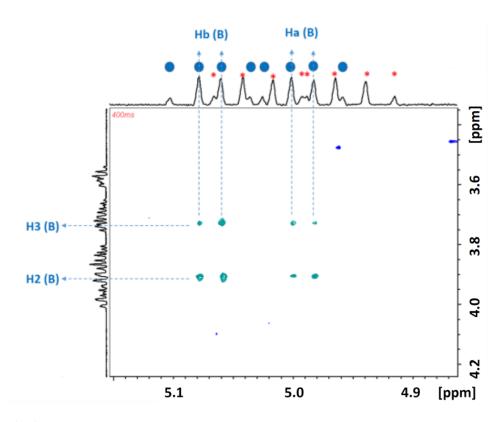
### **SUPPORTING INFORMATION**

- 1. Homo- and heteronuclear <sup>19</sup>F-<sup>19</sup>F and <sup>19</sup>F-<sup>1</sup>H correlation experiments and STD experiments
- 2. Theoretical electron density surfaces and charge distribution of (fluoro)acetamide moieties
- 3. Glycan array experiments
- 4. NMR spectra of new compounds
- 5. <sup>1</sup>H STD-NMR

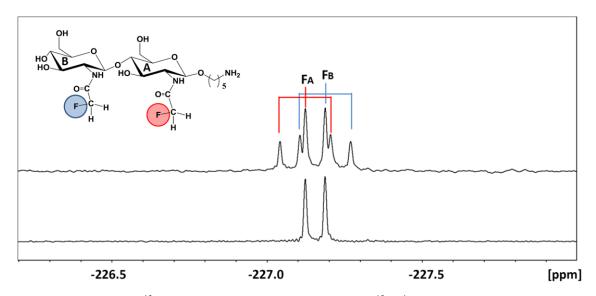
# 1. Homo- and heteronuclear <sup>19</sup>F-<sup>19</sup>F and <sup>19</sup>F-<sup>1</sup>H correlation experiments and STD experiments



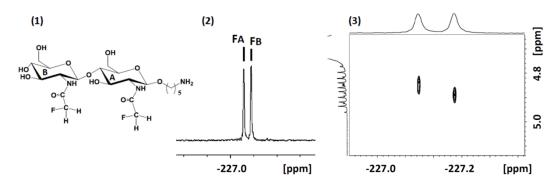
**Figure S1.** 600 MHz  $^{1}$ H NMR spectrum of **1**. The spectral region for the acetamide protons is shown in the inset. The homo- and hetero-nuclear ( $J_{HH}$ ,  $J_{HF}$ ) coupling constants for every residue are specified using a color code.



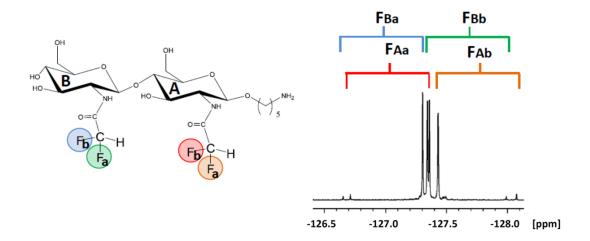
**Figure S2.**  $^{1}\text{H-}^{1}\text{H}$  NOESY spectrum (400ms mixing time) of compound **1**. Correlation between both protons of the  $^{-}\text{CH}_2\text{F}$  group (Hb and Ha) and H2 and H3 of the non-reducing end unit (B). Only crosspeaks with the strongest components of the multiplets are observed. The blue spots indicate the components of the multiplets for the non-reducing end (B), while red stars refer to the multiplets of the reducing end (A).



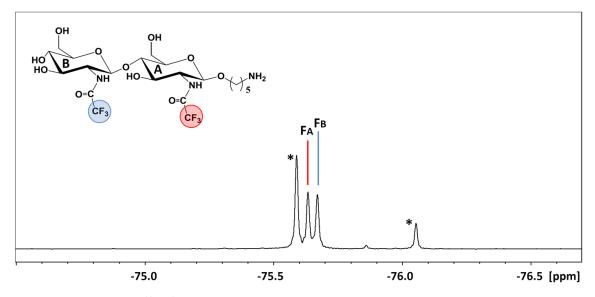
**Figure S3.** 564 MHz <sup>19</sup>F NMR spectrum of **1**. Bottom panel, <sup>19</sup>F-{<sup>1</sup>H} decoupled NMR spectrum. Top panel <sup>19</sup>F-{<sup>1</sup>H} coupled NMR spectrum, with the heteronuclear  $J_{FH}$  couplings for every residue specified in color code.



**Figure S4.** (1) Residue label for molecule **1**. (2) 546 MHz <sup>19</sup>F-{<sup>1</sup>H} decoupled NMR spectra. (3) <sup>19</sup>F-<sup>1</sup>H heteronuclear correlation spectrum of **1**. The correlation between the fluoroacetamide protons and their respective fluorine signals are evidenced.



**Figure S5.** 564 MHz <sup>19</sup>F-{<sup>1</sup>H}-decoupled NMR spectrum of **2**. The analysis of the multiplicity is shown.



**Figure S6.** 564 MHz <sup>19</sup>F-{<sup>1</sup>H} decoupled of **3**. The signals for every residue specified in a color code. Two impurities are labeled with (\*) symbol.

Table S1.  $^{1}$ H STD competition experiments for  $K_{D}$  determination of compounds 1 and 2.

<sup>1</sup> H-STD relative Intensity (%)				
CH <sub>3</sub> (4): CHxFy	CH <sub>2</sub> F (1)	CHF <sub>2</sub> (2)		
0	100	100		
1	60	85		
3	33	55		
6	25	33		

The  $K_D$  dissociation constants for the complexes of **1** and **2** with WGA were estimated by competitive STD experiments with the natural ligand, **4** ( $K_D = 0.19$  mM).<sup>[19]</sup> The absolute concentration of compounds **1** and **2** is 0.5 mM, while that of WGA protein is 10  $\mu$ M.

The target  $K_D$  values can be determined using the usual equation for competitive STD experiments:  $^{[28, 29]}K_D = (iK_i \cdot C_L)/(C_i - iC_i - iK_i)$ 

where  $K_i$  refers to the dissociation constant of **4**,  $C_i$  and  $C_L$  are the concentrations of **4** and the fluorinated analogue (**1** or **2**), respectively, and i is the fractional inhibition, defined as  $i = (L_0 - L_i)/L_0$ , where  $L_0$  and  $L_i$  are the <sup>1</sup>H-STD signal intensities of the corresponding CH protons at the fluoroacetamide moiety in the absence and presence of **4**, respectively.

Table S2.  $^{1}H$  STD competition experiments for  $K_{D}$  determination of compound 3.

<sup>1</sup> H-STD relative Intensity (%)			
[ -CF <sub>3</sub> (3) : -CH <sub>3</sub> (4) ]	CH <sub>3</sub> (4)		
0	100		
0,5	90		
0,75	85		
1	80		
2	70		
5	50		

The  $K_D$  dissociation constant for compound 3 has been estimated by competitive STD experiments on the WGA-4 complex. The absolute concentration of compounds 4 is 1.5 mM, while that of WGA protein is 30  $\mu$ M.

The target K<sub>i</sub> value can be determined using the equation for competitive STD experiments

$$K_i = C_i (K_D - i \cdot K_D) / i \cdot (C_L + K_D)$$

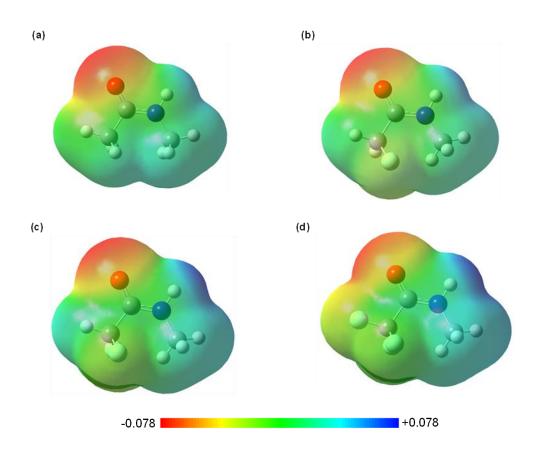
where  $K_D$  refers to the dissociation constant of **4**,  $C_i$  and  $C_L$  are the concentration of **3** and **4**, respectively, and i is the fractional inhibition, defined as  $i = (L_0 - L_i)/L_0$ , where  $L_0$  and  $L_i$  are the <sup>1</sup>H-STD signal intensities of the corresponding  $CH_3$  protons at the acetamide moiety of compound **4** in the absence and presence of **3**, respectively.

## 2. Theoretical electron density surfaces and charge distribution of (fluoro)acetamide moieties

**Table S3.** Charge distribution showing Mulliken-type atomic charge for model compounds N-methyl (fluoro)acetamides at M06-2X/6-31G(d,p) level of theory (including solvation). [30]

Molecule	π-interaction type	δ <sup>+</sup> <sub>Mulliken</sub> activated H	δ <sup>+</sup> <sub>Mulliken</sub> (-CX <sub>3</sub> )
N-Me-acetamide	СН-п	0.152	-0.219
N-Me-fluoroacetamide	СН-п	0.160	0.031
<i>N</i> -Me-difluoroacetamide	СН-п	0.162	0.427
N-Me-trifluoroacetamide	CF-π	_a	0.782

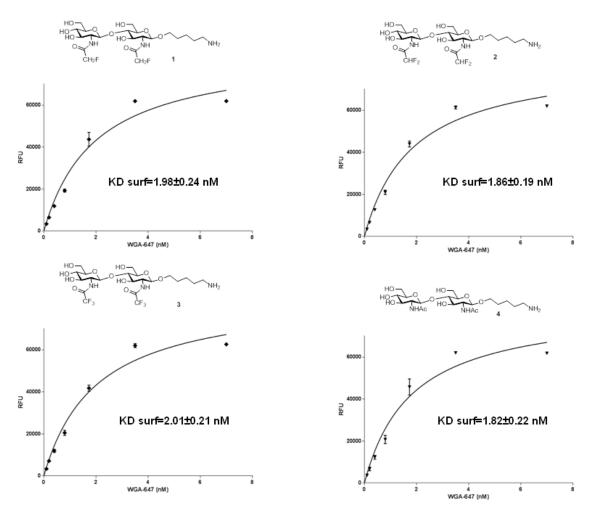
<sup>&</sup>lt;sup>a</sup>Calculated Mulliken charge for F atoms is -0.264



**Figure S7.** Electron density surface from total SFC density mapped with electrostatic potential at M06-2X/6-31G(d,p) level of theory (including solvation) for: (a) *N*-methyl acetamide; (b) *N*-methyl fluoroacetamide; (c) *N*-methyl difluoroacetamide and (d) *N*-methyl trifluoroacetamide.

### 3. Glycan array experiments

**Figure S8.** Binding curves for compounds 1, 2, 3 and 4 obtained after incubation with different concentrations of WGA-647 on the glycan microarray. The  $K_D$  surf values were obtained by fitting the curves to Langmuir isotherms.



### 4. NMR spectra of new compounds

**Figure S9.** <sup>1</sup>H NMR spectrum of compound **1** (D<sub>2</sub>O, 500 MHz)

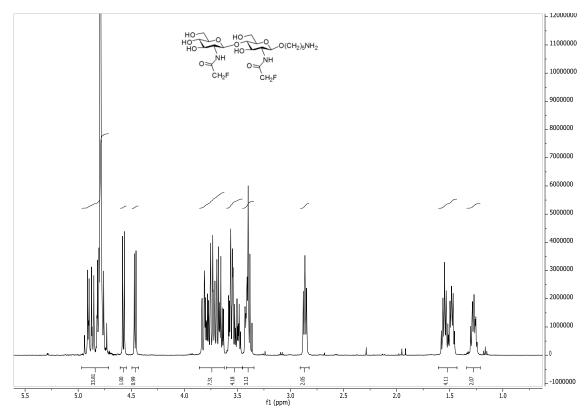
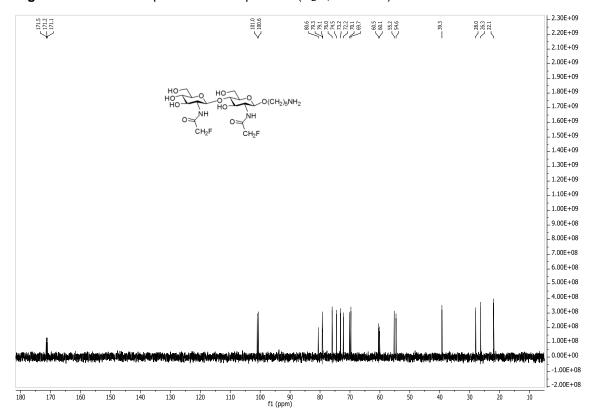


Figure S10.  $^{13}$ C NMR spectrum of compound 1 (D<sub>2</sub>O, 126 MHz)



**Figure S11.** <sup>1</sup>H NMR spectrum of compound **3** (D<sub>2</sub>O, 500 MHz)

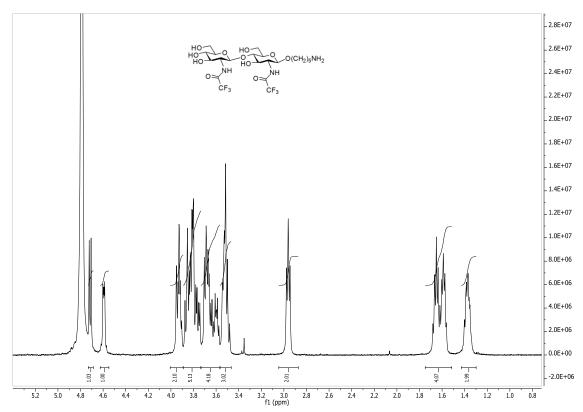
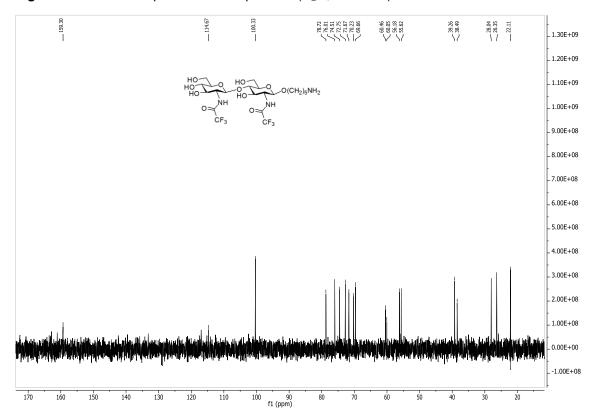
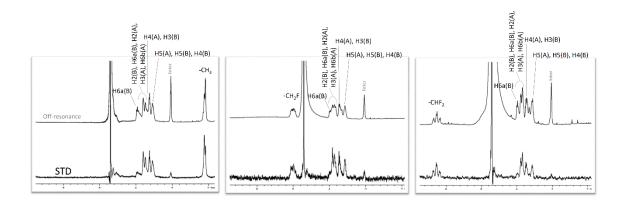


Figure S12. <sup>13</sup>C NMR spectrum of compound 3 (D<sub>2</sub>O, 126 MHz)





**Figure S13.**  $^{1}$ H-STD-NMR (top) and off-resonance (bottom) spectra for (A)  $\square$ -chitobioside (4), (B) fluorinated (-CH<sub>2</sub>F) ligand **2**, and (C) fluorinated (-CHF<sub>2</sub>) ligand **3**.

#### References

[19] G. Bains, R. T. Lee, Y. C. Lee, E. Freire, *Biochemistry* 1992, 31, 12624–12628.

[28] M. Mayer, B. Meyer, J. Am. Chem. Soc. 2001, 123, 6108-6117.

[29] J. P. Ribeiro, S. André, F. J. Cañada, H. J. Gabius, A. P. Butera, R. J. Alves, J. Jiménez-Barbero, *ChemMedChem* **2010**, *5*, 415–419.

[30] Gaussian 09, Revision C.01, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, Ö. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski, and D. J. Fox, Gaussian, Inc., Wallingford CT, 2009.