Supplementary data (Yarunin at al., 2004).

Yeast strains and plasmids. Microbiological and recombinant DNA work were performed according to Maniatis et al., (1982). The *rix19-1* strain (*MATα ura3 his3 leu2 rix19-1)* was isolated from a *ts* strains collection (Gadal et al., 2001). *RIX19, NBP35, NAR1* shuffle strains and *hcr1Δ* strain are derivatives from the corresponding EUROSCARF (Frankfurt, Germany) diploid knockout strains (*MAT***a**/α *his3Δ1/his3Δ1 leu2Δ0/leu2Δ0 lys2Δ0/LYS2 ura3Δ0/ura3Δ0*). *Tet-RLI1* and *RLI1-TAP* strains were purchased from Biocat GmbH (Heidelberg, Germany). Rli1-GFP strain was purchased from Invitrogen (Karlsruhe, Germany). Ret2-TAP strain was purchased from EUROSCARF (Frankfurt, Germany). *xpo1-1* temperature sensitive strain was obtained from the lab of Prof. Karsten Weis (Stade et al., 1997).

Rpl25-eGFP and Rps2-eGFP reporter plasmids were described previously (Gadal et al., 2001 and Milkereit et al., 2003). Plasmid pUN100-YIL003w was generated by cloning of 1528 bp fragment, containing *YIL003w* ORF (879 bp) and both untranslated regions (app. 320 bp each) to pUN100 (Elledge and Davis, 1988) vector using Sph I and Xba I restriction sites. A synthetic lethal (*sl*) screen with *rix19-1* was performed according to Wimmer et al. (1992). Plasmid pRS315-NBP35 was generated by cloning of a 1987 bp fragment, containing the *NBP35* ORF (987 bp) and both untranslated regions (app. 500 bp each) into pRS315 (Sikorski and Hieter, 1989) vector using BamH I and Xba I restriction sites. The generation of temperature sensitive mutants was performed according to Santos-Rosa et al., (1998) using Nsi I and Spe I sites in 5' and 3' untranslated regions respectively. Plasmid pRS315-NAR1 was generated by subcloning the Xho I – Xba I fragment containing *NAR1* from the pYCG_YNL240c plasmid (EUROSCARF) to pRS315 vector. The generation of temperature sensitive mutants was performed using Nde I and Eco 47III sites in 5' and 3' untranslated regions.

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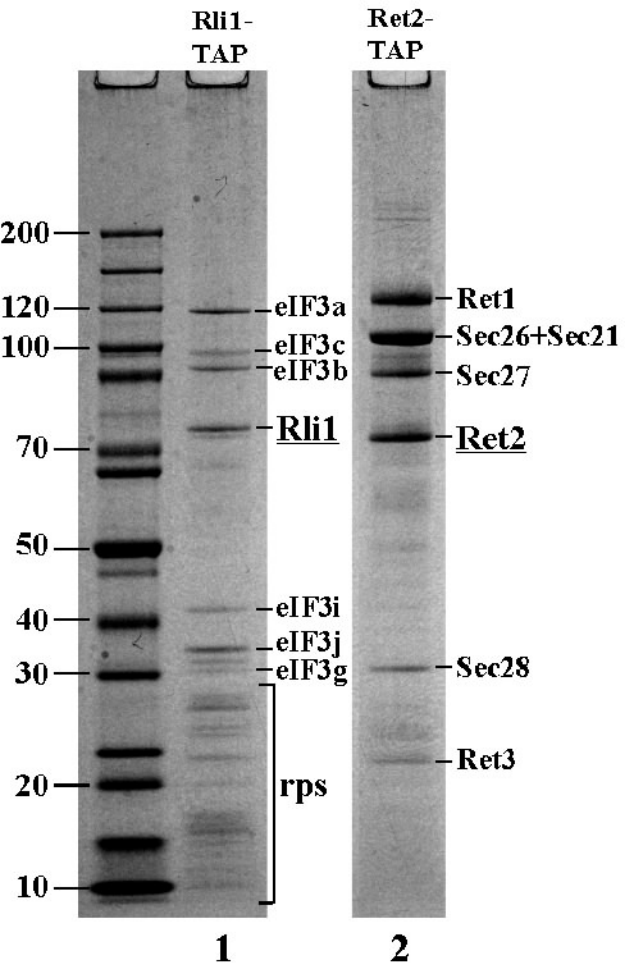
Legend to Supplementary Figure

Figure S1. Ribosomal S-proteins are specifically associated with Rli1.

TAP-purifications of Rli1 (lane 1) and Ret2 (lane 2) were performed under low-salt conditions (50 mM NaCl) as described in Fig. 6B. Ret2 is member of the coatomer (COPI) complex participating inthe secretory pathway (Hosobuchi et al., 1992).



Supplementary Figure 1

(Yarunin et al., 2004)